A Rare Intronic Variation of Presenilin-1 (rs201992645) is Associated with Alzheimer’s Disease and Down Syndrome Birth

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Rec Date: Jun 15, 2014, Acc Date: Sep 30, 2014, Pub Date: Oct 08, 2014

Abstract

**Background and objective:** Presenilin-1 (PSEN-1) gene is a potent candidate that relates Alzheimer’s disease (AD) to Down syndrome (DS). Genetic variation of PSEN-1 could be a risk factor that predisposes individual for both AD and DS and may contribute the common etiology of both the disorders.

**Methods:** We sequenced exon 8 of PSEN-1 with flanking introns in 136 DS patients with their parents, 96 AD patients, 173 age-matched controls. Results were analysed in-silico to anticipate the damaging effect at molecular level.

**Results:** A rare polymorphism rs201992645 was identified within intron 8 and in silico analysis revealed the variation as ‘potentially damaging’ at the transcript splicing level. The genotypic frequencies of mutant heterozygotes were 0.031, 0.029 and 0.029 for AD, DS and mother of DS respectively.

**Conclusions:** We have suggested that this variation may cause AD manifestation in mothers of DS patients and is the potential marker for predisposition testing of both disorders.

**Keywords:** Presenilin-1; Down syndrome; Alzheimer disease; Chromosomal segregation

Introduction

Alzheimer’s disease (AD) and Down syndrome (DS) probably share a common genetic factor, as suggested by their frequent co-occurrence. AD like neuropathology such as β-amyloid plaque and neurofibrillary tangles are seen in virtually all DS patients at about 35 to 40 years of age [1]. Families with AD patient exhibit frequent birth occurrence of DS child; vis-a-vis, higher incidence of AD has been reported among the relatives of DS child; vis-a-vis, higher incidence of AD has been reported [4]. The PSEN-1 encodes a protein component of gamma-secretase complex which is involved in the processing of amyloid precursor protein (APP). PSEN-1 protein is involved in many cardinal mechanisms of several molecular pathways which when impaired lead to the manifestation of AD. The pathways include APP processing [5], notch signaling [6], neuronal apoptosis [7], mitochondrial dysfunction [8], calcium homeostasis and synaptic function [9]. This protein is also localized at centromeres, nuclear envelope of dividing cells, in kinetochores at interphase and is involved in the process of faithful chromosome segregation [10]. Mutation in PSEN-1 leads to chromosomal instability and trisomy 21 mosaicism among AD patients [11]. The study of Petersen et al. [12], revealed an association between the intronic polymorphism of PSEN-1 and meiotic nondisjunction of chromosome 21(Ch21) among the women bearing DS children. In present study we explored the sequence of PSEN-1 exon 8 to find out any mutation or polymorphism that might explain the role of this gene in common etiology and co-occurrence of AD and DS in the Indian Bengali population. We chose the exon 8 and its flanking intronic sequence, as this region is known to carry mutational hotspots and many mutations within this region exhibit association with AD [11].

Materials and Methods

**Study subjects**

Blood samples were collected from 96 probable Alzheimer’s patients (mean age 62.26 ± 11.93 years with 70.48% male and 29.52% female subjects), 173 age matched healthy controls and 136 DS family trios (DS and parents) after obtaining their prior consent. Ethnic and...
demographic similarities were maintained between cases and controls through careful recruitment from hospitals within the same geographical localities. All participants were Bengali-speaking.

Dementia status of patients were diagnosed using DSM -IV (Diagnostic and Statistical Manual of Mental Disorders, Fourth Edition) criteria and NINCDS-ADRDA (National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer’s Disease and Related Disorders Association) criteria, both of which are generally used for diagnosis of Alzheimer’s disease by physicians. Severity of Dementia was measured by the MMSE (Mini Mental State Examination) score. Recruited controls were free from any recognizable neurodegenerative disorders.

Mutation analysis
Genomic DNA was isolated from blood using a Qiagen QIAamp DNA Blood Midi Kit (Catalogue No. 51185). A Polymerase Chain Reaction (PCR) was performed using oligonucleotide primer that has been described elsewhere [13]. The PCR amplicons were sequenced through ABI PRISM 3700 DNA Analyzer platform. The obtained sequence was thoroughly compared against the published genomic sequence (NCBI accession number NG_007386.2) of PSEN-1. The genotyping of APOE was done in all subjects and controls by direct DNA sequencing. The identified variation was submitted to NCBI (snp-sub@ncbi.nlm.nih.gov) to get its validity and a submitter SNP (ss) number (NCBI ss515119316) and reference SNP (rs) ID (rs201992645) were achieved in the process.

Functional prediction of detected variation
The online software ‘Human Splicing Finder’ Version 2.4.1 (http://www.umd.be/HSF/), ‘SpliceAid’ (http://www.introni.it/splicing.html) and ‘mutation t@sting’ (http://www.mutationtaster.org/) were used to predict the probable damaging effects of the mutant allele at splicing level and protein expression levels.

Results and Discussion
We found a rare polymorphism rs201992645 [(NG_007386.2:g.66696T>A or NM_000021.3:c.868+37T>A) ss515119316] within the intron 8 of PSEN-1 gene (Figure 1) at the 73664874th nucleotide position on chromosome 14 (GRCh37.p5 Assembly). The change involves nucleotide transition T>A. The mutant ‘A’ allele exists only as heterozygous condition (T/A) with genotype and allele frequencies of 0.031 and 0.02 respectively for AD cases and 0.029 and 0.01 respectively for DS. Within the parents of DS, the mutant allele was recorded only among the mother with genotypic and allelic frequencies of 0.029 and 0.01 respectively. But the father of the DS and controls were wild type homozygote (T/T). Statistical significance of the result was obtained by Fisher’s exact test (mutated AD + DS cases/total AD + DS cases vs. mutated control individual/total control individual = 7/232 vs. 0/173 = p value 0.02). The genotypes were not in a H-W equilibrium, as we did not find any AA mutant homozygous among the participating families.

The women who bear mutant allele exhibited lower age at conception of DS foetus from the control [(mean ± SD) 22.5 ± 1.29 vs. 32.02 ± 3.64; p value 0.045], as well as from the mothers of DS (32.42 ± 3.53; p value 0.037) who did not carry the mutant allele. This variation was not associated with APOE genotypic variations.

To anticipate the probable damaging effect of the detected intronic mutation on PSEN-1 expression and functions, we analyzed the mutation by in silico method, using three bioinformatics softwares.

Prediction of “human splicing finder” program
The outcome of Human Splicing Finder program shows that the said nucleotide change (NG_007386.2:g.66696T>A or NM_000021.3:c.868+37T>A) induces an alternation in acceptor splice site motif “tagtaatcagtgta” to a new splice site “tagtaatcagagTA” with +69% variations. Potentially new branch point has been generated (CV for wild type and mutant are 40.01and 69.64 respectively). The mutant allele also creates new binding site for essential sequence specific splicing factor protein SF2/ASF. Both RESCUE ESE hexamers matrix and intron-identity elements (EIEs) matrix show that a new enhancer motif is generated due to this SNP. Analysis of silencer motifs following the method by Sironi et al. [14] showed that motif-1 is changed to a new site in mutant sequence. New site also formed for binding of splicing regulatory SR protein 9G8 as well as heterogeneous nuclear ribonucleoprotein hnRNP A1. (Supplementary Table SI).
Prediction of “SpliceAid” software program

This database revealed that the said mutation rs201992645 generates (“gained”) an additional SC35 protein binding site. Due to this transition mutation (T>A) in DNA sequence the corresponding RNA sequence “AGUAG” is changed to “UGUAG” which is the binding site for this protein (Figure 2). The SC35 is a 35kDa mammalian splicing factor which plays an important role in an alternative splicing of genes involved in both AD and DS [15]. This variation may help in splicing of intron-8 of PSEN-1gene.

![Figure 2: The outcome of the online software 'SpliceAid' (http://www.introni.it/splicing.html): The Nucleotide variation rs201992645 [(NG_007386.2:g.66696T>A or NM_000021.3:c.868+37T>A) ss515119316] introduces a new binding site for a potent splicing factor “SC35”. In wild type pre-mRNA sequence of PSEN-1 intron-8, the nucleotide stretch “UGUAG” is changed to “AGUAG” in mutant, which is the binding site for this SC35 protein involved in intron slicing.]

Prediction of “mutation t@sting” program

The outcome of the program “mutation t@sting” suggested some alteration in the protein structure of PSEN-1 due to a splice site change. According to its prediction, this variation activates an additional splice site i.e. ’gained’ at the genomic DNA position 61720 (score 0.71). Moreover, the existing splice site also gets stronger, i.e. “increased” at the position 61715 (score 0.78 for wild type vs. 0.97 for mutant output by the software). Potential cytoplasmic and transmembrane helical domains of PSEN-1 protein as well as the PAL motif required for normal active site conformation were predicted to get lost due to this variation. This variation may affect some important protein features as newly generated splice sites may lead to whole exon skipping. “mutation t@sting” also suggested that the variation may alter certain amino acid sequences, resulting in impaired gamma-secretase activity, endoproteolytic cleavage, cleavage by caspase and progression of apoptosis. Both PKA and PKC mediated phosphorylation, interaction with CTNNB1, CTNN2D and MTC1, production of amyloid-beta, processing of APP, NOTCH1 and CDH2 and disassembly of N-cadherin/PSEN-1 complex at the cell surface. Summary of the results is presented in Table 1.

Conclusion

The significance of our study lies in finding that the polymorphism rs201992645 (ss515119316) is strongly associated with both the AD and DS and the PSEN-1 gene is a prospective ‘molecular link’ between these two disorders. This polymorphism has not been described yet by any workers as simultaneous risk for NDJ of chromosome 21 and AD. In this regard our result is absolutely noble. We have anticipated through in silico approach, the prospective damaging effects of this nucleotide change in PSEN-1 on RNA transcript processing (Figure 2) as well as on protein conformation which might have some deleterious effect on various interacting subcellular pathways (Table 1; Supplementary Table 1). Considering the outcome of bioinformatics analyses, we can interpret that the said nucleotide change imparts pleiotropic effects on various interactive molecular pathways that may include both the signals for proper chromosome segregation and processing of amyloid precursor protein.

Presence of mutant allele only in the mothers of DS and DS child (not among the fathers of DS) is consistent with the notion that mothers of DS and DS patients usually susceptible to early onset of AD due to variation of PSEN-1. The mutation carrying mothers of DS had a much younger age of conception [22.5 ± 1.29 years (mean ± SD)] of the trisomy 21 fetus. This fact suggests that the nucleotide change in PSEN-1 predisposes the women simultaneously for Ch21 nondisjunction and AD irrespective of maternal age of conception. The said variation imparts maternal age independent risk of nondisjunction according to the ‘DS risk model’ proposed in previous studies [16,17]. Further longitudinal follow up study is needed to know whether mutant allele bearing DS develop AD phenotype later at their more advanced age. Studies are also needed to reveal whether this polymorphism is endemic for this Indian population or whether it affects ethnically different populations as well. Moreover, functional validation through experimentation is needed for anticipated in silico results. To conclude it can be said that the polymorphism rs201992645 (ss515119316) (T>A) of PSEN-1 is a potential molecular marker for preconceptional genetic testing for screening DS pregnancy as well as to evaluate the risk of AD onset, particularly among the women.

Acknowledgement

We would like to thank all the AD patients, DS patients and their parents as well as the control individuals participating in the study.

Financial Support

This work was supported by the <University Grants Commissions (UGC), New Delhi, India> under Grant <F. No. 36-299/2008 (SR)>.
The mutation might alter the splicing pattern of the transcript, in downstream it alters the protein domain structure due to change in amino acid sequence as the whole exon may be skipped by splicing.

<table>
<thead>
<tr>
<th>Inferred Change in Presenilin-1 (PSEN-1) expression and its product</th>
<th>The interactive genes</th>
<th>Sub-cellular processes/Molecular pathways</th>
<th>Probable Damaging effect</th>
</tr>
</thead>
<tbody>
<tr>
<td>The mutation might alter the splicing pattern of the transcript, in downstream it alters the protein domain structure due to change in amino acid sequence as the whole exon may be skipped by splicing.</td>
<td>APP (Amyloid Precursor Protein)</td>
<td>Involved in the amyloid plaque production in brain.</td>
<td>Gamma secretase activity of PSEN-1 may be changed, altering amyloid beta production.</td>
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<td>Caspases</td>
<td>Involved in the proteolytic processing of PSEN-1.</td>
<td>Metabolism of PSEN-1 via proteolytic processing may be altered.</td>
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<td></td>
<td>PKC (Phospho kinase C)</td>
<td>Inhibits the caspase cleavage of PSEN-1 by phosphorylating at amino acid residue 346.</td>
<td>Proteolytic processing of PSEN-1 and progression of apoptosis may be altered.</td>
</tr>
<tr>
<td></td>
<td>CTNNB1 (Beta catenin)</td>
<td>Involved in regulation of apoptosis via cell signaling.</td>
<td>Neuronal apoptosis may be modulated.</td>
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<td></td>
<td>CTNNB2 (Delta catenin)</td>
<td>Critical protein for maintenance of neuronal structure and function.</td>
<td>Neuronal apoptosis may be modulated.</td>
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<td>CDH2 (Cadherin)</td>
<td>The cadherin-catenin-presenilin-1 complex triggers synaptic loss and make neurones vulnerable to apoptosis.</td>
<td>Neuronal apoptosis may be modulated.</td>
</tr>
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<td></td>
<td>MTCH1 (Mitochondrial carrier homolog 1)</td>
<td>Its over-expression is responsible for mitochondrial depolarization and apoptosis.</td>
<td>Apoptosis may be altered.</td>
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<td></td>
<td>NOTCH-1</td>
<td>PSEN-1-Notch1 interaction is necessary for neurogenesis.</td>
<td>Neurogenesis may be impaired.</td>
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Table 1: Summary of damaging effect of mutation rs201992645 (NG_007386.2:g.66696T>A or NM_000021.3:c.868+37T>A) (ss515119316) of PSEN-1 gene on subcellular processes as inferred from In Silico analysis using the program mutation t@sting.

References