Lipidomic Analysis of Serum from Horses with Strongyle Infection

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Abstract

The development of techniques capable of accurate diagnosis of strongyle infections is at the forefront of research in equine parasitology. The aim of this study was to evaluate the potential, for using lipidomics in the diagnosis of strongyle infection. Blood and fecal samples were collected from 30 horses. Fecal egg count (FEC) results were used to select the serum samples from six uninfected horses (negative controls) and from the five horses with the highest burdens. The lipid portion of serum samples was extracted and analyzed using High Performance Liquid Chromatography-Mass Spectroscopy. The concentrations of 66 lipid metabolites differed between infected and uninfected horses (p<0.025). Database comparison of mass/charge (m/z) ratios and retention times were used to tentatively identify 16 of these metabolites. The roles of these metabolites and reasons for the observable changes were discussed. These results demonstrate the potential for the use of high resolution lipidomic analysis, for the development of a diagnostic technique capable of detecting, and perhaps stratifying, equine strongyle infection.

Keywords: Strongyle; Biomarker; Metabolomics; Lipid profile; Lipidomics

Introduction

The family Strongylidae, which includes large strongyles (Strongylinae) and small strongyles (Cyathostominae), includes some of the most common and pathogenic parasites of horses. There are about 60 equine strongyle species [1], all capable of causing clinical disease termed strongylosis. Most of the species are so-called small strongyles or cyathostomes, for which infective third stage larvae migrate into the mucosa/submucosa of the large intestine. The large strongyle (Strongylus spp i.e. S. vulgaris) infective third stage larvae migration is within blood vessels. Strongylosis has a huge impact on the welfare of infected horses globally, with clinical effects ranging from mild depression and lethargy to profuse diarrhea, colic and a fatal wasting syndrome.

Diagnosis of strongyle infection in practice has been traditionally based on fecal analysis, with microscopic identification of eggs and parasite larvae. However, besides being labour-intensive and suffering from low diagnostic sensitivities, these techniques can’t detect prepatent infection. This is a major limitation in some strongyle infections such as Strongylus vulgaris and cyathostomin, where most of the damage is attributed to larval stages rather than adult worms. The increasing demand for accurate and reliable diagnostic tools has lead to the development of more sensitive and specific molecular diagnostic techniques, with the aim of eventually replacing the traditional techniques. New molecular techniques include a PCR-ELISA to amplify cyathostomins ribosomal intergenic spacer (IGS) region from fecal samples [2]; a reverse line blot assay that targets the IGS region of 13 species of cyathostomins and all 3 species of Strongylus [3]; a fluorescence-based real-time TaqMan PCR assay, to detect and semi-quantify the DNA from Strongylus vulgaris in fecal samples [4]. Despite their promising potential, none of these molecular tests is capable of detecting all important parasites of horses in one analysis. Further research and development is required, before a fully quantitative diagnostic test can become commercially available.

Lipidomics have been used to study and understand the biological mechanisms of parasites, often with the aim of developing more specific antiparasitic treatments [5,6]. However, the effect of strongyle parasitism per se on the equid host’s lipid profile has not been documented. This study aims to evaluate the use of metabolomics, to determine differences in the lipid profile of sera from strongyle infected versus non-infected horses. This will allow appraisal of this technique’s potential for future development of a diagnostic test.

Materials and methods

Sample collection

Samples of blood and feces were taken from horses being euthanased, at an equine abattoir, for purposes other than research. These horses had originated from different owners and therefore, had been kept under different management practices, with an unknown history with respect to their anthelmintic treatment. Although a formal BCS evaluation was not undertaken, animals that were obviously thin or very fat were excluded from the study (i.e., samples were collected from animals of a similar BCS). Thirty sets of samples were collected. A single blood sample was collected immediately post euthanasia from the jugular and carotid veins of each horse. Transport and storage in 50ml falcon tubes in the absence of anticoagulants allowed blood to clot, in preparation for serum collection. On return to the laboratory, samples were stored in an upright position at about 4°C in a cold room.

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Samples from several fecal balls were collected from the rectum of each horse within 10 min of the blood sampling. These were immediately put into 50ml sealed plastic containers, that were numbered to match with each horse's corresponding blood sample. Fecal samples were also stored at 4°C, to further minimize chances of egg development [7].

**Fecal egg count**

McMaster technique was used to detect and quantify strongyle eggs per gram (epg) in fecal samples as described [8]. Each sample was analyzed on three separate occasions using identical procedures and a mean egg count, ± the standard deviation (SD) for each horse was calculated, to be used for analysis.

**Lipidomic analysis**

**Serum Separation and Selection:** All serum was removed from the surface of clotted blood samples and transferred to 1ml sterile eppendorf tubes, using a pipette. Tubes were left overnight in the cold room to allow sedimentation of any remaining blood cell contamination. Serum was again pipetted off, transferred to clean eppendorfs and frozen at -20°C. Once all fecal samples had been analyzed to assess the worm burden of each horse, serum samples from six horses with negative FEC and from the five horses with the highest burdens (> 500epg) were selected for analysis. Selection was based primarily on FEC results. Before analysis, sera from the control horses with negative FEC were pooled to produce one control sera sample. Samples were packaged appropriately and transported on ice to the Centre for Analytical Bioscience (CAB), University of Nottingham, where they were kept at -80°C.

**Lipid Extraction:** Lipids were extracted from the serum using a chloroform: methanol liquid-liquid extraction method based on the technique described [9]. This method separated lipids from all non-lipid elements, including any lysed red blood cells. The lipid containing layer was removed and mixed with an equal volume of isopropanol, prior to analysis. 10μl of lipid extract from each sample was then used for analysis. Quality controls were prepared using equivalent volumes of plasma from the CAB plasma library.

**Liquid Chromatography:** Liquid chromatography (LC) was used to separate different components of the extracted lipid samples. A pump causes the mobile phase (comprised of solvent and analytes) to move through the stationary phase(s), within the column used. The time taken for the sample to pass down the column and be eluted is defined as the retention time (RT). RT varies between analytes and can therefore be used for identification purposes. It is dependent upon the strength of sample interaction with the stationary phase(s), the ratio and composition of solvents used, and the flow rate of the mobile phase. An Accela High Performance Liquid Chromatography (HPLC) system was used to introduce samples onto a Thermosil Gold C18 3mm x 2.1 mm column (1.9mm particle size). Ten microlitres were injected on to each occasion. Samples were eluted using a 5 minute water/acetoniitrile/ isopropanol gradient, modified with 10mM ammonium acetate at pH 6.8.

**Mass Spectroscopy:** The resulting eluent from the LC was ionised using an electrospray mode, in which the eluent is sprayed into a chamber at atmospheric pressure. A strong electric field causes further dissociation of analyte molecules, while a heated drying gas causes solvent in the droplets to evaporate. As droplets reduce in size, the internal charge concentration increases. Gaseous ions are ejected, once repulsive forces between ions with like charges exceed cohesive forces holding ions together. Gas ions enter the Exactive mass spectrometer. Samples and quality controls were injected 3 and 6 times respectively. Settings allowed ions with m/z ratios between 100 and 1500 to be detected, with the spectrometer being calibrated to within 1 ppm and 2 ppm, respectively. The ratio of the concentration of these metabolites in uninfected samples to infected samples was calculated and presented as a decimal. Ratios less than 1 indicate an increase of the relevant metabolite in horses with strongyle infection, compared to those that were uninfected. A ratio greater than 1 is indicative of a reduction in metabolite level in infected horses.

**Data analysis:** Statistical differences between the infected and uninfected sample groups were highlighted using Sieve 1.2 software. Results were then filtered using Microsoft Excel to produce a final list of ions that showed statistical differences, defined as p<0.025 (calculated using Student's T test) and strong MS signals (peak area >5000). Databases, including the Human Metabolome database and the laboratory's own accurate mass conversion of lipid maps, were screened for matching masses.

**Results**

**Fecal analysis**

Of the 30 horses whose fecal samples were analyzed, eight animals had consistently negative FEC results. 22 of 30 horses had strongyle burdens which ranged from barely detectable (<50 epg) to heavy (approx 1200 epg) showing a high prevalence of infection (73.3%). Other than eggs of ascarid nematodes, which were also seen in fecal samples from two horses, no parasitic elements other than strongyle eggs were identified. Blood samples from the five horses with the highest burden (602 ± 6.7, 825 ± 18.7, 1005 ± 119.5, 1140 ± 164.9 and 1200 ± 281.3 epg) were selected for serum analysis, and blood samples from six horses with negative FEC results were pooled and used as the negative control. This provided an estimation of normal metabolite concentrations in the lipid profile of equine serum.

**Serum analysis**

The concentrations of 66 metabolites were significantly different between infected horses and the pooled control sample (p<0.025). Tentative identification of 16 metabolites was achieved based on their LC retention times and MS m/z values (Table 1). The identifications could be grouped into cholesterol esters (including cholesteryl acetate, cholesteryl nitrolineate), phosphoglycerides (phosphoethanolamines, phosphocholines, diacyl-phosphoglycerides), triglycerides and sphingolipids (sphingomyelins, ceramide).

**Discussion**

The present study was conducted to determine, if there are detectable differences in the lipid metabolites in the serum of horses infected with strongyles, when compared to uninfected horses. This study found that the concentrations of 66 lipid metabolites were apparently changed when infection was present (p<0.025). The present study was also able to tentatively identify 16 of the 66 metabolites that differed between infected and uninfected horses. Full and specific identification of the lipid profile would require the existence of positive control standards, which were not available at this time. The 16 tentatively identified metabolites were grouped according to lipid class into sphingolipids, triglycerides, cholesterol esters and phosphoglycerides.

A large proportion of sphingolipids are found in cell membranes. Sphingolipid metabolism is involved in the regulation of cell growth, differentiation and apoptosis. Sphingolipids also have key roles in cell signaling [10]. Administration of exogenous sphingolipids has been
The high prevalence (73.3%) of strongyle infection seen in the horses used for this study, is supported by previous publications [19, 20]. Because there is substantial variation in strongyle burden during the course of disease, future studies to evaluate the correlation between concentrations of metabolites and degree of strongyle infection should use quantitative techniques, such as the TaqMan PCR assay described by [21], to stratify the infection levels in the tested animals. The encysted immature non-laying strongyle stages are unlikely to contribute to the parasitic burden, leading to increased production and release of cholesterol based hormones such as epinephrine and corticosteroids.

Phosphoglycerides are the most abundant phospholipids in the majority of cell membranes [16]. In the present study, the concentrations of the majority of phosphoglycerides were increased with strongyle infection when compared with the control. A previous study proposed that parasitic nematodes produce and secrete glycoconjugates, in order to evade host immune responses [17]. In particular, phosphocholine, a small haptenic molecule frequently found on the antigens of parasites, is known to have immunomodulatory activities. Immunomodulation of the host immune system allows persistence of the parasite and prevents the development of severe pathology, by inhibiting the host cellular immune responses [18].

Further validation of the findings from this study is required, to confirm that the observed alterations of the lipid profile were specifically due to the presence of strongyle infection, and is caused by a specific strongyle species. The level of significance chosen for this study suggests that the probability of these changes being due to chance alone was very low. However, there is a possibility that some alterations may not have been specific to strongylosis, and were instead linked to more general systemic responses such as inflammation. As these horses were being euthanased at a commercial abattoir for human consumption, they should have been healthy and generally free of overt systemic disease. For future development of this technique it may be advisable to concurrently measure, markers of inflammation and obtain a known clinical and management history of each animal. This would increase the probability that differences in lipid metabolite profile between horses with moderate- or high-intensity strongyle infections, compared with their uninfected counterparts are specific to strongyle infection and the pathology it causes.

A low number of ascarid eggs were found in two fecal samples from two different animals. Serum from one of these two horses was used for lipid extraction and analysis as a control animal, as this horse did not have a strongyle infection. It is possible that the ascarid infection could have interfered with results, affecting their reliability as an indicator of no strongyle infection. However, as the uninfected serum samples were pooled before HPLC-MS analysis, any effect of this minor ascarid infection noticed, should have been minimized by the presence of the five other ascarid-negative samples.

In conclusion, this study has successfully used for the first time, high resolution HPLC-MS technique, to detect and tentatively identify lipids in equine serum on the basis of accurate mass. Statistical differences in

<table>
<thead>
<tr>
<th>Mass/charge</th>
<th>Retention</th>
<th>Ratio control</th>
<th>Potential IDs</th>
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<tr>
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<td>2.25</td>
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<td>0.34</td>
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the concentrations of 66 lipid metabolites between strongyle infected and uninfected horses were observed. These preliminary results demonstrate the potential use of this type of lipid analysis technique, for diagnosis of strongyle infection in equids. Future research in this area should be aimed at accurate and full identification of the lipid profiles of strongyle infected and healthy horses. Further studies to assess correlation between lipid concentrations and the intensity of strongyle infection, taking into account exact strongyle species in horses, is also required if development of a quantitative diagnostic test is to be achieved.

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References