



# A New Approach to Drug Therapy: Fc-Fusion Technology

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## Abstract

Fc-fusion proteins have been successfully implemented in the treatment of many diseases. Advances in engineering and design of these therapeutic proteins have helped prolong the drug half-life, which in turn allows for longer dosing intervals (i.e. weekly or bi-weekly administration) and, thus, may improve patient adherence in a real world setting. In this review, we provide a brief summary of half-life extension technologies. Here, we focus on IgG-Fc fusion and the key roles that the Fc fragment plays in both physiology and drug therapy, and the potential to elicit immune responses in humans. This review provides examples of various recombinant Fc-fusion protein drugs, including, etanercept (Enbrel®) for the treatment of various forms of arthritis, aflibercept (Eylea®), for the treatment of neovascular age-related macular degeneration and dulaglutide (Trulicity™) for the treatment of type 2 diabetes. All of these fusion proteins can be administered at least weekly. Overall, Fc-fusion proteins have proven to be a successful alternative to improve pharmacological properties of therapeutic drugs, with more convenient utilization in the real world and with low immunogenic potential.

**Keywords:** Fc-Fusion; Half-life extension; Fc fragment; Fusion protein technology

## Introduction

There is evidence of the potential impact of less frequent dosages on quality of life and adherence to therapies for chronic diseases [1-3]. Therefore, improving the time a drug is available in circulation to exert its action is critical in pharmacotherapy. Recent advances in pharmacological engineering and design revealed the potential of fusion protein molecules to help prolonging intervals between doses. Among them, several fragment crystallizable (Fc)-fusion proteins have been successfully implemented in the treatment of various diseases, including autoimmune and inflammatory, cancer, infectious, cardiovascular and type 2 diabetes mellitus [4-8]. The current article intends to provide a brief review for clinicians on Fc-fusion protein technology.

## Half-life extension technologies

Biologically active proteins and peptides play a significant role in clinical management of human disease. In fact, more than 180 therapeutic proteins and peptides have been approved by the FDA [9]. However, many of these peptides and proteins have less than ideal pharmacokinetic properties, either because they are eliminated by kidney filtration due to their small size and/or due to proteolytic metabolism, thus imposing constant infusions or frequent subcutaneous administration to keep their circulating concentrations within the effective range, which is clinically undesirable [9-16].

In principle, two main strategies are commonly used to reduce the impact of the aforementioned clearance mechanisms on the time-action of a peptide or protein of interest.

The first strategy is depot formulations, which provide an extended drug payout from the site of administration into the systemic circulation, using polymeric and lipid microparticulate systems, in-situ depot-forming systems and implants (refer to reviews [17-19]) that allow proteins and peptides ("the active drug") to be continuously released from the subcutaneous tissue for extended periods of time at sufficiently high concentrations to exert pharmacological activity. In general, therapeutic peptides or proteins in this approach require limited or no molecular engineering, as ideally the depot formulation protects against

metabolism, and renal clearance is overcome by constantly releasing the peptide or protein into the circulation [17-19]. One example of an extended-release formulation of a small peptide is Exenatide extended-release (ER) (Bydureon®, Astra Zeneca, for type 2 diabetes mellitus) for once weekly subcutaneous administration. Exenatide ER is encapsulated in 60 µm diameter microspheres of poly-(D,L-lactide-co-glycolide) and is supplied as a dry powder with an aqueous diluent. Therefore, the injectable suspension is prepared by the patient just before use. This technique allows for a once a week administration, while exenatide by itself has a very short half-life and has to be administered twice a day (Byetta®, Astra Zeneca, for type 2 diabetes mellitus).

The second general approach consists of reducing renal filtration and elimination by increasing the size of the therapeutic peptide. This can be achieved by:

(a) Increasing the hydrodynamic radius of the therapeutic protein by chemical conjugation with a large polymer like methoxy polyethylene glycol (PEG) or, more recently, by recombinant techniques; or

(b) Increasing the molecular weight of the protein of interest to approximately 60 to 70 kilo Dalton (kDa), the renal threshold for renal filtration. This can be achieved by either non-covalent association of the therapeutic peptide to a larger carrier protein, such as circulating albumin or by covalent fusion of a therapeutic peptide to a carrier protein via genetic recombination [9,20-25].

In covalently fused proteins, the pharmacologically active moiety does not dissociate from the fusion partner, but works and interacts with the target as a large, fusion protein. The crystallizable fragment

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of a human immunoglobulin (IgG-Fc), albumin, or transferrin are commonly used as fusion partners. While fusion of a protein of interest with a carrier like albumin provides only half-life extension, fusion with an IgG-Fc can additionally achieve various therapeutic effects depending on the underlying disease [26-31].

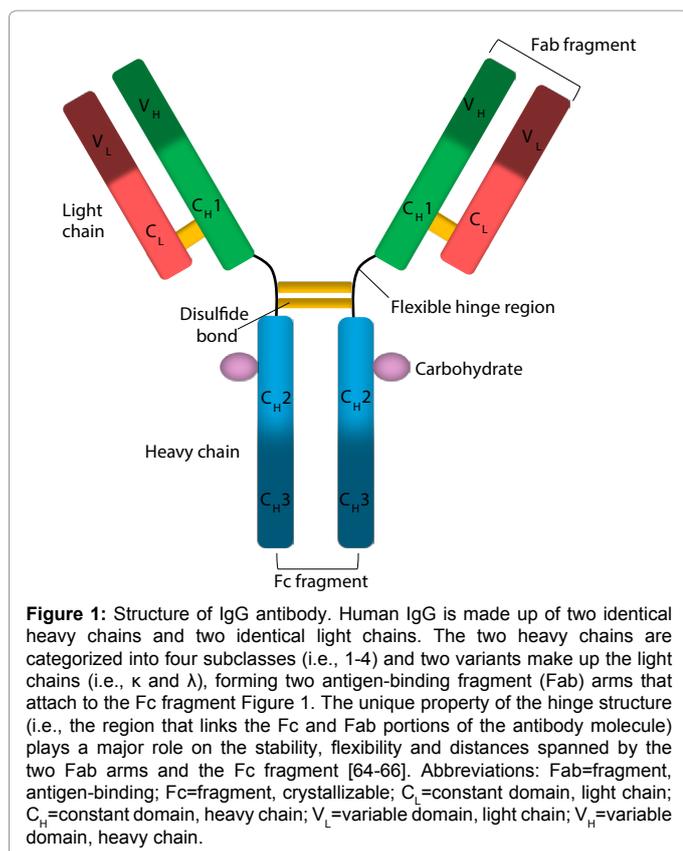
Due to increased circulating half-life over the native peptide or protein, engineering the therapeutic moiety to provide stability against proteolytic metabolism might be necessary.

The effects of depot formulation and increasing the size of a therapeutic are not mutually exclusive, as a component of the depot effect (delayed absorption) is also realized by subcutaneous administration of a large molecule, which reaches the systemic circulation via the lymphatic system [16].

### The Fc fragment in physiology

IgG represents approximately 85% of all immunoglobulins (IgA, D, E, G, M) in the serum and non-mucosal tissue [16] (Figure 1). Together with albumin, IgGs have the longest half-life among plasma proteins [32] (Table 1). Beyond reduced renal elimination due to large size, the IgG serum half-life may additionally be explained by the interaction with the *neonatal Fc receptor (FcRn)* in endothelial cells [33,34]. Binding to endothelial FcRn allows IgG, which is otherwise destined for endocytosis and further lysosomal degradation, to be recycled and released back into circulation [35] (Figure 2).

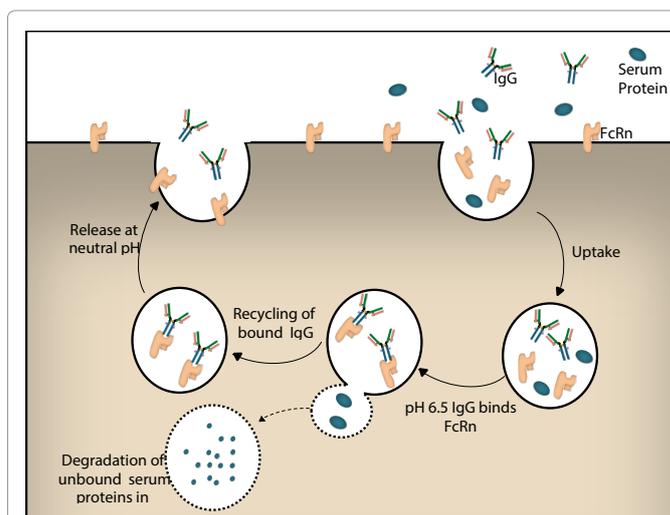
Additionally, IgG plays a critical role in the protective immunity against many pathogens and toxins by interacting with the cell surface *Fcγ receptor (FcγR)* on leukocytes, which mediate downstream effector mechanisms [30,36-38]. This process is known as cytotoxicity and is mediated through antibody-dependent cellular cytotoxicity (ADCC)



Ig Isotypes [16]	IgA	IgD	IgE	IgG	IgM
Serum concentration (mg/mL)	1.5-2.6	0.04	0.0003	9.5-12.5	0.7-1.7
Serum half-life (days)	6	3	2.5	23	5

This table provides a simplified comparison between each Ig isotype

**Table 1:** Characteristics of Ig Isotypes and IgG subclasses.



**Figure 2:** FcRn-mediated IgG recycling. FcRn-mediated IgG recycling plays an important role in extending plasma half-life of native IgG. IgG uptake into the cell, including endothelial cells, occurs via fluid phase pinocytosis with subsequent binding to FcRn in the acidified environment (~pH 6.5) of the endosomal compartment. IgGs are thought to be protected from degradation by recycling to the cell surface where the neutral pH facilitates dissociation from the FcRn and release back into circulation [67].

and complement-dependent cytotoxicity (CDC) and ultimately results in cell lysis, which is the basic protective response against pathogenic agents [5,39-41].

### The Fc fragment in therapeutics

IgG is the most commonly used antibody class for therapy, with a molecular weight of approximately 150 kDa [16]. In Fc proteins, the peptide or protein of interest (a ligand, an extracellular domain of a soluble receptor, etc.) is fused with an Fc fragment primarily to benefit from half-life extension [13]. Taking advantage of size increase and the natural recycling process of IgG, Fc proteins are thought to be protected from degradation by recycling IgG FcRn fused proteins into circulation.

The choice of the IgG isotype is critical in therapeutics: IgG1, IgG2 and IgG4 are often preferred to IgG3 due to their longer half-lives of approximately three weeks [42-45]. Also, IgG subtypes differ on their ability to exert effector functions depending on the binding affinity of IgG to FcγRs: IgG1 and IgG3 have the highest binding affinity and, therefore, are more cytotoxic [46]. In contrast, the binding affinity of IgG4 is approximately 10-fold less than the affinity of IgG1 and IgG3, whereas the binding affinity of IgG2 is undetectable [47,48]. The choice of either one or other IgG isotype as an Fc fusion partner will depend of the desired level of half-life extension and cytotoxic activity purchased for the final compound. Most of the approved therapeutic antibodies are indicated for the treatment of cancer and autoimmune diseases. They belong to the IgG1 subclass because of its potent ability to exert effector functions through *high affinity* binding to the Fc receptors, which is an important advantage for the treatment of those diseases [42,49,50]. Inversely, the IgG2 and IgG4 subclasses are the preferred

backbone of a therapeutic candidate when a lack of cellular activity is desired [40].

### Immunogenicity of protein therapeutics

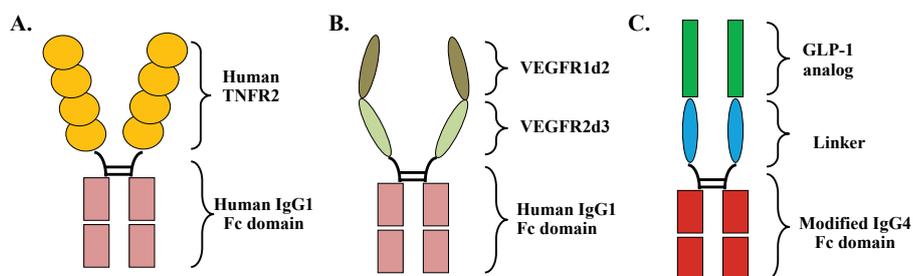
Despite the fact that the protein-based biologics might be similar or identical in sequence to the native human protein, immunogenicity is a major safety concern during the development of any therapeutic protein. Protein-based biologics have the potential to elicit immune responses in humans. Immune responses to therapeutic proteins are characterized by the generation of anti-drug antibodies, which may neutralize its pharmacodynamics effect, affect the pharmacokinetic profile of the biologic, cross-react with its natural counterpart, or have no negative consequences at all [51]. Factors influencing immunogenicity of a therapeutic protein are many, and could range from extrinsic factors, such as aggregates and adjuvant-like contaminants and the patient's immunological status and co-medication, to intrinsic factors, such as the presence of B-cell or T-cell epitopes (the part of an antigen that is recognized by the immune system) on the therapeutic protein itself. Of particular concern are sequences generated at the junction of the active moiety and the carrier protein in fusion proteins, as these are generally novel to the human immune system. The potential for T-cell mediated immunogenicity in humans can be reduced by examining the sequence of fusion proteins for potential T-cell epitopes using *in silico* algorithms; subsequently, lack of immunogenicity needs to be confirmed by clinical data. The presence of anti-drug antibodies is not predictive of anaphylaxis or other hypersensitivity reactions from occurring [52,53].

### Selected IgG-Fc fusion molecules in clinical use

To date, there are ten marketed Fc-fusion protein therapeutics that are approved by the FDA and EMA. We will describe three selected IgG-Fc fusion proteins in more detail below. For more information on other clinically used Fc-fusion therapeutics not described below, see reviews [9-16]. Etanercept (Enbrel<sup>®</sup>) was approved by the FDA in 1998 as a biweekly or weekly subcutaneous injection for the treatment of various forms of arthritis including rheumatoid arthritis, psoriatic arthritis and juvenile idiopathic arthritis, as well as, plaque psoriasis and ankylosing spondylitis. Etanercept is a human tumor necrosis factor (TNF) receptor p75 Fc fusion protein and is the first successful example of using a soluble receptor-Fc fusion protein as a therapeutic drug and the only TNF-blocker commercially available in a fusion protein [54]. Etanercept is the result from the fusion of the 75 kDa soluble extracellular ligand-binding domain of the TNF- $\alpha$  receptor II (TNFR II) to the Fc domain of human IgG1 (Figure 3A). The Fc fragment prolongs the half-life of the

compound, resulting in an extended and more profound biologic effect than its native form [54]. In addition, etanercept has less immunogenic potential compared to anti-TNF monoclonal antibodies [55]. The mechanism of action of etanercept is through the neutralization of both the membrane-bound and soluble forms of TNF- $\alpha$ , preventing TNF-mediated cellular responses and therefore modulating the concentrations of serum inflammatory cytokines, serum matrix metalloproteases, and adhesion molecules. Etanercept is considered safe and effective in both adults and children [55]. Antibodies to the TNF receptor portion or other protein components of the etanercept drug product were detected in approximately 6% of patients with adult rheumatoid arthritis, psoriatic arthritis, active ankylosing spondylitis or severe plaque psoriasis. These antibodies were all non-neutralizing. Treatment effects from juvenile patients with polyarticular idiopathic arthritis were similar to those seen in adult patients with rheumatoid arthritis. Commonly reported adverse events associated with etanercept include injection site reactions and infections, such as upper respiratory tract infection, sinusitis and influenza. Similar to other TNF blockers, there is an increased risk of more serious infections including, but not limited to, tuberculosis and bacterial sepsis [55]. However, TNF- $\alpha$  is a central regulator of inflammation, particularly in normal immune responses to certain pathogens. The increased risk in opportunistic infections observed in patients treated with anti-TNF- $\alpha$  therapy is explained by the immunomodulatory effects related to the blocking of TNF- $\alpha$  and the underlying disease and not related to the Fc component of the fusion protein [56,57].

Another example is the recombinant Fc fusion protein, aflibercept (Eylea<sup>®</sup>; Regeneron, Terrytown, NY, USA), approved in 2011 and administered every 4 to 8 weeks by intravitreal injection for the treatment of neovascular (i.e. wet) age-related macular degeneration (AMD) [58]. AMD is characterized by the growth of abnormal blood vessels in the eye (i.e., choroidal neovascularization) stimulated by angiogenic factors belonging to the VEGF family [59]. Aflibercept is a recombinant fusion protein associating portions of human VEGF receptors extracellular domains fused to the Fc portion of human IgG1 (Figure 3B) [58]. It has been specifically designed for high affinity to its ligands and for minimal interactions to the extracellular matrix, leading to an enhanced pharmacokinetic profile [60]. Aflibercept binds to the angiogenic factors thereby inhibiting the binding and activation of native VEGF receptors. Overall, the incidence of anti-drug antibodies following treatment with aflibercept remained low, ranging from 1% to 3% of patients across the neovascular age-related macular degeneration, retinal vein occlusion and diabetic macular edema studies and there were no differences in efficacy or safety between patients with or



**Figure 3:** Simplified diagram of the molecular structures of etanercept, aflibercept and dulaglutide. A. The molecular structure of etanercept is the result from the fusion of the extracellular ligand-binding domain of TNFR II to the Fc domain of human IgG1 [54] (data from Wallis 2008 [68]). B. The molecular structure of aflibercept consists of portions of human VEGF receptors extracellular domains that link to the Fc domain of human IgG1 [58] (data from Platania et al. 2015 [69]). C. The molecular structure of dulaglutide consists of a GLP-1 analog fused to the Fc domain of a modified IgG4 [4] (data from Kuritzky et al. 2014 [68]). Black bars represent disulfide bridges. Abbreviations: VEGFR1d2=domain 2 of vascular endothelial growth factor receptor VEGFR1; VEGFR2d3=domain 3 of vascular endothelial growth factor receptor VEGFR2

without immunoreactivity. Other common adverse reactions reported in patients treated with aflibercept included conjunctival hemorrhage, eye pain, cataract, vitreous detachment, vitreous floaters, and increased intraocular pressure [58].

Finally, glucagon-like peptide-1 receptor agonist, dulaglutide (Trulicity™, Eli Lilly and Company, Indianapolis, IN), was approved in 2014 as a once weekly subcutaneous injection to improve glycemic control in adults with type 2 diabetes [61]. The molecule is a recombinant fusion of glucagon-like peptide-1 (GLP-1) analog consisting of two identical disulfide-linked chains covalently linked to a modified IgG4 heavy chain-Fc fragment, which are fused together by a small peptide linker (Figure 3C) [4]. The GLP-1 analog portion of dulaglutide contains 3 mutations compared to the native GLP-1 peptide. The alanine change at position 8 to glycine confers resistance to inactivation by the enzyme dipeptidyl peptidase-IV and the glycine change at position 22 to glutamic acid increases solubility of dulaglutide. The arginine change at position 36 at the junction between the GLP-1 moiety and the linker sequence to glycine de-immunizes the fusion protein. The IgG4 Fc region of dulaglutide was optimized to reduce interaction with Fcγ receptor I and to eliminate half-antibody formation [4]. In addition, the C-terminal lysine was removed from the IgG-Fc. In the clinical program, 1.6% of dulaglutide-treated patients developed anti-drug antibodies with no differences in efficacy or safety between patients with or without antibodies. Similar to other drugs in the GLP-1 receptor agonist class, common adverse events related to dulaglutide are gastrointestinal in nature, including nausea, vomiting and diarrhea. These effects are generally mild to moderate in severity and transient [61].

### IgG-Fc fusion therapeutics in a real world setting

The main reason why this technology was developed was to enable therapy with short-acting proteins. The extension of a molecule's half-life offers additional benefits for patients. IgG-Fc fusion therapeutics, by extending the half-life of the drug, may provide clinically relevant advantages regarding patient preference and adherence. In general, patients suffering from chronic diseases are more likely to adhere to medications that require less frequent, intermittent dosing regimens. Studies have demonstrated an inverse relationship between medication adherence and dosing frequency in patients with chronic diseases. Up to a 19% decrease in taking adherence, up to a 23% decrease in regimen adherence, and up to a 54% decrease in timing adherence in chronically ill patients whose treatment required them to take multiple daily doses compared to patients who are on once-daily regimens [62]. Similarly, researchers found that adherence was up to 12% higher in patients on a once-weekly dosing regimen compared to patients on more frequently dosed agents for the same conditions and up to 96% of patients preferred an intermittent dosing regimen [63]. Therefore, a more favorable dosing regimen may enhance patient adherence and ultimately the patient's health.

### Concluding statements

In conclusion, Fc-fusion proteins have proven to be a successful alternative to improve pharmacological properties of therapeutic drugs with low immunogenic potential. Overall, these drugs have an acceptable safety profile with adverse events that are generally specific to the mechanisms of action of each drug class. By utilizing and manipulating Fc-Fc receptor interaction, developers are able to improve pharmacokinetics, significantly increase serum half-life and selectively enhance or disable effector functions, all while maintaining drug efficacy. Combined with an increased knowledge

of the complexity of antibody disposition, Fc-fusion has become an ideal platform for increasing half-life, thus reducing dosing frequency, reducing immunogenicity by providing optimal effects while avoiding undesired complications. Overall, advances in Fc-fusion technology have led to greater flexibility when developing therapeutics by selectively addressing the needs of various disease settings.

### References

1. Brundisini F, Vanstone M, Hulan D, DeJean D, Giacomini M (2015) Type 2 diabetes patients' and providers' differing perspectives on medication non-adherence: A qualitative meta-synthesis. BMC Health Serv Res 15: 516.
2. Richter A, Anton SF, Koch P, Dennett SL (2003) The impact of reducing dose frequency on health outcomes. Clin Ther 25: 2307-2335.
3. Saini SD, Schoenfeld P, Kaulback K, Dubinsky MC (2009) Effect of medication dosing frequency on adherence in chronic diseases. Am J Manag Care 15: e22-33.
4. Glaesner W, Vick AM, Millican R, Ellis B, Tschang SH, et al. (2010) Engineering and characterization of the long-acting glucagon-like peptide-1 analogue LY2189265, an Fc fusion protein. Diabetes Metab Res Rev 26: 287-296.
5. Chan AC, Carter PJ (2010) Therapeutic antibodies for autoimmunity and inflammation. Nat Rev Immunol 10: 301-316.
6. Weiner LM, Surana R, Wang S (2010) Monoclonal antibodies: Versatile platforms for cancer immunotherapy. Nat Rev Immunol 10: 317-327.
7. Reichert JM (2010) Antibodies to watch in 2010. MAbs 2: 84-100.
8. Reichert JM (2012) Which are the antibodies to watch in 2012? MAbs 4: 1-3.
9. Strohl WR (2015) Fusion proteins for half-life extension of biologics as a strategy to make Biobetters. BioDrugs 29: 215-239.
10. Baldo BA (2015) Chimeric fusion proteins used for therapy: Indications, mechanisms and safety. Drug Saf 38: 455-479.
11. Liu L (2015) Antibody glycosylation and its impact on the pharmacokinetics and pharmacodynamics of monoclonal antibodies and Fc-fusion proteins. J Pharm Sci 104: 1866-1884.
12. Czajkowsky DM, Hu J, Shao Z, Pleass RJ (2012) Fc-fusion proteins: New developments and future perspectives. EMBO Mol Med 4: 1015-1028.
13. Beck A, Reichert JM (2011) Therapeutic Fc-fusion proteins and peptides as successful alternatives to antibodies. MAbs 3: 415-416.
14. Huang C (2009) Receptor-Fc fusion therapeutics, traps and MIMETIBODY technology. Curr Opin Biotechnol 20: 692-699.
15. Schmidt SR (2009) Fusion-proteins as biopharmaceuticals--applications and challenges. Curr Opin Drug Discov Devel 12: 284-295.
16. Lobo ED, Hansen RJ, Balthasar JP (2004) Antibody pharmacokinetics and pharmacodynamics. J Pharm Sci 93: 2645-2668.
17. Pagels RF, Prud'homme RK (2015) Polymeric nanoparticles and microparticles for the delivery of peptides, biologics, and soluble therapeutics. J Control Release 219: 519-535.
18. Jaspert S, Piel G, Delattre L, Evrard B (2005) Solid lipid microparticles: Formulation, preparation, characterisation, drug release and applications. Expert Opin Drug Deliv 2: 75-87.
19. Shen J, Burgess DJ (2012) Accelerated *in vitro* release testing methods for extended-release parenteral dosage forms. J Pharm Pharmacol 64: 986-996.
20. Podust VN, Balan S, Sim BC, Coyle MP, Ernst U, et al. (2016) Extension of *in vivo* half-life of biologically active molecules by XTEN protein polymers. J Control Release 240: 52-66.
21. Hansen BF, Kurtzhals P, Jensen AB, Dejgaard A, Russell-Jones D (2011) Insulin X10 revisited: A super-mitogenic insulin analogue. Diabetologia 54: 2226-2231.
22. Beals JM, Cutler GBJ, Vick A, Koester A, Li S, et al. (2012) P-896: LY2605541: Leveraging hydrodynamic size to develop a novel basal insulin. Diabetes Care 61: A228.
23. Kurtzhals P (2007) Pharmacology of insulin detemir. Endocrinol Metab Clin North Am 36: 14-20.

24. Nauck MA, Petrie JR, Sesti G, Mannucci E, Courrèges JP, et al. (2016) A Phase 2, randomized, dose-finding study of the novel once-weekly human GLP-1 analog, semaglutide, compared with placebo and open-label liraglutide in patients with type 2 diabetes. *Diabetes Care* 39: 231-241.
25. Sockolosky JT, Szoka FC (2015) The neonatal Fc receptor, FcRn, as a target for drug delivery and therapy. *Adv Drug Deliv Rev* 91: 109-124.
26. Dumont JA, Low SC, Peters RT, Bitonti AJ (2006) Monomeric Fc fusions: Impact on pharmacokinetics and biological activity of protein therapeutics. *Bio Drugs* 20: 151-160.
27. Kratz F (2008) Albumin as a drug carrier: design of prodrugs, drug conjugates and nanoparticles. *J Control Release* 132: 171-183.
28. Widera A, Beloussow K, Kim KJ, Crandall ED, Shen WC (2003) Phenotype-dependent synthesis of transferrin receptor in rat alveolar epithelial cell monolayers. *Cell Tissue Res* 312: 313-318.
29. Widera A, Bai Y, Shen WC (2004) The transepithelial transport of a G-CSF-transferrin conjugate in Caco-2 cells and its myelopoietic effect in BDF1 mice. *Pharm Res* 21: 278-284.
30. Hogarth PM, Pietersz GA (2012) Fc receptor-targeted therapies for the treatment of inflammation, cancer and beyond. *Nat Rev Drug Discov* 11: 311-331.
31. Liang Y, Qiu H, Glinka Y, Lazarus AH, Ni H, et al. (2011) Immunity against a therapeutic xenoprotein/Fc construct delivered by gene transfer is reduced through binding to the inhibitory receptor FcγRIIb. *J Gene Med* 13: 470-477.
32. Frazer JK, Capra JD (1999) Immunoglobulins: Structure and function. In: *Fundamental immunology*. (4th Edition) ed. Philadelphia: Lippincott-Raven.
33. Challa DK, Velmurugan R, Ober RJ, Sally Ward E (2014) FcRn: from molecular interactions to regulation of IgG pharmacokinetics and functions. *Curr Top Microbiol Immunol* 382: 249-272.
34. Ward ES, Ober RJ (2009) Chapter 4: Multitasking by exploitation of intracellular transport functions the many faces of FcRn. *Adv Immunol* 103: 77-115.
35. Ober RJ, Martinez C, Vaccaro C, Zhou J, Ward ES (2004) Visualizing the site and dynamics of IgG salvage by the MHC class I-related receptor, FcRn. *J Immunol* 172: 2021-2029.
36. Roopenian DC, Akilesh S (2007) FcRn: The neonatal Fc receptor comes of age. *Nat Rev Immunol* 7: 715-725.
37. Guilliams M, Bruhns P, Saeys Y, Hammad H, Lambrecht BN (2014) The function of Fcγ receptors in dendritic cells and macrophages. *Nat Rev Immunol* 14: 94-108.
38. Peipp M, Beyer T, Dechant M, Valerius T (2007) Molecular engineering III: Fc Engineering, in *Handbook of Therapeutic Antibodies* (ed S. Dübel), Wiley-VCH Verlag GmbH, Weinheim, Germany.
39. Raju TS (2008) Terminal sugars of Fc glycans influence antibody effector functions of IgGs. *Curr Opin Immunol* 20: 471-478.
40. Jiang XR, Song A, Bergelson S, Arroll T, Parekh B, et al. (2011) Advances in the assessment and control of the effector functions of therapeutic antibodies. *Nat Rev Drug Discov* 10: 101-111.
41. Walport MJ (2001) Complement. First of two parts. *N Engl J Med* 344: 1058-1066.
42. Salfeld JG (2007) Isotype selection in antibody engineering. *Nat Biotechnol* 25: 1369-1372.
43. Reichert JM, Rosensweig CJ, Faden LB, CDM (2005) Monoclonal antibody successes in the clinic. *Nat Biotechnol* 23: 1073-1078.
44. Jefferis R, Lefranc M (2009) Human immunoglobulin allotypes. *MAbs* 1: 1-7.
45. Carter PJ (2006) Potent antibody therapeutics by design. *Nat Rev Immunol* 6: 343-357.
46. Bruhns P, Iannascoli B, England P, Mancardi DA, Fernandez N, et al. (2009) Specificity and affinity of human Fc-gamma receptors and their polymorphic variants for human IgG subclasses. *Blood* 113: 3716-3725.
47. Canfield SM, Morrison SL (1991) The binding affinity of human IgG for its high affinity Fc receptor is determined by multiple amino acids in the CH2 domain and is modulated by the hinge region. *J Exp Med* 173: 1483-1491.
48. Anderson CL, Abraham GN (1980) Characterization of the Fc receptor for IgG on a human macrophage cell line, U937. *J Immunol* 125: 2735-2741.
49. Reichert JM (2008) Monoclonal antibodies as innovative therapeutics. *Curr Pharm Biotechnol* 9: 423-430.
50. Jefferis R (2007) Antibody therapeutics: Isotype and glycoform selection. *Expert Opin Biol Ther* 7: 1401-1413.
51. Shankar G, Shores E, Wagner C, Mire-Sluis A (2006) Scientific and regulatory considerations on the immunogenicity of biologics. *Trends Biotechnol* 24: 274-280.
52. De Groot AS, Bosma A, Chinai N, Frost J, Jesdale BM, et al. (2001) From genome to vaccine: *In silico* predictions, *ex vivo* verification. *Vaccine* 19: 4385-4395.
53. Guidance for industry immunogenicity assessment for therapeutic protein products [Internet] August 2014.
54. Marotte H, Cimaz R (2014) Etanercept - TNF receptor and IgG1 Fc fusion protein: Is it different from other TNF blockers? *Expert Opin Biol Ther* 14: 569-572.
55. Enbrel product information. Rev March 2015. Amgen Inc. TO, CA; Immunex, Seattle, WA.
56. Ali T, Kaitha S, Mahmood S, Ftesi A, Stone J, Bronze MS (2013) Clinical use of anti-TNF therapy and increased risk of infections. *Drug Healthc Patient Saf* 5: 79-99.
57. Murdaca G, Spanò F, Contatore M, Guastalla A, Penza E, Magnani O, et al. (2015) Infection risk associated with anti-TNF-α agents: A review. *Expert Opin Drug Saf* 14: 571-582.
58. Eylea product information. Rev March 2015. Regeneron T, NY; Sanofi Aventis, Paris, Île-de-France.
59. Kwak N, Okamoto N, Wood JM, Campochiaro PA (2000) VEGF is major stimulator in model of choroidal neovascularization. *Invest Ophthalmol Vis Sci* 41: 3158-3164.
60. Holash J, Davis S, Papadopoulos N, Croll SD, Ho L, et al. (2002) VEGFTrap: A VEGF blocker with potent antitumor effects. *Proc Natl Acad Sci USA* 99: 11393-11398.
61. Trulicity [product information]. Rev March 2015. Eli Lilly and Company I, IN.
62. Coleman CI, Limone B, Sobieraj DM, Lee S, Roberts MS, et al. (2012) Dosing frequency and medication adherence in chronic disease. *J Manag Care Pharm* 18: 527-539.
63. Kruk ME, Schwalbe N (2006) The relation between intermittent dosing and adherence: Preliminary insights. *Clin Ther* 28: 1989-1995.
64. Liu H, May K (2012) Disulfide bond structures of IgG molecules: Structural variations, chemical modifications and possible impacts to stability and biological function. *MAbs* 4: 17-23.
65. Roux KH, Strelets L, Michaelsen TE (1997) Flexibility of human IgG subclasses. *J Immunol* 159: 3372-3382.
66. Tian X, Langkilde AE, Thorolfsson M, Rasmussen HB, Vestergaard B (2014) Small-angle x-ray scattering screening complements conventional biophysical analysis: comparative structural and biophysical analysis of monoclonal antibodies IgG1, IgG2 and IgG4. *J Pharm Sci* 103: 1701-1710.
67. Raghavan M, Bonagura VR, Morrison SL, Bjorkman PJ (1995) Analysis of the pH dependence of the neonatal Fc receptor/immunoglobulin G interaction using antibody and receptor variants. *Biochemistry* 34: 14649-14657.
68. Wallis RS. (2008) Tumour necrosis factor antagonists: structure, function, and tuberculosis risks. *Lancet Infect Dis* 8: 601-611.
69. Platania CB, Di Paola L, Leggio GM, Romano GL, Drago F, Salomone S, Bucolo C (2015) Molecular features of interaction between VEGFA and anti-angiogenic drugs used in retinal diseases: a computational approach. *Front Pharmacol* 6: 248.
70. Kuritzky L, Umpierrez G, Ekoé JM, Mancillas-Adame L, Fernández Landó L. (2014) Safety and Efficacy of Dulaglutide, a Once Weekly GLP-1 Receptor Agonist, for the Management of Type 2 Diabetes. *Postgraduate Med* 126: 60-72.

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