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5-Fluorouracil Induces Plasmonic Coupling in Gold Nanospheres: New Generation of Chemotherapeutic Agents

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Abstract

Loading 5- Fluorouracil (5-FU) into gold nanoparticles (AuNPs) could enhance its activity as anticancer drug hugely by enhancing its ability for penetration through the cell membrane. Accordingly, this work is devoted to loading 5-FU into AuNPs surface and studying the binding mechanism of the drug to the surface of the gold nanoparticles. Our finding indicates that new absorption band appears at longer wavelength upon loading 5-FU into gold nanospheres capped with citrate. This near IR band is due to induced surface plasmon coupling via hydrogen bonding between 5-FU and surface capping AuNPs. This leads to great enhancement of the drug action as chemotherapeutic as well as photothermal agents. Factors which affect the binding between 5-FU and the AuNPs such as pH, time after mixing the drug with AuNPs, concentration of the 5-FU, have been studied in detail. Accordingly, the binding interaction is proven 4uNPs showed high stability towards these factors. The spectral and morphological studies were measured via UV-VIS spectroscopy and Transmission Electron Microscopy (HR-TEM). Remarkable increases in the drug anticancer activity upon loading into AuNPs were observed for the cell viability test of human colon cancer (HCT16).

Keywords: Gold nanoparticles; Plasmonic coupling; Cytotoxicity; Colon cancer; 5-Flourouracil

Introduction

5-Fluorouracil has been in use for about 40 years as a chemotherapeutic agent against cancer. Some of its principal uses are in colorectal cancer, pancreatic cancer as well as inflammatory breast cancer. As a pyrimidine analogue, it is transformed inside the cell into different cytotoxic metabolites which are then incorporated into DNA and RNA, finally inducing cell cycle arrest and apoptosis by inhibiting the cell's ability to synthesize DNA. Some of the adverse reactions of taking 5-fluorouracil might include diarrhoea, nausea, vomiting, stomach/abdominal pain and general weakness [1,2].

Controlled drug-delivery strategies using nanomaterials have made a dramatic impact in medicine that develops highly selective and effective therapeutic and diagnostic modalities [3-5]. There are a number of advantages with nanoparticles in comparison to microparticles. For example, nanoscale particles can travel through the blood stream without sedimentation or blockage of the microvasculature. Small nanoparticles can circulate in the body and penetrate tissues such as tumors. In addition, nanoparticles can be taken up by the cells through natural means such as endocytosis.

Recently, AuNPs have been widely used in cancer therapy and imaging [6], also they have been used as contrast agents *in vitro* based on their ability to scatter visible light [7]. Au nanorods have been proposed and investigated as a new generation of photothermal agents for cancer therapy and imaging by El-Sayed group [8]. Sokolov et al. [7] used AuNPs conjugated to EGFR antibodies to label cervical biopsies for identification of precancerous lesions.

We believe that loading 5-FU into AuNPs could enhance its activity as anticancer drug hugely due to their ease for biofunctionalization, and penetration through the cell membrane. Accordingly, the aim of this work is to load 5-FU into AuNPs surface and study the nature of binding mechanism between the drug and the surface of AuNPs. Our

findings indicate that loading 5-FU upon gold nanospheres capped with citrate induces surface plasmon coupling and cause an assembly for the AuNPs. In the present work, the factors affecting the chemical interaction between the AuNPs and 5-FU such as pH, temperature and reaction time, have been investigated. In addition, the thermal and photostability of the drug loaded into AuNPs have been tested. All the prepared samples were characterized via Transmission Electron Microscopy (TEM) as a primary tool for measuring the particles size, structure and shape, and the plasmonic effect were detected via UV-VIS spectroscopy. The anticancer activity against colon carcinoma cells line has been investigated.

Materials and Instruments

All chemicals were reagent grade and were of highest available purity (A.P.); deionized water was used to prepare all the solutions. All glassware was thoroughly cleaned with aqua regia and rinsed with deionized water prior to use. Hydrogen tetrachloroaurate trihydrate (HAuCl $_4$ ·3H $_2$ O-98%) and trisodium citrate (99.99%) were purchased from Sigma–Aldrich. 5-Fluorouracil (5FU) was purchased from (Merck, 99%).

The UV-VIS absorption spectra were recorded on a PERKIN-

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ELEMER LAMBDA 40 spectrophotometer using 1 cm matched quartz cells. The transmission electron microscopy (HR-TEM) images were carried out in Nanotech Company for photo-electronic, Dreamland, 6-October, Egypt. The HR-TEM is JOEL JEM-2100 operating at 200 kV equipped with Gatan digital camera Erlangshen ES500. The pH measurements were adjusted by using JENWAY 4330 pH meter. The cell viability was recorded via ELISA reader (BIOTEK, Model: ELX 800) wavelength range: 400 to 750 nm with four built in filters. The particle size for the 5-FU@AuNPs was measured on Zetasizer, Malvern, UK, Model: Zetasizer nano series (Nano ZS), Size range (nm):0.6:6000 nm.

Experimental

Preparation of Spherical AuNPs and 5-FU@AuNPs: Au nanoparticles were prepared by the citrate reduction of $HAuCl_4$ 3 H_2O as described previously [9]. Tri-sodium citrate (0.0388 M, 10 mL) was added quickly to a boiling $HAuCl_4$ solution ($1x10^{-3}$ M, 100 mL). Strong pink color appears as an indication of the formation of AuNPs. The absorption spectrum of the obtained sample was measured via UV-VIS spectrophotometer, and the particle shape and size were characterized using TEM.

5-FU@AuNPs were prepared by adding an aqueous solution of (0.01 M, 0.0650 gm) 5-fluorouracil to 50 mL of the synthesized gold nanoparticles and stirred effectively under ultra-sonication to enhance the interaction between 5-fluorouracil and gold nanoparticles. The obtained composite has been characterized via absorption spectroscopy and HR-TEM.

Cytotoxicity and cellular uptake for the 5-Fluorouracil, AuNPs and 5-FU@AuNPs

Cytotoxicity of 5-Fluorouracil, AuNPs and 5-FU@AuNPs was assessed by cell culture method using established cell line derived from human colon cancer (HCT16).

Cell culture: HCT16 established cell line was commercially obtained from The Holding Company for Biological Products & Vaccines "VACSERA", Cairo, Egypt as a monolayer sheet in 20 ml of RPMI growth media supplemented with 10% fetal calf serum (Biowest, Germany). The cells were routinely propagated and maintained as well as the cells viability and plate preparation was carried out in Virology & Immunology Unit, National Cancer Institute, Cairo University, Egypt.

Cells viability and plate preparation: Cell viability and cell number were determined after staining of cells with trypan blue exclusion test (Biowest USA). Sterile 24 wells tissue culture plate was seeded with $1x10^5$ cells/ml and then was incubated at 37° C in 5% CO $_2$ incubator and left for 24 hrs to allow formation of confluent monolayer. After 24 hrs, the media was replaced with three different concentrations of 5-Fluorouracil ($1x10^{-4}$, $1x10^{-5}$ and $1x10^{-6}$ M), AuNPs ($1x10^{-4}$, $1x10^{-5}$ and $1x10^{-6}$ M), and of composite 5-FU@AuNPs ($1x10^{-4}$, $1x10^{-5}$ and $1x10^{-6}$ M), and incubated at 37° C, 5% CO $_2$ incubator for 6 hrs [10].

Cytotoxicity assay: It was estimated by using neutral red colorimetric assay performed according to Babichand and Borenfreund methodology [10]. After 6 hrs, 100 μ l of neutral red dye (Biochrom L6313; 0.003 % in 0.9% NaCl/ medium with 2% (FBS) Fetal Bovine Serum) was added to the plate and incubated for 3 hrs at 37°C. Then, the plate was washed with 200 μ l of Phosphate Buffer Saline (PBS) and the cells were lysed with 200 μ l of lysis buffer and placed on shaker for 20 min to extract dye from the viable cells. The extracted dye was then measured using ELISA reader. The relative percent of cell viability was calculated by [A]test/ [A]control x100, where [A]test is the absorbance

of the test sample and [A]control is the absorbance of control sample. The cell control represents 100% viability.

Results and Discussion

The visible absorption spectra of the wine red colloidal AuNPs showed a well-defined absorption band with $\lambda_{\rm max}$ =522 nm (Figure 2). This value is a characteristic for Surface Plasmon Resonance (SPR) absorbance for spherical AuNPs [11]. The wavelength was not significantly modified, during a year after preparation, thus suggesting the stability of the colloidal solution. The size and shape of the gold colloid particles have been measured by TEM imaging. A representative TEM image of these particles is given in Figure 2. The particles are mostly spherical. From the sizes of a great number of particles, measured on the TEM images, an average size (diameter) of the synthesized nanoparticles is 12-14 nm (histogram Figure 2).

Upon loading the 5-FU into AuNPs capped with sodium citrate, a gradual color change from red to purple then blue has been observed and a new band starts to appear at higher wavelength ~ 690 nm. The λ_{max} of this new absorption band continues its shift towards higher wavelengths with remarkable increase in intensity with time Figure 3B. The increase of absorbance for the new band at 690 nm was accompanied by a decrease in intensity of the SPR band of spherical AuNPs at 522 nm and a slightly shift was observed in its λ_{max} from 522 nm to 530 nm Figure 3A. This shift is due to the change in the dielectric constant of the solution [12]. The color change, as an effect of agglomeration (assembly of the particles), is a well-understood phenomenon [13]. When the interparticle distance in the assembly decreases to less than about the average particle diameter, the electric dipole-dipole interaction and coupling between the plasmons of neighboring particles in the assembly has resulted in the bathochromic shift of the absorption band which is called plasmon coupling band [14]. The diameter of the agglomerated particles was measured by using zetasizer analysis and most of them have ~ 48 nm, see supplemental information.

Increasing the concentration of 5-FU has a significant effect on the degree of the assembly and the nanocrystals agglomeration kinetics. As shown in Figure 4, an increase in the concentration of the drug leads to further agglomeration (assembly) of the gold nanocrystals.

We can assume that 5-FU bind to the citrate coating the AuNPs surface, inducing particles agglomeration. The interaction occurred through the formation of two hydrogen bonds; one between COO of citrate and the NH of the drug 5-FU and the other between carbonyl group in the drug and OH in sodium citrate as shown in Figure 5.

The degree of plasmonic coupling and AuNPs assembly coated by 5-FU were also found to be tunable by pH, where the pH of the medium is a vital factor that determines the stability of the composite.

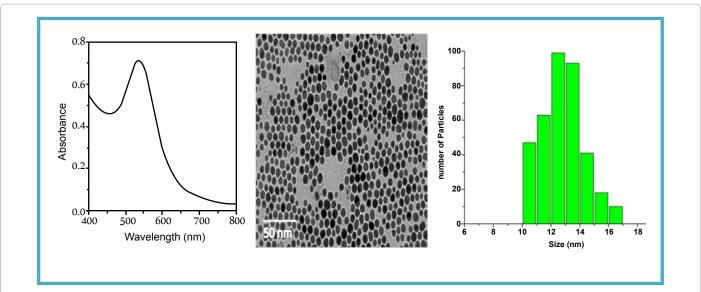


Figure 2: The UV-Vis spectrum of Au nanoparticles and transmission electron micrographic image of gold nanoparticles and their histogram of size distribution.

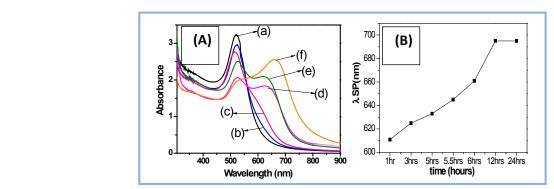


Figure 3: A)The time-dependent UV-Vis spectra of 5-FU@AuNPs: (a) pure AuNPs, (b-f) after mixing with 5-FU, b)10 mins, c) 30 mins, d) 2 hours, e) 4 hours, f) 6 hours, B) Time dependence for the shift of λ_{max} of the plasmon coupling band for 5-FU@AuNPs.

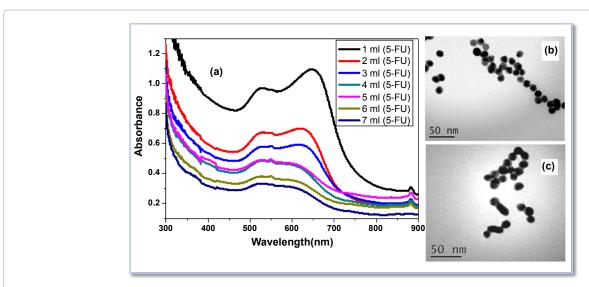


Figure 4: (a)UV-Vis spectra of 5-FU @AuNPs formed with different volume ratio of 5-FU, (b) TEM images for 1mL:0.5 mL of Au:5-FU show mostly linearly arranged assemblies of NPs, (c) TEM images for 1mL:1mL of Au:5-FU, while the nanoparticles assemblies are more complex.

For this reason, the 5-FU has been loaded into gold capped citrate nanocrystals at different pH values using 0.1 M NaOH and 0.1 M HCl. The absorption spectra of 5-FU @AuNPs at different pH values have been monitored (Figure 6) and the resultant composites have been characterized via HR-TEM (Figure 7).

Assembly of AuNPs can be produced by the surface acid-base reactions of the individual gold nanoparticles. Recent work of Kleinermanns et al. [16], reported the ensemble of AuNPs [15] modified with an ionizable and pH-sensitive monolayer of Thiobarbituric Acid (TBA). It was found that nanoparticle assembly can be produced in a controlled fashion by the variation of the pH of the solution [16]. Our results show that the interparticle spacing in Au assembly mediated by anticancer drug 5-FU was also found to be tunable by pH as shown with as TBA. One notable feature in Figure 6 is that, the plasmon coupling band of the assembled AuNPs at ~ 690 nm appeared gradually for the solutions of pH 3 to 6 with a color change from wine red to purple then blue with increase pH. This may be assigned to lowering of the pH of solution leads to protonation of the amine group in the drug (N=1, Figure 1), allowing hydrogen bond interactions between the drug and carboxylic group in the citrate inions that capping gold nanoparticles, thus leading to assembly of AuNPs. On the other hand, the absorption spectra of the solutions with high pH values (from pH =7 to pH =11), revealed no bands rather than the SPR band of AuNPs Figure 6(b).

A successive shift in SPR of AuNPs from 520 to 531 nm along with reduction in intensity was observed upon the addition of 5-FU for the solutions of pH from 2.0 to 11.0

All of these observations were also evident from TEM studies as shown in Figure 7. The pH-dependent in optical properties has implications for controlled release of molecules from the nanoparticle assembly, an area of importance in controlled drug release.

If gold nanoparticles are coated with a drug shell, the drug can be released from the carriers either via enzymatic activity or changes in physiological conditions such as pH, osmolality, or temperature [17] to be taken up by tumor cells. For this reason the thermal stability of 5-FU@AuNPs was investigated, by heating a solution of 5-FU@AuNPs to various temperature degrees in the range of 25-100°C. Furthermore, this study gives an impression to the decomposition temperature of the composite briefly. With increasing the temperature up to 90°C the electronic absorption spectrum Figure 8(a) shows no significant change in the absorption maximum of the two characteristic bands, the SPR band of AuNPs at 522 nm and the band due to plasmonic coupling

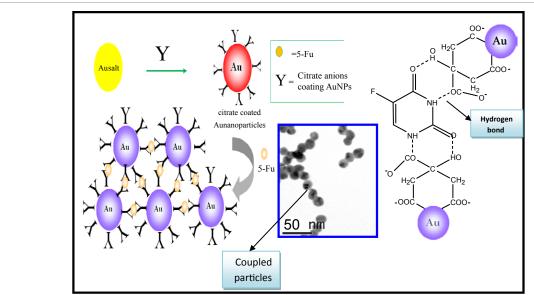


Figure 5: The chemical formula postulated for the linkage between 5-FU and AuNPs, and its TEM image of their assembly.

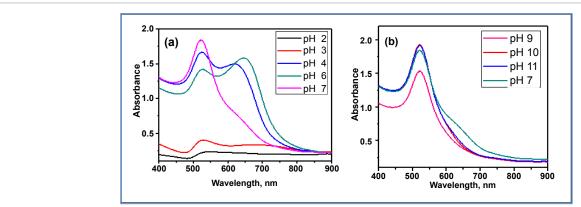


Figure 6: The UV-Vis spectra of 5-FU@AuNPs at different pH values: (a) acidic media pH=2-7, (b): basic media pH=7-11.

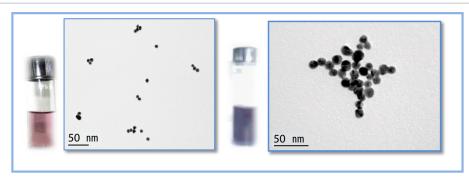


Figure 7: (a) The TEM images of 5-FU@AuNPs at pH=3 with the purple color showing the beginning of the assembly with two or three particles (b): at pH=6, while the assembly is more complex.

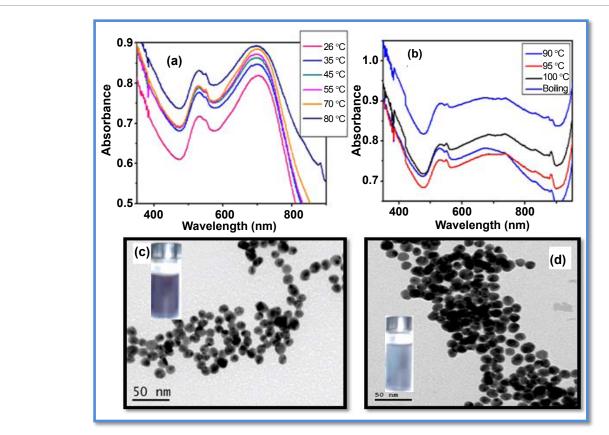


Figure 8: Effect of high temperature on absorption spectra of an aqueous solution of 5-FU@ AuNPs :(a) from 25-80°C (b) 90-100°C, (c) TEM image and photo at 25°C (d) TEM image and photo after boiling.

for the composite at 695 nm, which indicates the high stability of the formed composite up to this temperature.

Beyond 90°C, both of the SPR absorption band and plasmon coupling band of 5-FU@AuNPs nanocomposite are merged together Figure 8(b). This observation suggests that the micelles encapsulating the gold nanoparticles are relatively destroyed, and a black precipitate of aggregated gold powder is formed. This leads to the possibility that the decomposition temperature of the most stable micelles used in this work is in the range of 90-100°C.

The effect of UV-light on the 5-FU@AuNPs composite was observed using absorption spectrophotometer. To demonstrate the irradiation effect of UV-light on the sample solution, 5ml colloidal solution of the

5-FU@AuNPs were put in 5cm quartz cell under the UV-light source (λ =254/365nm) and exposed to different irradiation time.

The results obtained from absorption show that 5-FU@AuNPs before irradiation to light has two λ_{max} one at 522 nm due to the SPR of AuNPs and another band at 695 nm due to plasmon coupling of NPs. By irradiation, the 5-FU@AuNPs as a function of time and the optical density shows slight decrease during the first hour without any remarkable shift in the corresponding wavelengths of each band, Figure 9. This may be attributed to the capping strength of 5-FU which form a composite with AuNPs preventing the nanoparticles from aggregation during the irradiation time. On increasing the time of irradiation for 3 hours, a remarkable decrease in the intensity of both bands without

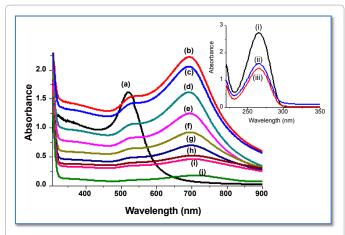


Figure 9: shows the effect of UV-light on AuNPs@5-FU: (a) AuNPs, (b) 5-FU@AuNPs without irradiation(c-j) after irradiation with time interval 1 hour until 6 hours. The inset figure (i) 5-FU (ii) 5-FU@AuNPs without irradiation and (iii) irradiated 5-FU@AuNPs.

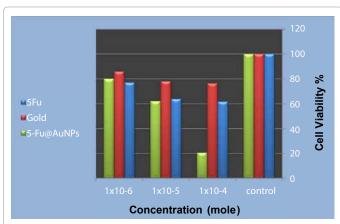


Figure 10: The percentage viability of cells at different concentrations of 5-Fu, AuNPs, and 5-Fu@AuNPs, respectively.

shift in the corresponding wavelengths was observed. The bands continue decaying as the time increase up to 6 hours of irradiation. This may be due to the regular release of the drug from the surface of AuNPs with the decomposition of the gold nanoparticles combined with an aggregation in the solution. The results suggest that the use of these particles in magnetic fluid hyperthermia with the exposure to UV radiation will be safe and the particles will do well up to 3 hours.

During the time of exposure, the $n-\pi^*$ transition band of the 5-FU molecule showed no changes as observed in the inset Figure 9.

Cytotoxicity test

In vitro cell viability (Cytotoxicity studies): Neutral red assay using the HCT16 (colon carcinoma cells line) was performed to analyze the potential toxicity of the AuNPs, 5-FU and 5-FU@AuNPs, in concentration (1x10⁻⁴, 1x10⁻⁵ and 1x10⁻⁶ M), After incubation time of 6 hours, viable cells have the ability to absorb neutral red whose absorption could be estimated at 570 nm; the value at which it is correlated to the number of living cells (Figure 10). It was found that at low concentration, 1x10⁻⁵ and 1x10⁻⁶ M of 5-FU and 5-FU@AuNPs nanocomposites show low cell viability while at slightly high concentration 1x10⁻⁴ M of 5-FU@AuNPs shows high cell viability (cells

death) about 20% compared with AuNPs and 5-FU which indicates the effectiveness for cell toxicity of this nano composites in cancer treatment and open the door for using anticancer drug loaded over nanoparticles. On the other hand, citrate stabilized AuNPs were shown to have little effect on this particular cell line as described by Connor et al. [18], and hence, we have used them for proper comparison. The results suggest that the 5-FU@AuNPs nanocomposites show excellent dispensability and long-term stability in physiological solution viability from 20 - 80% has been demonstrated compared to the control. Thus, on the basis of the *in vitro* data, the 5-FU@AuNPs nanocomposites are good potential candidates as a drug delivery for further *in vitro* studies as well as *in vivo* applications.

Conclusion and Future Perspective

The nature of binding the anticancer drug 5-FU with the prepared AuNPs has been established using UV-Vis spectroscopy and TEM imaging. The results revealed 5-FU bind to the citrate coating the AuNPs surface, inducing particles agglomeration i.e. assembly of the NPs, which increase gradually with time. The interaction occurred through the formation of two hydrogen bonds one between COO of citrate and the NH of the drug 5-FU and the other between carbonyl group in the drug and OH in citrate. The previous discussion also referred to the formation of 5-FU@AuNPs in acidic media due to protonation in 5-FU molecule that enhance interaction with citrate capped AuNPs. Furthermore, this investigation shows high thermal and photo stability for the 5-FU@AuNPs.

We have found substantial enhancement of the anti cancer effect of 5-FU@AuNPs against HCT16 (colon carcinoma cells line) compared to the same drug in typically administered free form or as prepared AuNPs only. The improvement was attributed to enhanced intracellular transport followed by the subsequent release in lysosomes. Enhanced activity and nanoparticle carriers will make possible the reduction of the overall concentration of the drug, renal clearance and side effects.

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Financial & competing interest disclosure

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