

Acute Toxicity of Glyphosate-Based Herbicide Glycot on Juvenile African Cat Fish *Clarias gariepinus* (Burchell 1822)

Ani LC¹, Nwamba HO¹, Ejilibe CO¹ and Nwani CD^{2*}

¹Department of Applied Biology and Biotechnology, Enugu State University of Science and Technology, Enugu, Nigeria

²Department of Zoology and Environmental Biology, University of Nigeria, Nsukka, Nigeria

Abstract

The present study was designed to evaluate the acute toxicity of glyphosate on the juvenile of *Clarias gariepinus*. The acute toxicity bioassay was conducted to determine the 96h LC₅₀ values following the probit analysis method while the safe level of the test pesticide was estimated by multiplying the 96 h LC₅₀ with different application factors (AF). The 24, 48, 72 and 96h LC₅₀ values (with 95% confidence limits) estimated by probit analysis were 34.72(31.02-37.20), 31.90 (28.12-33.89), 27.40 (24.98-29.30), 24.60 (21.95-26.54) mgL⁻¹ respectively. There were significant differences ($p < 0.05$) in the LC50 values obtained at different exposure time. The safe level for the herbicide varied from 2.46×10^{-1} to 2.46×10^{-4} mgL⁻¹. Exposed fish showed uncoordinated behavior such as erratic and jerky swimming, jumping out of water, gulping of air, loss of equilibrium status, hyperactivity, decreased opercula movement and subsequently death. Mortality increased with increase in concentration of glyphosate and time of exposure. The study indicated that glyphosate has toxic effects on the *Clarias gariepinus*. The herbicide should be prudently used in both terrestrial and aquatic eco systems to avoid ecotoxicological hazards.

Keywords: *Clarias gariepinus*; Glyphosate; Toxicity; LC₅₀; Behavioural changes; Safe level

Introduction

Glyphosate, a broad-spectrum weedicide is one of the most frequently applied pesticides in agriculture for the control of great variety of annual biennial and perennial grasses, sedges, broad leaved weeds and woody shrubs [1]. It is also used for aquatic weed control in fish ponds, lakes, canals and slow running water [2].

Glyphosate is formulated as an isopropylamine salt and can be described as an organophosphorous compound. Glyphosate is described by the manufacturer as pesticide of low toxicity and environmental friendliness [3]. But research has shown that higher concentrations of the product can be toxic, producing a number of physiological changes in organisms and in some cases resulting to death depending on the level, duration and route of exposure [4]. Various concentrations of glyphosate have been shown to be toxic to juveniles of cat fishes, producing mortality, low survival and various abnormal behavioural changes such as loss of equilibrium status, air gulping, hyper activity, decreased opercula movement, erratic swimming and jerky movements which have been shown to be deleterious to the survival rate of the affected species[5].

In the world today, glyphosate is the most widely used herbicide and its consumption has increased to about 95% in the period from the year 2000 to 2004 [6]. Its high water solubility, extensive usage (especially in shallow water systems) and effects on non-target aquatic organism is a concern [7]. Studies have revealed that the responses of aquatic organisms to acute concentrations of glyphosate, its products and surfactants were variably possible due to differences in the formulation of the herbicide, level of mixture of component herbicide, species-specific sensitivity and condition of the media which are known to modify toxicity of the herbicide to exposed organisms [8].

Environmental factors such as pH, turbidity, alkalinity, dissolved oxygen, temperature and conductivity influence the rate of reaction of the pollutants entering the water or the lethal effects on the aquatic organisms. Due to increase in the use of glyphosate herbicide in the recent times, it becomes necessary to study the lethal toxicity and stress

of the herbicide on local species which would help in formulating the strategies for safe guarding aquatic organisms. The present study thus aims at the determination of the acute toxicity, behavioural responses and possible safe levels of African catfish *Clarias gariepinus* exposed to glyphosate herbicide.

Materials and Methods

Three hundred juveniles of *Clarias gariepinus* with mean weight of 32.16 ± 0.22 g and length of 17 ± 0.10 cm were procured from Sacen fish farm Enugu State, Nigeria and used for the investigation. The fish were acclimatized for fourteen days in four plastic aquaria tanks of 200 L capacity each. During the acclimation period, juveniles were fed twice daily with Copen feed (2 mm) at 3% body weight. Feeding was terminated 24 h before the experiment to empty their stomach and avoid pollution of the water with their faeces. For the present study, commercial formulation of glyphosate 41% SL with trade name "Glycot" manufactured by Sabero Organics Gujarat Ltd India and supplied by Afcott Nigeria plc were used.

Acute toxicity bioassay was conducted to determine the 96 h LC₅₀ values of commercial formulation of glyphosate (Glycot). A range finding test was carried out to determine the concentrations of the test solutions for the definitive test. This was determined by subjecting juveniles of *Clarias gariepinus* to different concentrations of glyphosate herbicide. In the definitive test, a set of 30 fish specimen in three replicates of 10 fish each were randomly exposed to each of the five

***Corresponding author:** Nwani CD, Department of Zoology and Environmental Biology, University of Nigeria, Nsukka, Nigeria, Tel: +234 806 360 7400; E-mail: chris.nwani@unn.edu.ng

Received August 29, 2017; **Accepted** September 22, 2017; **Published** September 30, 2017

Citation: Ani LC, Nwamba HO, Ejilibe CO, Nwani CD (2017) Acute Toxicity of Glyphosate-Based Herbicide Glycot on Juvenile African Cat Fish *Clarias gariepinus* (Burchell 1822). J Fisheries Livest Prod 5: 252. doi: [10.4172/2332-2608.1000252](https://doi.org/10.4172/2332-2608.1000252)

Copyright: © 2017 Ani LC, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

selected concentrations of glyphosate herbicide (18.5, 21.5, 24.4, 27.4, 30.4 and 33.4 mgL⁻¹). Another set of 10 juveniles were simultaneously maintained in water without test chemical and considered as control. The experiment was conducted in plastic aquaria (60 cm × 30 cm × 30 cm) containing 10 litres of water. The test solution was changed on every 24 h to counter-balance the decreasing pesticide concentration. During the experiment, fish behavior and other external changes in the body of fish were observed daily. Mortality of the fish due to glyphosate exposure was recorded up to 96 h at every 24 h interval to obtain LC₅₀ values of the test pesticides. The median lethal concentration (LC₅₀) value was determined following the probit analysis method described by Finney [9]. The safe level of the test pesticide was estimated by multiplying the 96 h LC₅₀ with different application factors (AF) and was based on Hart et al. [10], Sprague [11], Committee on Water Quality Criteria (CWQC) [12], National Academy of Science/National Academy of Engineering (NAS/NAE) [13], Canadian council of Resources and Environmental Ministry (CCREM) [14], and the International joint commission (IJC) [15]. The physicochemical properties of test water namely temperature, pH, total hardness and conductivity were analyzed using standard Procedures [16].

Statistical Analysis

The data obtained were statistically analyzed by statistical package SPSS (Version 17). These data were subjected to one way Analysis of Variance (ANOVA) and Duncan's multiple range test to determine the significance difference at 5% probability level.

Results

Physico-chemical parameters of the test water

The results of physico-chemical characteristics of the test water are presented in Table 1. The pH of water ranged from 8.00-8.75. The water temperature ranged from 25.9-27.7°C. The conductivity varied from 100-260 ppm, whereas total hardness ranged from 0.160-0.395 during the experimental period.

Behavioral response of fishes to different test concentrations

The behavioral responses of fish were observed in the exposed fish as well as in the control (Table 2). Normal swimming behaviour and

natural colour were observed in the control throughout the exposure period and in the lowest (18.5 mgL⁻¹) concentration of the herbicide at 24 h exposure period. In tanks with higher concentration of the test chemical, the fish swam erratically with Jerky movements and hyperactivity while body pigmentation was greatly reduced. Faster opercula movement, surfacing and gulping of air were observed. With increase in duration of the exposure, swimming and body movements were retarded. Later, fish lost balance, became exhausted, lost consciousness owing to respiratory incumbency and finally settled down passively at the bottom of the tank with the operculum wide open and ultimately died.

Fish survival and mortality at different test concentration and time intervals in *Clarias gariepinus* exposed to glyphosate

The numbers of survived and dead fish were examined depending on the duration of exposure (24, 48, 72 and 96 h) in *Clarias gariepinus*. The herbicide concentration of 33.3 mgL⁻¹ showed the highest fish mortality of 100% and lowest survival of 0% while no mortality was recorded in the control throughout the experiment (Table 3). This shows that increase in mortality rate results to decrease in survival rate of fish.

Median Lethal concentrations of glyphosate herbicide

The concentration of glyphosate herbicide that would bring about 50% mortality of test organism at different time interval is referred to as LC₅₀. The LC₅₀ values of different concentrations of glyphosate on *Clarias gariepinus* were found to be 34.72 (31.02-37.20), 31.90 (28.12-33.89), 27.40 (24.98-29.30) and 24.60 (21.95-26.54) mgL⁻¹ for 24, 48, 72 and 96 h exposure time, respectively (Table 4). Our results indicate that as the exposure time increases from 24 to 96 h, the median lethal concentration (LC₅₀) reduces.

Estimation of safe level

The concentration of glyphosate herbicide that is harmless to fish is referred to as safe level. The safe level of glyphosate estimated by different methods at 96 h exposure are presented in Table 5. The values of safe level of glyphosate in *Clarias gariepinus* varied from 24.6 × 10⁻¹ to 2.46 × 10⁻⁴.

Discussion

The study was carried out to evaluate the acute toxicity of glyphosate on the juvenile of *Clarias gariepinus*. Acute toxicity studies are the very first step in determining the water quality requirements of fish. This present study showed that exposure of the juvenile catfish to glyphosate resulted in increased mortality rate and decreased survival rate at different level of concentrations. The result is in agreement with the

Parameters	Range	Mean S.E
PH	8.00-8.75	8.375 ± 0.375
Temperature (°C)	25.9-27.7	26.80 ± 0.9
Conductivity (ppm)	100-260	180 ± 80
Total hardness	0.160-0.395	0.278 ± 0.118

Table 1: The physico-chemical parameters of the test water.

Exposure time	24 h							48 h							72 h							96 h													
	0	18.5	21.5	24.4	27.4	30.4	33.3	0	18.5	21.5	24.4	27.4	30.4	33.3	0	18.5	21.5	24.4	27.4	30.4	33.3	0	18.5	21.5	24.4	27.4	30.4	33.3							
Behaviour																																			
Hyper activity	-	-	-	+	+	+	+	-	-	-	+	+	+	+	-	-	-	+	+	+	-	-	-	+	+	+	+	+	+	+	+	+	+	+	
Equilibrium status	+++	+++	++	++	+	+	+	+++	+++	+++	++	+	+	+	++	++	++	++	++	+	-	+++	+	+	+	+	-	-							
Swimming rate	+++	+++	+++	+++	+	+	+++	+++	+++	+++	++	++	+	+	++	++	++	++	++	+	-	+++	+	+	+	+	-	-							
Fin movt	+++	+++	+++	+++	+	+	+++	+++	+++	+++	++	++	+	+	++	++	++	++	++	+	-	+++	+	+	+	+	-	-							
Jerky Movt	-	-	-	+	++	+++	+++	-	-	-	-	++	++	++	-	+	+	++	++	++	-	+	+	+	+	+	-	-							

Key=none -, mild +, moderate ++, Strong +++

Table 2: Behavioral changes at different test concentration and time intervals observed in *Clarias gariepinus* exposed to glyphosate.

Exposed concentration (mgL ⁻¹)	Number exposed	Periods (Hours)				% Survival	% Mortality
		24	48	72	96		
Control 0.00	30	0	0	0	0	100	0
18.5	30	0	3	3	3	90	10
21.5	30	0	6	6	9	70	30
24.4	30	3	6	9	15	50	50
27.4	30	6	6	12	18	40	60
30.4	30	9	12	18	24	20	80
33.3	30	12	21	27	30	0	100

Table 3: Fish survival and mortality at different test concentrations and time intervals in *Clarias gariepinus* exposed to glyphosate.

Concentration	24	48	72	96
LC ₁₀	25.57 ^a (21.08-27.02)	19.73 ^a (17.24-22.06)	19.60 ^c (17.24-21.86)	18.69 ^a (16.20-20.24)
LC ₂₀	28.40 ^a (23.42-30.92)	23.27 ^b (20.89-26.12)	21.99 ^c (20.00-23.02)	20.54 ^d (17.72-23.01)
LC ₃₀	30.60 ^a (27.20-33.02)	26.21 ^b (23.14-28.90)	23.89 ^c (21.02-25.69)	21.98 ^d (19.02-23.98)
LC ₄₀	32.68 ^a (28.40-34.60)	29.01 ^b (25.15-33.02)	25.65 ^c (23.10-27.61)	23.30 ^d (21.02-25.20)
LC ₅₀	34.72 ^a (31.02-37.20)	31.90 ^b (28.12-33.89)	27.40 ^c (24.98-29.30)	24.60 ^d (21.95-26.54)
LC ₆₀	36.88 ^a (32.10-38.27)	35.08 ^b (33.20-40.28)	29.28 ^c (25.02-33.98)	25.97 ^d (23.01-27.12)
LC ₇₀	39.35 ^a (35.08-41.95)	38.84 ^b (36.20-40.28)	31.43 ^c (27.10-35.28)	27.53 ^d (25.13-29.20)
LC ₈₀	42.45 ^a (38.17-44.01)	43.74 ^b (40.76-45.79)	34.15 ^c (30.48-38.12)	29.46 ^d (26.55-32.64)
LC ₉₀	47.15 ^a (44.19-50.12)	57.59 ^b (48.92-59.27)	38.32 ^c (35.12-41.27)	32.38 ^d (30.10-34.72)

Values with different alphabetic superscripts differ significantly ($p < 0.05$) between exposure time within lethal concentration.

Table 4: Lethal concentrations of glyphosate herbicide for *Clarias gariepinus* at different intervals.

Chemical	96 h LC ₅₀ (mgL ⁻¹)	Method	AF	Safe level (mgL ⁻¹)
Glyphosate	24.6	Hart et al. (1948)	-	8.08×10^{-1}
		Sprague (1971)	0.1	24.59×10^{-1}
		CWQC (1972)	0.01	24.59×10^{-2}
		NAS/NAF (1973)	0.1-0.00001	24.60×10^{-1} - 2.60×10^{-4}
		CCREM (1991)	0.05	1.23
		IJC (1977)	5% of 96hLC ₅₀	1.23

$C = 48 \text{ hLC}_{50} \times 0.03/S^2$, where C is the presumable harmless concentration and $S = 24 \text{ hLC}_{50}/48 \text{ hLC}_{50}$.

Table 5: Estimate of safe levels of glyphosate at 96h exposure time.

report of Olele and Zelibe [17] that fish and other aquatic organisms are harmed by herbicide contaminated water. Behavioural changes are the most sensitive indicators of potential toxic effects in fishes [18]. The observed behavioral alteration in the studied formulation of glyphosate are consistent with previous report on glyphosate based herbicides [19,20] and other pesticides like chloropyriphos [21], Profenofos [22], butachlor [23] and fenthion [24]. Fish exposed to higher concentrations of glyphosate exhibited faster swimming with jerky movement, gulping of air, faster opercula movement, loss of equilibrium, skin discoloration and subsequently death.

The median lethal concentration (LC₅₀) value reported in the present study for commercial formulation of glyphosate is higher than 1.05 mgL⁻¹ and 13.6 mgL⁻¹ reported by Ayoola [19] and Langiano and Martinez [25] when *Oreochromis niloticus* and *Prochilodus lineatus* were exposed to glyphosate and glyphosate-based herbicides respectively. The LC₅₀ obtained in the present study for glyphosate is lower than 108 mgL⁻¹ and higher than 3.74 mgL⁻¹ obtained by Clements et al. [26] and Shiogiri et al. [27] when bull frog tadpoles and neotropical fish (*Piarctus mesopotamicus*) were exposed to glyphosate-based herbicide roundup for 48 h. The LC₅₀ value however is lower than 620 mgL⁻¹ and 975 mgL⁻¹ 96h LC₅₀ reported by Shiogiri et al. [28] when *Cyprinus carpio* and *Paloceroscaudi maculatus* were exposed to glyphosate and glyphosate commercial formulation (Rodeo) herbicides respectively. The LC₅₀ value of 24.60 mgL⁻¹ reported in the present study is also lower than the 32.54 mgL⁻¹ obtained by Nwani et al. [29] when the same fish species (*Clarias gariepinus*) was exposed to similar glyphosate based

herbicide roundup. Previous literature indicates that the toxicity of glyphosate-based herbicides varies from one species to another species and even in strains of the same species. Toxicity is both concentration and time dependent, this account for the differences in the values obtained at different concentration and exposure times. Toxicity of chemicals to fish have been reported to be affected by water quality parameter (such as pH, temperature etc.), size and age, type of species, water quality, concentration and formulation of test chemicals [30].

The safe level obtained for glyphosate in the present study varied from 2.460 to 2.460×10^{-4} . However, due to large variation in safe levels as determined by different methods, the estimates of safe levels cannot be guaranteed [31]. Extrapolation of laboratory data to field is not always meaningful value and hence, it is difficult to decide on acceptable concentration based on laboratory experiments that may be considered safe in the field [32]. From this experiment, it is evident that glyphosate is toxic to fish and its application should be monitored and controlled to avoid possible eco-toxicological hazards.

Conclusion

From the present research it was evident that glyphosate is less toxic to *Clarias gariepinus* than other herbicide widely used in Nigeria. Most of the behavioral and physiological abnormalities were recorded mainly at higher concentrations. However, the herbicide should be prudently used in both terrestrial and aquatic eco-systems to avoid eco-toxicological hazards. More studies on the toxicity of glyphosate and its

formulations on catfishes are necessary to understand the mechanisms of actions of the herbicide.

Acknowledgements

The authors wish to thank the Head of Department of Applied Biology and Biotechnology, Enugu State University of Science and Technology for the support received during the conduct of the work.

Conflict of Interest

The authors declare that there are no conflicts of interest.

References

- Luschak, OV, Kubrak OI, Storey JM, (2009) Low toxic herbicide Roundup induces mild oxidative stress in goldfish tissue. *Chemosphere* 76: 932-937.
- Tsui MT, Chu IM (2008) Environmental fate and non-target impact of glyphosate-based herbicide (Roundup) in a subtropical wetland. *Chemosphere* 71: 439-446.
- Cox C (2001) Environmental contamination and ecological effects of Atrazine. *Journal of Pesticide Reform* 21: 12-20.
- Olaifa FE, Olaifa AK, Lawis OO (2003) Toxic stress of Lead on *Clarias gariepinus* (African catfish) fingerlings. *African Journal of Biomedical Research* 6: 101-104.
- Abdelgani AA, Tchounwou BB, Anderson AC, Monkiedje A (1997) Toxicity evaluation of single and mixture of roundup (garlon 3A, 2,4-1) and syndets surfactant to channel cat fish (*Ictalurus punctatus*), blue fill (*Lepomis macrochirus*) and crawfish (*Procambarus spp*). *Environmental Toxicology and Water Quality* 12: 237-244.
- Gluszczak L, Miron DS, Pedron FA, Vieira VLP (2006) Effect of glyphosate herbicide on acetylcholinesterase activity and metabolic and haematological parameters in Piava (*Laporinus obtusidens*). *Ecotoxicology and Environmental Safety* 65: 237-241.
- Peixoto F (2005) Comparative effects of the roundup and glyphosate on mitochondrial oxidative phosphorylation. *Comparative Biochemistry and Physiology* 147: 222-231.
- Gabriel UU, Mark P, Orlu EE (2009) Toxicity and behavioural changes in *heterobranchus bidorsalis* fingerlings treated with a micronutrient fertilizer, agrolyser. *Research Journal of Environmental and Earth Sciences* 1: 34-38.
- Finney DT (1971) Probit analysis. Cambridge University Press, Cambridge: 333.
- Hart WB, Weston RF, Dermann JG (1948) An apparatus for Oxygenating test solution which fish are used as test animals for evaluating toxicity. *Journal of the American Fisheries Society* 75: 228-236.
- Sprague JB (1971) Measurement of pollutant toxicity to fish. *International Bioassay methods for acute toxicity*. *Water Resources* 3: 793-821.
- Committee on Water Quality Criteria (1972) A report of the committee on water quality and research series, US Environmental Protection Agency Report; Cincinnati, OH, USA, EPA – R₃ – 73 – 003.
- National Academy of Science/National Academy of Engineering (NAS/NAE) (1973) Water quality criteria, EPA –R3-033; us Government Printing Office: Washington DC, USA.
- Canadian Council of Resources and Environmental Ministry (1991) Canadian Water Quality Guidelines; Canadian Council of Resources and Environmental Ministry, Inland waters Directorate, Environment Canada: Ottawa, On, Canada.
- International Joint Commission (IJC) (1977) New and Revised Great Lakes Water Quality Objectives. Great lake basin, Windsor; IJC: Ottawa, ON, Canada.
- APHA, AWWA, WPCF (2005) Standard methods for the examination of water and waste water 21st, 401 ed. Washington (DC): American Public Health Association.
- Olele NF, Zelibe SA (2013) Acute toxicity and behavioural changes on African catfish (*Clarias gariepinus*) exposed to dizensate (*glyphosate herbicide*). *International Journal of Scientific and Engineering Research* 4: 1-5.
- Banaee M, Sureda AR, Ahmadi K (2011) Effects of diazinon on biochemical parameters of blood in rainbow trout (*Oncorhynchus mykiss*). *Pesticide Biochemistry and Physiology* 99: 1-6.
- Ayoola SO (2008) Toxicity of glyphosate herbicide on Nile Tilapia (*Oreochromis niloticus*) juvenile. *African Journal of Agricultural Resources* 3: 825-834.
- Nwani CD, Lakra WS, Kumar R, Kushwaha B, Sirvastava SK (2010) Mutagenic and genotoxic effects of carbosulfan in fresh water air-breathing fish *Channa punctatus* (Bloch) using micronucleus assay and alkaline single cell gel electrophoresis. *Food and Chemical Toxicology* 48: 202-208.
- Ali D, Nagpure NS, Kumar R , Kushuwaha B (2009) Assessment of genotoxic and mutagenic in Fresh water fish *Channa punctatus* using micronucleus assay and alkaline single cell gel electrophoresis. *Food and Chemical and Toxicology* 47: 650-656.
- Pandey AK, Trivedi PS, Lakra WS (2011) Investigation on acute toxicity and behavioural changes in *Channa punctatus* due to organophosphate pesticide (Profenofos). *Drug Chemistry and Toxicology* 34: 424-428.
- Chang J, Liu S, Zhou S, Wang M, Zhu G. (2013) Effects of butachlor on reproduction and hormone levels in adult zebra fish (*Danio rerio*). *Experimental Toxicology Pathology* 65: 205-209.
- Nwani CD, Somdare JA, Ukonze VC, Ejere AO, Nwadingwe JC, et al. (2016) Subchronic exposure to fenthion induce hematological changes in African catfish *Clarias gariepinus* (Burchell 1822). *Journal of Aquatic Animal Health* 28: 229-234.
- Langiano VC, Martinez CB (2008) Toxicity and effects of glyphosate-based herbicide on the Neotropical fish *Prochilodus lineatus*. *Comparative Biochemistry and Physiology* 147: 222-231.
- Clements C, Ralph S, Petras M (1997) Genotoxicity of select herbicides in (*Rana catesbeiana*) tadpoles using the alkaline single cell gel electrophoresis (Comet) Assay. *Environ Mol Mutagen* 29: 277-288.
- Shiogiri NS, Paulino MG, Carraschi SP, Baraldi FG, da Cruz C, et al. (2012) Acute exposure to glyphosate-based herbicide affects the gills and liver of the Neotropical fish, *Piaractus mesopotamicus*. *Environmental Toxicology Pharmacology* 34: 388-396.
- Shiogiri NS, Cubo P, Schiavetti C, Pitelli RA (2010) Ecotoxicity of glyphosate and aterbane ® br surfactant on guaru (*Phalloceros caudimaculatus*). *Journal of Animal and Biological Science* 32: 285-289.
- Nwani CD, Nagpure NS, Kumar R, Kushwaha B, Lakra WS (2013) DNA damage and oxidative stress modulatory effects of glyphosate-based herbicide in freshwater fish *Channa punctatus*. *Environmental Toxicology Pharmacology* 36: 539-547.
- Kumar R, Nagpure B, Srivastava SK, Lakra WS (2009) Investigation of the genotoxicity of malathion to freshwater teleost fish (*Channa punctatus*) using the micro nucleus test and comet assay. *Environmental Contamination and Toxicology* 58: 123-130.
- Buikema JR, Naider-lehner AL, Cairns JR (1982) Biological monitoring Part IV - toxicity testing. *Water Research* 16: 239-262.
- Elmegaard N, Jagers GA, Akkerhuis OP (2000) Ministry of Environment and Energy, National Research Institute, Safety Factors in Pesticide Rise Assessment. Differences in Species sensitivity and Acute chronic Relations NERI Technical Report.