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Heading to the Origins – Rice Microbiome as Functional Extension of the Host

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Commentary Article

Plant has evolved along microbial symbionts, including bacteria, archaea, fungi, and protista. These symbionts, i.e. microorganisms living in close association with the host, with their collection of intricate genetic components have improved plant fitness by adding essential and functional capabilities. Therefore, one might continuously see plant as a superorganism composed of an amalgam of both selected symbiont microbiota and host cells. As single unit, both partners interact closely and often operate by complementarity of their metabolic capabilities. The nature of these interactions range from mutualism, where cooperation between symbionts evolved by enhancing all participants' inclusive fitness, to pathogenesis, where the symbiont increases its own growth and fitness selfishly, and trigger the host response leading to disease development and ultimately to terminate the interaction. Thus, plants have evolved to detect molecular patterns of symbionts and are capable to discriminate them by either reinforcement of beneficial cooperation [1] or sanctions of detrimental interactions [2].

Endophyte is a category of microbial symbionts that interacts intimately with the host cells. Endophytes ('endo', inside; 'phyte', plant) are mainly fungi and bacteria that thrive inside the host tissues and are detected in all vegetative stages of the host development. They invade the host plant from surrounding soil and are under direct influence of root exudates. Rice, as other plants, attracts distinct community of endophytes by releasing specific compounds into the soil [3]. A rich source of 'readily-available' (e.g. low molecular weight compounds) and recalcitrant nutrients diffuses into the rhizosphere (the soil which is directly affected by plant roots), attracting diverse heterotrophic microorganisms. Competent heterotrophic microorganisms first colonize the rhizoplane (i.e. the epidermal cells of plant roots) and later a selected fraction of these organisms may occupy the internal root tissues to become endophytic. Hence, most microbial colonization traits that are observed in rhizophere are expected to be present in the endosphere [4]. Furthermore, microbes equipped with traits for efficient nutrient uptake, versatile nutrient metabolism, stress resistance and competitive fitness might be at an advantage to become endophytic. The rhizodeposition as mechanism of plant selection is poorly understood and further research is needed to clarify which molecules are discriminated by each microbe among the rich source of photo-assimilated compounds released by the host. Once inside the roots, endophytes might migrate to aerial tissues, including seeds [5,6]. Indeed, vertical transmission via seeds is another important mechanism for the establishment of endophytes. Various endophytes have been observed in rice seedlings growing aseptically [7-12] and it has been estimated that almost half of the early bacterial community from rice seedlings is originated from vertical transmission [9]. Thus, one might not exclude the selection of specific microbial communities to perpetuate mutualistic symbiosis [13]. The importance of vertical transmission becomes even more evident when beneficial endophytes improve host fitness upon adverse environmental conditions [14] or under conditions without induced stress [15].

A complete understanding of the community composition and

their putative functions inside the plant are necessary to foster the efficient use of endophytes as plant growth promoters. Many surveys of the prokaryotic community inside rice plants have been reported. For instance, culture-dependent and -independent surveys of the rice roots cv. APO identified an astonishing diversity of prokaryotic endophytes that were encompassed by 16 phyla/classes [16]. In both approaches, members of Gammaproteobacteria were the most abundant class, followed by Alphaproteobacteria. Members of Epsilon- and Delta-proteobacteria, Bacteroidetes, Fibrobacteres, Planctomycetes, Nitrospirae, Tenericutes, Clostridia, Negativicutes, candidatus division TM7, Cyanobacteria and Crenarchaeota were exclusively identified in the clones, whereas Actinobacteria were only found as isolates. Although diverse, the prokaryotic community was largely dominated (i.e. one third of the sequences) by members of the recently described genus Kosakonia [17]. The investigated Kosakonia strains revealed various in vitro plant growth-promoting properties such as production of siderophores, indole-3-acetic acid (IAA), acetoin and 1-aminocyclopropane-1-carboxylate (ACC) deaminase, solubilisation of inorganic phosphate, reduction of nitrate, and fixation of atmospheric N₂ as well as plant adaptation properties such as catalase production, oxidation of methanol, and production of extracellular cellulase, amylase and protease [17].

The first functional metagenome analysis from rice roots was carried out using the same cultivar described above. The results collected from the non-PCR-amplified DNA fragments also revealed high diversity of prokaryotic endophytes [18]. The identified genes encoding proteins suggest that the endophytic bacteria have special properties to live inside the host tissues, which often includes osmotic protection and resistance to oxidative stresses, production of hydrolytic enzymes and outer-membrane proteins involved in host recognition and adhesion, dedicated communication systems via autoinducer molecules, diverse transcriptional regulators to rapidly respond to host cues, metabolic adaptation to cope in oxic and anoxic niches, and degradation of aliphatic, aromatic and carbon storage compounds as well as ecological and nutritional versatility to use plant metabolites as carbon sources. In addition, many of the plant growth promotion properties described earlier and even more were detected in the rice metagenome [18]. The authors also showed that almost all enzymes involved in the N-cycle are represented in the endophyte metagenome. Genes encoding proteins

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for nitrification (occurring in aerobic condition), denitrification (anaerobic), and nitrogen fixation (semi-anaerobic) were detected in high abundance. Moreover, the *nifH* gene involved in nitrogen fixation was actively transcribed inside rice roots, suggesting that some bacterial species might contribute to the overall nitrogen pool inside the host [18-20]. This is an energetically highly demanding process, which only occurs when certain conditions are met. Under flood conditions and in the presence of reliable nutrient sources, qualified bacteria might actively incorporate atmospheric N₂ into the rice host metabolism, like rhizobia do inside leguminous plants.

Rice plants inoculated with endophytic bacteria show significant improvement in plant growth and grain yields. Vigorous seedling development is important for plant establishment and grain production and inoculation of rice seeds with endophytic bacteria Herbaspirillum seropedicae, Sinorhizobium meliloti, Achromobacter xylosoxidans, Pseusomonas spp., Sphingomonas trueperi, and Serratia marcescens promotes seedlings' growth [21-26]. Nitrogen, one of the most limiting nutrients for rice cultivation, is also significantly increased inside rice plants inoculated with diazotrophic endophyte strains Herbaspirillum seropedicae, Burkholderia spp., Rhizobium sp. and Corynebacterium sp. as well as fungal endophytes Phomopsis liquidambari [22,25,27-29]. Increased in grain yields have been reported for rice plants inoculated with endophytes Rhizobium leguminosarum bv. trifolii, Rhizobium sp. IRBG74, Burkholderia cepacia/vietnamiensis complex, Pseudomonas sp., Herbaspirillum seropedicae, Gluconacetobacter diazotrophicus, Azospirillum brasilense, and Pantoea agglomerans strains [30-33]. Endophytes can also improve plant growth via biocontrol mechanisms. Those with excellent antagonistic properties against plant pathogens might be used to reduce pathogen growth. Rice endophytic bacteria Acidovorax sp. S20, Acinetobacter baylyi, Acinetobacter calcoaceticus, Acinetobacter junii, Acinetobacter sp. B, Actinomycetes spp., Agrobacterium larrymoorei, Agrobacterium radiobacter, Arthrobacter citreus, Bacillus amyloliquefaciens, Bacillus cereus, Bacillus megaterium, Bacillus mycoides, Bacillus polyfermenticus, Bacillus pumilus, Bacillus simplex, Bacillus sp. WL-3, Bacillus sphaericus, Bacillus subtilis, Brevibacillus brevis, Brevibacterium epidermidis, Burkholderia cepacia, Burkholderia gladioli, Burkholderia glumae, Burkholderia kururiensis, Burkholderia sp. KJ006, Curtobacterium flaccumfaciens, Deinococcus aquaticus, Enterobacter cloacae, Gluconacetobacter diazotrophicus, Klebsiella mobilis, Microbacterium esteroaromaticum, Micrococcus lylae, Paenibacillus cineris, Paenibacillus favisporus, Paenibacillus lentimorbus, Pantoea agglomerans, Pantoea ananatis, Pantoea dispersa, Pantoea stewartii, Pseudomonas aeruginosa, Pseudomonas entomophila, Pseudomonas fluorescens, Pseudomonas oryzihabitans, Pseudomonas putida, Pseudomonas sp. PALXIL12, Pseudomonas stutzeri, Sphingomonas capsulate, Staphylococcus arlettae, Staphylococcus simulans, Stenotrophomonas maltophi, and Xanthomonas sp. LMG 20137 and fungi Fusarium moniliforme, Trichoderma, Harpophora oryzae, Phialemonium curvatum, Phaeosphaeriopsis musae have been recommended for their biocontrol potential [7,34-43].

Until recently, microbial community detected inside the host plants has been associated with diseases development. We learned from human microbiome the importance of high diversity community to maintain the health and functioning stability. As in human microbiome, plant disease outbreaks are often associated with shifts of the whole microbial community originated from the imbalance growth of specific symbiont community. The symbionts mentioned above exemplify the importance of endophytes for the host fitness and grain production. It also demonstrates that host genome and its symbiont microbiome are intimately interconnected creating the rice holobiome, the genome content of this superorganism. As the study of Gregor Johann Mendel on genetic inheritance in garden peas revolutionized biology by introducing the laws of inheritance, the concept of plant holobiome will allow us to understand even further the factors affecting plant production. Sustainable and responsible agricultural production will be guided by our knowledge on how plant holobiome operates.

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References

- 1. Kiers ET, Denison RF (2008) Sanctions, cooperation, and the stability of plantrhizosphere mutualisms. Annu Rev Ecol Evol Syst 39: 215-236.
- 2. Jones JDG, Dangl JL (2006) The plant immune system. Nature 444: 323-329.
- Bacilio-Jimenez M, Aguilar-Flores S, Ventura-Zapata E, Perez-Campos E, Bouquelet S, et al. (2003) Chemical characterization of root exudates from rice (*Oryza sativa*) and their effects on the chemotactic response of endophytic bacteria. Plant Soil 249: 271-277.
- Hardoim PR, van Overbeek LS, van Elsas JD (2008) Properties of bacterial endophytes and their proposed role in plant growth. Trends Microbiol 16: 463-471.
- Chi F, Shen SH, Cheng HP, Jing YX, Yanni YG, et al. (2005) Ascending migration of endophytic rhizobia, from roots to leaves, inside rice plants and assessment of benefits to rice growth physiology. Appl Environ Microbiol 71: 7271-7278.
- Compant S, Kaplan H, Sessitsch A, Nowak J, Barka EA, et al. (2008) Endophytic colonization of *Vitis vinifera* L. by *Burkholderia phytofirmans* strain PsJN: from the rhizosphere to inflorescence tissues. FEMS Microbiol Ecol 63: 84-93.
- Cottyn B, Debode J, Regalado E, Mew TW, Swings J (2009) Phenotypic and genetic diversity of rice seed-associated bacteria and their role in pathogenicity and biological control. J Appl Microbiol 107: 885-897.
- Fisher PJ, Petrini O (1992) Fungal saprobes and pathogens as endophytes of rice (*Oryza sativa* L.). New Phytol 120: 137-143.
- Hardoim PR, Hardoim CCP, van Overbeek LS, van Elsas JD (2012) Dynamics of seed-borne rice endophytes on early plant growth stages. PLoS ONE 7.
- Mano H, Tanaka F, Nakamura C, Kaga H, Morisaki H (2007) Culturable endophytic bacterial flora of the maturing leaves and roots of rice plants (*Oryza* sativa) cultivated in a paddy field. Microbes Environ 22: 175-185. DOI: 10.1264/ jsme2.22.175
- Mukhopadhyay K, Garrison NK, Hinton DM, Bacon CW, Khush GS, et al. (1996) Identification and characterization of bacterial endophytes of rice. Mycopathologia 134: 151-159.
- Ruiza D, Agaras B, de Werrab P, Wall LG, Valverde C (2011) Characterization and screening of plant probiotic traits of bacteria isolated from rice seeds cultivated in Argentina. J Microbiol 49: 902-912.
- Bragina A, Cardinale M, Berg C, Berg G (2013) Vertical transmission explains the specific *Burkholderia* pattern in *Sphagnum mosses* at multi-geographic scale. Front Microbiol 4.
- Puente M E, Li CY, Bashan Y (2009) Endophytic bacteria in cacti seeds can improve the development of cactus seedlings. Environ Exp Bot 66: 402-408.
- Mastretta C, Taghavi S, van der Lelie D, Mengoni A, Galardi F, et al. (2009) Endophytic bacteria from seeds of *Nicotiana tabacum* can reduce cadmium phytotoxicity. Int. J. Phytoremediation 11: 251-267
- Hardoim P (2011) Bacterial endophytes of rice their diversity, characteristics and perspectives. Ridderprint BV, Ridderkerk, The Netherlands.
- 17. Hardoim PR, Nazir R, Sessitsch A, Elhottova D, Korenblum E, et al. (2013) The new species *Enterobacter oryziphilus* sp. nov. and *Enterobacter oryzendophyticus* sp. nov. are key inhabitants of the endosphere of rice. BMC Microbiol 13.
- Sessitsch A, Hardoim P, Doering J, Weilharter A, Krause A, et al. (2012) Functional characteristics of an endophyte community colonizing rice roots as revealed by metagenomic analysis. Mol Plant Microbe Interact 25: 28-36.
- Egener T, Hurek T, Reinhold-Hurek B (1999) Endophytic expression of *nif* genes of *Azoarcus* sp strain BH72 in rice roots. Mol Plant Microbe Interact 12: 813-819.
- 20. Elbeltagy A, Ando Y (2008) Expression of nitrogenase gene (nifH) in roots and

stems of rice, *Oryza sativa*, by endophytic nitrogen-fixing communities. Afr J Biotechnol 7: 1950-1957.

- Adhikari TB, Joseph CM, Yang GP, Phillips DA, Nelson LM (2001) Evaluation of bacteria isolated from rice for plant growth promotion and biological control of seedling disease of rice. Can J Microbiol 47: 916-924.
- Baldani VLD, Baldani JI, Dobereiner J (2000) Inoculation of rice plants with the endophytic diazotrophs *Herbaspirillum seropedicae* and *Burkholderia* spp. Biol. Fertil. Soils 30: 485-491.
- Chi F, Yang P, Han F, Jing Y, Shen S (2010) Proteomic analysis of rice seedlings infected by *Sinorhizobium meliloti* 1021. Proteomics 10: 1861-1874.
- Gyaneshwar P, James EK, Mathan N, Reddy PM, Reinhold-Hurek B, et al. (2001) Endophytic colonization of rice by a diazotrophic strain of *Serratia marcescens*. J Bacteriol 183: 2634-2645.
- James EK, Gyaneshwar P, Mathan N, Barraquio QL, Reddy PM, et al. (2002) Infection and colonization of rice seedlings by the plant growth-promoting bacterium *Herbaspirillum seropedicae* Z67. Mol Plant Microbe Interact 15: 894-906.
- 26. Jha P, Kumar A (2009) Characterization of novel plant growth promoting endophytic bacterium Achromobacter xylosoxidans from wheat plant. Microb Ecol 58: 179-188.
- Gyaneshwar P, James EK, Reddy PM, Ladha JK (2002) Herbaspirillum colonization increases growth and nitrogen accumulation in aluminium-tolerant rice varieties. New Phytol 154: 131-145.
- Naher UA, Othman R, Shamsuddin ZHJ, Saud HM, Ismail MR (2009) Growth enhancement and root colonization of rice seedlings by *Rhizobium* and *Corynebacterium* spp. Int J Agric Biol 11: 586-590.
- 29. Yang B, Ma H-Y, Wang X-M, Jia Y, Hu J, et al. (2014) Improvement of nitrogen accumulation and metabolism in rice (*Oryza sativa* L.) by the endophyte *Phomopsis liquidambari*. Plant Physiol Biochem 82: 172-182.
- Biswas JC, Ladha JK, Dazzo FB, Yanni YG, Rolfe BG (2000) Rhizobial inoculation influences seedling vigor and yield of rice. Agronomy J 92: 880-886.
- Feng Y, Shen D, Song W (2006) Rice endophyte *Pantoea agglomerans* YS19 promotes host plant growth and affects allocations of host photosynthates. J Appl Microbiol 100: 938-945.
- 32. Jha B, Thakur MC, Gontia I, Albrecht V, Stoffels M, et al. (2009) Isolation, partial identification and application of diazotrophic rhizobacteria from traditional Indian rice cultivars. Eur J Soil Biol 45: 62-72.

- 33. Yanni YG, Rizk RY, Corich V, Squartini A, Ninke K, et al. (1997) Natural endophytic association between *Rhizobium leguminosarum* bv. *trifolii* and rice roots and assessment of its potential to promote rice growth. Plant Soil 194: 99-114.
- Seenivasan N, David PMM, Vivekanandan P, Samiyappan R (2012) Biological control of rice root-knot nematode, *Meloidogyne graminicola* through mixture of *Pseudomonas fluorescens* strains. Biocontrol Sci Technol 22: 611-632.
- Cho H-S, Park S-Y, Ryu C-M, Kim JF, Kim J-G, et al. (2007) Interference of quorum sensing and virulence of the rice pathogen *Burkholderia glumae* by an engineered endophytic bacterium. FEMS Microbiol Ecol 60: 14-23.
- Le HTT, Padgham JL, Sikora RA (2009) Biological control of the rice root-knot nematode *Meloidogyne graminicola* on rice, using endophytic and rhizosphere fungi. Int J Pest Manage 55: 31-36.
- Saravanan VS, Madhaiyan M, Osborne J, Thangaraju M, Sa TM (2008) Ecological occurrence of *Gluconacetobacter diazotrophicus* and nitrogen-fixing Acetobacteraceae members: their possible role in plant growth promotion. Microb Ecol 55: 130-140.
- Sikora RA, Pocasangre L, zum Felde A, Niere B, Vu TT, et al. (2008) Mutualistic endophytic fungi and *in-planta* suppressiveness to plant parasitic nematodes. Biol. Control 46: 15-23.
- Su Z-Z, Mao L-J, Li N, Feng X-X, Yuan Z-L, et al. (2013) Evidence for biotrophic lifestyle and biocontrol potential of dark septate endophyte *Harpophora oryzae* to rice blast disease. PLoS ONE 8.
- Suada IK, Suhartini DMWY, Sunariasih NPL, Wirawan IGP, Chun KW, et al. (2012) Ability of endophytic fungi isolated from rice to inhibit *Pyricularia oryzae* induced rice blast in Indonesia. J Fac Agr Kyushu U 57: 51-53.
- Yang JH, Liu HX, Zhu GM, Pan YL, Xu LP, et al. (2008) Diversity analysis of antagonists from rice-associated bacteria and their application in biocontrol of rice diseases. J Appl Microbiol 104: 91-104.
- 42. Zhang S-M, Wang Y-X, Meng L-Q, Li J, Zhao X-Y, et al. (2012) Isolation and characterization of antifungal lipopeptides produced by endophytic *Bacillus amyloliquefaciens* TF28. Afr J Microbiol Res 6: 1747-1755.
- 43. Huong LTT (2010) Activity of fungal and bacterial endophytes for the biological control of the root-knot nematode *Meloidogyne graminicola* in rice under oxic and anoxic soil conditions. University of Bonn, Bonn, Germany.

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