Associated Factors and Liver Disease Severity for Decreased Bone Mineral Density in HIV Mono- and HIV/HCV Co-infected Patients

Valentina Li Vecchi1, Maurizio Soresi1, Lydia Giannitrapi1, Giovanni Mazzola2, Pietro Colletti2, Ilaria Domenica Amico3, Fabio Tramuto3, Walter Granà1, Massimo Midiri2, Giuseppe Caruso2, Giuseppe Montalto3 and Paola Di Carlo4

1Biomedical Department of Internal Medicine and Specialties, University of Palermo, Italy
2Department of Clinical Medicine and Emerging Pathologies, University hospital P. Giacone-Palermo, Italy
3Department of Diagnostic Imaging, University of Palermo, Italy
4Department of Medicine and Specialties, University of Palermo, Italy

Abstract

Objective: We assessed the prevalence and risk factors of decreased bone mineral density (BMD) in patients mono-infected with human immunodeficiency virus (HIV) or co-infected with hepatitis C virus (HIV/HCV). We also evaluated whether bone loss was linked to lipid asset in both groups and to severity of liver fibrosis in the co-infected group.

Methods: We consecutively enrolled 194 HIV-patients (129 mono-infected and 65 co-infected). All HIV-patients underwent dual-energy X-ray absorptiometry (DXA), while co-infected patients underwent transient elastography. Advanced liver fibrosis was defined as a median liver stiffness ≥ 9.5 kPa. Fibrosis was also assessed in all the HIV-patients using FIB-4.

Results: The overall prevalence of low BMD and osteoporosis was 26.8% and 26.0%, respectively. It was significantly higher among HIV/HCV co-infected than mono-infected patients in lumbar/femoral sites (P<0.04 and P<0.05, respectively). HDL-cholesterol levels correlated independently with lumbar DXA Z-score (P=0.03) in HIV mono-infected subjects. Liver stiffness correlated negatively and independently with femoral Z- and T-scores among co-infected patients (P=0.003; P=0.01, respectively). Stratifying co-infected subjects by sex, liver stiffness and lumbar/femoral Z-scores (P<0.04) or T-scores (P<0.05; P<0.04, respectively) correlated negatively only in the females. Longer PI exposure was negatively and independently correlated with BMD.

Conclusion: Our HIV-infected patients appeared at high risk for low BMD and osteoporosis. Severity of liver fibrosis was an independent predictor of bone loss in co-infection, although other factors could affect the skeletal system in HIV/HCV co-infection. Further research into the impact of liver fibrosis and lipid asset on bone disease in HIV-infection is necessary.

Keywords: HIV; HCV; Coinfection; Bone mineral density; Osteoporosis; Liver fibrosis

Introduction

Osteoporosis and fragility fractures are increasingly being recognized in people with human immunodeficiency virus (HIV) infection [1,2]. A meta-analysis of cross-sectional studies shows that the prevalence of osteoporosis in HIV-infected patients is approximately 15%, which is more than three times greater than figures reported in HIV-uninfected controls [3].

In the last few years, many efforts have been made to identify the several and multifactorial mechanisms that may be involved in skeletal disorders in the HIV-setting [1,4].

Highly active antiretroviral therapy (HAART) has led to a significant reduction in morbidity and mortality in HIV-patients, although its long-term toxicity has been largely recognized [3,5,6]. The resulting aging of the HIV-population and a variety of effects related to specific antiretroviral drug exposure have been associated to the skeletal disorders [3,7,8]. A number of the traditional osteoporosis risk factors may be more prevalent in HIV-infected patients: vitamin D deficiency [9], low body weight [7,10], lifestyle risk factors (including alcohol consumption [7], smoking, corticosteroid use [11] and drug addiction [12]), as well as the HIV infection per se (due to immunodeficiency). In addition, the chronic pro-inflammatory status and the direct action of HIV proteins may also play a role in decreasing bone mineral density (BMD) [4].

Hepatitis C virus (HCV) infection is a major health problem in the HIV-infected population [13]. A recent systematic review and meta-analysis suggests that HIV/HCV co-infection is associated with a greater risk of osteoporosis than HIV mono-infection [14], although many divergences exist regarding osteoporosis prevalence in co-infected patients, with values ranging from 5-45%. Moreover, it is debated whether liver disease severity may indeed have a relevant role in increasing the risk of osteoporosis in co-infection [15].

The relationship between lipid abnormalities and BMD has been little investigated in the setting of HIV-infection [16].

In this study we analyzed the prevalence of low BMD and frank osteoporosis in the whole HIV-population; we also studied both the...
HIV/HCV co-infected and HIV mono-infected groups to investigate (i) what potential metabolic, lifestyle and/or HIV-related risk factors are associated with decreased BMD, and (ii) whether bone loss is more prevalent in HIV/HCV co-infected patients and if there is a link with severity of liver fibrosis.

**Methods**

**Study design**

This was a cross-sectional study, including consecutively enrolled HIV mono- and HIV/HCV co-infected patients who were being followed-up at the AIDS Center of the University of Palermo, Italy between January 2011 and May 2013.

Exclusion criteria included acute liver events, hepatocellular carcinoma, chronic hepatitis B infection, kidney disease, gastrointestinal disorders and steroid or sex steroid therapy.

The study protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki and was approved by the local Ethics Committee. Informed consent was obtained from all subjects.

**Data collection**

Data on age, gender and ethnicity, total time on antiretroviral therapy (ART), as well as exposure to protease inhibitors (PIs), non-nucleoside reverse-transcriptase inhibitors (NNRTIs) and tenofovir were collected, and recorded in a database designed for this study. Specific exposure >1 year to tenofovir and some other antiretroviral drugs (such as the first generation of HIV-1 integrase strand transfer inhibitors, raltegravir, or the NNRTIs, etravirine and PIs, darunavir) was also recorded. Exposure to PIs was graded in the following score (PI score): 0-3 (0=no exposure; 1: <five years; 2: from five to nine years; 3: ≥ ten years).

HIV diagnosis date, medical and drug abuse history, lifestyle habits (including smoking status and alcohol consumption) and bone fractures were also recorded through patient interviews. Smoking status was categorized as non-smokers, former smokers and current smokers. Alcohol intake >20 gm/day either at the time of the study or in the past was also recorded.

Body mass index (BMI) was calculated as weight (in kg) divided by height squared (m²). Diabetes mellitus or Impaired Fasting Glucose (IFG) is defined in accordance with the Expert Committee on the Diagnosis and Classification of Diabetes Mellitus criteria [17]. Diabetes and IFG were analyzed together because of the small number of patients.

**Laboratory methods**

CD4+ T-cell count, and nadir, plasma HIV-RNA levels, hepatitis B virus surface antigen and HCV antibody were assessed within two weeks after DXA. In the HIV/HCV co-infected patients HCV-genotype and plasma HCV-RNA levels were also recorded. Complete blood cell count, alanine aminotransferase (ALT) and aspartate aminotransferase (AST) were assessed within two weeks after DXA in the HIV mono- and HIV/HCV co-infected patients, respectively. Total-cholesterol, high-density lipoprotein (HDL)-cholesterol, low-density lipoprotein (LDL)-cholesterol, triglycerides and glycemia were determined at baseline in both the HIV-groups. The plasma concentration ratio of triglycerides to HDL (triglycerides/HDL ratio) was also calculated as a surrogate marker of insulin resistance [18]. Serum bone alkaline phosphatase, 25-hydroxyvitamin D, parathyroid hormone, phosphorus and calcium levels were assayed at the time of DXA scan in both the HIV-groups. All the blood tests were performed on samples from fasting patients.

**Bone mineral density assessment**

BMD was assessed in all patients by dual-energy X-ray absorptiometry (DXA), using a QDR Discovery Hologic DXA in the femoral neck and DXA in the lumbar spine by total body DXA, as previously reported [12]. For each scan, BMD, T-scores and Z-scores were recorded. T-score compares BMD with the mean of a healthy young (age 20-30 years) reference population, matched for sex and ethnicity, and is expressed as the number of standard deviations above or below the reference mean. Z-score is the number of standard deviations below the reference mean BMD of a gender- and ethnicity-matched population of the same age. Osteopenia was defined when at least one of the two DXA T-scores was less than -1. Osteoporosis was diagnosed when either femoral neck or lumbar spine DXA T-scores were less than -2.5, as recommended by the World Health Organization (WHO) and the National Osteoporosis Foundation guidelines [19,20]. Low BMD was defined as a lumbar and/or femoral neck Z-score of -2 or less.

**Liver fibrosis assessment**

Transient elastography (FibroScan; EchoSens, Paris, France), which assesses liver fibrosis by measuring liver stiffness, was performed in the HIV/HCV co-infected patients. Details of the technical background and examination procedure have previously been described [21]. Liver cirrhosis was defined as liver stiffness ≥ 12.5 kPa [22]. Fibrosis was assessed biologically using the FIB-4 index. It was calculated as follows: age × AST [IU/L]/(platelet count [10⁹/L] × (ALT [IU/L])¹/²) [23]. Advanced liver fibrosis was estimated using an FIB-4 index >3.25 as a reference.

**Statistical analysis**

Data were expressed as mean ± standard deviation when distribution was Gaussian. Differences were calculated using Student's t-test. Otherwise, data were expressed as median and range and analyzed with the Mann-Whitney U test. Fisher's exact and χ² tests, Pearson's correlation and Spearman's rank correlation were used where appropriate. Multiple linear regression analysis was performed to estimate the independence of the association between lumbar spine and femoral neck T-scores as well as variables significant at univariate analysis in all the HIV-infected subjects. Variables contributing significantly to fit the logistic equation were then selected by a step-wise procedure. P<0.05 was considered significant.

All analyses were performed using the SPSS software package (version 22.0; Chicago, IL, SA).

**Results**

**Study population**

From 260 eligible HIV-positive we enrolled 194 patients (121 M, 73 F), 129 of which were HIV mono-infected (66.5%). Mean age was 48 ± 8 years; 89/194 (45.9%) patients were >50 years old.

Table 1 lists the main demographic, lifestyle and clinical features of the whole population and of the two study groups.

No significant differences were found in mean age or BMI values between the HIV mono-infected and co-infected patients. Analysis of life style risk factors showed that more than 50% of the study patients were smokers or ex-smokers (P<0.03). In addition, alcohol abuse (P<0.001) and drug addiction (P<0.0001) were significantly more prevalent in the co-infected vs. HIV mono-infected patients.
Most HIV mono- and HCV co-infected patients (95%) were following HAART: 9 patients (7 HIV mono- and 2 HIV/HCV co-infected) were treatment-naive, while only 1 HIV mono-infected patient had interrupted treatment due to poor compliance. Time since HIV diagnosis and duration of ART exposure were both significantly longer in the co-infected than HIV mono-infected patients (P<0.0001). 

Numerous HIV mono- and co-infected patients (46/121 vs. 34/63 patients) had received PIs for more than 10 years, although no significant difference in duration of PI exposure was observed between the two groups when the PI treatment score was calculated (P=ns). Moreover, no significant difference was found either in the prevalence of exposure to tenofovir or to the newer antiretroviral drugs (data not shown). 

Table 2 shows the main hematological and virological parameters in the study population. All of the HCV co-infected patients had detectable HCV-RNA and 41 patients (63%) had HCV-RNA >700 000 IU/ml. Serum ALT and AST levels were significantly higher in co-infected than in mono-infected patients (P<0.0001). Total-cholesterol, LDL- and HDL-cholesterol levels were higher in the mono-infected than in co-infected patients (P<0.0001; P<0.05, respectively). The median level of vitamin D was lower in HIV mono-infected than HIV/HCV co-infected patients (P<0.0001). CD4+ T-cell count was lower in patients with co-infection than in those with mono-infection (P<0.01).

Liver stiffness ≥ 12.5 kPa was found in 27.6% of HIV/HCV co-infected cases, with a higher percentage among the males [14/47 (29.7%) males vs. 4/16 (25%) females]. In 2 HIV/HCV co-infected females liver

**Table 1**: Baseline demographic, lifestyle and clinical characteristics of the 194 study patients.

| CHARACTERISTICS | Total n=194 | HIV n=129 | HIV/HCV n=65 | P<  
|----------------|-----------|-----------|---------------|------|
| Age years ± SD | 48.3 ± 8.5 | 48.5 ± 9.4 | 47.7 ± 5.9 | ns  
| Male/Female    | 121/73    | 74/55     | 47/18         | ns   
| BMI (Kg/m²) ± SD | 23.5 ± 3.1 | 23.8 ± 3.4 | 22.8 ± 2.7 | ns   
| Fracture (n, %) | 15 (7.7)   | 7 (5.4)    | 8 (12.3)      | ns   
| Ethnicity (n, %): Caucasian | 181 (93.3) | 119 (92.2) | 62 (95.3)  | ns   
| African        | 13 (6.7)   | 10 (7.7)   | 3 (4.6)       | ns   
| Smoking status (n, %): Non-smokers | 62 (32.0) | 50 (38.7)  | 12 (18.4)  |       
| Smokers        | 97 (50.0)  | 57 (44.2)  | 40 (61.5)     | 0.03 
| Former smokers | 35 (18.0)  | 22 (17.7)  | 13 (20)       |       
| Alcohol (n, %) | 20 (10.3)  | 6 (4.6)    | 14 (21.5)     | 0.01 
| Drug addiction (n, %) | 56 (28.8) | 14 (10.8)  | 42 (64.6)    | 0.0001 
| IFG/Diabetes (n, %) | 45 (23.2) | 28 (21.7)  | 17 (26.1)    |       
| Time since HIV diagnosis (years) (range) | 15 (0.1-29) | 12 (0.1-28) | 18 (0.1-29) | 0.0001 
| Treatment-naive (n, %) | 9 (4.7) | 7 (5.4)    | 2 (3.0)       |       
| Previous antiretroviral therapy (years) (range) | 9 (0.5-26) | 8 (0.5-23) | 13 (0.5-26) | 0.0001 
| Genotype 1/others (n, %) | - | - | 40 (61)       | -    


**Table 2**: Main biological parameters.

| Characteristics | Total n=194 | HIV n=129 | HIV/HCV n=65 | P<  
<table>
<thead>
<tr>
<th></th>
<th></th>
<th></th>
<th></th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>ALT U/L (range)</td>
<td>28.0 (10-175)</td>
<td>24 (10-171)</td>
<td>43 (13-175)</td>
<td>0.0001</td>
</tr>
<tr>
<td>AST U/L (range)</td>
<td>24.0 (10-281)</td>
<td>22 (10-80)</td>
<td>41 (13-281)</td>
<td>0.0001</td>
</tr>
<tr>
<td>Total-cholesterol (mg/dL) ± SD</td>
<td>195.2±51.7</td>
<td>208.8 ± 47.1</td>
<td>165.7 ± 49.9</td>
<td>0.0001</td>
</tr>
<tr>
<td>HDL-cholesterol (mg/dL) ± SD</td>
<td>50.7±17.4</td>
<td>51.7 ± 14.8</td>
<td>46.5 ± 19.3</td>
<td>0.05</td>
</tr>
<tr>
<td>LDL-cholesterol (mg/dL) ± SD</td>
<td>124.2±45.4</td>
<td>133.5 ± 42.8</td>
<td>90.3 ± 36.1</td>
<td>0.0001</td>
</tr>
<tr>
<td>Triglycerides (mg/dL) (range)</td>
<td>132 (35-637)</td>
<td>134 (35-637)</td>
<td>121.5 (52-429)</td>
<td>ns</td>
</tr>
<tr>
<td>Triglycerides/HDL ratio</td>
<td>2.7 (0.5-14)</td>
<td>2.5 (0.5-20)</td>
<td>3.2 (0.7-14)</td>
<td>ns</td>
</tr>
<tr>
<td>Calcium (mg/dL)</td>
<td>9.2±0.51</td>
<td>9.1 ± 0.6</td>
<td>9.2 ± 0.5</td>
<td>ns</td>
</tr>
<tr>
<td>Phosphorus (mg/dL)*</td>
<td>3.2±0.6</td>
<td>3.2 ± 0.6</td>
<td>3.2 ± 0.8</td>
<td>ns</td>
</tr>
<tr>
<td>Parathyroid hormone (pg/ml)*</td>
<td>37.0 (11-362)</td>
<td>37.1 (12-119)</td>
<td>36 (11-362)</td>
<td>ns</td>
</tr>
<tr>
<td>Bone alkaline phosphatase (IU/L)†</td>
<td>46.5 (17.2-170)</td>
<td>46.8 (17.2-116)</td>
<td>46.2 (17.4-170)</td>
<td>ns</td>
</tr>
<tr>
<td>25-Hydroxyvitamin D (ng/mL)</td>
<td>18.4 (3-88.3)</td>
<td>16.7 (3-68)</td>
<td>27.2 (4.8-88.3)</td>
<td>0.0001</td>
</tr>
<tr>
<td>HCV-RNA (IU/mL)=700.000 (n, %)</td>
<td>-</td>
<td>-</td>
<td>41 (63)</td>
<td>-</td>
</tr>
<tr>
<td>CD4+ T-cell count (cells/μL) (range)</td>
<td>480 (34-1338)</td>
<td>533.5 (34-1082)</td>
<td>413.5 (10-1338)</td>
<td>0.01</td>
</tr>
<tr>
<td>Nadir CD4+ T-cell count &lt;200 (cells/μL)</td>
<td>136 (70.1)</td>
<td>88 (68)</td>
<td>49 (75)</td>
<td>ns</td>
</tr>
<tr>
<td>Detectable HIV-RNA (n, %)</td>
<td>54 (27.6)</td>
<td>30 (23.2)</td>
<td>24 (36.9)</td>
<td>ns</td>
</tr>
</tbody>
</table>

HIV: Human Immune Deficiency Virus; HCV: Hepatitis C Virus; ALT: Alanine Aminotransferase; AST: Aspartate Aminotransferase; HDL: High-Density Lipoprotein; LDL: Low-Density Lipoprotein; ns: Not Significant.*Missing in 10 HIV mono-infected and 2 HIV/HCV co-infected patients; °Missing in 6 HIV mono-infected and 3 HIV/HCV co-infected patients; Missing in 5 HIV mono-infected patients.

stiffness assessment was not performed because of decompensated liver disease.

Prevalence of low bone mineral density/osteopenia/osteoporosis

135/194 study patients (69.5%) had BMD values lower than the reference values for a healthy Caucasian population. In detail, significant skeletal disease, i.e., low BMD and frank osteoporosis, was found in 52 (26.8%) and 50 (26%) HIV-infected patients, respectively. Comparison between the two groups showed that overall prevalence of low BMD and osteoporosis in both the lumbar and femoral sites was significantly higher in HIV/HCV co-infected than in mono-infected patients (P<0.04 and P<0.05, respectively) (Table 3).

As regards the analysis of BMD in the single sites, although lumbar and femoral DXA Z- and T-scores were significantly lower in HIV/HCV co-infected than in mono-infected subjects (P<0.02; P<0.05), only in the lumbar site was the prevalence of low BMD and osteoporosis higher in co-infected vs. mono-infected subjects (P<0.03 and P<0.02, respectively) (Table 3).

Correlation between BMD and extent of liver fibrosis

In the HIV mono-infected group no significant correlations were observed between BMD measured with either lumbar or femoral DXA Z- and T-scores and liver fibrosis evaluated by FIB-4. In co-infected patients FIB-4 values negatively correlated either with lumbar and femoral DXA Z-scores (r=-0.32 and -0.33; P<0.01, respectively) or lumbar and femoral DXA T-scores (r=-0.3; P<0.03 and r=-0.31; P<0.01, respectively).

When HIV/HCV co-infected patients were stratified by sex, a significant and negative correlation between extent of liver stiffness and both the lumbar and femoral DXA Z-scores [r=-0.5; (P<0.04)] or lumbar and femoral neck DXA T-scores [r=-0.45 (P<0.05); r=-0.54 (P<0.04), respectively] was found only in the female group (Figures 1 and 2).

<table>
<thead>
<tr>
<th>BONE MINERAL DENSITY</th>
<th>HIV+</th>
<th>HIV/HCV+</th>
<th>P&lt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>Overall (femoral neck or lumbar spine)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Osteopenia (n, %)</td>
<td>56 (44.9)</td>
<td>27 (41.5)</td>
<td>ns</td>
</tr>
<tr>
<td>Low BMD (n, %)</td>
<td>27 (20.9)</td>
<td>23 (33.8)</td>
<td>0.05</td>
</tr>
<tr>
<td>DXA T-score ± SD</td>
<td>-0.3 (-2.5-2.2)</td>
<td>-0.8 (-3.3-1.7)</td>
<td>0.02</td>
</tr>
<tr>
<td>DXA Z-score ± SD</td>
<td>8 (6.2)</td>
<td>7 (10.7)</td>
<td>ns</td>
</tr>
<tr>
<td>DXA T-score</td>
<td>-0.6 (-2.8-1.8)</td>
<td>-1 (-3.6-1.4)</td>
<td>0.02</td>
</tr>
<tr>
<td>Osteopenia (n, %)</td>
<td>47 (36.4)</td>
<td>24 (36.9)</td>
<td>ns</td>
</tr>
<tr>
<td>Osteoporosis (n, %)</td>
<td>3 (2.3)</td>
<td>5 (7.7)</td>
<td>ns</td>
</tr>
<tr>
<td>Lumbar spine</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>DXA T-score ± SD</td>
<td>-0.9 (-3.7-2.7)</td>
<td>-1.3 (-4.5-1.3)</td>
<td>0.02</td>
</tr>
<tr>
<td>DXA Z-score ± SD</td>
<td>27 (20.9)</td>
<td>24 (36.9)</td>
<td>0.03</td>
</tr>
<tr>
<td>Osteopenia (n, %)</td>
<td>-1.3 (-3.9-2.3)</td>
<td>-1.7 (-5.2-1.1)</td>
<td>0.05</td>
</tr>
<tr>
<td>Osteoporosis (n, %)</td>
<td>50 (38.7)</td>
<td>24 (36.9)</td>
<td>ns</td>
</tr>
</tbody>
</table>

Table 3: Bone mineral density measurements of the 194 study patients.

Figures 1 and 2 show the correlation between liver fibrosis, measured by transient elastography and lumbar spine DXA Z- and T-scores, respectively, in HIV/HCV co-infected patients divided according to sex.

Correlation between BMD and other study parameters in both the HIV-groups

Tables 4 and 5 show the correlations between lumbar/femoral DXA Z- and T-scores and the variables analyzed in the two groups by univariate and multivariate analysis.

HIV mono-infected group: BMD negatively correlated with nadir CD4+ <200 (cells/μl) in lumbar/femoral DXA Z-scores (P<0.05; P<0.01, respectively), Bone alkaline phosphatase (P<0.01) and PI score (P<0.02) negatively correlated with femoral DXA Z-score (Table 4).

Lumbar/femoral DXA T-scores negatively correlated with age (P<0.01), bone alkaline phosphatase (P<0.02, respectively), nadir CD4+ <200 (cells/μl) (P<0.03; P<0.01, respectively), and PI score (P<0.02; P<0.01, respectively) (Table 5). In addition, both BMI and HDL-cholesterol levels positively correlated with lumbar DXA Z-score (P<0.01; P<0.05) (Table 4). A positive correlation was also found between BMI and femoral DXA Z-score (P<0.001) (Table 4).

HIV/HCV co-infected group: Bone alkaline phosphatase levels negatively correlated with lumbar/femoral DXA Z-scores (P<0.05;
Among the variables significant at univariate analysis, low BMD was negatively and independently correlated with PI use score in HIV mono-infected patients (β=0.22; P<0.02). No significant independent correlation was found between low BMD and HIV/HCV co-infection. On multiple linear regression analysis an independent and positive correlation was found between BMI and DXA Z-score measured in both sites (lumbar spine and femoral neck) in HIV mono-infected patients (β=0.30; P<0.01; β=0.45; P<0.0001, respectively). Lumbar spine DXA Z-score was also independently correlated with HDL-cholesterol (β=0.22; P<0.02) (Table 4). Lumbar/femoral DXA T-scores were negatively and independently correlated with PI use score in the HIV mono-infected group (β=-0.33; P<0.02; β=-0.50; P<0.0001, respectively) (Table 5).

In HIV/HCV co-infected patients an independent and positive correlation was found between BMI and both femoral DXA Z- and
T-scores (β=0.33; P<0.02; β=0.36; P<0.03, respectively) (Tables 4, 5); in the same group PI score negatively correlated with femoral DXA Z-score (β=-0.32; P<0.02) (Table 4). Extent of liver stiffness was an independent predictor of bone loss in HIV/HCV co-infected patients in the femoral DXA Z- and T-scores (β=-0.43; P<0.003; β=-0.41; P<0.01, respectively). FIB-4 values were negatively and independently correlated with lumbar DXA Z- and T-scores in HIV/HCV co-infected patients (β=-0.43; P=0.004; β=-0.46; P<0.002, respectively).

Discussion

In this cross-sectional study we found a very high prevalence of low BMD and osteoporosis among predominately Caucasian HIV-infected patients with or without chronic hepatitis C, with long-term HIV-infection and prolonged ART. The overall 26% prevalence of frank osteoporosis in our HIV-cohort was higher than the percentages reported in a previous meta-analysis and in other European case studies [1,3,24]. In our HIV/HCV co-infected patients osteoporosis prevalence was significantly higher than among HIV mono-infected subjects, in agreement with previous findings [25]. Many divergences persist regarding the prevalence of osteoporosis in HIV/HCV co-infection [14]. The 33.8% osteoporosis prevalence in our HIV/HCV co-infected patients was higher than the pooled estimate of 22% taken from a recent meta-analysis [14]. Likewise, the prevalence of low BMD in our co-infected patients was higher than the result reported by El-Maouche et al. [15] (36.9% vs. 26%, respectively), as well as being considerably higher than the 16% observed in a large cohort of HIV/HCV co-infected patients from Modena (northern Italy) [26]. Although the proportion of enrolled co-infected males and females and sample size in the two Italian HIV cohorts were similar, other co-morbidities, as well as the heterogeneity of liver disease may account for the discrepancy in the low BMD prevalences.

Although controversial, a direct correlation between extent of liver fibrosis and low bone density has been reported in non-cirrhotic HCV mono-infected patients [27,28]. No association between liver disease severity and BMD at any site was reported in co-infected patients by El-Maouche et al. [15], despite the high histological evidence of cirrhosis (26%). Liver cirrhosis, assessed by liver biopsy or non-invasive methods (FibroScan and FibroTest), was also not found to be associated with decreased BMD in co-infected patients by Lawson-Ayay et al. [25]. In our study the extent of liver fibrosis, measured by transient elastography, was an independent predictor of bone loss in HIV/HCV co-infected patients in the femoral neck DXA Z- and T-scores. Of clinical interest, a significant negative correlation between BMD in both the lumbar spine/femoral neck sites and severity of hepatic fibrosis was found in our HIV/HCV co-infected females, but not males, suggesting that viral hepatitis co-infected women are even at risk for vertebral and femoral fractures. Although we did not assay sexual hormones, the correlation between menopause and lumbar/femoral DXA Z- and T-scores on univariate analysis suggests a role of estrogen deficiency for bone loss in our HIV/HCV co-infected women. This hypothesis is furthermore corroborated by the evidence of liver stiffness ≥ 12.5 kPa in a higher percentage of co-infected males than females (29.7% vs. 25%, respectively), which in addition to the lack of a significant correlation between the extent of liver fibrosis and BMD in the males of the same co-infected group, suggests that bone abnormalities are more likely to be the result of HIV/HCV co-infection and female gender. Among the multifactorial mechanisms underlying skeletal disorder in the HIV and HCV setting, vitamin D deficiency is considered a risk factor for osteoporotic fracture [14]. Vitamin D levels were found to be lower in our HIV mono-infected patients than in subjects with HIV/HCV co-infections. In addition, the median vitamin D level was >30 ng/mL in HIV/HCV co-infected patients. Our results reflect those reported in an Italian study, which found that median vitamin D serum levels >25 ng/mL were similar in the HIV mono-infected, HIV/HCV co-infected and healthy controls [29]. Moreover, in a recent, large study on a general healthy population in Central Europe, Pudlowski et al. [30] reported an average vitamin D concentration of >30 ng/mL. However, since the Hormone Foundation’s Patient Guide to Vitamin D Deficiency suggests that patients with chronic (long-term) liver disease are at high risk of deficiency [31], further randomized controlled trials are warranted to determine what level of vitamin D insufficiency and deficiency places an individual at risk for cirrhosis evolution.

The mechanisms underlying osteoporosis in HIV-infected patients with viral hepatitis have not been completely understood to date. The evidence gathered suggests a close link between bone disease and a state of chronic inflammation [4]. In this respect, the HIV-related immune activation induces cytokine changes that may increase liver inflammation and fibrosis, which in turn may result in an increase in osteoclastic activity, mediated by the osteoclastogenic pro-inflammatory cytokines [4,32,33]. HIV-associated chronic inflammation may also cause endothelial dysfunction, atheroma formation and acute thrombosis, which are determinants of cardiovascular diseases in HIV-infection [34], and this may represent a potential link between bone disease, liver disease and cardiovascular risk in HIV/HCV co-infection. Since cardiovascular risk has been negatively associated to low HDL-cholesterol levels, a relationship between HDL-cholesterol and BMD has been postulated [35]. Our data showed an independent and positive correlation between lumbar spine DXA Z-score and HDL-cholesterol levels in HIV mono-infected patients. The link between lipid abnormalities and low BMD has been little investigated in the HIV-setting [16,36].

Several HIV-related factors negatively correlated with BMD in the lumbar/femoral DXA Z- and T- scores of our mono-infected and co-infected patients on univariate analysis. The correlation between time of HIV diagnosis, as well as nadir CD4+ T-cell count <200 cells/μL and BMD suggest that HIV infection per se and in particular the more advanced stages of the disease might affect the skeletal system. The negative and independent correlation between longer PI exposure and BMD supports earlier evidence on the unfavorable role of PI exposure in the progression of bone loss [12].

Low BMI values were independently correlated with low BMD. These findings support the report in a meta-analysis of 10 studies on HIV-infected patients that low body weight may largely account for the high prevalence of low BMD in the setting of HIV-infection [10].

Some limitations of the present study need to be mentioned. The small size and homogeneity of the sample do not allow a generalization of our results. In addition, the cross-sectional study design does not permit us to establish a cause-effect relationship in our results or to evaluate the impact of liver disease progression on bone metabolism. Our analysis is also limited by the lack of sexual hormone assessment and of the absence of markers of systemic inflammation, which would be useful in evaluating the impact of female sex or inflammation on bone health. Finally, the numerous variables evaluated are potentially confounding and the multivariate analyses may not entirely eliminate the residual confounding factors.

In conclusion, our Caucasian HIV mono- and HIV/HCV co-infected patients appeared at high risk for low BMD and osteoporosis. The extent of liver fibrosis, measured by transient elastography, was
an independent predictor of bone loss in HIV/HCV co-infection. When patients were stratified by sex, a negative correlation between BMD and severity of liver fibrosis was found in co-infected women only, suggesting that other factors (i.e., sexual hormones) may affect the skeletal system in HIV/HCV co-infection more than liver fibrosis. Additional research into the impact of chronic hepatitis C on osteoporosis and the interaction between lipid asset, cardiovascular risk and bone disease effects are required in the HIV infection setting.

Acknowledgements

We would like to thank Dr. Carole Greenall, BA, for the revision of the English text.

References


