

Dengue Viral Replication and Cytokine Secretion are Suppressed by Cepharanthine

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Abstract

Dengue fever, a mosquito-borne viral complaint, is a major public health problem worldwide. It's aboriginal in numerous tropical and tropical corridor of the world, especially Southeast Asia and the Western Pacific. Annually, roughly 390 million people are at threat for dengue contagion (DENV) infection, including, dengue hemorrhagic fever (DHF) cases and 1000 deaths, substantially children. There remains no specific treatment for DENV-infected cases; treatments only give relief of symptoms by close monitoring of vital signs in the critical period between days two and seven of fever, and may include both probative treatment and broad-diapason antiviral agents. Therefore, the development of anti-DENV agents for the treatment of DENV-infected cases are a high precedence.

Keywords: Dengue fever; Antiviral agents; Cepharanthine

Cytokine secretion are suppressed by Cepharanthine

Cepharanthine (CEP) is a natural isoquinoline alkaloid emulsion uprooted from *Stephanie Cepharanthine* Hayata and has been used in Japan since the 1950s for treating colorful acute and habitual conditions similar as leukopenia, snake mouthfuls, xerostomia and alopecia. Cell societies and Cepharanthine (CEP) mortal myelogenous leukemia K562 cells (RIKEN cell bank, Tsukuba, Japan) were dressed in Roswell Park Memorial Institute medium (RPMI- 1640; Fujifilm Wako, Osaka, Japan) supplemented with 10 FBS, penicillin (100 IU/ml) and streptomycin (100 U/ml) at 37°C in a humidified atmosphere containing 5 CO₂ and 95 air. mortal lung melanoma (A549) cells and monkey order epithelial (Vero) cells were attained from JRCB cell bank (Osaka, Japan), dressed in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10 FBS, penicillin (100 IU/ml) and streptomycin (100 U/ml) at 37°C in a humidified atmosphere containing 5 CO₂ and 95 air [1]. Enzyme-linked immunosorbent assay (ELISA) A549 cells were planted at 2.5 × 10⁵ cells per well in 12-well plates and incubated overnight [2]. Cells were co-treated with CEP at 6.67, 10, 15, and 22.5 µg/ml and infected with DENV- 2 for 4 h. At 24 h post-infections, the culture supernatant was collected to determine IL-6 stashing using a marketable ELISA tackle in agreement with the manufacturer's instructions (R&D Systems, Minneapolis, MN) [3]. Dengue contagion (DENV) infection is a mosquito-borne complaint affecting tropical and tropical areas. A correlation between high viral cargo and inflexibility has been observed in numerous studies. The characteristics of inflexibility in DENV infection (DHF/DSS) are increased pro-inflammatory cytokines, immunosuppressive cytokines, and chemokines [4]. The condition of massive cytokine product is known as a "cytokine storm". NF-κB plays a central part in the signaling pathways, enabling the product of pro-inflammatory cytokines. Inhibition of DENV infection and cytokine stashing would reduce the inflexibility of the complaint, so the development of antiviral agents for reducing DENV product and cytokine stashing in all four DENV serotypes is urgently demanded [5]. In the present study, we determined the antiviral and anti-inflammatory goods of CEP in DENV-infected K562 and A549 cells. K562 cells warrant the function of interferon response and are susceptible to DENV infection. CEP inhibited viral E protein and viral product for all four DENV serotypes in infected K562 cells [6]. CEP demonstrated a advanced inhibitory effect on DENV- 2 infection than on DENV- 4, -3 and -1. The different degrees of inhibition may be due to variations in viral

replication rate among the DENV serotypes. CEP inhibited original way of DENV replication, which are DENV infection and internalization step [7], performing in imperfect downstream replication way similar as viral RNA replication, viral protein conflation, and viral product. A drop of DENV RNA is due to a dwindling viral infection at the original state and consequent lower viral product [8].

Discussion

CEP is substantially an inactivating or virucidal agent for DENV. The inhibitory effect of CEP treatment was also observed in DENV-infected A549 cells [9-10]. A selectivity indicator (SI) value is a parameter which measures the correlation between effective attention of medicines and cytotoxicity, calculated as 50 cytotoxic attention/50 effective attention (CC₅₀/EC₅₀). The high SI value is considered as high efficacy with low toxin composites with SI ≥ 10 are generally defined to be active in vitro. still, some composites linked to be effective in beast model with SI < 10. Samples with SI < 10.

References

1. Dell'Agnola C, Biragyn A (2007) Clinical utilization of chemokines to combat cancer: the double-edged sword. *Expert Rev Vaccines* 6: 267-283.
2. Rottman JB (1999) Key role of chemokines and chemokine receptors in inflammation, immunity, neoplasia, and infectious disease. *Vet Pathol* 36: 357-367.
3. Speyer CL, Ward PA (2011) Role of endothelial chemokines and their receptors during inflammation. *J Invest Surg* 24: 18-27.
4. Koizumi K, Hojo S, Akashi T, Yasumoto K, Saiki I, et al. (2007) Chemokine receptors in cancer metastasis and cancer cell-derived chemokines in host immune response. *Cancer Sci* 98: 1652-1658.
5. Chow MT, Luster AD (2014) Chemokines in cancer. *Cancer Immunol Res* 2: 1125-1131.

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6. Bernardini G, Ribatti D, Spinetti G, Morbidelli L, Ziche M, et al. (2003) Analysis of the role of chemokines in angiogenesis. J Immunol Methods 273: 83-101.
7. Tanaka T, Bai Z, Srinoulprasert Y, Yang BG, Hayasaka H, et al. (2005) Chemokines in tumor progression and metastasis. Cancer Sci 96: 317-322.
8. Szekanecz Z, Koch AE (2001) Chemokines and angiogenesis. Curr Opin Rheumatol 13: 202-208.
9. Strieter RM, Burdick MD, Mestas J, Gomperts B, Keane MP, et al. (2006) Cancer CXC chemokine networks and tumour angiogenesis. Eur J Cancer 42: 768-778.
10. Slettenaar VI, Wilson JL (2006) The chemokine network: a target in cancer biology? Adv Drug Deliv Rev 58: 962-974.