

## Biochemical Pathways of CO<sub>2</sub> Fixation and the Evolution of Autotrophy: A Comparative Study

Sheng Jeyaraj\*

Department of biochemistry, Deakin University, Australia

### Abstract

The evolution of autotrophy, characterized by the ability of organisms to fix carbon dioxide (CO<sub>2</sub>) into organic compounds, is a cornerstone of life on Earth. This study provides a comparative analysis of the biochemical pathways involved in CO<sub>2</sub> fixation across various autotrophic organisms. By examining the Calvin-Benson-Bassham (CBB) cycle, the reverse Krebs cycle, the reductive acetyl-CoA pathway, and the 3-hydroxypropionate cycle, we aim to elucidate the evolutionary trajectories and adaptations that have led to the diversification of autotrophic mechanisms. Understanding these pathways not only sheds light on the evolution of life but also has implications for bioengineering and sustainability.

### Introduction

Autotrophy is a fundamental metabolic strategy that enables organisms to convert inorganic carbon (CO<sub>2</sub>) into organic matter, forming the basis of the food web and driving the global carbon cycle. Various biochemical pathways have evolved to facilitate CO<sub>2</sub> fixation, each with distinct enzymatic mechanisms and energy requirements. This comparative study investigates the major CO<sub>2</sub> fixation pathways, their distribution among different life forms, and their evolutionary significance [1].

### Major CO<sub>2</sub> fixation pathways

The ability to fix carbon dioxide (CO<sub>2</sub>) into organic compounds is a fundamental process for autotrophic organisms, forming the basis for the biosphere's primary productivity. Over evolutionary history, several distinct biochemical pathways have evolved to achieve CO<sub>2</sub> fixation, each suited to the specific environmental and metabolic requirements of the organisms that utilize them. The major CO<sub>2</sub> fixation pathways include the Calvin-Benson-Bassham (CBB) cycle, the reverse Krebs cycle, the reductive acetyl-CoA pathway, and the 3-hydroxypropionate cycle. Here, we explore the mechanisms, key enzymes, and ecological significance of each pathway [2].

### The calvin-benson-bassham (CBB) Cycle

The Calvin-Benson-Bassham (CBB) cycle, commonly known as the Calvin cycle, is the most prevalent CO<sub>2</sub> fixation pathway and is primarily associated with oxygenic photosynthetic organisms such as cyanobacteria, algae, and terrestrial plants.

### Mechanism

The Calvin cycle operates in the chloroplasts of photosynthetic cells and consists of three main phases: carbon fixation, reduction, and regeneration of the CO<sub>2</sub> acceptor. The cycle begins with the enzyme ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO) catalyzing the carboxylation of ribulose-1,5-bisphosphate (RuBP) to form two molecules of 3-phosphoglycerate (3-PGA) [3].

### Key enzymes

- RuBisCO: Catalyzes the first step of the cycle and is the most abundant enzyme on Earth.
- Phosphoglycerate kinase and glyceraldehyde-3-phosphate dehydrogenase: Facilitate the reduction of 3-PGA to glyceraldehyde-

3-phosphate (G3P).

- Ribulose-5-phosphate kinase: Regenerates RuBP from ribulose-5-phosphate [4].

### Ecological significance

The Calvin cycle is vital for the global carbon cycle, contributing to the majority of organic carbon production on Earth. It is highly efficient under moderate light and temperature conditions, making it the cornerstone of plant and algal productivity.

### The reverse krebs cycle

The reverse Krebs cycle, also known as the reductive citric acid cycle, is employed by some anaerobic and microaerophilic bacteria and archaea [5]. This pathway is essentially the citric acid cycle operating in reverse, using CO<sub>2</sub> to synthesize organic compounds.

### Mechanism

In the reverse Krebs cycle, CO<sub>2</sub> is fixed by various enzymes to form acetyl-CoA and other intermediates that are further processed to synthesize essential biomolecules.

### Key enzymes

- ATP citrate lyase: Converts citrate to oxaloacetate and acetyl-CoA.
- 2-oxoglutarate:ferredoxin oxidoreductase: Reduces 2-oxoglutarate to succinyl-CoA.
- Fumarate reductase: Catalyzes the reduction of fumarate to succinate [6].

**\*Corresponding author:** Sheng Jeyaraj, Department of biochemistry, Deakin University, Australia, Email: Sheng@edu.in

**Received:** 01-Jan-2024, Manuscript No. bcp-24-136714; **Editor assigned:** 03-Jan-2024, PreQC No. bcp-24-136714 (PQ); **Reviewed:** 17-Jan-2024, QC No. bcp-24-136714; **Revised:** 23-Jan-2024, Manuscript No. bcp-24-136714 (R); **Published:** 31-Jan-2024, DOI: 10.4172/2168-9652.1000450

**Citation:** Jeyaraj S (2024) Biochemical Pathways of CO<sub>2</sub> Fixation and the Evolution of Autotrophy: A Comparative Study. Biochem Physiol 13: 450.

**Copyright:** © 2024 Jeyaraj S. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

## Ecological significance

This pathway is considered ancient and is believed to have been used by some of the earliest autotrophic organisms. It allows for CO<sub>2</sub> fixation in environments where oxygen is limited or absent, demonstrating the metabolic flexibility of early life forms.

## The reductive acetyl-coa pathway

The reductive acetyl-CoA pathway, also known as the Wood-Ljungdahl pathway, is utilized by a variety of anaerobic bacteria and archaea, including methanogens and acetogens.

## Mechanism

This pathway involves the reduction of CO<sub>2</sub> to formate and subsequently to acetyl-CoA. The process is coupled with energy conservation mechanisms, making it highly efficient under anaerobic conditions.

## Key enzymes

- Carbon monoxide dehydrogenase/acetyl-CoA synthase (CODH/ACS): Catalyzes the final steps of CO<sub>2</sub> reduction to acetyl-CoA.
- Formate dehydrogenase: Reduces CO<sub>2</sub> to formate.
- Methylenetetrahydrofolate reductase: Involved in the transfer of methyl groups during the pathway.

## Ecological Significance

The reductive acetyl-CoA pathway is one of the most energy-efficient CO<sub>2</sub> fixation mechanisms, allowing organisms to thrive in anaerobic environments [7]. It also plays a crucial role in global carbon cycling, particularly in the production of methane and acetate.

## The 3-hydroxypropionate cycle

The 3-hydroxypropionate cycle is a less common CO<sub>2</sub> fixation pathway found in certain extremophiles, including some green non-sulfur bacteria and archaea.

## Mechanism

This pathway fixes CO<sub>2</sub> into 3-hydroxypropionate through a series of carboxylation and reduction reactions, ultimately producing glyoxylate and pyruvate, which are key intermediates for biosynthesis.

## Key enzymes

- Acetyl-CoA carboxylase: Catalyzes the carboxylation of acetyl-CoA to malonyl-CoA.
- Propionyl-CoA carboxylase: Converts propionyl-CoA to methylmalonyl-CoA.
- Methylmalonyl-CoA epimerase: Facilitates the conversion between different isomers of methylmalonyl-CoA.

## Ecological significance

The 3-hydroxypropionate cycle enables organisms to fix CO<sub>2</sub> in

environments with extreme conditions, such as high temperatures or salinity. This adaptability highlights the evolutionary innovation of autotrophic pathways to diverse ecological niches [8].

## Evolutionary implications

The diversity of CO<sub>2</sub> fixation pathways reflects the adaptability of life to various environmental conditions. The evolution of these pathways likely involved horizontal gene transfer, gene duplication, and the recruitment of pre-existing metabolic enzymes [9,10]. By comparing these pathways, we can trace the evolutionary history of autotrophy and understand how different organisms have optimized CO<sub>2</sub> fixation for their ecological niches.

## Conclusion

The comparative study of biochemical pathways for CO<sub>2</sub> fixation highlights the evolutionary ingenuity of autotrophic organisms. Each pathway presents unique adaptations that have enabled life to thrive in diverse environments, from oxygen-rich atmospheres to extreme anaerobic conditions. The diversity of CO<sub>2</sub> fixation pathways illustrates the evolutionary versatility and adaptability of autotrophic organisms. Each pathway has evolved to meet specific environmental challenges and metabolic needs, ranging from oxygen-rich to anaerobic and extreme environments. Understanding these pathways not only provides insights into the evolution of life on Earth but also offers potential applications in biotechnology, such as enhancing carbon capture and developing sustainable biofuel production strategies. Understanding these pathways not only provides insights into the early evolution of life but also offers potential applications in biotechnology and sustainable practices, such as the development of carbon capture strategies and biofuel production.

## References

1. Le Quéré C, Raupach MR, Canadell JG, Marland G, Bopp L, et al. (2009) Trends in the sources and sinks of carbon dioxide. *Nat Geosci* 2: 831–836.
2. Pan Y, Birdsey RA, Fang J, Houghton R, Kauppi PE, et al. (2011) A large and persistent carbon sink in the world's forests. *Science* 333: 988–993.
3. Lal R (2004) Soil carbon sequestration impacts on global climate change and food security. *Science* 304: 1623–1627.
4. Tilman D (1998) The greening of the green revolution. *Nature* 396: 211–212.
5. Fargione JE, Hill JD, Tilman D, Polasky S, Hawthorne P (2008) Land clearing and the biofuel carbon debt. *Science* 319: 1235–1238.
6. Searchinger T, Heimlich R, Houghton RA, Dong F, Elobeid A, et al. (2008) Use of U.S. croplands for biofuels increases greenhouse gases through emissions from land-use change. *Science* 319: 1238–1240.
7. Melillo JM, Reilly JM, Kicklighter DW, Gurgel AC, Cronin TW (2009) Indirect emissions from biofuels: How important. *Science* 326: 1397–1399.
8. Fargione JE, Plevin RJ, Hill JD (2010) The ecological impact of biofuels. *Ann Rev Ecol Evol Syst* 41: 351–377.
9. Donner SD, Kucharik CJ (2008) Corn-based ethanol production compromises goal of reducing nitrogen export by the Mississippi River. *Proc Natl Acad Sci USA* 105: 4513–4518.
10. Hill J, Polasky S, Nelson E, Tilman D, Huo H (2009) Climate change and health costs of air emissions from biofuels and gasoline. *Proc Nat Acad Sci USA* 106: 2077–2082.