

Antagonistic Effects of Insulin Signaling and Glucagon Signaling on Controlling Hepatic Gluconeogenic Gene Expression

Evan Chang and Ling He*

Department of Pediatrics, Division of Metabolism, Johns Hopkins University School of Medicine, Baltimore, MD 21287, USA

*Corresponding author: Ling He, Division of Metabolism, 600 N. Wolfe St, Baltimore, MD 21287, USA, Tel: 410-502-5765; Fax: 410-502-5779; E-mail: heling@jhmi.edu

Received date: Feb 08, 2014, Accepted date: Apr 09, 2014, Published date: Apr 11, 2014

Copyright: © 2014 Chang E, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

With the worldwide epidemics of obesity and diabetes, there is an urgent need for action at the global and national levels to prevent and to treat these metabolic disorders. Inappropriate hepatic glucose production is the major cause of hyperglycemia in obese and diabetic patients. In this mini-review, we summarize the antagonistic effects of insulin signaling and glucagon signaling on controlling gene expression related to hepatic glucose production through CREB co-activators and FOXO1. In the fasted state, phosphorylation of CREB at S133 recruits CBP/P300 and CRTC2 to CREB, leading to the formation of CREB co-activators complex. CREB and P300 also up-regulate FOXO1 gene expression. CREB co-activators together with FOXO1 drive gluconeogenic gene expression to maintain euglycemia. In the fed state, insulin suppresses gluconeogenic gene expression through the phosphorylation of CBP, which results in the disassembly of CREB-CBP-CRTC2 complex, furthermore, phosphorylation of CRTC2 and FOXO1 by insulin excludes CRTC2 and FOXO1 from nucleus and promotes their degradation in cytoplasm.

Keywords: Glucagon; Gene expression; Metabolic disorder

The Epidemics of Obesity and Diabetes Cause Serious Health Problems

Over the past three decades, the number of children with obesity has more than doubled, and children obese rate at age of 6-11 years has increased from 7% in 1980 to 18% in 2010 in the United States, accounting for one third of all children [1,2]. The World Health Organization estimates that nearly 43 million children under the age of five are obese worldwide [3].

The obesity epidemic, over nutrition and increasing sedentary lifestyles among young people are the major contributors to the increase in type 2 diabetes among children and adolescences. Currently, 215,000 individuals under the age of 20 in the United States have diabetes [4]. Data from the International Diabetes Federation show that diabetes affects at least 382 million individuals worldwide [5]. Hyperglycemia is the hallmark metabolic abnormality of diabetes and is the major health concerns in those with type 2 diabetes mellitus patients. Elevated glucose levels lead to the severe adverse effect of non-enzymatic glycosylation of many cellular proteins that causes them to function improperly or lose function completely [6,7]. These molecular effects are often seen in diabetes patients evident by microvascular tissue damage to the kidney, retina and nerves, which lead to the commonly observed outcomes of end-stage renal failure, cardiovascular disease, loss of visual acuity, and loss of extremities through amputation [8].

Thus, the prime clinical goal in the treatment of type 2 diabetes mellitus patients is to maintain blood glucose levels as close to the normal blood glucose levels as possible so as to reduce the occurrence of these complications.

Opposing Actions of Insulin Signaling and Glucagon Signaling Maintains Normal Blood Glucose Levels

Blood glucose levels are tightly regulated by the opposing actions of insulin signaling and glucagon signaling pathways. In the fed and postprandial states, glucose is absorbed in the gastrointestinal tract and enters the blood circulation. The elevation in blood glucose levels triggers the secretion of insulin from pancreatic β cells. This, in turn, stimulates glucose utilization in peripheral tissues such as muscle and adipose tissue, and suppresses hepatic glucose production [9].

Eventually, blood glucose levels return to the normal defined range due to insulin signaling. In the fasting state, blood glucose levels drop, and glucagon is secreted from α cells of the pancreas. During early fasting, glucagon stimulates hepatic glycogenolysis, in which stored glycogen in the liver is broken down into glucose and released into the bloodstream to maintain euglycemia.

Moreover, glucagon-stimulated hepatic gluconeogenesis plays a dominant role in maintaining euglycemia during prolonged fasting [10]. The maintenance of normal blood glucose levels (70 -110 mg/dL) is essential in the protection against hypoglycemia during fasting, because hypoglycemia is detrimental to the health of an organism due to the fact that glucose is the main and only energy source for neurons and erythrocytes [11].

Glucagon Stimulates Hepatic Gluconeogenic Gene Expression in the Fasted State

In the fasted state, glucagon stimulates hepatic glucose production through the cAMP-PKA (protein kinase A) signaling pathway (Figure 1).

Suppression of Hepatic Gluconeogenic Gene Expression by Insulin Signaling in the Fed State

Insulin signaling is crucial for the suppression of glucose production in the liver. Mice with liver specific insulin receptor knockout exhibited marked increase of hepatic glucose production and extreme hyperglycemia [17]. In the fed and postprandial states, the suppression of hepatic glucose production by insulin is complicated and involves many transcription factors and signaling mediators that have been reported. The gluconeogenic engine, CREB co-activator complex, first needs to be turned off to reduce endogenous glucose production. When assembled, the CREB co-activators complex upregulates the transcription of hepatic gluconeogenic related genes, such as *G6pc* and *Pck1*, and increases hepatic glucose production. To sufficiently control the blood glucose levels in the fed and postprandial states, several mechanisms have been proposed. We have proposed that phosphorylation of CBP at Ser436 by insulin leads to the disassembly of CREB co-activator complex [13]. This phosphorylation event is mediated by *aPKC δ* (atypical protein kinase C), which is activated by insulin through the PI3K-PDK1 pathway (Figure 2).

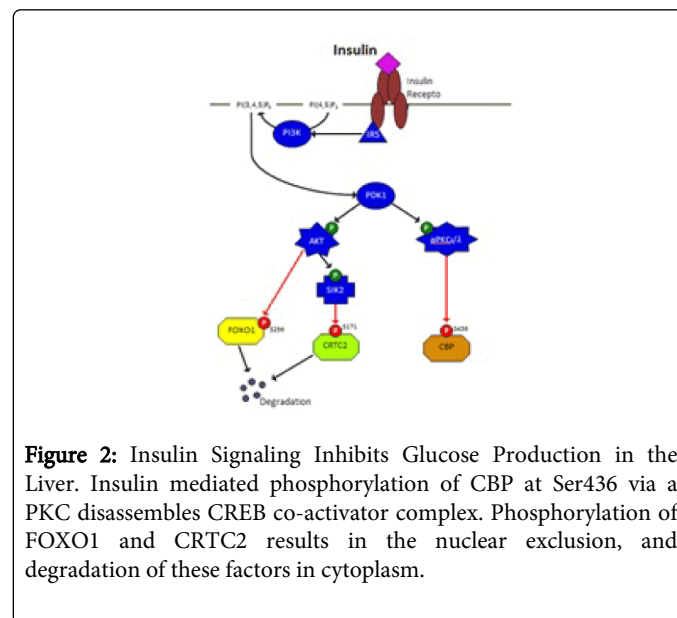


Figure 2: Insulin Signaling Inhibits Glucose Production in the Liver. Insulin mediated phosphorylation of CBP at Ser436 via a PKC disassembles CREB co-activator complex. Phosphorylation of FOXO1 and CRTC2 results in the nuclear exclusion, and degradation of these factors in cytoplasm.

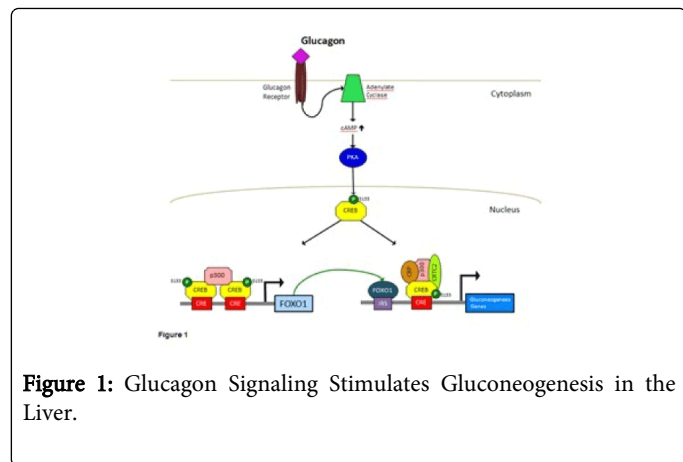


Figure 1: Glucagon Signaling Stimulates Gluconeogenesis in the Liver.

Phosphorylation of CREB at Ser133 leads to the recruitment CBP, P300 and CRTC2 to CREB and the formation of CREB co-activator complex. CREB and P300 also drive *Foxo1* gene expression; FOXO1 binds to insulin responsive sequence (IRS) on gluconeogenic gene to fully activate the gluconeogenic program.

Activated PKA by glucagon phosphorylates CREB at Ser133. This event recruits the formation of the CREB (cAMP response-element-binding protein) co-activators complex and initiates the transcription of gluconeogenic genes containing CRE sites (cAMP response elements), including *G6pc* (glucose-6-phosphatase) and *Pck1* (phosphoenolpyruvate carboxykinase1). In addition to PKA mediated CREB phosphorylation, PKA also stimulates the dephosphorylation of CRTC2 (CREB regulator transcription coactivator 2) and CBP (CREB binding protein) [12,13]. The salt-inducible kinases (SIKs), especially SIK2, play a critical role in regulating CRTC2 activity. Phosphorylation of CRTC2 by SIK2 excludes CRTC2 from nucleus. However, activated PKA phosphorylates SIK2 at Ser587 and suppresses SIK2 activity, therefore negates its inhibition on CRTC2 [14,15]. Hence, CRTC2 re-localizes into the nucleus. Furthermore, dephosphorylation of CBP also leads to its association with CREB [13,16]. Together, these events result in the formation of the CREB co-activator complex to promote gluconeogenic gene expression and hepatic glucose production.

Recently, we found that fasting led to the marked increase of *Foxo1* mRNA and FOXO1 protein levels in the liver [11]. A non-hydrolyzable cAMP analog-dibutyrylcAMP (Bt-cAMP) activated the cAMP-PKA signaling pathway and also increased *Foxo1* gene expression, suggesting that the cAMP-PKA pathway may up-regulate *Foxo1* gene expression. Using adenoviral shRNAs to deplete co-activators in hepatocytes, we found that depletion of P300 blocked the induction of FOXO1 by Bt-cAMP. Moreover, depletion of CREB abolished the induction of *Foxo1* mRNA levels by Bt-cAMP. These data demonstrate that *Foxo1* gene expression stimulated by Bt-cAMP is mediated by CREB and P300 [11]. After characterization of the *Foxo1* gene promoter, we found that *Foxo1* gene expression is driven by CREB and P300, which bind to tandem CRE sites in the proximal promoter region of *Foxo1* gene. In addition, inhibition of P300 histone acetyl transferase activity decreased hepatic FOXO1 protein levels as well as blood glucose levels. Since FOXO1 also up-regulates gluconeogenic expression by binding to insulin response sequences located in the promoters of *G6pc* and *Pck1*, the induction of the *Foxo1* gene by cAMP-PKA, thus, fully activates the gluconeogenic program and maintains euglycemia in the fasted state [11].

event triggers the export of FOXO1 from nucleus to the cytoplasm and promotes its ubiquitinylation and degradation [20,21].

Perspective

Insulin and glucagon actions function in concert at the molecular level to maintain the blood glucose levels in a defined normal range. However, diabetic patients often have elevated serum glucagon levels, which should stimulate gluconeogenic gene expression and excessive production of glucose in the liver. In addition, the impairment of insulin signaling due to insulin resistance in diabetic patients weakens the insulin-mediated suppression of gluconeogenic gene expression, suppression of glucose production in the liver, and uptake of glucose in peripheral tissues. Together, these effects result in the development of hyperglycemia. The elucidation of the signaling cascade and transcriptional activity in both the glucagon and insulin pathways will provide us with valuable information to identify and create more efficient and robust therapeutic compounds to better combat obesity and type 2 diabetes mellitus in children and adolescents. For example, the inhibition of P300 histone acetyltransferase activity may be a target for the treatment of diabetes because inhibition of P300 histone acetyltransferase activity decreases fasting blood glucose levels [11,22].

References

1. Ogden CL, Carroll MD, Kit BK, Flegal KM (2012) Prevalence of obesity and trends in body mass index among US children and adolescents, 1999-2010. *JAMA* 307: 483-490.
2. National Center for Health Statistics (2012) Health, United States, 2011: With Special Feature on Socioeconomic Status and Health. Hyattsville, USA.
3. World Health Organization (2014) Obesity and Overweight (No. 311).
4. Centers for Disease Control and Prevention (2011) National diabetes fact sheet: national estimates and general information on diabetes and prediabetes. Atlanta, GA; USA.
5. International Diabetes Federation (2013) IDF Diabetes Atlas, (6th edn), Brussels, Belgium.
6. Einarsdóttir AB, Stefánsson E (2009) Prevention of diabetic retinopathy. *Lancet* 373: 1316-1318.
7. Bellentani S, Marino M (2009) Epidemiology and natural history of non-alcoholic fatty liver disease (NAFLD). *Ann Hepatol* 8 Suppl 1: S4-8.
8. Reinehr T (2013) Type 2 diabetes mellitus in children and adolescents. *World J Diabetes* 4: 270-281.
9. Muoio DM, Newgard, CB (2008) Mechanisms of disease: molecular and metabolic mechanisms of insulin resistance and B-cell failure in type 2 diabetes. *Nat Rev Mol Cell Biol* 9: 193-205.
10. Habegger KM, Heppner KM, Geary N, Bartness TJ, DiMarchi R, et al. (2010) The metabolic actions of glucagon revisited. *Nat Rev Endocrinol* 6: 689-697.
11. Wondisford AR, Xiong L, Chang E, Meng S, Meyers DJ, et al. (2013) Control of Foxo1 gene expression by co-activator p300. *J Biol Chem* 289: 4326-4333.
12. Koo SH, Flechner L, Qi L, Xhang X, Sreaton RA, et al. (2005) The CREB coactivator TORC2 is a key regulator of fasting glucose metabolism. *Nature* 49: 1109-1114.
13. He L, Sabet A, Djedjos S, Miller R, Sun X, et al. (2009) Metformin and Insulin Suppress Hepatic Gluconeogenesis by Inhibiting cAMP Signaling Through Phosphorylation of CREB Binding Protein (CBP). *Cell* 137: 635-646.
14. Patil S, Unterman TG (2005) TORCs rev up gluconeogenesis. *Cell Metab* 2: 210-212.
15. Muraoka M, Fukushima A, Viengchareun S, Lombés M, Kishi F, et al. (2009) Involvement of SIK2/TORC2 signaling cascade in the regulation of insulin-induced PGC-1 α expression in brown adipocytes. *Am J Physiol Endocrinol Metab* 296: E1430-1439.
16. He L, Naik K, Meng S, Cao J, Sidhaye AR, et al. (2012) Transcriptional coactivator p300 maintains basal hepatic gluconeogenesis. *J Biol Chem* 287: 32069-77.
17. Fisher, SJ, Kahn CR (2003) Insulin signaling is required for insulin's direct and indirect action on hepatic glucose production. *J Clin Invest* 111: 463-468.
18. Zhou XY, Shibusawa N, Naik K, Porras D, Temple K, et al. (2004) Insulin regulation of hepatic gluconeogenesis through phosphorylation of CREB-binding protein. *Nat Med* 10: 633-637.
19. He L, Cao J, Meng S, Ma A, Radovick S, et al. (2013) Activation of basal gluconeogenesis by coactivator p300 maintains hepatic glycogen storage. *Mol Endocrinol* 27: 1322-1332.
20. Brent, MM, Anand R, Marmorstein R (2008) Structural basis for DNA recognition by FoxO1 and its regulation by posttranslational modification. *Structure* 16: 1407-1416.
21. Tsai, KL, Sun YJ, Huang CY, Yang JY, Hung MC, et al. (2007) Crystal structure of the human FOXO3a-DBD/DNA complex suggests the effects of post-translational modification. *Nucleic Acids Res* 35: 6984-6994.
22. Liu Y, Dentin R, Chen D, Hedrick S, Ravnskjaer K, et al. (2008) A fasting inducible switch modulates gluconeogenesis via activator/coactivator exchange. *Nature* 456(7219): 269-273.