

## "OUJDBODFS 1PUFOUJBMTPG 1IZUPDIFNJDBMT .FEJDJOBM 1MBOU TP G 8FTU "GSJDB

Joseph O Nwankwo\*

Department of Medical Biochemistry, Faculty of Basic Medical Sciences, Federal University, Ndufu-Alike Ikwo, Ebonyi State, Nigeria

### Abstract

A selected number of potential anticancer agents in phytochemicals isolated from some indigenous food and medicinal plants in the traditional medicine of the region, and as nutritional sources for countless generations [1-3]. Isolation and characterization of the chemical structures of phytochemicals from useful indigenous plants have however been undertaken only for a minimal fraction, largely because the modern techniques of chromatography and mass spectroscopy have been exploited by African scientists relatively recently [4-6]. The recent development of the latter has been to greatly enhance an understanding of the molecular mechanisms of action for these isolated compounds as pharmacological agents. In particular, such knowledge has been applied in the present discussion to identify phytochemicals from indigenous West African plants with potential anticancer activities, based on the structure-activity relationships to known active compounds. An outcome totally unanticipated by the organic chemists who merely isolated and characterized these phytochemicals, or whose main focus was on other biological activities.

**Keywords:** Anticancer; phytochemicals; West Africa; Alkaloids; phytochemical investigation of Nigerian plant species may yield flavonoids; Terpenoids; maytansinoids, as is the case with its near relatives in East Africa.

### Introduction

Many indigenous West African plants have been employed as local remedies for various human ailments in the traditional medicine of the region, and as nutritional sources for countless generations [1-3]. Isolation and characterization of the chemical structures of phytochemicals from useful indigenous plants have however been undertaken only for a minimal fraction, largely because the modern techniques of chromatography and mass spectroscopy have been exploited by African scientists relatively recently [4-6]. The recent development of the latter has been to greatly enhance an understanding of the molecular mechanisms of action for these isolated compounds as pharmacological agents. In particular, such knowledge has been applied in the present discussion to identify phytochemicals from indigenous West African plants with potential anticancer activities, based on the structure-activity relationships to known active compounds. An outcome totally unanticipated by the organic chemists who merely isolated and characterized these phytochemicals, or whose main focus was on other biological activities.

At a selected few plants would yield the interesting examples discussed here, underscores the fact that a programmed, methodical investigation of the vast flora of the region, in particular, the prodigious rich tropical rain forests, would uncover a resource of an unimaginable magnitude and inestimable value as anticancer agents. After all, it is on record, that the first universally acclaimed and potent anticancer agents were discovered from plants of the African continent and include such famous examples as vincristine and vinblastine, both isolated from the Madagascar periwinkle plant, *Catharanthus roseus* L. Don (Synonym: *Madagascar periwinkle*). These compounds were touted as: "the most successful of the plant-derived antitumor agents" [7]. Another significant example is maytansin, isolated from the Ethiopian shrub *Maytenus serrata* (Celastraceae) and was one of the most promising antitumor agents discovered by the commendable program for extensive screening of antitumor principles from plants, by the National Cancer Institute of the USA [8]. The Kenyan plant species *Maytenus buchananiana* reported to be a rich source of maytansin, yielding more than seven times the amount from *M. serrata* [7]. A *Maytenus* species indigenous to West Africa and commonly found in Nigerian states (Senegalensis) *Gymnosporia senegalensis* (Celastraceae). It is very likely that a

selected number of potential anticancer agents in phytochemicals isolated from some indigenous food and medicinal plants in the traditional medicine of the region, and as nutritional sources for countless generations [1-3]. Isolation and characterization of the chemical structures of phytochemicals from useful indigenous plants have however been undertaken only for a minimal fraction, largely because the modern techniques of chromatography and mass spectroscopy have been exploited by African scientists relatively recently [4-6]. The recent development of the latter has been to greatly enhance an understanding of the molecular mechanisms of action for these isolated compounds as pharmacological agents. In particular, such knowledge has been applied in the present discussion to identify phytochemicals from indigenous West African plants with potential anticancer activities, based on the structure-activity relationships to known active compounds. An outcome totally unanticipated by the organic chemists who merely isolated and characterized these phytochemicals, or whose main focus was on other biological activities.

### Cancer Chemopreventive Agents

#### A. Induction of carcinogen metabolizing enzymes

From fresh seeds of an indigenous plant, *Bitter Melon*

\*Corresponding author: Joseph O Nwankwo, Department of Medical Biochemistry, Faculty of Basic Medical Sciences, Federal University, Ndufu-Alike Ikwo, Ebonyi State, Nigeria, Tel: +234 706 267 7888; E-mail: joseph.nwankwo@funai.edu.ng

Received August 09, 2017; Accepted October 12, 2017; Published October 22, 2017

Citation: Nwankwo JO (2017) Anticancer Potentials of Phytochemicals from Some Indigenous Food and Medicinal Plants of West Africa. Adv Cancer Prev 2: 124. doi: 10.4172/2472-0429.1000124

Copyright: © 2017 Nwankwo JO. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

kola Heckel (Guttiferae), was isolated kolaviron (KV), a mixture of three flavonoids; GB-11, GB-2 and kolaavanone, KF3, [12]. Previous studies had demonstrated that kolaviron reduced hepatotoxicity in experimental animals [12-14]. A subsequent study by this author, indicated that it could inhibit a toxin B1 (AFB1)-induced genotoxicity in the human liver-derived HepG2 cells [11]. Kolaviron, at the highest tested concentration of 90 µM reduced AFB1-induced unscheduled DNA synthesis, measured by <sup>3</sup>H-thymidine incorporation into HepG2 cells, by 53%. The latter study further demonstrated that kolaviron's mechanism of action involved an induction of the AFB1-detoxifying enzymes such as cytochrome P450 3A4, and glutathione S-transferases A1-1, A2-2, M1-B, indicated by a three-fold increase in the transcripts for the enzymes, and a two-fold increase for GST A1-1 and A2-2 protein. Kolaviron therefore appeared to have a chemopreventive potential in inhibition of human AFB1-induced genotoxicity and possibly, carcinogenesis (Figures 1 and 2).

Bitter kola (Garcinia kola) seed from which kolaviron was obtained, is an edible nut frequently consumed as "kola", or guest snack both at homes and at social gatherings in the southern parts of Nigeria. Two flavonoids structurally similar to kolaviron were also isolated from the stem bark of *G. densivenia* [15] and named, morellavone, and its methyl ether derivative, O-methylfukugenein. Many other flavonoids have been isolated from various species of the Douala-Edea Forest Reserve of West Cameroon, some of which could display cancer chemopreventive activities similar to kolaviron [16]. These compounds are yet to be investigated for possible anticancer activity.

In a follow-up study, the potentials of kolaviron as a pleiotropic inducer of hepatoprotective genes, especially in a toxin B1 toxicant on human hepatocytes, was evaluated by gene microarray assay [17]. This work employed human primary hepatocytes (HPH) separately treated with AFB1 (2.0 µM/L) and KV (90 µM/L) then a combined pretreatment with kolaviron before AFB1, while gene expression analysis employed the Affymetrix Gene 1.0ST array containing the entire human genome. Results suggested a central role for the human "Inhibitor of DNA binding 1, ID1 gene" (Gene ID: 3397; MIM: 600349), which was significantly up-regulated to about three-fold message levels, in all three treatments, as well as a few other genes that were separately regulated [17].

Figure 2: 4/5  
4: R = OH  
5: R = OCH3

## Cancer Chemotherapeutic Agents

### A. Cytotoxicity to tumor cells

**Alkaloids: Benzophenanthridine alkaloids:** Benzophenanthridine alkaloids, nortidinechloride, (and a new fagaron derivative, 6-methoxy-5,6-dihydro nortidine, were isolated from *Fagara macrophylla* [18]. Both compounds were shown to be about equipotent in the P-388 mouse leukemia system, giving high T/C values of 240% and 260% respectively, at doses of 30-50 mg/kg. Another derivative, fagaronine, was isolated from the Ghanaian and Nigerian plant, *Fagara zanthoxyloides* (Rutaceae) [19]. Fagaronine, like its relative, nortidine, is active in both the P-388 and L-1210 leukemia test systems [20]. A latter study suggests that the structural determinants for antitumor activities in benzophenanthridines, include the existence of a planar, catenar resonance hybrid between the keto-amine and the ether on forms in neutral media, and a phenolic OH at C-7 [21]. It therefore becomes necessary to review the activities of the subsequent discovered derivatives as described below (Figures 3 and 4).

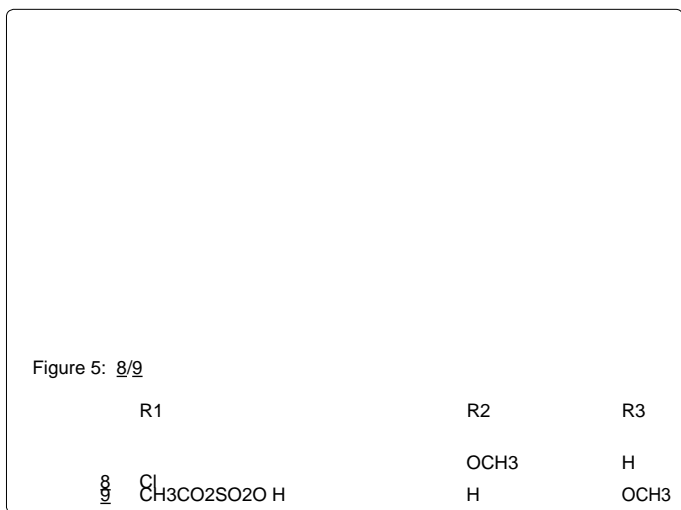
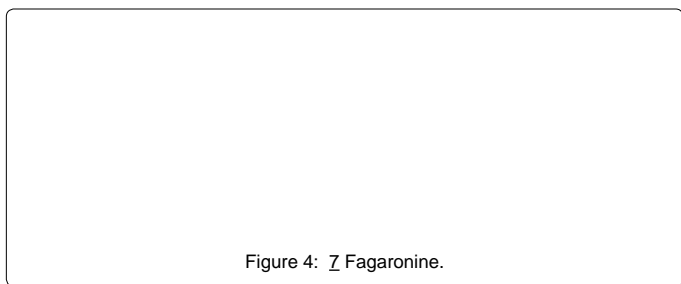
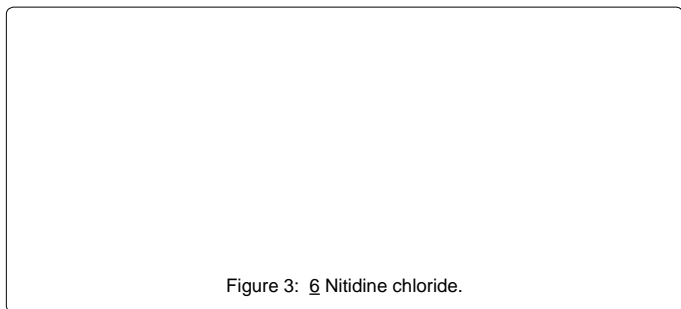
One such alkaloid bearing structural similarities to a known antitumor compound, palmatine (extracted among other alkaloids, from roots of the West African climbing shrub *Corymbium racemosum*) [22]. Palmatine, is structurally similar to coralyne sulfacetate, (the latter being one of the related alkaloids of nortidinechloride, which showed antileukemic activity [23].

A decoction of the stem bark of *Fagara macrophylla* taken as an antiseptic for the genitourinary tract, and is also applied as a rubefacient in toothaches, in the folk medicine of Nigeria [16]. Extracts of the roots are also used for coughs and colds. *Corymbium racemosum* (Menispermaceae), is a climbing shrub indigenous to the forests of Ghana and other parts of West Africa, whose roots are added to palm wine, and extracts of the plant used medicinally as nasal drops, and as an aphrodisiac (Figure 5) [22].

**Peptide alkaloids:** For peptide alkaloids, the isolation of 2 new cyclic octapeptidides designated phakellastanin 10 and phakellastanin 11 from the marine sponge *Phakellia sphaera* has been reported [24]. Both compounds demonstrated significant cytotoxic activity against the P-388 leukemia cell line, having ED50 values of 2.1 and 0.2 µg/mL, respectively. A structurally related cyclic octapeptide named cyclogossin B1, was subsequently isolated from latex of the plant

Figure 1: 1/2/3

- 1: GB1 (R1 = R2 = R3 = R5 = H, R4 = OH)
- 2: GB2 (R1 = R2 = R4 = R5 = H, R3 = OH)
- 3: KF (R3 = R5 = H, R1 = R4 = OH, R2 = OCH3)

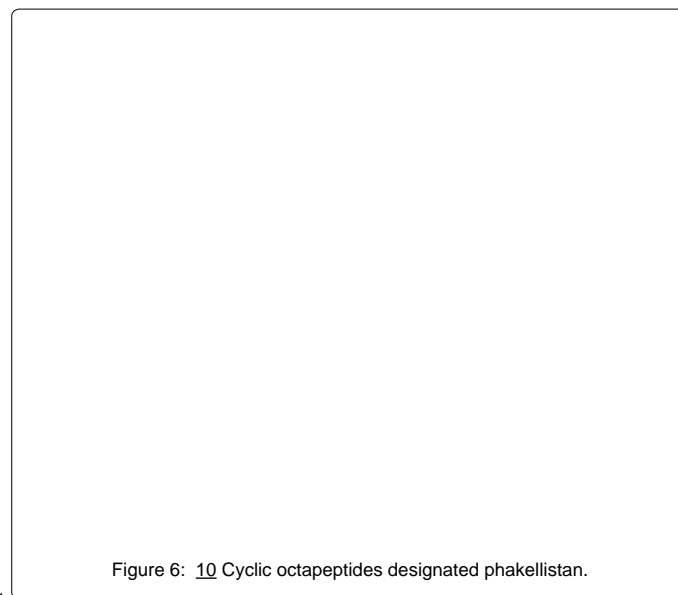


It had been reported that the methanol extract from the same part of the plant exhibited cytotoxic activity against a panel of human tumor cell lines [28].

Hymenocardia acida (Phyllanthaceae) is a shrub or small tree indigenous to the African savannah where the leaves and roots of the plant are used to treat malaria in traditional African medicine [29]. The roots of the plant are also said to be used as antimalarial in Nigeria [30].

) Terpenoids: Of the numerous reports of tumor cytotoxic terpenoids, one example selected from indigenous West African plants would serve to illustrate the potency of anticancer activity that may be found in this group.

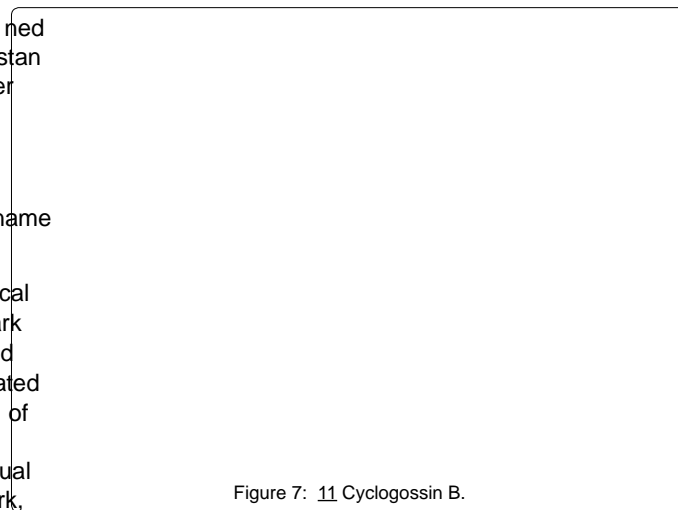
a) *Sesquiterpenoids*: The cytotoxic and antitumor sesquiterpenes have been found almost exclusively among the germacrane, guaiane, eudesmane and elemane sub-groups. Almost without exception, a common structural factor for these active compounds appears to be the possession of an  $\alpha$ -methylene- $\gamma$ -lactone ring. The latter chemical group, alone, or in addition to other reactive groups such as  $\alpha$ -enones,  $\beta$ -unsaturated lactones, ketones, and esters or epoxides, have been hypothesized to be subject to nucleophilic attack, resulting in the



Jatropha gossypifolia (Euphorbiaceae) [25]. Cyclogossin B is yet to be investigated for possible cytotoxic activity. Structure-activity relationships of cytotoxic cyclic octapeptides have not been determined but on the basis of the close structural similarity between phakellistan 10 (10) and cyclogossin B (1), it is reasonable to expect that the latter may also display some cytotoxic activity.

The fresh latex of Jatropha gossypifolia reportedly applied to infected wounds and ulcers and a decoction of the plant is used to treat fever in Togo and Senegal, where the plant bears the vernacular name of "lumulum" (Figures 6 and 7) [25].

Other cyclopeptide alkaloids have been isolated from local plants, including a number of such compounds from the root bark of Hymenocardia acida beginning with the first such isolate named hymenocardine [26]. Subsequently, hymenocardine was re-isolated along with its derivatives including one with a hydroxyl in place of a carbonyl group that was named hymenocardinol, a hymenocardine N-oxide, and a new cyclopeptide alkaloid containing an unusual histidine moiety named hymenocardine-H [27]. In a previous work,



alkylation of biologically significant macromolecules, and consequently leading to cancer cell death [31].

A typical example is the active compound vernodalol (12), from the elemanolide sub-group isolated from the dried leaves of *Vernonia amygdalifolia*. [32] and having significant in vitro cytotoxic activity against the Walker carcinoma 256 with an ED50 of 1.8  $\mu\text{g/ml}$ . It was re-isolated from *Vernonia amygdalifolia* along with two other elemanolide lactones, vernodalol and a new compound dihydrovernodalol (13), [33]. The possible anticancer activities of the latter two compounds have not yet been investigated (Figure 8).

A new anticancer sesquiterpene lactone has been isolated from *Vernonia amygdalifolia* and named:

“vernodalol” [34]. The compound which possesses the requisite  $\alpha$ -methylene-lactone ring postulated as the structural requirement for anticancer activity in the sesquiterpene lactones, displayed modest growth inhibitory activity (inhibition of DNA synthesis) in the human mammary adenocarcinoma cell line MCF-7, where values of 34% and 40% ( $P < 0.025$ ) growth inhibition were obtained at concentrations of 10 and 50  $\mu\text{g/ml}$ , respectively [34].

*Vernonia amygdalifolia* (bitter leaf) is a highly valued food item as the leaves are a popular vegetable for soups among the Igbo of southern Nigeria. Medicinal applications of *Vernonia amygdalifolia* include use of the twigs as “chewing stick”, or local toothbrush, which are chewed as a stomach tonic and appetizer. A decoction of the leaves is taken as an antipyretic and laxative, and for coughs [16].

v) Miscellaneous: this category covers phytochemicals possessing significant in vitro cytotoxicities against tumor cell lines, and which, for reasons of proper nomenclature, may not be included in the other classical groups. For reasons of space and brevity, only one example is discussed here.

**Gamma-linolenic acid:** Patients with malignant primary gliomas or mammary adenocarcinomas have a poor prognosis as common therapeutic approaches are relatively ineffective. Interestingly, an edible, essential polyunsaturated fatty acid (PUFA),  $\gamma$ -linolenic acid (GLA, 14) was found to be selectively cytotoxic to gliomas and other tumor types [35,36], while normal cells are largely unaffected. Although the mechanism for this selective cytotoxicity is poorly understood, free radical generation with lipid peroxidation from PUFA supplementation is generally believed to play a role [35,36]. However, the latter assumption is flawed on many counts, not least of which is the fact that higher members of the PUFAs, containing higher unsaturation and presumably better able to generate the assumed toxic free radicals, are ineffective (Figure 9).

The author proposed a plausible mechanism for GLA-induced cytotoxicity to tumor cells, particularly gliomas and mammary

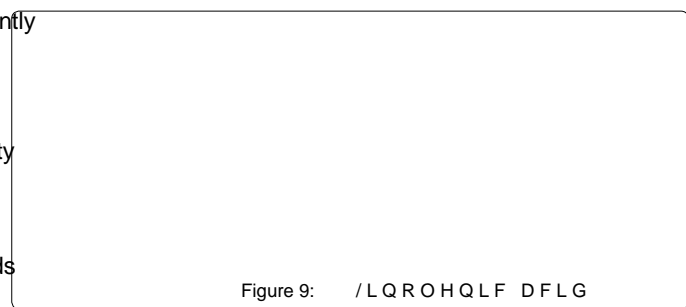


Figure 9: /LQROHQFLDFLGL

adenocarcinomas [37]. The article was published in 2001 and titled: “Repression of cellular anaplasia as the hypothesized mechanism for GLA-induced toxicity to tumor cells” [37]. About four years later, workers in another laboratory [38] independently provided laboratory experimental support for this hypothesis, thus strengthening the theory and conferring scientific legitimacy on the postulate. One hopes that this postulate would serve as a basis for the design of new therapeutic agents against the dreaded gliomas and metastatic mammary carcinomas.

GLA, the tumor cell cytotoxic agent, was subsequently shown to be present in large amounts, as its biochemical precursor, linoleic acid, in the commonly eaten (South-Eastern Nigeria) oil bean seed (‘ugba’ or ‘upkaka’; *Pentaclethra macrophylla*) in a related article co-published by the author [39].

In a related anticancer activity of a dietary linolenic acid obtainable from indigenous plants, dietary supplementation with 0.01% and 0.1% chlorophyllin from bitter melon (*Momordica charantia*) significantly reduced the incidence of azoxymethane (AOM)-induced colon aberrant crypt foci (ACF) in male 344 rats [40]. The authors suggested that the mechanism of CLN suppression of AOM-induced colon carcinogenesis might, in part, be through modification of lipid composition in the colon and liver and/or increased expression of PPAR $\gamma$  in the colon mucosa.

*Momordica charantia* (Cucurbitaceae) is a plant of common use in the traditional medicine of West Africa as an antimalarial [41], in the treatment of measles [42] in Nigeria, and for the treatment of gastroenteritis and viral disease in Togo [43].

## B. Reversal of multi-drug resistance (MDR)

An illustrative example will be cited here, from the alkaloid class of phytochemicals.

Two known benzylisoquinoline alkaloids: Figure 10 Two known benzylisoquinoline alkaloids, sotrolobine (15), and trilobine (16), were isolated from an extract of the *Stephanandra japonica*. Using the benzonitrile assay, both compounds were shown to be active in reversing doxorubicin resistance in the Adriamycin-resistant human breast adenocarcinoma cell line, MCF-7/ADR, with sotrolobine (15) being more active than trilobine (16) and equipotent with the reference compound verapamil [44]. Since the only structural difference between (15) and (16) is the methyl group at N-2', the greater efficacy of (15) suggests that a tertiary amine is preferred at this position to a secondary amine. The authors also suggested that the increased lipophilicity induced by presence of an extra methyl group may contribute to the enhanced activity, since MDR inhibitors are known to be lipophilic.

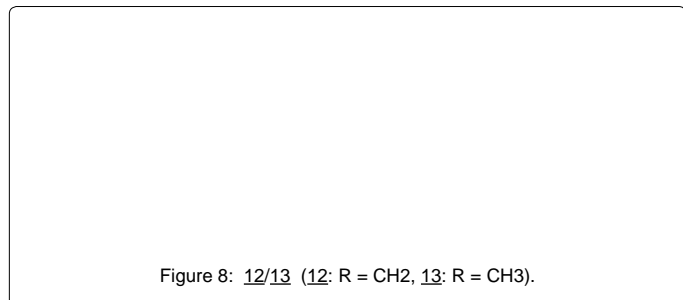
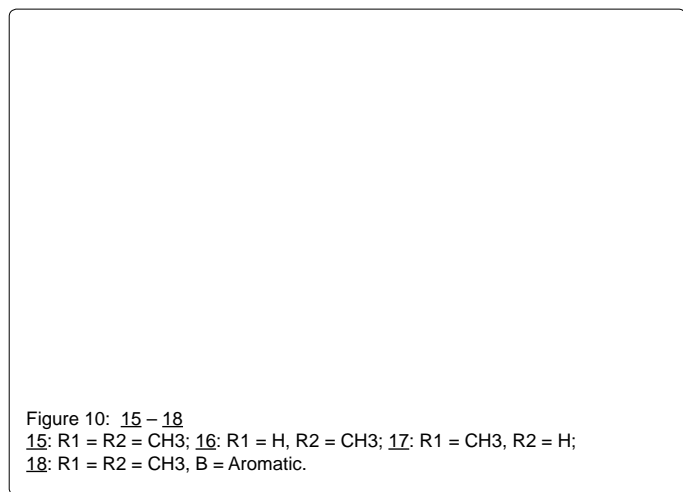


Figure 8: 12/13 (12: R = CH<sub>2</sub>, 13: R = CH<sub>3</sub>).



Other *isobenzylisoquinoline* alkaloids, *cocculine* (17) and *triglochin* (18), have been reported from extracts of the whole plant of *Trichostema* species (Menispermaceae), [45], and from leaves of *T. glaberrimum* (DeW.) Staner, were isolated four more such compounds named *obamegline*, *stebamine*, *glaberrine* and *soglaberrine-N-oxide*, the last two described as new *isobenzylisoquinoline* *N*-benzoyl-*N*-oxide alkaloids [46]. The remarkable structural similarity between these *Trichostema* isolates and the MDR-actives compounds *sotrolobine* (15) and *trilobine* (16) raises the hope that some may be found active, if tested. Of particular interest in this regard is *cocculine* (17), which is almost identical with the highly active *sotrolobine* (15) (and deriving only in the possession of a hydroxyl, rather than a methoxy substituent, as in position 12). Similarly, *triglochin* (18) may display a higher activity than *trilobine* (16) since the only structural difference between the two is that the "B" ring in the former is aromatic which should stand in good stead. For instance, possession of a terminal aromatic group, as well as an internal  $\pi$ -environment are two of the structural requirements for reversal of MDR-activity in these compounds [44].

A *isobenzylisoquinoline* alkaloid named 'cycleanine' has been isolated from ethanol extract of

*Trichostema subcordata* and shown to be potent (IC<sub>50</sub> < 2.4 μM) in inhibiting tumor cell growth in the human cancer, Ovarian-8 and A2780 cell lines, using the Sulforhodamine B assay [47]. The IC<sub>50</sub> of cycleanine on human normal ovarian surface epithelial cells was 3.0 μM, thus suggesting modest selectivity toward cancer cells. Mechanism of action for cycleanine was further indicated to be through apoptosis as shown by activation of caspases 3/7 and cleavage of poly(ADP-ribose) polymerase [47].

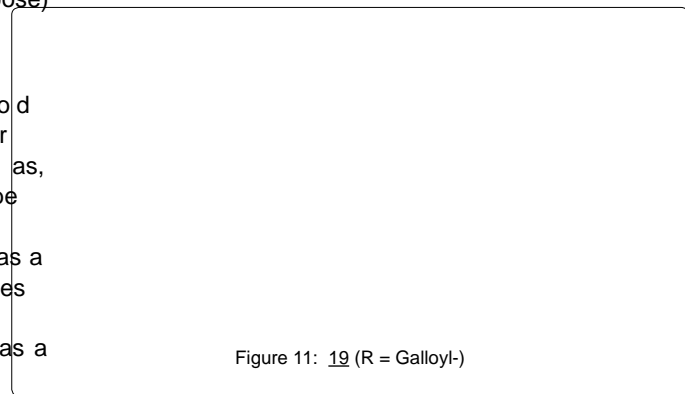
From leaves of another indigenous West African plant, *Pithecellobium* *funferia* was isolated a new *isobenzylisoquinoline* alkaloid named, 'lafunmine' [48]. The authors also described two other *isobenzylisoquinoline* alkaloids isolated from the same source as, *sotetrandrine* and *thalugosine*. These compounds are yet to be investigated for possible multidrug resistance activity. *Trichostema subcordata* (Menispermaceae) was described by the authors as a medicinal plant traditionally used for the treatment of various diseases in West Africa [47], while *Pithecellobium funferia* (Menispermaceae) is a woody climber of Ghana and West Africa that has been used as a medicinal in the treatment of numerous ailments [48].

### C. Inhibition of metastasis

A significant example in this category would be the report of work involving the phytochemical, (+)-catechin (19). This compound has been, almost exclusively, the only advanced investigated for its effects on cancer. Prominent among the anticancer effects reported are its antioxidant effect in inhibiting the auto-oxidation of lipoic acid, and methyl lipoic acids [49], and the *in vitro* inhibition of metastasis as described below. In the latter study, the anti-metastatic potential of some phytochemicals, including (+)-catechin, against the highly invasive, Dunn rat prostate adenocarcinoma cells (R3327-5A), was investigated [50]. Among the invasive parameters studied, were matrigel membrane invasion by the trans-well assay; gel zymography assay and Northern analysis, for the matrix metalloproteinases (MMP-2 and MMP-9)'s activity and gene transcription respectively; as well as Western immunoblot assay, for gene expression analysis of the membrane-associated proteins,  $\beta$ -catenin and  $\gamma$ -catenin. Results indicated that catechin at 25 μM, significantly decreased matrigel membrane invasion of the prostate tumor cells by 24% and also inhibited  $\beta$ -catenin protein levels by 58% (P < 0.01), (Figure 11) [50].

A significant Nigerian food source from which (+)-catechin (19), and its derivatives have been isolated is described. It was obtained from five edible species of *Dioscorea* (yams), indigenous to Nigeria [51]. The five yam species, and the cultivars or Nigerian (Igbo) names were given as *Dioscorea alata*, two cultivars (Omnelu and UM 680); *D. bulbifera* L. (Adu); *D. cayana* Lam. (Oku); *D. dumetorum* Pax. (Ona); and *D. rotundata* Poir., five cultivars (Ab, Ekpe, Nwopoke, Obaturugo and Okwocha), [51]. Yams are a starchy staple and highly prized foodstuff in Nigeria and neighboring countries. Often, the wealth of a farmer in the Igbo-speaking areas of Nigeria is determined by the quantity of yams stocked.

In an analysis for secondary plant metabolites contents of kola nuts (*Cola nitida*) popularly consumed in Nigeria and other West African countries, (+)-catechin was shown to be present in the highest amount among the phytochemicals found [52]. The varieties of kola nuts (*Cola nitida alba*, *Cola nitida rubra* A. Chev. and *Cola acuminata* Schott & Endl) were analyzed and the phytochemical contents given in decreasing order of quantity as: (+)-catechin (27-37 g/kg), caffeine (18-24 g/kg), (-)-epicatechin (20-21 g/kg), procyanidin B1 [epicatechin-(4β-->8)-catechin] (15-19 g/kg), and procyanidin B2 [epicatechin-(4β-->8)-epicatechin] (7-10 g/kg) [52,53]. Kola nuts would therefore constitute a significant source of the anticancer compound (+)-catechin, especially for people in the northern parts of Nigeria where kola nut consumption is a very popular pastime.



## Conclusion

The objective of this review has been to primarily highlight the existence of promising anticancer potential inherent in phytochemicals from indigenous West African plants of common usage, as local food or in the traditional medicine of the area. Ethnopharmacologists and other scientists in related fields on the African continent are hereby exhorted to spear-head more of such studies as a means of exploiting the immense benefits accruable from a revival of interest in this relatively neglected pool of valuable resource.

It is however important to stress that efforts should be made to isolate and characterize the specific active principles responsible for anticancer activity. The common practice of ascribing activities to crude extracts while understandably attributable to a paucity of research facilities, would hamper a desirable appreciation of mechanisms and deter necessary contribution to the international knowledge base on this subject. A discussion on which the chemical structures of active agents are presented and evaluated, illustrates the advantages to be gained in the prescribed approach.

Finally, the author once more wishes to draw attention to the imminent threat posed by rapid deforestation of tropical forests, and the looming danger that rare curative plants would be lost forever, if immediate preventive actions are not taken.

## References

- Sawadogo WR, Schumacher M, Teiten MH, Dicato M, Diederich M (2012) Traditional West African pharmacopeia, plants and derived compounds for cancer therapy. *Biochem Pharmacol* 84: 1225-1240.
- Lifongo LL, Simoben CV, Ntie-Kang F, Babiaka SB, Judson PN (2014) A bioactivity versus ethnobotanical survey of medicinal plants from Nigeria, west Africa. *Nat Prod Bioprospect* 4: 1-19.
- Tsouh Fokou PV, Nyarko AK, Appiah-Opong R, Tchokouaha Yamthe LR (2015) Ethnopharmacological reports on anti-Buruli ulcer medicinal plants in three West African countries. *J Ethnopharmacol* 172:297-311.
- Avoseh O, Oyediji O, Rungqu P, Nkeh-Chungag B, Oyediji A (2015) Cymbopogon species; ethnopharmacology, phytochemistry and the pharmacological importance. *Molecules* 20: 7438-7453.
- Boucherle B, Haudecoeur R, Queiroz EF, De Waard M, Wolfender JL, et al. (2016) *Nauclea latifolia*: biological activity and alkaloid phytochemistry of a West African tree. *Nat Prod Res* 33: 1034-1043.
- Costa R, Albergamo A, Pellizzeri V, Dugo G (2017) Phytochemical screening by LC-MS and LC-PDA of ethanolic extracts from the fruits of *Kigelia africana* (Lam) Benth. *Nat Prod Res* 31: 1397-1402.
- Snedden AT (1984) Novel antitumor agents from plants. In: Ottenbrite RM, Butler GB (editors) *Anticancer and Interferon Agents: Synthesis and Properties*. Marcel Decker Inc.: New York and Basel.
- Komoda Y, Kishi T (1980) *Maytansinoids*. In: Cassady JM, Douros JD (editors) *Anticancer Agents based on Natural Products Models*. Academic Press.
- Iwu MM, Igboko OA, Onwuchekwa UA, Okunji CO (1987) Evaluation of W KH D Q W L K H S D W R W R [ L F D F W L Y L W \ R I W K H E L A D C A R I A R L K O I F I U Z I N S H U P ( 1 9 8 8 ) D i s c r i m i n a t i n g H e r c o n o l i d e l a c t o n e s f r o m V e r n o n i a a m y g d a l i n a P h y t o c h e m . 2 2 : 2 5 2 5 - 2 5 2 6 .
- Farombi EO (2000) Mechanisms for the hepatoprotective action of kolaviron: studies on hepatic enzymes, microsomal lipids and lipid peroxidation in carbon tetrachloride-treated rats. *Pharmacol Res* 42: 75-80.
- 1 Z D Q N Z R - 2 7 D K Q W H Q J - \* ( P H U R O H \* 2 J H Q R W R [ L F L W \ L Q X P D Q O L Y H U G H U L Y H G + H S \* molecular mechanisms of action. *Eur J Cancer Prev* 9:351-361.
- Iwu MM (1985) Antihepatotoxic constituents of *Garcinia kola* seeds. *Experientia* 41:699-700.
- Iwu MM, Igboko OA, Elekwa OK, Tempesta MS (1990) Prevention of W K L R D F H W D P L G H L Q G X F H G K H S D W R W R [ L F L W \ E \ B e a d i n g o f p o s t m e m o r i a l i n e i c \* a n d i n d u c e d D o x i n e C D T u m o r c e l l s . M e d H y p o t h e s e s 5 6 : 5 8 2 - 5 8 8 .
14. % U D L G H 9 % \$ Q W L K H S D W R W R [ L F E L R F K H P L F D O of *Garcinia kola* seeds. *Phytother Res* 5:35-37.
15. : D W H U P D Q 3 \* & U L F K W R Q ( \* ; D Q W K R Q H V D Q G d e n s i v e n i a s t e m b a r k . P h y t o c h e m 1 9 : 2 7 2 3 - 2 7 2 6 .
16. Nwankwo JO (2011) In: Potential anticancer and antiviral agents from West African phytochemicals. *Flavonoids* pp: 56-59
17. Nwankwo JO, Fitzgerald MP, Domann FE (Unpublished results).
18. Itokawa H, Oshida Y, Ikuta A, Inatomi H, Ikegami S (1981) Flavonol glycosides I U R P W K H A R Z H U V R I & X F X U E L W D S H S R 3 K \ W R F K H P
19. Wall ME, Wani MC (1987) Plant antitumor agents, 27. Isolation, structure, and structure activity relationships of alkaloids from *Fagara macrophylla*. *J Nat Prod* 60: 1095-1099.
20. Cordell GA (1977) Recent experimental and clinical data concerning antitumor and cytotoxic agents from plants. In: "New Natural Products and Plant Drugs with Pharmacological, Biological or Therapeutical Activity" Proc 1st Intl Congress Med Plant.
21. Res Section A. Wagner H, Wolff P (editors) Springer-Verlag, Berlin, Heidelberg, New York.
22. Nakanishi T, Suzuki M, Saimoto A, Kabasawa T (1999) Structural Considerations of NK109, an Antitumor Benzof[*c*]phenanthridine Alkaloid. *J Nat Prod* 62:864-867.
23. Dwuma-Badu D, Ayim JSK, Withers SF, Agyemang NO, Ateya AM, et al. (1980) Constituents of West African Medicinal Plants. XXVII. Alkaloids of *Rhigiocarya racemifera* and *Stephania dinklagei*. *J Nat Prod* 43: 123-129.
24. Suffness M, Douros J (1980) In: Cassady JM, Douros JD (editors). *Anticancer agents based on natural products models*. Acad Press: NY; p:14.
25. Pettit GR, Tan R, Ichihara Y, Williams MD, Doubek DI, et al. (1995) Antineoplastic Agents, 325. Isolation and Structure of the Human Cancer Cell Growth Inhibitory Cyclic Octapeptides Phakellistatin 10 and 11 from *Phakellia* sp. *J Nat Prod* 58:961-965.
26. Auvin-Guette C, Baraguey C, Blond A, Pousset JL, Bodo B (1997) Cyclogossin B, a cyclic octapeptide from *Jatropha gossypifolia*. *J Nat Prod* 60: 1155-1157.
27. Pais M, Marchand J, Ratle G, Jarreau FX (1968) Peptidic alkaloides. VI. Hymenocardine, alkaloid from *Hymenocardia acida* Tul. *Bulletin de la Société Chimique de France* 7:2979-2984.
28. Tuenter E, Exarchou V, Baldé A, Cos P, Maes L, et al. (2016) Cyclopeptide alkaloids from *Hymenocardia acida*. *J Nat Prod* 79: 1746-1751.
29. Muanza DN, Euler KL, Williams L, Newman DJ (1995) Screening for antitumor and anti-HIV activities of nine medicinal plants from Zaire. *Int J Pharmacognosy* 33:98-106.
30. Tuenter E, Bijttebier S, Foubert K, Breynaert A, Apers S, et al. (2017) In vitro and in vivo study of the gastrointestinal absorption and metabolism of Hymenocardine, a cyclopeptide alkaloid. *Planta Med* 83: 790-796.
31. Ainslie J (1937) A list of plants used in native medicine in Nigeria. Imperial Forestry Institute, University of Oxford p: 107
32. Cassady JM, Suffness M (1980) In: Cassady JM, Douros JD (editors) *Antitumor Agents based on Natural Product Models*. Academic Press:NY; pp: 201-269.
33. Kupchan SM, Hemingway RJ, Karim A, Werner D (1969) Tumor inhibitors. XLVII. Vernodaline and vernomygdin, two new cytotoxic sesquiterpene lactones from *Vernonia amygdalina* del. *J Org Chem* 34: 3908-3911.
34. Sanyal BK, Chatterjee I, Kundu SK (1988) Isolation and characterization of a novel sesquiterpene lactone from *Vernonia amygdalina* *Phytochem* 22:2525-2526.
35. Luo X, Jiang Y, Fronczek FR, Lin C, Izevbogie EB, et al. (2011) Isolation and structure determination of a sesquiterpene lactone (vernodaline) from *Vernonia amygdalina* extracts. *Pharm Biol* 49: 464-470.
36. Hsueh WL, BRQ (1989) In: A. D. W. H. L. S. (editor) *Carcinogenesis and dietary fat*. Food and Academic Publishers, Boston, MA 247-262.
37. Jiang WG, Bryce RP, Horrobin DF (1998) Essential fatty acids: molecular and cellular basis of their anti-cancer action and clinical implications. *Crit Rev Oncol Hematol* 27:179-209.
38. Nwankwo JO (2001) Repression of cellular anaplerosis as the hypothesized mechanism of gamma-linolenic acid-induced Doxorubicin CD tumor cells. *Med Hypotheses* 56: 582-588.

39. Menendez JA, Colomer R, Lupu R (2005) Inhibition of fatty acid synthase-G H S H Q G H Q W Q H R S O D V W L F O L S R J H Q H V L V D V W K H P H I R A D O G R U V P I C I S I A D I C Y P A O J H O P F D 4 1 6 0 1 6 5 Q G X F H G toxicity to tumor cells: an extension to Nwankwo's hypothesis. Med Hypotheses 64: 337-341.
40. Onwuliri VA, Attah I, Nwankwo JO (2004) Anti-nutritional factors, essential and non-essential fatty acids composition of Ugba (*Pentaclethra macrophylla*) seeds at different stages of processing and fermentation J Biol Sci. 4: 671-675.
41. Kohno H, Yasui Y, Suzuki R, Hosokawa M, Miyashita K, et al. (2004) Dietary seed oil rich in conjugated linolenic acid from bitter melon inhibits azoxymethane-induced rat colon carcinogenesis through elevation of colonic PPARgamma expression and alteration of lipid composition. Int J Cancer 110: 896-901.
42. Olasehinde GI, Ojurongbe O, Adeyeba AO, Fagade OE, Valecha N, et al. (2014) In vitro studies on the sensitivity pattern of *Plasmodium falciparum* to anti-malarial drugs and local herbal extracts. Malar J 13: 63.
43. Sonibare MA, Moody JO, Adesanya EO (2009) Use of medicinal plants for the treatment of measles in Nigeria. J Ethnopharmacol 122: 268-72.
44. Beloin N, Gbeassor M, Akpagana K, Hudson J, de Souza K, et al. (2005) Ethnomedicinal uses of *Momordica charantia* (Cucurbitaceae) in Togo and relation to its phytochemistry and biological activity. J Ethnopharmacol 96: 49-55.
45. Hall AM, Chang CJ (1997) Multidrug-Resistance Modulators from *Stephania japonica*. J Nat Prod 60: 1193-1195.
46. Spiff AI, Zabel V, Watson WH, Zemaitis MA, Ateya AM, et al. (1981) Constituents of West African Medicinal Plants. XXX. Tridictyophylline, A New Morphinan Alkaloid from *Triclisia dicarpa*. J Nat Prod 44: 160-165.
47. Owusu PD, Slatkin DJ, Knapp JE, Schiff PL (1981) Constituents of West African Medicinal Plants. XXVIII. Additional Alkaloids of *Triclisia gilletti*. J Nat Prod 44: 61-66.
48. Uche FI, Drijfhout FP, McCullagh J, Richardson A, Li WW (2016) Cytotoxicity effects and apoptosis induction by bisbenzylisoquinoline alkaloids from *Triclisia subcordata*. Phytother Res 30: 1533-1539.
49. Ayim JS, Dwuma-Badu D, Fiagbe NY, Ateya AM, Slatkin DJ, et al. (1977) Constituents of West African medicinal plants. XXI. Tiliapunimine, a new imino bisbenzylisoquinoline alkaloid from *Tiliacora funifera*. Lloydia 40: 561-565.
50. 7 R U H O - & L O O D U G - & L O O D U G 3 \$ Q W L R [ L G D Q V with peroxy radical. Phytochem 25: 383-385.
51. Nwankwo JO (2002) Anti-metastatic activities of all-trans retinoic acid, indole-3-carbinol and (+)-catechin in Dunning rat invasive prostate adenocarcinoma cells. Anticancer Res 22: 4129-4135.
52. 2 J R 2 1 & D \ J L O O - & R X U V H \ ' \* 3 K H Q R O L F V F species. Phytochem 23: 329-331.
53. Atawodi SE, Pfundstein B, Haubner R, Spiegelhalder B, Bartsch H, et al. (2007) Content of polyphenolic compounds in the Nigerian stimulants *Cola nitida* ssp. *alba*, *Cola nitida* ssp. *rubra* A. Chev, and *Cola acuminata* Schott & Endl and their antioxidant capacity. J Agric Food Chem 55: 9824-9828.