

# Amyloid- $\beta$ Oligomers and Aluminum Co-Aggregate to Form Toxic Amyloid Channels in Alzheimer's Disease Brain: A New "Amyloid- $\beta$ Channel-Aluminum Hypotheses"

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## Abstract

Large numbers of senile plaques are thought to be characteristic of Alzheimer's disease (AD), but these deposits are also a by-product of normal senescence. In AD and normal brains, senile plaques are primarily composed of amyloid- $\beta$  peptides (A $\beta$  P) with aluminum (Al). Evidence suggests the oligomerization of A $\beta$  P is part of the molecular mechanism of AD pathogenesis by forming neurotoxic amyloid channels. However, the relationship between Al and AD has been a subject of scientific debate for many years. The complex nature of Al bioavailability has made it difficult to evaluate its toxicity to the human brain. In 2004, Al concentration in CSF of AD patients analyzed to be  $1.6 \pm 0.4$  times higher than normal people. Importantly, AD patients with more Al in CSF showed less MMSE score, indicating Al may decrease cognitive ability. Recently, Al accumulations in sporadic AD and familial AD brains were reported to be much higher than in normal control brains. Above its neurotoxicity, Al<sup>3+</sup> has a crucial role as a cross-linker in  $\beta$ -amyloid oligomerization. Therefore, I propose a hypothesis that  $\beta$ -amyloid oligomerizes with Al, forming non-specific cation amyloid channels in cell membranes, which allows calcium to enter cells and finally causes neuronal death by together with Al's own neurotoxicity.

**Keywords:** Beta amyloid; Senile plaque; Aluminum; Amyloid channel; Calcium influx; Oligomer; Alzheimer's disease

## Abbreviations:

A $\beta$  P: Amyloid  $\beta$  Peptides; Al: Aluminum; AD: Alzheimer's Disease; ALS: Amyotrophic Lateral Sclerosis; NFTs: Neurofibrillary Tangles

## Introduction

Alzheimer's disease (AD), Parkinson's disease, and amyotrophic lateral sclerosis (ALS), such neurodegenerative diseases are characterized by the loss of neurons, cognitive decline and motor impairment. These diseases share the common assemblage of misfolded or aggregated intracellular or extracellular peptides or proteins, known as inclusion bodies. Senile plaques are extracellular deposits of amyloid- $\beta$ -peptides (A $\beta$  P) in the brain. According to the prevailing original "amyloid cascade hypothesis" [1], senile plaques are responsible for the pathology of AD; this hypothesis used to be accepted by the majority, but is not yet conclusively established. An alternative hypothesis is that some types of amyloid oligomers rather than senile plaques are responsible for the disease pathogenesis [2,3]. Recent evidence together with old ones, showed a need to put aluminum (Al) on the patho-physiology and patho-chemistry of AD.

## Oligomers of A $\beta$ P form neurotoxic channels

A $\beta$  P is inclined to aggregate with Al to make various types of oligomers. Some oligomers of A $\beta$  P are inserted into neural cell membranes to form amyloid- $\beta$  channels that are neurotoxic [4-6]. These A $\beta$  P deposits as senile plaque which are also a by-product of

normal aging process. A $\beta$  P (1-40) and A $\beta$  P (1-42) also seem to feature highly different conformational states [7], with the C-terminus of A $\beta$  P (1-42) being more structured than that of the 1-40 peptide fragment.

A $\beta$  P may damage cells and finally do neurons to death. The mechanism starts by generating non-specific cation channels (like pores in the membrane). As a result, the amyloid- $\beta$  channel promotes depolarization of the synaptic membrane, to cause excessive calcium influx through the channel, and mitochondrial impairment. Dissociated hippocampal and cerebral cortical neurons from embryonic brain form many functional synapses in culture. It appears that Al promoted the aggregation of A $\beta$  P and enhanced its neurotoxicity in the cortical neurons, as well [8]. The abnormal aggregation of A $\beta$  P and Al own neurotoxicity in neuronal cells is critical for the onset of Alzheimer's disease. It was shown that toxic A $\beta$  P (1-40) aggregates formed channels in GM1-ganglioside-containing membranes. A $\beta$  P formed without membranes were thinner and much less toxic, because of weaker binding to cell membranes and a less surface hydrophobicity [9].

Increases in either total A $\beta$  P levels or the relative concentration of both A $\beta$  P (1-40) and A $\beta$  P (1-42) [10] have been implicated in the pathogenesis of both familial and sporadic AD. Due to its more hydrophobic character, the A $\beta$  P (1-42) is the most amyloid-like form of the peptide. Indeed, data indicated that, when in soluble intracellular form, the oligomers of A $\beta$  P (1-42) (a toxic species of A $\beta$  P), acutely inhibited synaptic functions, various types of patho-physiology that characterizes AD [11-13]. Genetically engineered mice to express oligomers but not plaques (APPE693Q) develop AD-like symptoms. On the other hand, mice engineered to convert oligomers

into plaques (APPE693Q X PS1 $\Delta$ E9), are no more cognitively impaired than the oligomer-only AD-like mice [14].

### Accumulation of neurotoxic aluminum (Al) in AD brain

Although distributed environmentally abundant, aluminum is never essential for life, but is recognized as cell-toxic, especially as neurotoxic. Al inhibits thousands of biologically important functions and causes a lot of adverse effects in plants, animals, and, of course, in humans. The relationship between Al exposure and neurodegenerative diseases has been suggested, including dialysis-encephalopathy, AD [15] and ALS, and Parkinson-dementia complex in the Kii Peninsula of Japan and in Guam.

It was reported an accumulation of Al in neurofibrillary tangles (NFTs)-bearing neurons of AD brains [16]. An accumulation of Al in both senile plaques and NFTs has been reported in renal failure patients [17]. In 2004, Al accumulation was also analyzed in cerebrospinal fluid (CSF) of AD patients and Pick disease patients. Shoda et al. reported that Al concentration in CSF of AD patients were a  $1.6 \pm 0.4$  higher, and in Pick disease case  $2.5 \pm 1.0$  times higher than normal people. Al concentration of CSF in cerebrovascular dementia patients is not significantly different from normal levels. Importantly, the more Al in CSF of dementia patients shows the less MMSE score, indicating Al may decrease recognition ability [18].

Recently, Yumoto et al. analyzed Al using energy-dispersive X-ray spectroscopy with transmission electron microscopy. Their detailed careful analysis demonstrated that Al was present in cores of senile plaques [19]. In 2012, sporadic AD brains suggested that a diagnosis of AD could be predicted by a combination of A $\beta$  P pathology and ratio of brain concentration of Al to copper [20]. Very recently Exley's group published new paper, stating that "the first ever measurements of Al in brain tissue from 12 donors diagnosed with familial AD. The concentrations of Al were extremely high, for example, there were values in excess of 10  $\mu$ g/g tissue dry wt. in 5 of the 12 individuals. Overall, the concentrations were higher than all previous measurements of brain Al except cases of known Al-induced encephalopathy" [21].

### Effect of Al on the forming oligomers of A $\beta$ P

Many studies on biochemistry, cell biology, toxicology, and genetics have favored a hypothesis, namely, that the oligomerization of A $\beta$  P and its neurotoxicity play a central role in the pathogenesis of AD [22].

Genetic studies of familial AD indicated that APP mutations and A $\beta$  P metabolism are associated with AD [23]. The first 40 amino acid residues of A $\beta$  P (1-40) caused the cell-death of cultured rat hippocampal neurons or neurodegeneration in the brains [24].

A $\beta$  P is a hydrophobic peptide with a tendency to self-assemble and form SDS-stable oligomers in aqueous solution. The monomeric form of A $\beta$  P has a random coiled structure. Oligomeric A $\beta$  Ps have  $\beta$ -pleated sheet structures and form amyloid channels in cell membranes, but in the normal aging process finally form insoluble aggregates, mainly termed senile plaques. Using size-exclusion chromatography, gel electrophoresis, and atomic force microscopy, it is demonstrated that the soluble oligomers are neurotoxic and impair synaptic plasticity [25].

A $\beta$  P is secreted in the CSF of young normal individuals as well as in aged or dementia patients. Factors to accelerate or inhibit the

oligomerization can have fundamental roles in the pathogenesis of AD [26].

Intriguingly, rodent A $\beta$  P showed the less tendency to oligomerize than human A $\beta$  P *in vitro* [27] and deposits of A $\beta$  P are rarely observed in the brains of rodents as compared to humans. The amino acid sequences of rodent A $\beta$  P differ from human with 3 amino acids, namely Arg5, Tyr10, and His13. All three amino acids have the strong ability to bind metals. Therefore, trace metals, especially Al<sup>3+</sup>, are particularly of interest as potential accelerators and may play important roles in the accumulation of A $\beta$  P in the human brain.

Exley et al. demonstrated by CD spectroscopy that Al<sup>3+</sup> induces a conformational change in A $\beta$  P (1-40) [28]. We have demonstrated that Al enhances polymerization of A $\beta$  P (1-40) and forms SDS-stable oligomers *in vitro* [29]. Oligomerization induced by Al<sup>3+</sup> is stronger than that induced by other metals. Furthermore, Al-aggregated A $\beta$  P binds tightly to the surface of cultured neurons of rat cerebral cortex and forms fibrillar deposits [30]. Secreted A $\beta$  P is usually degraded by various proteases such as neprilysin within a short time. Down regulation of neprilysin induced by Al<sup>3+</sup> can cause the accumulation of A $\beta$  P [31]. Indeed, Al<sup>3+</sup> has been shown to inhibit degradation of A $\beta$  P as a result of conformational changes [32]. More interestingly, A $\beta$  P combined with Al is more toxic than normal A $\beta$  P, and may easily form amyloid channels, causing membrane disruption and interruption of neural Ca<sup>2+</sup> homeostasis and mitochondrial functions [33,34].

Rat orally administered Al<sup>3+</sup> caused a marked increase in the amount of A $\beta$  P both in its secreted and accumulated forms, and increased deposition of senile plaques in AD-model mice genetically transfected with the human APP gene (Tg 2576) [35]. These results are consistent of other studies demonstrating that oral Al<sup>3+</sup> exposure causes the accumulation of A $\beta$  P and impairs spatial learning memory in AD-model mice [36]. Very recently, it was reported that only A $\beta$  P (1-42) oligomers, but not significantly A $\beta$  P (1-40) oligomers, appeared to form 3 types of amyloid channels in neuronal cells [37]. From the standpoint of "Amyloid- $\beta$  channel-Al hypotheses", the results seem to be quite possible, because so little Al<sup>3+</sup> exist in the medium under the *in vitro* experimental condition, compare to considerable amounts of Al<sup>3+</sup> exist *in vivo* human CSF [18] to form a stable and convenient chemical structure for the channel formation in the membrane.

### Conclusion

The etiology of AD seems to be dependent on the interaction of two neurotoxic substances, A $\beta$  P and Al<sup>3+</sup> ions. A new "Amyloid- $\beta$  channel-Al hypothesis" will follow. A $\beta$  P oligomers with Al<sup>3+</sup> are readily incorporated into cell membranes, resulting in the formation of Ca<sup>2+</sup>-permeable amyloid channels. A following influx of Ca<sup>2+</sup> through these amyloid channels leads to the phosphorylation of tau, depletion of neurotrophic factors and the formation of free radicals, and so on, with final neuronal death. Al<sup>3+</sup> neurotoxicity also blocks various Ca<sup>2+</sup> channels and influences Ca<sup>2+</sup> homeostasis.

Further research is necessary to understand fully about AD.

### References

1. Selkoe DJ (1991) The molecular pathology of Alzheimer's disease. *Neuron* 6: 487-498.
2. Neri LC, Hewitt D (1991) Aluminium, Alzheimer's disease, and drinking water. *The Lancet* 338: 390.
3. Castorina A, Tiralongo A, Giunta S, Carnazza ML, Scapagnini G, et al. (2010) Early effects of aluminum chloride on beta-secretase mRNA

- expression in a neuronal model of  $\beta$ -amyloid toxicity. *Cell Biol Toxicol* 26: 367-377.
4. Arispe N, Pollard HB, Rojas E (1996)  $Zn^{2+}$  interaction with Alzheimer amyloid beta protein calcium channels. *Proc Natl Acad Sci USA* 93: 1710-1715.
  5. Kawahara M, Kuroda Y, Arispe N, Rojas E (2000) Alzheimer's beta-amyloid, human islet amylin, and prion protein fragment evoke intracellular free calcium elevations by a common mechanism in a hypothalamic GnRH neuronal cell line. *J Biol Chem* 275: 14077-14083.
  6. Arispe N, Diaz J, Durell SR, Shafir Y, Guy HR (2010) Polyhistidine peptide inhibitor of the Abeta calcium channel potently blocks the Abeta-induced calcium response in cells. Theoretical modeling suggests a cooperative binding process. *Biochemistry* 49: 7847-7853.
  7. Sgourakis NG, Yan Y, McCallum SA, Wang C, Garcia AE (2007) The Alzheimer's peptides A $\beta$ 40 and 42 adopt distinct conformations in water: A combined MD/NMR study. *J Mol Biol* 368: 1448-1457.
  8. Kuroda Y, Kobayashi K, Ichikawa M, Kawahara M, Muramoto K (1995) Application of long-term cultured neurons in aging and neurological research: Aluminum neurotoxicity, synaptic degeneration and Alzheimer's disease. *Gerontology* 1: 2-6.
  9. Okada T, Ikeda K, Wakabayashi M, Ogawa M, Matsuzaki K (2008) Formation of toxic Abeta(1-40) fibrils on GM1 ganglioside-containing membranes mimicking lipid rafts: Polymorphisms in Abeta (1-40) fibrils. *J Mol Biol* 382: 1066-1074.
  10. Luo Y, Niu F, Sun Z, Cao W, Zhang X, et al. (2009) Altered expression of A metabolism-associated molecules from D-galactose/AlCl<sub>3</sub> induced mouse brain. *Mechan Ageing Develop* 130: 248-252.
  11. Lukiw Z, Percy ME, Kruck TP (2005) Nano molar aluminum induces pro-inflammatory and pro-apoptotic gene expression in human brain cells in primary culture. *J Inorg Biochem* 99:1895-1898.
  12. Lin R, Chen X, Li W, Han Y, Liu P, et al. (2008) Exposure to metal ions regulates mRNA levels of APP and BACE1 in PC12 cells: Blockage by curcumin. *Neurosci Lett* 440: 344-347.
  13. Walton JR, Wang MX (2009) APP expression, distribution and accumulation are altered by aluminum in a rodent model for Alzheimer's disease. *J Inorganic Biochem* 103: 1548-1554.
  14. Gandy S, Simon AJ, Steele JW, Lublin AL, Lah JJ, et al. (2010) Days to criterion as an indicator of toxicity associated with human Alzheimer amyloid-beta oligomers". *Annals of Neurol* 68: 220-230.
  15. McLachlan DR (1995) Aluminium and the risk for Alzheimer's disease. *Environmetrics* 6: 233-275.
  16. Bouras C, Giannakopoulos P, Good PF, Hsu A, Hof PR, et al. (1997) A laser microprobe mass analysis of brain aluminum and iron in dementia pugilistica: Comparison with Alzheimer's disease. *Euro Neurol* 38: 3-58.
  17. Edwardson JA, Candy JM (1989) Aluminium and the pathogenesis of senile plaques in Alzheimer's disease, Down's syndrome and chronic renal dialysis. *Ann Med* 21: 95-7.
  18. Shoda T, Sasaki K, Kuroda Y, Kawahara M, Onti M (2004) Micro metallic elements (Al, Cu, Zn, Fe) in cerebrospinal fluid. *Nihon Rinsho* (in Japanese) 62: 183-189.
  19. Yumoto S, Kakimi S, Ohsaki A, Ishikawa A (2009) Demonstration of aluminum in amyloid fibers in the cores of senile plaques in the brains of patients with Alzheimer's disease." *J Inorgan Biochem* 103: 1579-1584.
  20. Exley C, House E, Polwart A, Esiri MM (2012) Brain burdens of aluminium, iron and copper and their relationships with amyloid- $\beta$  pathology in 60 human brains. *J Alzheimers Dis* 31: 725-730.
  21. Mirza A, King A, Troakes C, Exley C (2017) Aluminium in brain tissue in familial Alzheimer's disease. *J Trace Elem Med Biol* 40: 30-36.
  22. Yankner BA, Duffy LK, Kirschner DA (1990) Neurotrophic and neurotoxic effects of amyloid  $\beta$  protein: reversal by tachykinin neuropeptides. *Science* 250: 279-282.
  23. Goate A, Chartier-Harlin MC, Mullan M (1991) Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* 349: 704-706.
  24. Chaussidon M, Netter P, Kessler M, Membre H, Fener P, et al. (1993) Dialysis associated arthropathy: Secondary ion mass spectrometry evidence of aluminum silicate in  $\beta$ -microglobulin amyloid synovial tissue and articular cartilage. *Nephron* 65: 559-563.
  25. Selkoe DJ (2008) Soluble oligomers of the amyloid  $\beta$ -protein impair synaptic plasticity and behavior. *Behav Brain Res* 192: 106-113.
  26. Fukuyama R, Mizuno T, Mizuno T et al. (2000) Age-dependent change in the levels of A $\beta$ 40 and A $\beta$ 42 in cerebrospinal fluid from control subjects, and a decrease in the ratio of A $\beta$ 42 to A $\beta$ 40 level in cerebrospinal fluid from Alzheimer's disease patients. *Eur Neurol* 43: 155-160.
  27. Dyrks T, Dyrks E, Masters CL, Beyreuther K (1993) Amyloidogenicity of rodent and human  $\beta$ A4 sequences. *FEBS Lett* 324: 231-236.
  28. Exley C, Price NC, Kelly SM, Birchall JD (1993) An interaction of  $\beta$ -amyloid with aluminium in vitro." *FEBS Lett* 324: 293-295.
  29. Kawahara M, Muramoto K, Kobayashi K, Mori H, Kuroda Y (1994) Aluminium promotes the aggregation of Alzheimer's amyloid  $\beta$ -protein in vitro. *Biochem Biophys Res Com* 198: 531-535.
  30. Kawahara M, Kato M, Kuroda Y (2001) Effects of aluminum on the neurotoxicity of primary cultured neurons and on the aggregation of  $\beta$ -amyloid protein. *Brain Res Bull* 55: 211-217.
  31. Luo Y, Niu F, Sun Z, (2009) Altered expression of A $\beta$  metabolism-associated molecules from d-galactose/AlCl<sub>3</sub> induced mouse brain. *Mech Ageing Dev* 130: 248-252.
  32. Sakamoto T, Saito H, Ishii K, Takahashi H, Tanabe S, et al. (2006) Aluminum inhibits proteolytic degradation of amyloid  $\beta$  peptide by cathepsin D: A potential link between aluminum accumulation and neuritic plaque deposition. *FEBS Lett* 580: 6543-6549.
  33. Ricchelli F, Drago D, Filippi B, Tognon G, Zatta P (2005) Aluminum-triggered structural modifications and aggregation of  $\beta$ -amyloids. *Cell Mol Life Sci* 62: 1724-1733.
  34. Drago D, Cavaliere A, Mascetra N, Ciavardelli D, di Ilio C, et al. (2008) Aluminum modulates effects of  $\beta$ -amyloid1-42 on neuronal calcium homeostasis and mitochondria functioning and is altered in a triple transgenic mouse model of Alzheimer's disease. *Rejuvenation Res* 11: 861-871.
  35. Pratic OD, Uryu K, Sung S, Tang S, Trojanowski JQ, et al. (2002) Aluminum modulates brain amyloidosis through oxidative stress in APP transgenic mice. *The FASEB J* 16: 1138-1140.
  36. Rodella LF, Ricci F, Borsani E, Stacchiotti A, Foglio E, et al. (2008) Aluminium exposure induces Alzheimer's disease-like histopathological alterations in mouse brain. *Histol Histopathol* 23: 433-439.
  37. Bode DC, Baker MD, Viles JH (2017) Ion channel formation by amyloid- $\beta$ 42 oligomers but not amyloid- $\beta$ 40 in cellular membranes. *J Biol Chem* 292: 1404-1413.