

# Prothymosin Alpha (ProT $\alpha$ ) and Thymosin Beta (T $\beta$ ): Conserved Immunerelated Genes from Teleost to Mammals

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#### Abstract

Prothymosin alpha ( $ProT\alpha$ ) and thymosin beta ( $T\beta$ ), members of the thymosin family, consist of a series of highly conserved peptides capable of stimulating immune responses in different species. Our recently published report discussed an issue of the conserved roles of  $ProT\alpha$  b and  $T\beta$ -like ( $T\beta$ -l) in common carp (Cyprinus carpio L), which suggested that the highly-conserved carp  $ProT\alpha$  b and  $T\beta$ -l can ultimately enhance immune response during viral infection and modulate the development of T lymphocytes in Teleost. These results enthusiastically pilled more evidences of evolutionary conservation of thymosins.

Keywords: Prothymosin alpha; Thymosin beta; Teleost; Immune responses

#### Introduction

Thymosin, a mixture of small polypeptides ranging from 1 to 15 kDa [1], were classified into three categories: a-thymosins (Ta) (PH<5.0),  $\beta$ -thymosins (T $\beta$ ) (5.0<PH<7.0), and  $\gamma$ -thymosins (T $\gamma$ ) (PH>7.0) according to different isoelectric points [2]. ProTa is the precursor of  $\alpha$ -thymosins, and is a highly acidic, intrinsically disordered protein containing 109-113 amino acids varying among the species. It was characterized by consisting mainly of aspartic and glutamic acid residues (more than 50%) with few hydrophobic amino acids and without any aromatic or sulfur amino acids [3]. TBs are highly conserved polar 5-kDa polypeptides consisting of 40-44 amino acid residues. ProT $\alpha$  and T $\beta$  genes have been well studies in mammals, but little is known in zeleost. ProTa was reported to be duplicated in Zebrafish (Danio renio) (ProTa a and ProTa b) as they showed different expression patterns [4]. Two thymosins of ProTa b and Tβ-l were studied in carps. It has been clearly demonstrated that ProTa b presented a thymosin  $\alpha$  domain and a KKQK nuclear location signal motif well conserved from fish to mammals. T<sub>β</sub>-l gene revealed the existence of conserved actin binding LKKTET motif and two helix motifs indicated in all known T $\beta$ s [5]. The conserved motifs are supposed to anchor conserved functions of thymosins in different species.

Although thymosins were firstly discovered during the investigation of thymus in mammals, they were later found to be widely distributed in various organs [6] and played important roles in regulation of immunity. Studies, especially in mammals, gradually revealed that ProTa functioned well in regulating immune response and apoptosis, while T $\beta$ 4, one member of T $\beta$ s, may promote lymphocyte proliferation and differentiation [7]. Our study in carps filled the knowledge gaps of their immune roles in teleost. By examining the expression levels of T $\beta$ -l in different carp organs, highest expression was detected in skin and intestine [5], which were important entry sites of pathogens [5]. In SVCV-infected carp models, with introperitoneal injection of virus we successfully demonstrated elevated ProTa b expression in kidney, peripheral blood, spleen, and intestine. At the same time T $\beta$ -l shared the similar expression changes in kidney, peripheral blood, and liver [5]. These results suggested that carp ProTa b and T $\beta$ -l played important roles in antiviral defense, with T $\beta$ -l perhaps functioning as a key activator of NK cell cytotoxicity similar to T $\beta$ 4 [7]. Besides, over-expression of carp ProTa b and T $\beta$ -l genes in zebrafish obviously induced the expression of Rag 1, TCR- $\gamma$ , CD4, and CD8 [5], all of which were essential for cell-mediated immune response and the development of T lymphocytes. Thus, consistent with their conserved structures, ProTa b and T $\beta$ -l indeed exhibited conserved immune-related roles from fish to mammals.

Besides regulation of immunity, ProT $\alpha$  and T $\beta$  also played conserved roles in development. Knockdown of ProTa was reported to lead to apoptosis and developmental defects in zebrafish embryos early in 2013 [8], which was considered as the first evidence that ProTa regulate early embryogenesis. By analyzing the expression levels of ProTα b and Tβ-l at early developmental stages of carps, ProTα b was found to gradually increase starting from 4 h pf, reach maxmium at 16 h pf, and then decrease and keep at a certain stable level, whereas Tβ-l started to increase at 24 h pf and gradually increased up to 72 h pf, and then began to decrease at later developmental stages [5]. Therefore, ProT $\alpha$  b and T $\beta$ -l played important roles in carp's development, with ProTa b exhibiting a clearer role in cell proliferation and/or differentiation, for ProTa b showed the highest expression in carp liver which was considered as an organ with a strong regeneration and recovery capabilities [9]. Moreover, early studies have already pointed out that higher expression of ProTa was found in proliferative cells than in quiescent cells, and more proliferative tissues exhibited higher expression of ProTa in adults [10]. Similar changes were also found in zebrafish as transient expression of ProTa on zebrafish epidermal cells promoted cell proliferation and attunated UBV-induced apoptosis [11]. However, little is known about the role of T $\beta$ -l in development. In consideration of the highly-conserved structures, zeleost can be an effective model for further studying the role of thymosins in development.

Studies of thymosin genes have already progressed from labs to clinics in mammals. As a proliferative protein during development, ProTa is now considered as an oncoprotein and can be an indicator of poor prognosis in human cancers [12]. For example, ProTa was found to protect hepatocellular carcinoma cells against sorafenib-induced apoptosis [13], and targeting ProTa by miR-1 successfully induced apoptosis in nasopharyngeal carcinoma cells [14]. Tβ4, as one member of Tßs, was also reported as an oncoprotein in glioblastoma to promote mesenchymal signature by modulating P53 and TGFB signaling networks [15]. And it was also found to be co-localized with CD133 in ovarian cancers to induce the patterns of cancer stemness [16]. However, not all thymosins function as oncoproteins during carcinogenesis. Ta 1 was reported to exert its anti-cancer effects through PTEN-mediated inhibition of PI3K/Akt/mTOR signaling pathway to suppress proliferation and induce apoptosis in breast cancer [17]. And a successful treatment case of stage Wilms tumor with integrative medical therapy of hyperthermia and Ta 1 combined with herbal remedy was reported in Korean, with liver metastasis successfully disappeared and lung metastasis stably maintained after nine months of treatment [18]. Suppression of T $\beta$ 10 was reported to activate Ras and ERK1/2, upregulate Snail and MMP2 [19], and finally induce metastasis of cholangiocarcinoma, which also indicates the anticancer role of T $\beta$ 10. Besides the carcinogenic roles, T $\beta$ 4 was reported to be successfully used in several clinical trials involving tissue repair and regeneration [20], and could reduce dryness in autoimmune disease of dry eye syndrome [21], which was consistent with its conserved immune-regulating roles and diversely expanded its clinical application. And Serum Ta 1 was supposed to expand its clinical application of regulating disordered immune system in patients with chronic inflammatory autoimmune diseases [22].

## Conclusion

In conclusion, thymosins are extremely conserved during evolution, and share similar functions of immune-regulation and developmental modification in different species. Now the clinically therapeutic and prognostic importance of thymosins has been admitted by more and more studies.

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# References

- Goodall GJ, Dominguez F, Horecker BL (1986) Molecular cloning of cDNA for human prothymosin alpha. Proc Natl Acad Sci 83: 8926-8928.
- Hannappel E, Huff T (2003) The thymosins. Prothymosin alpha, parathymosin, and beta-thymosins: structure and function. Vitam Horm 66: 257-296.
- Gast K, Damaschun H, Eckert K, Schulze-Forster K, Maurer HR, et al. (1995) Prothymosin alpha: a biologically active protein with random coil conformation. Biochemistry, 34: 13211-13218.
- Donizetti A, Liccardo D, Esposito D, Del Gaudio R, Locascio et al. (2008) Differential expression of duplicated genes for prothymosin alpha during zebrafish development. Dev Dyn 237: 1112-1118.

- Xiao Z, Shen J, Feng H, Liu H, Wang Y, et al. (2015) Characterization of two thymosins as immune-related genes in common carp [Cyprinus carpio L.]. Dev Comp Immunol. 50: 29-37.
- 6. Eschenfeldt WH, Berger SL (1986) The human prothymosin alpha gene is polymorphic and induced upon growth stimulation: evidence using a cloned cDNA. Proc Natl Acad Sci U S A. 83: 9403-9407.
- Lee HR, Yoon SY, Kang HB, Park S, Kim KE, et al (2009) Thymosin beta 4 enhances NK cell cytotoxicity mediated by ICAM-1. Immunol Lett. 123: 72-76.
- Emmanouilidou A, Karetsou Z, Tzima E, Kobayashi T, Papamarcaki T, et al. (2013) Knockdown of prothymosin alpha leads to apoptosis and developmental defects in zebrafish embryos. Biochem Cell Biol 91: 325-332.
- 9. Fausto N, Campbell JS, Riehle KJ (2006) Liver regeneration. Hepatology 43: 45-53.
- Haritos AA, Salvin SB, Blacher R, Stein S, Horecker BL (1985) Parathymosin alpha: a peptide from rat tissues with structural homology to prothymosin alpha. Proc Natl Acad Sci U S A, 82: 1050-1053.
- 11. Pai CW, Chen YH (2010) Transgenic expression of prothymosin alpha on zebrafish epidermal cells promotes proliferation and attenuates UVB-induced apoptosis. Transgenic Res 19: 655-665.
- Ha SY, Song DH, Hwang SH, Cho SY, Park CK (2015) Expression of prothymosin alpha predicts early recurrence and poor prognosis of hepatocellular carcinoma. Hepatobiliary Pancreat Dis Int 14: 171-177.
- Lin YT, Lu HP, Chao CC (2015) Oncogenic c-Myc and prothymosinalpha protect hepatocellular carcinoma cells against sorafenib-induced apoptosis. Biochem Pharmacol 93: 110-124.
- Wu CD, Kuo YS, Wu HC, Lin CT (2011) MicroRNA-1 induces apoptosis by targeting prothymosin alpha in nasopharyngeal carcinoma cells. J Biomed Sci 18: 80.
- Wirsching HG, Krishnan S, Florea AM, Frei K, Krayenbuhl N (2014) Thymosin beta 4 gene silencing decreases stemness and invasiveness in glioblastoma. Brain 137: 433-448.
- Ji YI, Lee BY, Kang YJ, Jin-Ok J, Lee SH, et al. (2013) Expression patterns of Thymosin beta4 and cancer stem cell marker CD133 in ovarian cancers. Pathol Oncol Res 19: 237-245.
- 17. Guo Y, Chang H, Li J, Xu XY, Shen L, et al. (2015) Thymosin alpha 1 suppresses proliferation and induces apoptosis in breast cancer cells through PTEN-mediated inhibition of PI3K/Akt/mTOR signaling pathway. Apoptosis 20: 1109-1121.
- Lee D, Kim SS, Seong S, Cho W, Yu H, et al. (2016) Stage IV Wilms Tumor Treated by Korean Medicine, Hyperthermia and Thymosinalpha1: A Case Report. Case Rep Oncol 9: 119-125.
- Sribenja S, Sawanyawisuth K, Kraiklang R, Wongkham C, Vaeteewoottacharn K, et al. (2013) Suppression of thymosin beta10 increases cell migration and metastasis of cholangiocarcinoma. BMC Cancer 13: 430.
- Goldstein AL, Kleinman HK (2015) Advances in the basic and clinical applications of thymosin beta4. Expert Opin Biol Ther 15: 139-145.
- 21. Sosne G, Kim C, Kleinman HK (2015) Thymosin beta4 significantly reduces the signs of dryness in a murine controlled adverse environment model of experimental dry eye. Expert Opin Biol Ther 15: 155-161.
- 22. Pica F, Chimenti MS, Gaziano R, Bue C, Casalinuovo IA et al. (2016) Serum thymosin alpha 1 levels in patients with chronic inflammatory autoimmune diseases. Clin Exp Immunol