In-Vitro Release and Pharmacokinetics of Anti-tubercle Drug Ethionamide in Healthy Male Subjects

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Abstract

The aim of study was to assess the pharmacokinetics of ethionamide in the local population of healthy human subjects. Serum samples were taken from each of the selected subject at different time intervals. These samples were analyzed by using High Performance Liquid Chromatography consisting of reverse phase C18 column, UV detector set at 291nm. The mobile phase was consisted of 0.02 M disodium hydrogen phosphate and acetonitrile (75:25) and delivered at a rate of 1.5ml/min. The value of $C_{\text{max}}$ was found to be 1.941 ± 1.487 µg/ml (mean ± SEM) and $T_{\text{max}}$ was 1.75 ± 1.487 hours (mean ± SEM). The area under the curve (AUC) was 8.745 ± 0.536 (mean ± SEM). The elimination half life ($t_{1/2}$) was found out as 1.995 ± 1.157 hours (mean ± SEM). The total body clearance (CI) was determined as 32.591 ± 0.298 ml/hr/kg (mean ± SEM). It was concluded that ethionamide (Ethomid® Schazoo-Lahore, Pakistan) found in consistent with the values reported in the available literature. The study will be beneficial and valuable in designing dosage regimen for the patients on ethionamide therapy and can be utilized as guideline in accessing the bioavailability and pharmacokinetics parameters in clinical situations.

Keywords: Ethionamide (ETA); Pharmacokinetics; Serum; High performance liquid Chromatography (HPLC)

Introduction

Ethionamide is an isonicotinic acid derivative of thioamide class. It is the second line orally administered drug used in the treatment of clinical tuberculosis that has failed to respond to adequate first line therapy. Ethionamide is often combined with other anti tuberculous agents for the treatment of multidrug resistant organisms. Ethionamide is active against tubercle bacilli that are growing with in human macrophages (Auclair et al., 2001; Conte et al., 2000). It is used to cure tuberculosis, a disease that infects more than a third of the world’s population (Vannelli et al., 2002). Infections caused by Mycobacterium avium intracellulur complex (MAI) and drug resistant mycobacterium are increasingly common in different parts of the world and are fueled increasing by decrease in absolute number of patients treated with clofazimine, Cylcoserine, Paraminosalicylic acid or pyridoxine, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Subjects, Materials and Methods

Subjects

Twelve healthy adult male human volunteers participated in this study. The ages of volunteers ranged from 21 to 30 years and their body weights in the range of 56-70kg. On the basis of medical history, clinical examination and laboratory investigations (hematology, blood biochemistry and urine analysis), none of the participants had any revealed medical abnormality. In addition, the included subjects had no history of hospitalization and involvement in any medical trial within twelve weeks prior to the initiation of the study, and none of them had received any regular course of drug therapy within the preceding four weeks.

Subjects were excluded from the study if they had a history of serious illness, were taking any medications (including OTC) within one week prior to the start of the study, or having a history of any kind of allergy, history of hypersensitivity to ETA, Clofazimine, Cycloserine, Paminosalicylic acid or pyridoxine, had donated blood within 30 days prior to the study. Written consent was obtained from the volunteers. The Pharmacy ethical committee of Faculty of Pharmacy and Alternative Medicine, the Islamia University of Bahawalpur approved the protocols of this study.

In-vitro studies

The drug formulation was tested and passed the Compendial requirements with respect to its content uniformity and weight variation. Dissolutions studies were performed in 0.1 N HCl (Dissolution media) by placing six tablets in USP apparatus-I at 37°C. The apparatus was adjusted at 100 rpm. Samples were then drawn at designated time intervals at 5, 15, 30 and 45 minutes and analyzed for their Ethionamide content spectrophotometrically at 290 nm against the dissolution medium as blank (USP-NF, 2004). Concentrations were determined with reference to a standard curve. The values are given in Table 1 and shown in Figure 1.

In vivo studies

The study was an open, single dose, complete one period of treatment dosing. Subjects received single dose of Ethionamide hydrochloride (Ethomid® Schazoo Laboratories (Pvt.) Ltd., Lahore–Pakistan) Tablets of 250 mg (Batch No. ETT-011). Drug administration began approximately at 7.30 a.m. on the study day after an overnight fast of 10 h. Written informs consent was obtained from each subject before the study. An indwelling cannula was inserted into a forearm vein before the ingestion of Ethionamide tablet with 250 ml of water. The subjects were asked to remain fasting for 4 hours into the study. No water was permitted 2 hours after dosing. A light breakfast was allowed at 2 hours after dosing. A light breakfast was allowed at 4 hours followed by standardized lunch and evening tea with refreshment. Cigarettes and food or beverage containing caffeine were not allowed over 24 hour.

Sample collection

A 20-gauge venous cannula was inserted into a forearm for the collection of blood samples and 3 ml blood samples were collected through the syringe before dosing (zero time) and at 0.5, 1.0, 1.5, 2.0, 2.5, 3.0, 4.0, 6.0, 8.0, 12.0 and 24.0 hours after dosing of Ethionamide tablet. Following clot retraction, the samples were centrifuged at 4000 rpm for 15 min. The obtained serum was frozen at -20°C pending analysis. Samples were kept refrigerated during the period of collection.

High performance liquid chromatography

Analyses were performed by using Shimadzu Liquid Chromatography, with a pump LC 10 A.S, SPD 10 A. Shimadzu U.V. Visible Detector set at 291 nm. A reverse phase system was used consisting of a 5-µm Hypersil ODS (C18) column (250nm x 4.6 mm I.D.) preceded by 5-µm Hypersil ODS (C18) cartridge Guard (10mm x 4.6 mm I.D.) column. The mobile phase was 0.02 M disodium hydrogen phosphate buffer–Acetonitrile (HPLC) method (Charles et al., 1991). The mobile phase consisted of disodium hydrogen phosphate 0.02 M and Acetonitrile in the ratio of 75:25. 0.02 M disodium Hydrogen phosphate was prepared by dissolving 3.588 g of di sodium hydrogen phosphate in about 200 ml of distilled water and mix thoroughly until complete solubility in distilled water, then make the volume to 1000 ml with distilled water. Now take 750 ml of this buffer solution and 250 ml of Acetonitrile to make 1000 ml with Acetonitrile. Filter the mobile phase by passing through filtration assembly under vacuum pressure of 150-200 torr using 0.45µm membrane filter (Sartorius). Now degas the mobile phase by placing it in the sonicator for 2-3 min. until complete degassing of the mobile phase is ensured.

Stock solution (100µg/ml) were prepared fresh daily by dissolving the drug in the acetonitrile. Extraction procedure was simple based on precipitation method. In the extraction procedure 1 ml of the drug solution was spiked with 1 ml of the blank serum in the 8 ml glass centrifuge tube and mixes well, then centrifuge for 10 min. Separate the upper layer (organic layer) by micropipette, filter by using of the filtration syringe and take the filtrate in 1.5 ml eppendorf tubes. Evaporate the sample by using vacuum evaporator to dryness under the steam of nitrogen. Reconstitute the sample with 70µL of the mobile phase; vortex mix and Inject in to the HPLC injection port 20µl of the volume by injection syringe.

Preparation of standard curve

Standard curve was constructed to encompass the anticipated range of serum ethionamide concentration found in healthy Subjects. Blank serum was spiked with ethionamide drug solution to give the concentrations of 0.1, 0.5, 1.0, 2.0, 3.0, and 4.0µg/ml. The extraction procedure was same as described earlier. The standard curve is constructed for each of the subjects to avoid variation and to ensure precision and accuracy in the results, also average of the twelve volunteer is also calculated by taking mean of the twelve subjects.

Injections of 20µl were injected and spectra were taken of each concentration. The peak areas are noted for each concentration. The absolute recovery of ETA was 86%. The within day precision [coefficient of variation (CV)] of validation samples was 0.42-5.26%, and the overall validation precision was 0.65-3.89 pattern was determined from samples assayed over the course of study. The specificity of the HPLC assay for ethionamide was determined by testing spiked samples (Auclair et al., 2001; Charles et al., 1991).

Safety Analysis

Health assessment, including vital signs, physical examination and clinical laboratory testing was performed before and seven days after the study. Subjects were interviewed at the beginning and end of each study period and were monitored

Table 1: Dissolution profile of Ethionamide tablets (ETT-011).

<table>
<thead>
<tr>
<th>Sr. No.</th>
<th>Time (min)</th>
<th>Percent Release</th>
<th>SEM</th>
</tr>
</thead>
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<tr>
<td>1</td>
<td>5</td>
<td>18</td>
<td>0.128</td>
</tr>
<tr>
<td>2</td>
<td>15</td>
<td>42</td>
<td>0.249</td>
</tr>
<tr>
<td>3</td>
<td>30</td>
<td>72</td>
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</tr>
<tr>
<td>4</td>
<td>45</td>
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<td>0.194</td>
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Figure 1: Percentage release of Ethionamide Tablets determined by the dissolution studies.

ISSN:1948-593X JBABM, an open access journal

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Table 1: Dissolution profile of Ethionamide tablets (ETT-011).
throughout of the confinement period to determine any adverse events potentially related to study medication of procedures (Auclair et al., 2001).

**Pharmacokinetic Analysis**

Pharmacokinetics analyses were performed by non-compartmental method of analysis. Maximum concentration of Ethionamide in serum (Cmax) and times to these concentrations (Tmax), area under the concentration time curve (AUC) and other pharmacokinetic parameters were calculated by using Kinetica Software.

**Results**

The in vitro results indicate that the Ethionamide formulation met the specifications for content uniformity and weight variation stipulated by compendial standards. Ethionamide concentration and peak height ratios of Ethionamide were linear throughout the concentration range of 0.1 to 4µg/ml. Standard curve of Ethionamide in serum was constructed to determine the slope. The limit of detection of the assay was calculated to be 10 ng of Ethionamide per ml serum. Mean serum concentrations following the oral administration of Ethionamide to 12 volunteers are presented in Table 2 and plotted on rectangular co-ordinate graph & on a semi logarithmic scale in Figure 2 and Figure 3, respectively. The ratio of AUC_{0-12h}/AUC_{0-∞} for all individuals was > 98% indicating an adequate sampling time, since the extrapolated portion of the total AUC is less than 2%. Maximum concentration in all the twelve subjects found in the range of 1.229 to 2.7527µg/ml with a mean value of 1.941 ± 1.3857µg/ml at time ranging from 1 to 2.5 hours with a mean value of 1.75 ± 1.487 hours.

<table>
<thead>
<tr>
<th>Sr. No.</th>
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<th>Concentration (µg/ml)</th>
<th>SEM</th>
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<tr>
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</table>

Table 2: Mean ± SEM (n=12) Serum concentration of Ethionamide HCl administered in the oral dose of 250 mg in Healthy Human Volunteers.

![Figure 2: Serum Concentration vs time profile of Ethionamide HCl plotted on rectangular co-ordinate graph, administered in the oral dose of 250 mg in healthy human volunteers (n=12).](image)

![Figure 3: Concentration vs time of Ethionamide HCl plotted on semi-log graph, administered in the oral dose of 250 mg in healthy human volunteers (n=12).](image)

The bioavailability and pharmacokinetic parameters of Ethionamide are calculated by using non-compartmental model of pharmacokinetic analysis for accessing Cmax, Tmax and Area under the curve (AUC). The absorption of Ethionamide was excellent with mean serum concentration (Cmax) of 1.941µg/ml obtained at average maximum time (Tmax) of 1.75 hours. The geometric mean (± SEM) of bioavailability and pharmacokinetic parameters (AUC_{0-∞},AUMC_{0-∞}, Ka, t_{1/2(abs)}, t_{1/2(elim)}, MAT, t_{1/2(elim)}, MRT, Vd and Cl) following the administration of Ethionamide are presented in Table 3.

**Discussion**

Although less information is available in literature on the bioavailability and pharmacokinetics of ethionamide in healthy human subjects as well as in patients with tuberculosis. However, the bioavailability and pharmacokinetic parameters observed in this study are in accordance with the earlier reported values.

The absorption of Ethionamide is rapid and complete after oral administration and more than 90% of the given dose absorbed from gastro-intestinal tract. The absorption rate constant (Ka) in this study found in the range of 0.272 to 0.598 hr⁻¹ with a mean value of 0.384 ± 3.185 hr⁻¹ which shows that absorption is rapid and measurable quantities of drug reaching blood in 15 to 20 minutes. Absorption rate constant (Ka) is an important parameter having an influence on the Cmax, Tmax and AUC. Shorter the value of Tmax faster will be the absorption rate. Cmax also increases with higher values of Ka. The Mean absorption time (MAT) 1.671 to 3.554 hours (2.758 ± 1.23) shows that absorption of the drug completed within 2 to 3 hours. The maximum
parameter having effect on $C_{\text{max}}$ with mean values of 8.745 ± 0.566 µg/ml. Area under the curve (AUC) reflects the total amount of drug that reaches the systemic circulation. AUC is directly proportional to the dose of the drug. As dose of the drug increases, AUC also increases. Ethionamide is rapidly and widely distributed throughout the body. The values for AUC$_{\text{max}}$ are calculated in this study ranging from 5.231-15.615 µg.h/ml with mean values of 8.745 ± 0.566µg.h/ml. This value is in accordance with the previously reported value ranging from 5.4 to 17µg.h/ml with a mean value of 10 ± 0.360µg.h/ml (Auclair et al., 2001). The elimination rate constant (ke) is an important parameter having effect on $C_{\text{max}}, T_{\text{max}}$ and AUC. With an increase in the elimination rate, the values of $C_{\text{max}}, T_{\text{max}}$ and AUC will decrease and vice versa. The mean ± SEM values of elimination rate constant and elimination half life were calculated to be 0.39 ± 2.864 hr$^{-1}$ and 1.8 ± 1.362 hours, respectively. The mean ± SEM values reported by Zhu et al. (2001) for elimination rate constant (ke) and elimination half life ($t_{1/2}$) is 0.43 ± 2.105 hr$^{-1}$ and 1.94 ± 1.362 hours which are in line with the present findings. The values of clearance reported in literature lies in the range of 27.5 to 66.5 L/hr/Kg (32 ± 0.698 L/hr/Kg, mean ± SEM). The clearance is a useful index for the measurement of drug removal from the body. Faster the clearance, lower will be volume of distribution and elimination half life. If elimination half life is decreased the accumulation of the drugs in the body will also decreased and the clearance will be increased. Volume of distribution is also related with the dose of the drug. As dose of the drug increases, the volume of distribution ($V_{d}$) also increased. The volume of distribution $V_{d}$ estimated in this study ranging from 52.78 to 75.70 L/kg. Volume of distribution relates plasma concentration to the amount of drug in the body. A large value of volume of distribution shows that the drug is readily soluble and completely absorbed from the routes of administration. Volume of steady state shows the transfer of the drug from the tissue to central compartment. The volume of steady state, $V_{s}$ worked out in the range of 4.128-38.388 L/kg with mean value of 18.154 L/kg ± 0.2958 L/kg.

The bioavailability and pharmacokinetic parameters assessed in the present study shows a similarity with the previous reported parameters. The drug shows better absorption profile and more than 90% of the drug absorbed. The drug also shows excellent distribution and elimination profiles. These parameters will be beneficial and valuable in designing dosage regimen for the patients on ethionamide therapy and can be utilized as guideline for assessing the bioavailability and pharmacokinetic parameters in clinical situations. These studies must be conducted in the real clinical conditions in local population to determine its safety and tolerability.

Acknowledgement

The Authors wish to thank to the Management of Schazoo Laboratories (Pvt.) Ltd. Lahore-Pakistan for providing all kind facilities for the completion of this research project.

References