

Human Interleukin 2 (IL-2) Promotion of Immune Regulation and Clinical Outcomes: A Review

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Abstract

Interleukin 2 (IL-2) is a monomeric glycoprotein that is primarily produced by activated CD4+ T cells, CD8+ T cells and dendritic cells. It is characterized as a proinflammatory cytokine that is secreted by Th1 cells. IL-2 plays a central role in the activation of regulatory T cells to produce the cytokines tumor necrosis factor alpha (TNF- α) and interferon gamma (IFN- γ). IL-2 may also enhance the cytolytic activity of natural killer cells, thereby ensuring their significance in the control of the immune response, and effectively participate in the pathogenesis of several pathological conditions, such as cancer and metabolic, infectious, autoimmune and inflammatory diseases. We emphasize the importance of studies of IL-2 and discuss perspectives resulting from our increasing understanding of genetic diversity and its role in the immune response.

Keywords: IL-2; Immune response; Polymorphism; Parasitic infectious diseases

Interleukin 2 and the Immune Response

Cytokines are produced by various immune system cells and perform several functions, including mediation of the immune and inflammatory responses. The effects of cytokines on the immune response depend on a number of factors, such as their local concentrations, receptor expression patterns and the integration of multiple signalling pathways in response to immune cells [1]. The immune system includes proinflammatory cytokines that can enhance the functions of other cytokines and the immune response and antiinflammatory cytokines that suppress this response; various interleukins (ILs) stand out in these responses [2]. ILs are small protein molecules that signal specific cells to regulate the immune systems of organisms. They are primarily synthetized by T cells, monocytes, macrophages and endothelial cells. The functions of ILs include the facilitation of communication among immune system cells, regulation of transcription factors, and control of inflammation, cell differentiation, proliferation and antibody secretion [3]. The characterization of interleukin 2 (IL-2) as a T-cell growth factor was consolidated in 1975 at the Second International Lymphokine Workshop. The number of studies on this molecule increased quickly; by 1983, the IL-2 gene was cloned, and in 1992, the IL-2 crystal structure was described [1]. Analysis of the three-dimensional structure of the IL-2 molecule shows that it is composed of four "packed" α-helices. The first and fourth helices are connected by a long upper loop to form a typical structure known as "up-up-down-down". Within this configuration, the first two a-helices are turned upward,

and the last two helices are turned downward. Importantly, the disulphide bond between the cysteines at positions 58 and 105 (Cys 58-105) of the second helix and the inter-helix region of the third and fourth α -helices are necessary to ensure the stability of the protein [3]. IL-2 is a monomeric glycoprotein with a molecular weight of approximately 15 kDa that is primarily produced by activated CD4+ T cells, CD8+ T cells and dendritic cells [4]. IL-2 is a proinflammatory cytokine that is secreted by Th-1 cells, and it effectively participates in the activation of T cells to produce the cytokines tumor necrosis factor alpha (TNF- α) and interferon gamma (IFN- γ); IL-2 can also enhance the 3 cytolytic activity of natural killer cells (NK) [5,6]. Therefore, IL-2 is used therapeutically to stimulate the immune system [7]. IL-2 also contributes to the development of regulatory T cells, which control the expansion and apoptosis of activated T cells [5,8]. Furthermore, IL-2 influences cell survival, differentiation [1] and the formation of immune memory cells [1,9] and acts as a negative regulator of immune activation [8]. Recent studies showed that IL-2 played a critical role in the differentiation and survival of regulatory T cells, thereby ensuring their significance in the control of the immune response [10]. Cytokines effectively participate in the pathogenesis of several pathological conditions, such as cancer and metabolic, infectious, autoimmune and inflammatory diseases [11,12]. Thus, IL-2 plays multiple roles in immune functions by contributing to the generation and propagation of antigen-specific immune responses [13]. Studies conducted with animals and humans showed that low doses of IL-2 induced expansion of regulatory T cells in vivo and suppressed autoimmune diseases; this phenomenon is representative of a novel therapeutic approach to modulate the immune response for the treatment of these types of illnesses [14]. Anti-humoral therapy associated with IL-2 administration led to the remission of metastatic

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renal cell carcinoma in up to 30% of patients and increased the survival of patients with melanoma and acute myeloid leukemia [15]. In these situations, the administration of high doses of IL-2 was associated with improved survival, although the related adverse effects were considerably severe in most patients [16]. Cases of cellular and humoral immunodeficiency exhibited satisfactory outcomes following IL-2 administration [17].

Genetic Variability of Il-2 and the Physiopathogenesis of Parasitic Infections and Autoimmune Diseases and Cancer

Individual genetic variability is an essential component of the immune response in general. This fact has become increasingly evident in recent years because genetic variability contributes to the susceptibility to, progression and outcomes of infectious and autoimmune diseases and cancer. While attempting to map its role in multifactorial and polygenic diseases, several studies revealed the true extension of human genetic variation [18]. Cytokines and their receptors emerged as intriguing targets of therapeutic intervention. Cytokine receptor subunit expression differs among various cell types. These subunits may be shared by different cytokines, which increases the complexity of these molecules [19]. The IL-2 receptor (IL-2R) is comprised of three subunits (α , β and γ) [20]. The The gene that encodes subunit a is located on chromosome 10; this subunit (also known as CD25 or Tac antigen) is the subunit to which IL-2 selectively binds. The main function of the IL-2R α chain is to mediate the receptor's affinity for its ligand. Different from the other two IL-2R subunits, subunit a is located in special regions of the plasma membrane [21]. Subunit a simply requires IL-2 binding to help subunits β and γ come closer and consequently, trigger the signaling cascade. The gene that encodes subunit β (p75 or CD122) is located on chromosome 22. The β and γ chains together constitute an intermediate affinity receptor that is sufficient to trigger the IL-2 signaling pathways. Subunits β and γ are located in regions close to subunit a, which is the subunit that selectively binds IL-2. The gene that encodes the γc chain of the IL-2R (common γ or p64) is located on chromosome X. Several cytokines share IL-2Ry, such as IL-4, IL-7, IL-9, IL-15 and IL-21 [1]. When IL-2 receptor alpha (IL-2Ra) is activated, its soluble form is released into the serum. Thus, IL-2Ra protein concentration can be measured. In patients with colorectal and breast cancer, high serum IL-2Ra concentrations are indicative of disease progression and distant metastasis [22]. According to studies that analyzed genes encoding cytokines, several polymorphisms in the regulatory regions of these genes might be responsible for the changes in the production of the corresponding cytokines [23]. Several studies noted the importance of single-nucleotide polymorphisms (SNPs) in the occurrence of infectious and autoimmune diseases [24], the transplantation course [25] and the allele frequencies of populations [26]. The association of SNPs with human diseases has great potential for clinical applications because it provides new genetic markers for diagnosis and prognosis and possibly new therapeutic targets.

The IL-2 gene (3558) is located in chromosome 4, region q26-q27. It contains four exons separated by three introns, with a total extension of approximately 5 kb [27]. Several IL-2 gene polymorphisms are known, including those at positions -330 T/G and +114 T/G. Polymorphism -330 T/G (rs2069762) is located in the gene promoter region and is associated with increased susceptibility to inflammatory diseases and cancer, including rheumatoid arthritis and myeloid leukemia [28-30]. Additionally, studies show that changes in the serum

IL-2 levels were found when this polymorphism was analyzed [29-31]. This finding was confirmed because the presence of allele G in gene position -330 was correlated with reduced IL-2 production *in vivo* [32]. Studies showed that the T lymphocytes of patients with SNP -330 G/G were able to produce larger amounts of IL-2 than the lymphocytes from patients with SNP -330 T/G or T/T, which suggests that the presence of -330T/G (rs2069762) in the aforementioned promoter region may influence IL-2 production in healthy individuals [33]. Studies on prostate cancer also found an association between genetic variants within IL-2 and the risk for this type of cancer, revealing a significant contribution of the IL-2 exon 1 variant rs2069763 G/T to disease susceptibility [34].

Several studies noted the possible association of asthma with genes that encoded components of the immune response, including cytokine genes, due to their roles in the pathophysiology of the disease [35]. One study investigated the association of the IL-2 gene polymorphism +114 T/G (rs2069763) and its genetic variants with asthma; however, no significant positive correlation was found [36]. IL2 +166 G/T is another polymorphism of the IL-2 gene that encodes a silent mutation that does not affect the amino acid sequence [37].

Studies on leishmaniasis showed that the presence of variants in the IL-2R β gene predisposed individuals infected with *Leishmania donovani* to the development of visceral leishmaniasis (VL), which indicated that the IL-2 signaling pathway participated in the occurrence of leishmaniasis [38]. Additionally, susceptibility to VL (caused by *L. donovani*) in Sudan (Aringa ethnic group) was controlled by locus 22q12 [39]. Another study showed that mutations in the IL-2R β gene located in locus 22q12 might be at least partially responsible for the genetic linkage with VL [38], thus demonstrating the importance of this IL-2 signaling pathway.

The immune response to malaria involves innate and adaptive mechanisms, including the participation of several cell types and soluble components that lead to the elimination of the etiologic agent or to immunopathology [40-42]. *Plasmodium vivax* elicits a specific immune response in the host that is mediated by humoral (Th2 lymphocytes) and cellular (Th1 lymphocytes) mechanisms. The humoral mechanisms are characterized by the involvement of antibodies that confer protection through the opsonization of merozoites, thereby blocking the invasion of erythrocytes. This response is characterized by the participation of T lymphocytes, which, after recognizing agents processed by antigen-processing cells (APCs), release interleukins that regulate macrophages, dendritic cells and even B lymphocytes to activate the immune response against the etiological agent.

B lymphocytes are activated by released IL-2, IL-4 and IL-5, which determine the type of antibodies that will be produced, as well as the immunoglobulin isotype switch. Regarding the cell-mediated response, ILs released by T lymphocytes increase the phagocytic activity of macrophages, neutrophils, dendritic cells, monocytes and NK lymphocytes to combat infections [40,42-44].

T lymphocytes are activated when the T-cell receptor (TCR) and CD4 or CD8 co-receptors recognize the major histocompatibility complex (MHC) expressed on APCs and bind to peptide antigens processed by lysosomal or proteasomal mechanisms [40,45]. However, binding alone does not suffice to trigger clonal expansion, which needs a second co-stimulatory signal provided by the APCs themselves through glycoproteins known as B7.1 (or CD80) and B7.2 (or CD86).

These glycoproteins bind to the corresponding receptor on T cells (the CD28 molecule) [45].

Several studies revealed an association between polymorphisms of human genes involved in the invasion of erythrocytes by parasites and the susceptibility to vivax malaria. Additionally, the variability in genes that encode molecules involved in the immune response is associated with changes in the pattern of parasitemia, clinical aspects and susceptibility to disease [18,46].

Concerning *P. vivax* erythrocyte invasion receptors and ligands, individuals heterozygous for the Duffy blood group antigens (FyA/ FyB) were found to be more susceptible to malaria in Brazil [47]. However, Duffy-negative individuals may become infected with *P. vivax* via other receptors involved in erythrocyte recognition [47,48].

The role of the human leukocyte antigen (HLA) system in the immune response to Plasmodium antigens has been widely investigated. Allelic differences in these genes exhibited contradictory results among the various investigated endemic areas, including Brazil; therefore, it seems unlikely that they are the only mechanism responsible for deviations of the immune response [49-51].

The current knowledge about the genes involved in the immune response and the implications of their variability for the causes of disease has been greatly advanced by data from human genome sequencing. In addition to the genes and alleles involved in the pathogenesis of several diseases, association studies have allowed the frequencies of these alleles to be determined in these populations. Because genetic polymorphisms vary among different population groups, the heterogeneity of the Brazilian populations will contribute relevant information to the understanding of the causes of complex diseases, such as malaria. These studies also contribute knowledge on the evolution a functional implications of genetic polymorphisms.

Concluding Remarks

The development of the immune response depends on a cellular and molecular complex that is essential for the protection of humans against infectious agents, autoimmune diseases and tumors. For the immune response to adequately occur, a balance is needed in the ability of the cells to respond to infectious agents and to suppress autoimmunity [52]. Thus, polymorphisms associated with modulation of the expression of genes that encode immune response costimulatory molecules might influence the occurrence of various diseases; indeed, several recent studies demonstrated this association [53].

Variability in genes that encode molecules involved in the immune response is associated with changes in the pattern of parasitemia, clinical aspects and susceptibility to disease. The current knowledge about the genes involved in the immune response and the implications of their variability for the cause of disease has been greatly advanced by data resulting from human genome sequencing. IL-2 is a cytokine that contributes to the differentiation and survival of regulatory T cells, thereby ensuring their significance in the control of the immune response and their effective participation in the pathogenesis of several pathological conditions, such as cancer and metabolic, infectious, autoimmune and inflammatory diseases [11,12]. Therefore, IL-2 plays multiple roles in immune functions by contributing to the generation and propagation of antigen-specific immune responses [13]. Although the relevant role of this molecule in the immune response is known, more studies on its function are needed, especially concerning parasitic infectious diseases.

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References

- 1. Gaffen SL, Liu KD (2004) Overview of interleukin-2 function, production and clinical applications. Cytokine 28: 109-123.
- 2. Shankaran V, Ikeda H, Bruce AT, White JM, Swanson PE, et al. (2001) IFNgamma and lymphocytes prevent primary tumour development and shape tumour immunogenicity. Nature 26: 410.
- Salazar-Onfray F, Lopez MN, Mendoza-Naranjo A (2007) Paradoxical effects of cytokines in tumor immune surveillance and tumor immune escape. Cytokine & growth factor reviews 18: 171-182.
- 4. Nelson AJ, Staines WR, McIlroy WE (2004) Tactile stimulus predictability modulates activity in a tactile-motor cortical network. Exp Brain Res: 154(1):22-32.
- Lenardo MJ (1991) Interleukin-2 programs mouse alpha beta T lymphocytes for apoptosis. Nature 353: 858-861.
- Sakaguchi S, Yamaguchi T, Nomura T, Ono M (2008) Regulatory T cells and immune tolerance. Cell 133: 775-787.
- Smith AJP, Humphries SE (2009) Cytokine and cytokine receptor gene polymorphisms and their functionality. Cytokine & Growth Factor Reviews 20:43-59.
- D'Souza WN, Lefrancois L (2003) IL-2 is not required for the initiation of CD8 T cell cycling but sustains expansion. Journal of Immunology 171: 5727-5735.
- 9. Malek TR, Bayer AL (2004) Tolerance, not immunity, crucially depends on IL-2. Nature Reviews/ Immunology 4: 665-674.
- Malek TR, Yu A, Zhu L, Matsutani T, Adeegbe D, et al. (2008) IL-2 family of cytokines in T regulatory cell development and homeostasis. J Clin Immunol. 28: 635-639.
- 11. Mok CC, Lau CS (2003) Pathogenesis of systemic lupus erythematosus. JClinPathol.56: 481-490.
- 12. Scheller J, Ohnesorge N, Rose-John S (2006) Interleukin-6 transsignalling in chronic inflammation and cancer. Scand J Immunol. 63: 321.
- 13. Raeburn CD, Sheppard F, Barsness KA, Arya J, Harken AH (2002) Cytokines for surgeons. Am J Surg183:268-273.
- 14. Yu HB, Yurieva M, Balachander A, Foo I, Leong X, et al. (2015) NFATc2 mediates epigenetic modification of dendritic cell cytokine and chemokine responses to dectin-1 stimulation. Nucleic Acids Res 43: 836-847.
- 15. Waxman J, Balkwill F (1992) Interleukin 2. Black-well Sci Publ.
- Halama N, Zoernig I, Jaeger D (2010) Advanced malignant melanoma: immunologic and multimodal therapeutic strategies. Journal of oncology 689893.
- 17. Hatakeyama M, Tsudi M, Minamoto S, Kono T, Doi T, et al. (1989) Interleukin 2 receptor beta chain gene: generation of the three receptor forms by cloned human alpha and beta chain DNAs. Science 244: 551-556.
- Driss A, Hibbert JM, Wilson NO, Iqbal SA, Adamkiewicz TV, et al. (2011) Genetic polymorphisms linked to susceptibility to Malaria. Malaria Journal 10: 271.

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- Bluestone JA, Bour-Jordan H, Cheng M, Anderson M (2015) T cells in the control of organ specific autoimmunity. J Clin Invest 125: 2250-2260.
- Yu A, Malek TR (2001) The Proteasome Regulates Receptor-mediated Endocytosis of Interleukin-2. The Journal of Biological Chemistry 276: 381-385.
- 21. Marmor MD, Julius M (2001) Role for lipid rafts in regulating interleukin-2 receptor signaling. Immunology 98: 1489-1497.
- 22. Saito H, Tsujitani S, Katano K, Ikeguchi M, Maeta M, et al. (1998) Levels of serum-soluble receptor for interleukin-2 in patients with colorectal cancer. Surgery today 28(10):1115-1117.
- 23. Pravica V, Asderakis A, Perrey C, Hajeer A, Sinnott PJ, et al. (1999) In vitro production of IFN-gamma correlates with CA repeat polymorphism in the human IFN-gamma gene. Eur J Immunogenet 26: 1-3.
- Maxwell JR, Potter C, Hyrich KL (2008) Association of the tumour necrosis factor-308 variant with differential response to anti-TNF agents in the treatment of rheumatoid arthritis. Hum Mol Genet. 17: 3532-3538.
- 25. Wilson AG, di Giovine FS, Blakemore AI, Duff GW (1992) Single base polymorphism in the human tumour necrosis factor alpha (TNF alpha) gene detectable by NcoI restriction of PCR product. Hum Mol Genet 1: 353.
- Visentainer JE, Sell AM, da Silva GC, Cavichioli AD, Franceschi DS, et al. (2008) TNF, IFNG, IL6, IL10 and TGFB1 gene polymorphisms in South and Southeast Brazil. Int J Immunogenet. 35: 287-293.
- 27. Fujita T, Takaoka C, Matsui H, Taniguchi T (1983) Structure of the human interleukin 2 gene. Proc Natl Acad 80: 7437-7441.
- Amirzargar AA, Bagheri M, Ghavamzadech A, Alimoghadam K, Khosravi F, et al. (2005) Cytokine gene polymorphism in Iranian patients with chronic myelogenousleukaemia. International Journal Immunogenetic 32:167-171.
- 29. Shin WG, Jang JS, Kim HS, Kim SJ, Kim KH, et al. (2008) Polymorphisms of interleukin-1 and interleukin-2 genes in patients with gastric cancer in Korea. Journal of Gastroenteroly and Hepatology 23: 1567-1573.
- 30. Wu J, Lu Y, Ding YB, Ke Q, Hu ZB, et al. (2009) Promoter polymorphisms of IL2, IL4, and risk of gastric cancer in a high-risk Chinese population. Molecular Carcinogenesis 48: 626-632.
- 31. Togawa S, Joh T, Itoh M, Katsuda N, Ito H, et al. (2005) Interleukin-2 gene polymorphisms associated with increased risk of gastric atrophy from Helicobacter pylori infection. Helicobacter 10: 172-178.
- 32. Matesanz F, Fedetz M, Leyva L, Delgado C, Fernandez O, et al. (2004) Effects of the multiple sclerosis associated - 330 promoter polymorphism in IL2 allelic expression. Journal of Neuroimmunology 148: 212-217.
- 33. Hoffmann SC, Stanley EM, Darrin Cox E, Craighead N, DiMercurio BS, et al. (2001) Association of cytokine polymorphic inheritance and in vitro cytokine production in anti-CD3/CD28- stimulated peripheral blood lymphocytes. Transplantation. 72(8):1444-1450.
- 34. Wu HC, Chang CH, Wan L, Wu CI, Tsai FJ, et al. (2006) IL-2 gene C/T polymorphism is associated with prostate cancer. Journal of clinical laboratory analysis 20: 245-249.
- 35. Ober C, Hoffjan S (2006) Asthma genetics 2006: the long and winding road to gene discovery. Genes Immun 7: 95-100.
- Padrón-Morales J, Sanz C, Dávila I, Muñoz-Bellido F, Lorente F, et al. (2013) Polymorphisms of the IL12B, IL1B, and TNFA genes and susceptibility to asthma. J Investig Allergol Clin Immunol 23: 487-494.
- 37. John S, Myerscough A, Marlow A, Hajeer A, Silman A, et al. (1998) Linkage of cytokine genes to rheumatoid arthritis Evidence of genetic heterogeneity. Ann Rheum Dis 57: 361-365.
- Bucheton B, Argiro L, Chevillard C, Marquet S, Kheir MM et al. (2007) Identification of a novel G245R polymorphism in the IL-2 receptor β

membrane proximal domain associated with human visceral leishmaniasis. Genes Immun. 8: 79-83.

- 39. Bucheton B, Abel L, El-Safi S, Kheir MM, Pavek S, et al. (2003) A major susceptibility locus on chromosome 22q12 plays a critical role in the control of kala-azar. Am. J. Hum. Genet73: 1052-1060.
- 40. Rummel T, Batchelder J, Flaherty P, Lafleur G, Nanavati P, et al. (2004) CD28 Costimulation Is Required for the Expression of T-Cell-Dependent Cell-Mediated Immunity against Blood-Stage Plasmodium chabaudiMalaria Parasites. Infection and Immunity72: 5768-5774.
- Walther M, woodruff J, Edele F, Jeffries D, tongren JE, et al. (2006) Innate Immune Responses to Human Malaria: Heterogeneous Cytokine Responses to Blood-Stage Plasmodium falciparum Correlate with Parasitological and Clinical Outcomes. Journal of Immunology177: 5736-5745.
- 42. Medina TS, Costa Sp, Oliveira MD, Ventura AM, Souza JM, et al. (2011) Increased interleukin-10 andinterferon-g levels in Plasmodium vivax malaria suggest a reciprocal regulation which is not altered by IL-10 gene promoter polymorphism. Malaria Journal 10: 264.
- 43. Jain V, Singh PP, Silawat N, Patel R, Saxena A, et al. (2010) A preliminary study on pro- and anti- inflammatory cytokine profiles in Plasmodium vivax malaria patients from central zone of India. Acta Tropica 113: 263-268.
- 44. Wipasa J, Okell L, Sakkhachornphop S, Suphavilai C, Chawansuntati K, et al. (2011) Short-Lived IFN- γ Effector Responses, but Long-Lived IL-10 Memory Responses, to Malaria in an Area of Low Malaria Endemicity. PLoS Pathogens 7.
- 45. Elias RM, Sardinha LR, Bastos KR, Zago CA, Silva AP, et al. (2005) Role of CD28 in Polyclonal and Specific T and B Cell Responses Required for Protection against Blood Stage Malaria. Journal of Immunology 174: 790-799.
- 46. Sohail M, Kaul A, Bali P, Raziuddin M, Singh MP, et al. (2008) Allels -308A and -1031C in the TNFa gene promoter do not increase the risk but associated with circulating levels of TNFa and clinical features of vivax malarian in Indian patients. Molecular Immunology 45:1682-1692.
- 47. Cavasini CE, De Mattos LC, Couto AA, Couto VS, Gollino Y, et al. (2007) Duffy blood group gene polymorphisms among malaria vivaxpatients in four areas of the Brazilian Amazon region. Malaria Journal.6: 167.
- 48. Ménard D, Barnadas C, Bouchier C, Henry-Halldin C, Gray LR, et al. (2010) Plasmodium vivax clinical malaria is commonly observed in Duffy-negative Malagasy people. Proceedings of the National Academy of Sciences of the United States of America107: 5967-5971.
- 49. Oliveira-Ferreira J, Pratt Riccio LR, Arruda M, Santos F, Ribeiro CT, et al. (2004) HLA class II and antibody responses to circumsporozoite protein repeats of P. vivax (VK210, VK247 and P. vivax-like) in individuals naturally exposed to malaria. ActaTropica 92: 63-69.
- Duah NO, Weiss HA, Jepson A, Tetteh KK, Whittle HC, et al. (2009) Heritability of Antibody Isotype and Subclass Responses to Plasmodium falciparum Antigens. PLoS One 4: 7381.
- 51. Storti-Melo LM, Da Costa DR, Souza-Neiras WC, Cassiano GC, Couto VS, et al. (2012) Influence of HLA-DRB-1 alleles on the production of antibody against CSP, MSP-1, AMA-1, and DBP in Brazilian individuals naturally infected with Plasmodium vivax. Actatropica 121: 152-155.
- 52. Greenwald RJ, Freeman GJ, Sharpe AH (2005) The B7 family revisited. Annu. Rev. Immunol 23:515-548.
- 53. Zayed RA, Sheba HF, Abo Elazaem MA, Elsaadany ZA, Elmessery LO, et al. (2013) B-cell activating factor promoter polymorphisms in egyptian patients with systemic lupus erythematosus. Ann Clin Lab Sci 43:289-294.