

An Essential Oil Blend Prevents *Pseudomonas aeruginosa* PA01 from Forming Biofilms

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Abstract

Essential oils are natural ingredients exhibiting antimicrobial and antioxidant effects. The properties of a mix of *Allium sativum*, *Cinnamomum cassia* and *Mentha piperita* essential oils are established in this study on a biofilm forming pathogen, *Pseudomonas aeruginosa* PA01.

Minimal inhibitory and bactericidal concentrations were first determined on planktonic cells for the pure essential oils, then for the blend. A crystal violet assay provided the active concentration of a blend for bacterial biofilm. *Pseudomonas aeruginosa* PA01 planktonic and sessile cells were affected by a concentration of an essential oil blend of 0.125%. The active concentration of 0.1% was confirmed using a protocol developed on glass slides, for its activity on adhesion and biofilm formation. The experiment was then repeated in flow cells to monitor the structural biofilm development using confocal laser scanning microscopy. *Pseudomonas aeruginosa* PA01 adhesion was severely inhibited by the blend concentration at 0.5%. Its biofilm was strongly affected from a 0.1% blend concentration and was totally eradicated when the concentration reached 1%.

The essential oil blend has an enlarged spectrum of activity and is effective on early biofilm development stages, with a concentration that does not involve bactericidal activities. These results are predictive for potential use of this blend as a cleanser.

Keywords: Essential oils; Adhesion; Biofilm; *Pseudomonas aeruginosa*; Structural analyses; Confocal laser scanning microscopy

Introduction

Biofilms are complex structures involving different species of microorganisms, embedded in an exopolysaccharidic matrix [1]. Bacteria are able to communicate due to a quorum sensing system-nutrient, DNA and proteins being exchanged between the different protagonists [2]. Due to this structure, biofilms are resistant to disinfectants or detergents and lead to complications in many fields, such as the food industry or medical surgery [1,3]. Well known *Pseudomonas aeruginosa* is a pathogen able to contaminate medical devices like dentures, catheters or prosthesis, but also corporal tissue, leading to severe infections and exhibit multi-drug resistance [4,5].

Although surface cleaning is strongly controlled by chemical or physical methods (mechanical scrubbing, brushing, scraping or high pressure spraying for example), it is rarely effective for total eradication of biofilms [2]. In fact, biofilms are able to adapt themselves to chemical products, leading to high resistance and new solutions have to be investigated. Enzymes, phages, interspecies competition or natural plant extracts - like volatile compounds - have been studied and offer alternatives to obtain new anti-biofilm products [6-9].

Terrestrial plants are sessile organisms, a status which forces them to adapt to biotic or abiotic stresses through the production of secondary metabolites. Biofilm development on their surface is one of the biotic stresses they have to deal with, leading to the excretion of essential oils

[10]. Essential oils are lipophilic and complex chemical mixes with high terpenic and phenolic contents. Antibacterial activities of essential oils and their components have been widely studied over the past few years [11-13]. Due to their lipophilic character, they are able to pass through cell membranes, disrupt the different phospholipids, polysaccharides and fatty acid layers, and finally permeabilize cells causing a loss of integrity. Clove, thyme or oregano essential oils among others are also able to affect bacterial biofilms specifically by interfering with quorum sensing, inhibiting the peptidoglycan synthesis or reducing cell adherence [14].

Allium sativum essential oil has proved its effectiveness against *P. aeruginosa* planktonic cells, due to the presence of diallyl sulphides, compounds containing sulphuric bounds. In their studies, Sandasi and Valeriano also showed the effectiveness of *Mentha piperita* essential oils against biofilms [2,3]. Kim et al. reported the particular ability of cinnamaldehyde, the main compound of *Cinnamomum cassia* essential oil to down regulate quorum sensing in *P. aeruginosa* biofilms [15]. As the effectiveness of these three essential oils has been reported in different publications, we associated them in order to develop products that are active during early and late biofilm formation stages. As bacteria within a biofilm are 10 to 1000 times more resistant than planktonic ones [16], we also combined essential oils in order to enlarge the antimicrobial activity spectrum. Furthermore, a biofilms' morphology is often adaptable against external growth conditions, linked to differences in susceptibility. To prove specific anti-biofilm activity, we tested the blend against both planktonic and sessile bacteria.

Materials and Methods

Plant extracts

The essential oils (EOs) used in this study were purchased from BioArmor Développement SARL (Plaintel, France). Gas chromatography coupled with mass spectrometry (GC-MS) reports detail the major constituents, and were joined with each EO. *Cinnamomum cassia* essential oil (CEO) was distilled from leaves and branches, *Allium sativum* essential oil (AEO) was extracted from pods and *Mentha piperita* essential oil (MEO) was extracted from leaves. All EOs were extracted by a steam distillation process and were stored in the dark at 4°C.

Bacteria and culture conditions

Pseudomonas aeruginosa PAO1 was grown aerobically in Lysogeny broth (LB) medium (g l⁻¹: NaCl 10; Tryptone, 10; Yeast Extract, 5) at 37°C overnight. NaCl was purchased from Fischer, France; Tryptone and Yeast Extract were purchased from Dutscher, France. The bacterial strain was stored at -80°C in 20% (v/v) glycerol.

Minimal inhibitory concentrations (MIC) and minimal bactericidal concentrations (MBC) of the EOs and blend

The antibacterial activity of the EOs and the blend was measured using a broth dilution method in microplates, adapted from a previously described protocol [17]. CEO, AEO and MEO and the blend were diluted in LB medium within a range of 0.5% to 0.005% (v/v). The same amount of an Ethoxylated Castor Oil (ECO) based emulsifier was added, as for the EO. A volume of 195 µl of each dilution was placed in the wells of a flat bottomed 96 well microplate and bacterial suspension were added to obtain a final Optical Density (OD) of 0.0125 at 600 nm (Shimadzu UV-1800 spectrophotometer Shimadzu Europa GmbH, Duisburg, Germany). Microplates were incubated for 24 h at 37°C without agitation. After cell growth, a tetrazolium salt (MTT, Sigma Aldrich, France) solution in DMSO (Fisher, France) was added to each well at a final concentration of 90 µg ml⁻¹ for 30 min. This allowed viable microorganisms to metabolize the yellow MTT into purple formazan crystals. An aliquot of each well which did not present a formazan production was then spread on LB agar plates, incubated for 24 h at 37°C for the determination of minimal inhibitory concentrations (MIC) or minimal bactericidal concentrations (MBC). MIC and MBC corresponded to the lower concentrations leading to growth or absence of growth, respectively, after spreading. Each test was carried out independently in triplicate.

Anti-biofilm crystal violet assay

Crystal violet assay is a rapid method for the observation of biofilm inhibition [2,10]. EOs was diluted in a LB medium as before. Solution of Gentamycin (Sigma Aldrich, France) at 50 µg ml⁻¹ was used as a positive control. LB medium was used as a negative control. Wells without bacteria inoculation were also used to control the final washing step. For the attachment step, overnight cultures of bacteria were centrifuged for 5 min at 2500 g and the cells were then suspended in sterile physiological saline (SPS 0.9% NaCl) and adjusted at an OD of 0.1 at 600 nm. A volume of 200 µl of this suspension was first placed in a 96 well microplate for adhesion. After 2 h, the wells were rinsed twice with SPS and filled with the different tested conditions. After 24 h incubation at 37°C without agitation, the medium was removed and the wells were rinsed twice with SPS in order to eliminate planktonic

cells. In each well, 200 µl of a 1% crystal violet staining solution in water (Reactif RAL purchased by Servilab, Le Mans, France) was added and rinsed with SPS after 15 minutes until the well which was not inoculated with bacteria was totally colourless. Crystal violet contained in the bacteria was then solubilized with 70% ethanol (Fischer, France) and the OD was read at 600 nm with a Multiskan™ GO Microplate Spectrophotometer (ThermoFisher, Illkirch, France). Each test was conducted in triplicate.

Confirmation of active concentration on glass slides

Cell adhesion and biofilm culture

For an adhesion step, clean and sterile glass slides were immersed in SPS (control), in SPS with 0.1% ECO or in SPS with 0.1% ECO and 0.1% EO blend. For each condition, a working solution was inoculated at 0.1 OD at 600 nm with overnight culture of bacteria. After 2 h, glass slides were carefully rinsed once in SPS to remove non-attached bacteria.

For biofilm formation, bacterial adhesion was performed in SPS as previously described. After 2 h, the glass slides were carefully washed once with SPS, and immersed in LB, LB with 0.1% ECO or LB with 0.1% ECO and 0.1% EOB. After 24 h of growth at 37°C without agitation, the slides were rinsed once with SPS before staining. Each experiment was conducted in triplicate.

Fluorescence microscopy

Attached cells and biofilm were observed with an Olympus BX43 fluorescence microscope (Olympus microsystems, Hamburg, Germany) powered by the X-cite 120q led fluorescence illuminator (Lumen dynamics) using a 100x oil immersion lens. Bacteria were detected by immersion in a 28.55 µM DAPI solution, for 10 min in the dark. After staining, the slides were washed once in SPS to remove the excess of dye and covered with cover glasses to enable observation. Three images were taken per slide. The glass surfaces covered by attached bacteria were evaluated by using the ImageJ software (National Institutes of Health, USA).

Anti-biofilm activity tested in flow cells

Cell adhesion and biofilm culture

Biofilm was grown at 37°C under dynamic conditions in a three channel flow cell (1 × 4 × 44 mm; Biocentrum DTU, Denmark) [18]. The flow system was assembled, prepared and sterilized as described before [19]. The substratum consisted of a microscope glass coverslip (24 × 50 st1; KnittelGlasser, Braunschweig, Germany).

For the adhesion step each channel was inoculated with 250 µl of an overnight bacterial culture diluted to an OD of 0.1 at 600 nm in SPS, or in SPS containing 0.5% or 1% ECO, or in SPS containing 0.5% or 1% EOB (with the same amount of ECO). ECO (10% mother solution in SPS) was filtered on a 20 µm sterile membrane to enhance the quality of the images. Bacteria were allowed to attach to the glass substratum during 2 h at room temperature without flow. A constant flow (2.5 ml h⁻¹) of SPS using a Watson Marlow 205U peristaltic pump (Watson Marlow, Falmouth, United Kingdom) was then applied for 15 min in order to remove non-attached cells. Attached bacteria were observed by confocal laser scanning microscopy (CLSM) as described below. This experiment was conducted in duplicate and three images were taken per channel.

For biofilm formation, each channel of the flow cell chamber was connected to LB medium, or LB medium supplemented with ECO (0.1% or 1%) or else with the EOB (0.1% or 1%). The channels were inoculated with 250 µl of a bacterial suspension diluted in SPS to an OD of 0.1 at 600 nm. A two-hour attachment step was performed as described before, and a flow (2.5 ml h⁻¹) of the medium was then applied for 24 h. Biofilms were observed by CLSM as described below. Each experiment was conducted in triplicate and three images were taken per channel.

Confocal Laser Scanning Microscopy (CLSM)

Attached bacteria and biofilm observations were performed with a TCS-SP2 microscope (Leica Microsystems, Heidelberg, Germany), using a 63x oil immersion lens. Bacteria were detected after cell attachment by staining with 5 µM of SYTO9 green or after biofilm growth by double-labelling with 5 µM SYTO9 green and 0.3 µM propidium iodide (PI) (Live/Dead BacLight Bacterial Viability Kit, Invitrogen, Carlsbad, USA). The staining was performed by injecting 250 µl of fluorescent dye prepared in SPS into a flow cell channel, incubating at room temperature for 15 min in the dark and washing for 15 min with a flow (2.5 ml h⁻¹) of medium, immediately before CLSM observation. SYTO9 green was excited at 488 nm and fluorescence emission was detected between 500 and 550 nm. PI was excited at 543 nm and fluorescence emission was detected at 620 nm. The fluorescence signal of double-labelled specimens was acquired simultaneously.

The glass surfaces covered by attached bacteria were evaluated by using the ImageJ software. For three-dimensional (3D) visualization, images were taken every micrometer throughout the whole biofilm depth. For the processing of 3D image data (volume rendering with shadow projection), the Leica LAS AF software (Leica Microsystems, Heidelberg, Germany) was used. Quantitative analyses of image stacks

were performed using the COMSTAT software (<http://www.imageanalysis.dk/>) [20]. At least three image stacks from each independent experiment were used for each analysis.

Statistical analysis

Results are expressed as means ± standard deviation (SD). The statistical analysis was carried out with Microsoft® Excel 2013 using one-way analysis of variance (ANOVA) followed by a two-tailed T-test at the 5% level to evaluate differences between the samples. A *p* value of <0.05 was considered significant.

Results and Discussion

Essential oil analysis

The EOs selected here were each constituted of a different family of molecules. As MEO was rich in terpenes, AEO was principally composed of sulphonates and the major compounds of CEO derive from cinnamic acid. The whole composition of the EOs, determined by GC-MS, is presented in Table 1. If their global composition was in accordance with literature, the fine composition was slightly different from published data. MEO is quasi exclusively composed of terpenes (>90%). Main compounds were menthol (32.5%) and its derivate (DL)-menthone (26.5%), menthylacetate (4.9%), isomenthone (4.1%), (D)-neomenthol (3.2%), menthofuran (2.4%) and pulegone (1.6%). Other terpenes like eucalyptol (4.7%), D-limonen (3.2%) and alpha/beta-pinen (2.6%) and the sesquiterpene β-caryophyllen (3.7%) also make up its composition. The qualitative and quantitative compositions are similar to those presented in further published data [21]. The plants from genus *Allium* are well known to be rich in sulphinates (69.8%) and the main components of AEO were identified as diallyl disulphides (53.5%) diallyl trisulphides (9.4%) and diallyl sulphides (6.9%).

Plant	<i>Cinnamomum cassia</i> (CEO)		<i>Mentha piperita</i> (MEO)		<i>Allium sativum</i> (AEO)	
Organ	Leaves, Branches		Leaves		Pods	
Essential oil Composition	Cinnamaldehyde	79.9%	Menthol	32.5%	Diallyl disulphides	53.5%
	o-Methoxy cinnamaldehyde	8%	(DL)-Menthone	26.5%	Diallyl trisulphides	9.4%
	Cinnamyl acetate	2.5%	Menthyl acetate	4.9%	Diallyl sulphides	6.9%
	Coumarin	1.5%	Eucalyptol	4.7%	Propene	6.7%
	Benzaldehyde	0.8%	Isomenthone	4.1%		
	Styrene	0.2%	β-caryophyllen	3.7%		
	Cinnamic alcohol	0.1%	D-neomenthol	3.2%		
	Cinnamic acid	0.1%	D-limonen	3.2%		
	D-limonen	0.04%	α/β-pinen	2.6%		
			Menthofuran	2.4%		
		Pulegone	1.6%			

Table 1: Main components (%) detected by GC-MS in the three EOs tested.

A low presence of propene (6.7%) was also detected. Reported concentrations of disulphides are quite low (from 29.1% to 36.5%), and the rate of trisulphides is higher in other published data (from 21.6% to 30%) [22,23]. In CEO, a majority of components derive from cinnamic acid (0.1%) like cinnamaldehyde (79.9%) which was identified as a major compound, o-methoxycinnamaldehyde (8%), cinnamyl acetate (2.5%), coumarin (1.5%), benzaldehyde (0.8%) and also styrene (0.2%) and cinnamic alcohol (0.1%). A low concentration of terpenes is present in the form of d-limonen (0.04%). This composition is in accordance with most of the published data [24] but diverge from data published by Murbach Teles Andrades et al. [25], where the main compound was eugenol (72.1%). This change in main compound can be explained by the use of leaves instead of branches.

An EO's activity is generally related to the main compounds present (menthol, diallyl sulphides and cinnamaldehyde here), even if the activity of the totum should be taken into account. The cinnamaldehyde and eugenol molecular family are extremely different, the presence of a phenolic cycle leads to various activities: as cinnamaldehyde has been reported not to cause the disintegration of the outer membrane, eugenol leads to high degree of cell lysis [26]. Compounds present in minor concentrations may contribute to bioactivity, but are not estimated as active neither tested. Variations in EO compositions obtained from two of the same plants can generally be explained by the effect of differences in where they are sourced (climatically changes, presence of biotic or abiotic stresses), changes in treated plant parts or the extraction protocol. Several parameters can influence the EO compositions, which leads to modifications in their biological properties [27]. As the main compounds presented here belong to different molecular families, we assume that their blend could target diverse cellular mechanisms.

Antibacterial activity of EOs and blend

MIC and MBC against PAO1 were evaluated for each EO and the blend (Table 2). EOs was diluted from 0.5 to 0.005% (v/v), a concentration range considered as the most usable regarding the literature. MEO was not active in the tested concentrations but Tsai et al. [28] obtained a MIC of 0.92% on PAO1. The activity of monoterpenes and especially menthol has been studied by Trombetta et al. on models. They proved that this molecule and other terpenes were able to enhance membrane fluidity, leading to expansion, altered respiration and destabilisation of intramembranous proteins [29]. AEO inhibited the growth of PAO1 at a concentration of 0.5%. As described by Casella et al. [23], the activity of AEO is due to the presence of diallyl sulphides and diallyl disulphides. These molecules derive from allacin, naturally present in garlic, which acts on planktonic bacteria and on biofilms [30,31]. The presence of sulphuric bounds is a key for sulphhydryl dependent enzyme inactivation [32]. Through their work, Chen et al. [33] also highlighted an N-acetyl transferase inhibition activity induced by diallyl sulphides and diallyl disulphides on *Klebsiella pneumoniae*. As PAO1 also possesses this enzyme, it can be assumed that both molecules act due to the same mechanism [34]. CEO was the most active of the three tested EOs and was bactericidal at a concentration of 0.125%. The activity of CEO and mostly cinnamaldehyde has already been reported for *P. aeruginosa* and other bacteria like *Escherichia coli* and *Listeria monocytogenes* [35,36]. Among these previous results, cinnamaldehyde was found to inhibit the Z-ring formation during cell division, by targeting FtsZ protein. Contrary to terpenic compounds, cinnamaldehyde has not been reported to disintegrate the membrane or interfere with ATP formation [37].

The minor differences in active concentrations compared to published data could be explained by slight composition changes as described above (section 3.1) and the use of ECO in our test leading to physico-chemical changes. Glover et al. [38] observed an enhancement of bacterial membrane fluidity when surfactants were added to the growth medium, which was not correlated with high mortality in the case of alcohol ethoxylates.

The three EOs were mixed at an equal concentration (EOB) to evaluate a potential synergistic effect. An inhibiting activity on PAO1 was observed at 0.125% concentration and it became bactericidal at 0.25%. As the EOs were three-times less concentrated in the blend, the appearance of an inhibiting activity at 0.125% and the conservation of a bactericidal effect suggests a synergistic activity between the EOs. An eventual synergistic activity between Cinnamon, Garlic and Peppermint EOs has not been reported yet. It is unusual to study a blends' activity on planktonic and sessile bacteria and the results highlight the benefit of forming an EOs blend.

	CMI % (v/v)	CMB % (v/v)
AEO	0.500	>0.500
CEO	-	0.125
MEO	>0.500	>0.500
EOB	0.125	0.250

Table 2: CMI and CMB of AEO (*Allium sativum* Essential Oil), CEO (*Cinnamomum cassia* Essential Oil), MEO (*Mentha piperita* Essential Oil) and their blend (EOB) on PAO1.

Anti-biofilm concentration of EOB

The crystal violet assay was performed, in order to screen the activity of the blend (EOB). As a first result, we observed that the ECO emulsifier at a maximal concentration of 0.5% had no effect on biofilm formation (data not shown). Using the minimal blend concentration of 0.005%, a slight enhancement of PAO1 biofilm formation was observed, corresponding to a non-significant increase (Figure 1a). Increasing the concentration at 0.25% led to a total biofilm inhibition ($101 \pm 4\%$). Inhibition approximately became stable when the blend's concentration reached 0.1%, an amount of EOB that we conserved for the next steps of our study. In their publication, Kim et al. also showed a 49% reduction of *P. aeruginosa* biofilm formation at 0.1% concentration of CEO according to a 96 well microplate assay [15], suggesting a synergistic effect of EOB.

Confirmation of active concentration on glass slides

Biofilm formation is a process described in different stages, including reversible cell adhesion, non-reversible adhesion, formation of micro-colonies and macro-colonies, leading to the detachment of planktonic cells [4].

Results obtained for cell adhesion on glass slides are presented in Figure 1b. Under control conditions, PAO1 adhered to the glass slides and covered respectively $12.9 \pm 6.7\%$ of the surface. When ECO was added to SPS, attached bacteria seemed to be fewer and less uniform among the slide, but the covered area decreased non-significantly to $10.4 \pm 6.5\%$. This result confirmed the absence of the activity of ECO. After a 2 h treatment with the EOB, the adhesion rate decreased significantly ($P < 0.001$), reaching only $1 \pm 1.1\%$. As EOB was active at a

sublethal concentration, we can consider that it has an anti-adhesive activity, maybe by disrupting the complex cellular processes leading to the development of the adhesive structure [39]. The activity of the other essential oils or components like carvacrol or thymol on early biofilm formation steps has been reported before, and can be related to an inhibition of flagella formation observed on *E. coli* [39,40].

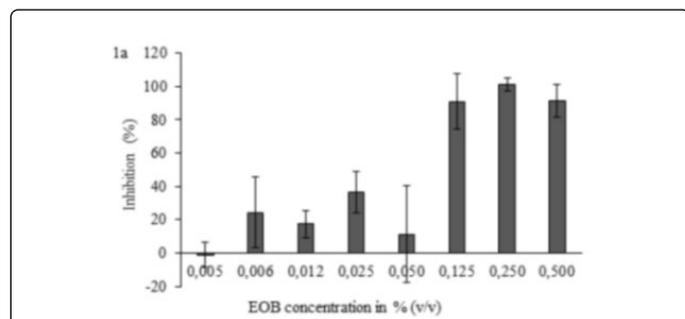


Figure 1a: Impact of a 24 h EOB treatment, with different concentrations ranging from 0.005% to 0.5%, on PAO1 biofilm formation. Biofilms were grown in the wells of a microplate, using the LB medium and increasing concentrations of EOB. In each well ECO was used as emulsifier in same concentrations related to EOB. Use of ECO at a maximal concentration of 0.5% did not cause any inhibition. Biofilm was quantified after a 24 h incubation using crystal violet. Experiments were carried out in triplicate (n=3) and error bars indicate the standard deviation from the mean.

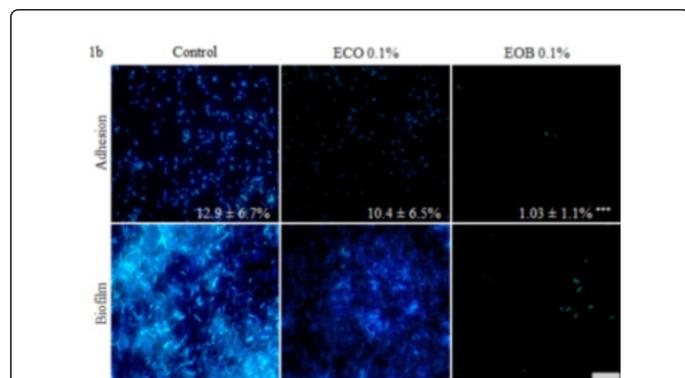


Figure 1b: Fluorescence microscopic visualisation of PAO1 cell adhesion and biofilm formation on glass slides in different conditions. Adhesion and biofilm growth were performed in control condition, in presence of 0.1% ECO or in presence of 0.1% EOB emulsified with 0.1% ECO, according to the respective protocols. Attached cells and biofilms were stained with DAPI (28.5 μM) and observed at magnification x1000. Scale represents 8μm. Covered area fractions of adhered cells were obtained after treatment with IMAGEJ software. Experiments were carried out in triplicate and three images were taken pro slides. Error bars represent standard deviation from the mean (n=9) and ***P<0.001.

When biofilm was grown under control conditions, bacteria were able to colonize the total glass surface, leading to macrocolony formations (Figure 1b, Biofilm control). When ECO was added at 0.1% to the growth medium, biofilm formation was altered (Figure 1b, Biofilm ECO 0.1%). In appearance, the bacteria which adhered to the

glass surface did not form identical biofilm as those observed for the control conditions (Figure 1b). After 24 h of EOB treatment, only few attached bacteria were still present (Figure 1b, Biofilm EOB 0.1%).

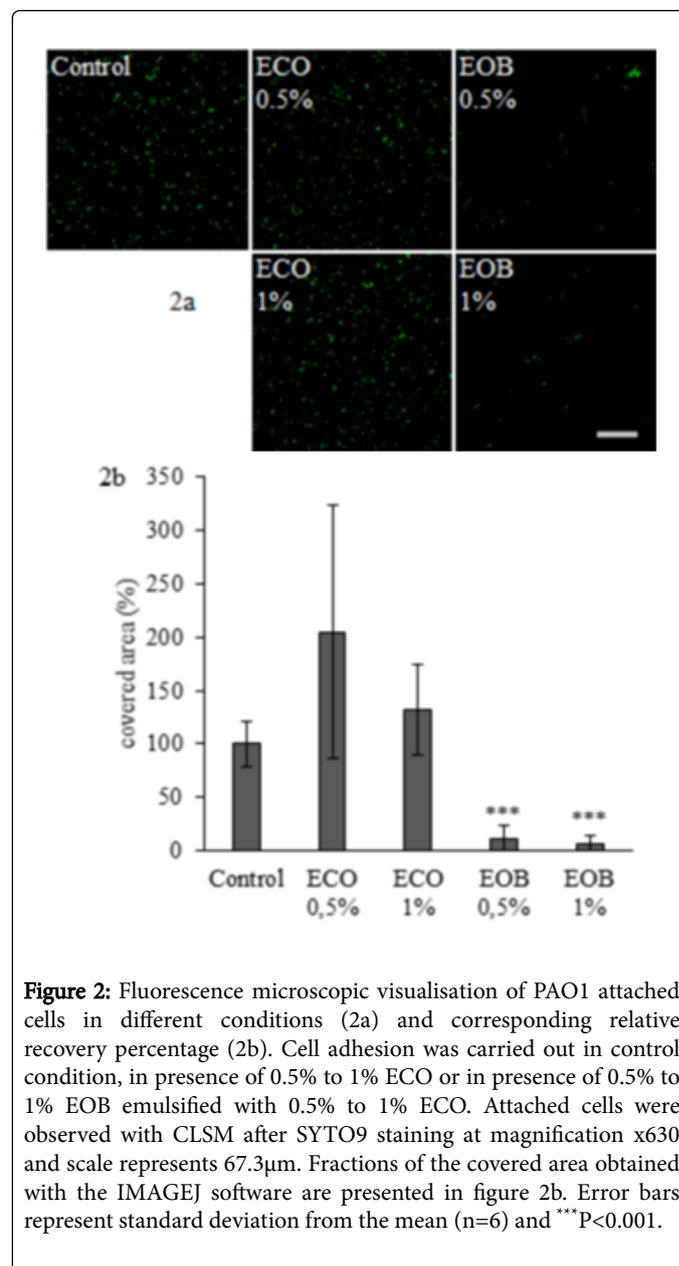


Figure 2: Fluorescence microscopic visualisation of PAO1 attached cells in different conditions (2a) and corresponding relative recovery percentage (2b). Cell adhesion was carried out in control condition, in presence of 0.5% to 1% ECO or in presence of 0.5% to 1% EOB emulsified with 0.5% to 1% ECO. Attached cells were observed with CLSM after SYTO9 staining at magnification x630 and scale represents 67.3μm. Fractions of the covered area obtained with the IMAGEJ software are presented in figure 2b. Error bars represent standard deviation from the mean (n=6) and ***P<0.001.

Bacterial adhesion and biofilm formation in dynamic conditions

Biofilm formation was followed in a flow cell chamber (Figure 2). As previously for the adhesion step, bacteria were allowed to adhere to glass slides for 2 h without flow (Figures 2a and 2b). In the control condition, PAO1 covered a mean area of $1.3 \pm 1.1\%$ that we considered as the 100% adhesion rate. The recovery value order of magnitude was slightly lower in these tests than those observed on glass slides (Section 3.4). As we observed no effect on bacterial adhesion with a 0.1% EOB treatment, we tested both 0.5% and 1% concentrations of EOB. When ECO was added at a concentration of 0.5%, the covered area was non-

significantly doubled and increased by a factor of 1.3 when the emulsifier was added at 1%. Statistical testes indicate that bacterial attachment was not significantly changed by the addition of ECO ($P>0.05$) (Figure 2b).

Treatment with 0.5% and 1% of EOB significantly reduced the attachment of PAO1 to the glass surface with only $10 \pm 12\%$ and $6 \pm 7\%$ of the surface remaining covered when compared to the control (Figures 2a and 2b). The EOB remained active at low concentrations while an enhancement of the adhesion phenomenon was observed when ECO was added to SPS. It is recognised that microorganisms are able to secrete bio-surfactants in order to enhance their motility on surfaces [41], a fact that could explain the better cell repartition when 0.5% ECO was used. As cell adhesion is a complex mechanism including the presence of motility and attachment appendix like pili, the surface tension modification can lead to the inhibition of adhesion, observed when a higher dose of ECO was added.

Biofilm growth and mortality were followed as biovolume estimation with COMSTAT software of SYTO9 and PI labelled cells respectively (Figures 3a and 3b). In control conditions, we observed a thin biofilm (maximal thickness (Tm) of $11.4 \pm 1.2 \mu\text{m}$) associated with the filamentous organization of cells (Figure 3a, Control), damaged cells were evenly distributed in all biofilm layers. In this

condition, the biofilm had a total biovolume of $1.6 \pm 1.1 \mu\text{m}^3 \mu\text{m}^{-2}$ with a mortality rate of $1.5 \pm 2.6\%$ (Figure 3a and 3b). When 0.1% ECO was added to the medium, no significant change was observed in the total biovolume, and the mortality rate was not significantly reduced to $0.2 \pm 0.2\%$ (Figure 3a). The biofilm structure was however more heterogeneous since the thickness was increased (maximal thickness of $19.5 \pm 6.9 \mu\text{m}$) and we did not observe any filamentous consortium (Figure 3a, ECO 0.1%). Overall, this result is in total opposition with the observation carried out on glass slides. When the EOB was added in a 0.1% concentration to LB, the total biovolume significantly increased to $2.6 \pm 0.94 \mu\text{m}^3 \mu\text{m}^{-2}$ and mortality reached $1.5 \pm 2.7\%$ (Figure 3 EOB 0.1%). Thickness was close to that observed with 0.1% ECO ($18.8 \pm 2.8 \mu\text{m}$) but all cells grew and formed filamentous consortia (Figure 3a, EOB 0.1%). Filaments are a known response to sublethal stresses in some bacteria, involving the stoppage of cell division mechanism, like the FtsZ ring formation [42]. As described by Domadia et al. [37] cinnamaldehyde bind FtsZ proteins and inhibit their polymerization, which could lead to filament formation. Furthermore, the fact that filamentous cells were all stained by PI proved the limit of Live/Dead staining: in fact filamentous cells multiply in a rapid manner, which leads to a fault in the membrane integrity and a higher permeability of the PI, as the cells are still alive [42].

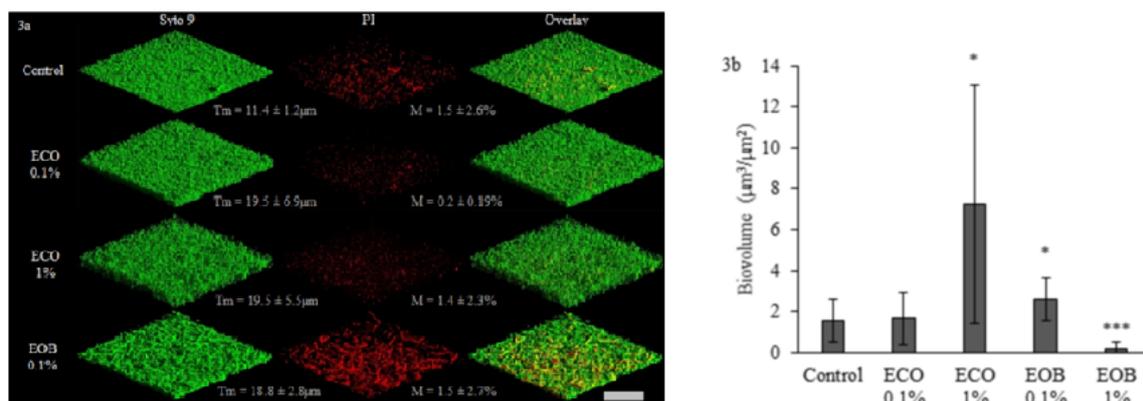


Figure 3: 3D fluorescence microscopic visualisation of PAO1 biofilms grown in different conditions (3a) and corresponding biovolumes (3b). PAO1 biofilms were performed in 3 well chambered cover glasses under 2.5 ml h^{-1} flow. LB (Control), LB with 0.1% or 1% ECO or LB with 0.1% or 1% EOB emulsified with ECO in the same amount, in relation to the EOB concentrations, were used as growth media. No biofilm was observed at 1% EOB concentration (image not shown). Total biofilm cells were detected after SYTO9 ($5 \mu\text{M}$) staining and appear in green, as altered cells stained with Propidium Iodid (PI, $0.3 \mu\text{M}$) appear in red. The overlay represents the superposition of SYTO9 and PI stained cells images. Scale represents $67.3 \mu\text{m}$. Total biovolume and maximal thickness (Tm) were obtained from SYTO9 stained cell images treated with COMSTAT. Mortality rates (M) were obtained by calculating the ratio: total biovolume (SYTO9) / altered biovolume (PI) x 100. Error bars represent standard deviation from the mean (n=9) and * $P<0.05$, *** $P<0.001$.

As 1% ECO was added to the LB medium, a high augmentation of total biovolume was observed and correlated to a constant mortality rate ($7 \pm 5 \mu\text{m}^3 \mu\text{m}^{-2}$ total biovolume for $1.4 \pm 2.3\%$ mortality, Figure 3b). Again the maximal thickness observed was $19.5 \pm 5.5 \mu\text{m}$ but as we can observe on corresponding pictures, the cells were closely organized (Figure 3a, ECO 1%). As in early biofilm formation stages, Biosurfactants play an important role while enhancing *Pseudomonas* Quinolone System (PQS) solubility and therefore its activity, explaining the dose dependent augmentation of biofilm formation when using ECO [41].

Finally, when the EOB was added at 1%, the biofilm was totally destabilized. The total biovolume observed for this condition was $0.15 \pm 0.27 \mu\text{m}^3 \mu\text{m}^{-2}$ and mortality reached 0.25%. According to these results, the blend's anti-biofilm activity seems to depend on a bactericidal activity but, in a recent publication, Kim et al. also proved the efficiency of cinnamaldehyde to affect the Quinolone Signal, pyocyanin production and swarming motility of *P. aeruginosa* [15]. Another work showed the efficiency of a 1 mg ml^{-1} peppermint extract on *P. aeruginosa* bacterial adhesion and provided a 38% inhibition on preformed biofilms [2].

Conclusion

Due to the three methods, the effectiveness of a blend of three essential oils against a multidrug-resistant pathogen, PA01 has been proved. This work first highlighted the increased effectiveness of a blend, in comparison to pure essential oil, on planktonic cells and on biofilm formation. As expected, each essential oil contributes to global activity by targeting different mechanisms, which enlarge the spectrum of activity. CLSM enabled us to observe filamentous consortia, revealing high stress within PA01 biofilms treated with 0.1% EOB. Total biofilm eradication was finally obtained when the concentration reached 1% under a constant flow. The different active concentrations obtained in each conditions highlighted the importance of varying the methods in order to obtain representative results.

These properties could be used in order to work out a preventive surface treatment using molecular grafting, or a sanitizer active on preformed biofilm. Additionally, further experiments have to be carried out in order to understand the effect of the blend at a genetic scale, to have a better understanding of the mechanism. This could lead to a combination of molecules with known antibiotics in order to obtain new treatments against multi-drug-resistant bacteria like PA01.

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