Behavior of Hair Follicles in Vitiligo: Clinical Presentation and Discussion
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Abstract
The color of the hair is dependent on the existence and function of melanocytes, the melanin-producing cells, in the hair matrix of the bulb. In vitiligo, the loss of color of the affected skin is due to loss of epidermal melanocytes, which may also be accompanied by loss of the bulbar melanocytes with subsequent loss of the hair color. Lymphocytic immune-cytotoxic process may be involved in the destruction of epidermal or bulbar melanocytes. Hair depigmentation is said to follow surface depigmentation in the disease. This delay in development of leukotrichia in vitiligo could be claimed to the difference in melanization nature of hair follicle in comparison to epidermal melanization process. In rare cases, follicular melanocytes can be primarily destroyed in vitiligo without destruction of epidermal melanocytes. Apart from mature melanocytes, the hair follicles possess also immature form of melanocytes residing in the bulge area, outer root sheath, dermal papilla and sometimes the hair matrix. These immature cells usually escape the vitiligo immune destructive process.

The existence of leukotrichia used to be a sign of disease recalcitrance and expected treatment failure. In such cases several surgical procedures were introduced to help repigmenting the skin.

It was found that activation of the immature melanocytic precursors in hair follicles is responsible for the repigmentation of epidermis and hair follicles due to the supply with new mature melanocytes.

Keywords: Vitiligo; Hair follicle; Melanocytes; Stem cells

Biology of Hair Coloring
The hair bulb is the only site of pigment production for the hair shaft and contains highly melanogenic melanocytes and a minor subpopulation of poorly differentiated pigment cells. In the adult hair follicle, pigmentation results from interactions between follicular melanocytes, matrix keratinocytes, and Dermal Papilla (DP) fibroblasts.

The "follicular-melanin unit" is residing in the "immune privileged" proximal hair bulb. This follicular-melanin unit consists of one melanocyte for every five keratinocytes in the hair bulb as a whole, the ratio is 1:1 in the basal epithelial layer next to the dermal papilla. Hair bulb melanogenic melanocytes differ from epidermal melanocytes in being larger, with longer and more extensive dendrites, producing two- to four-fold larger melanosomes (Figure 1). Melanocytes of the hair bulb are known to have potential proliferation power and are not as indolent as epidermal melanocyte lacking completely this power of renewal. The hair bulb melanocyte system has been perceived as self-perpetuating whereby melanocytes involved in the pigmentation of one generation are also involved in the pigmentation of the next wave. Follicular melanogenesis is characteristically cyclic in nature, as opposed to the continuous melanogenesis of epidermal pigmentation.

The increase in number of "white" hairs with or without vitiligo is time dependent due to extrinsic and intrinsic elements affecting the follicular melanocytes as well as epidermal melanocytes. Loss (mostly probably via apoptosis) may occur in both sun-exposed and covered skin with 10% reduction per decade after 30 years of age until 80 years, followed by more dramatic cell loss thereafter. Again, oxidative stress can affect follicular melanocytes over time and may be a major factor in the loss of hair pigment, the reduction in overall numbers of melanocytes per follicle, and ultimately for induction of white hairs.

Vitiligo and the Follicle
In Vitiligo, destruction and loss of melanocytes from the skin epidermis is the cause of leukodermia. This destructive process may occasionally affect the active bulbar melanocytes too leading to the development of leukotrichia (Figure 2). T-lymphocytes are well known pathological element during the development of vitiligo, expressing cytotoxic/apoptotic factors to induce melanocytic destruction. These lymphocytes were also demonstrated perifollicularly in considerable density during disease activity in biopsies including black hairs within vitiligo patches. This density of perifollicular infiltrate was not found in cases with white hair follicles lacking the targeted antigen.

Induction of Leukotrichia
Distinct compartmentalization of skin melanocyte sub-populations

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can be evidenced clinically by the selective or preferential targeting of epidermal melanocytes (but not follicular melanocytes) in vitiligo [5,13].

Hair depigmentation usually follows surface depigmentation in long standing disease. Hair bulb melanocytes are said to be targeted first only in acute alopecia areata, whereas poliosis develops later in vitiliginous patches because of the loss of melanocytes from hair bulbs after the primary immunologic destruction of epidermal melanocytes [13].

This delay in vitiligo follicular depigmentation could be claimed to the stable condition of melanin contents in the follicles, whereas in the epidermis, the release of melanosomes from the melanocyte is followed by their transport to keratinocytes and subsequent exfoliation and biodegradation of melanin. The functioning melanocytes keep the equilibrium between release and exfoliation of melanin in normal situations. With the loss of melanocytes in the epidermis this equilibrium is lost too with subsequent biodegradation and exfoliation of melanin through epidermis. In hair follicles, melanin does not exfoliate or biodegrade as fast as within the epidermis. While melanin degrades almost completely in the differentiating layers of the epidermis, eumelanin granules transferred into hair cortical keratinocytes remain minimally digested [5].

Again, being within immune privileged hair bulb, the follicular-melanin unit can escape the autoimmune cytotoxic process which destroyed other epidermal melanocytes.

All these points may explain why hair can remain pigmented even during vitiligo activity with the considerable lymphocytic dermal or perifollicular infiltration.

Long standing disease duration is not the trigger for induction of poliosis as the development of follicular depigmentation is not a must sequence after certain duration of vitiligo. Neither the severity of disease extension nor duration of the vitiligo can be considered cause, as many vitiligo patients can retain the hair color even after many years of the disease (Figure 3).

Also, the hypothesis that hair color loss only follows the loss of skin pigment is not overall, although a common sequence, as some vitiligo patients can present with merely leukotrichia over normal looking pigmented skin surface with persistence of epidermal melanocytic function during the course of their disease (Figure 4). This means that mature bulbar melanocytes can be primarily destroyed in vitiligo without destruction of epidermal melanocytes.

Fully differentiated bulbar melanocytes undergo apoptosis during catagen [14], a similar process of destruction is expected during active immune process in vitiligo. Still, some less differentiated hair bulb melanocytes appear to survive catagen [7], and may possibly survive this immune/cytotoxic vitiliginous effects and be capable of differentiation and maintaining hair color within active vitiliginous patches.

**Follicular Melanocytes Reservoir**

The hair follicles contain several kinds of melanocytes, among them the non-mature amelanotic cells. Melanocyte stem cells survive catagen and are involved in restoration of the HF pigmentary unit after the HF enters into new anagen [7,15,16].

The "re-differentiating" melanocytes of early anagen are likely newly recruited immature melanocytes derived from a melanocyte reservoir located in the upper, "permanent," outer root sheath. This view is supported by the observation that melanocyte stem cells located at the base of the permanent part of hair follicle are not only immature, but also slow cycling, self-maintaining and are fully competent to...
amelanotic bipolar sub-population [9]. This detected proliferation of melanocytes, less-differentiated tripolar cells, and an undifferentiated distinct sub-populations, including highly pigmented dendritic bulbar melanocytes in culture by Kauser et al. stating that HFMs may show at least three reaction. These forms of follicular melanocytes were also described in vitiliginous skin after ultraviolet radiation [22]. These cells were demonstrated in the infundibular and isthmus area of hair follicles [1].

Melanocyte Related Proteins in Hair Follicle

Several proteins and receptors were described and correlated to the melanogenesis process in the hair follicles. The famous c-Kit is required during the hair growth cycle for activation of melanocytes, although in stem cell compartment appears to exhibit stem cell factor SCF/c-Kit independence [15]. Some amelanotic dopa-negative melanocytes may also be distributed in the periphery of the bulb and the most proximal matrix [1,19]. All the dopa-positive cells, and also some dopa-negative melanocytes of the mid outer root sheath are (pre)melanosome gp100 positive [19]. Amelanotic hair follicle melanocytes are devoid of dopa-oxidase activity, although low levels of the tyrosinase protein itself may be detected in some cells. Similarly, c-Kit and Bcl-2 reactive amelanotic melanocytes are present in this hair follicle compartment [20], but do not express the melanogenic enzymes TRP (Tyrosinase-Related Protein)1 and TRP2 [19].

Yet, Slominski et al. stated that melanocytes localized to the hair follicle bulge (site of presumptive reservoir) express only TRP2, lacking TRP1, c-kit, and K667 immunoreactivities. Melanocytes from the fully developed anagen V1 hair follicles express TRP2 only when located in the hair bulge. TRP2 and c-Kit only when located in the outer root sheath, and all three proteins, together with c-Kit, when located in the hair bulb matrix [21].

Stimulation of Melanocyte Stem Cells

New population of amelanotic spindle cells were clearly demonstrated in the infundibular and isthmus area of hair follicles in vitiligious skin after ultraviolet radiation [22]. These cells were presumed stimulated melanocyte precursors and showed bipolar and tripolar morphology with minimal silver staining and absent Dopa reaction. These forms of follicular melanocytes were also described in culture by Kauser et al. stating that HFMs may show at least three distinct sub-populations, including highly pigmented dendritic bulbar melanocytes, less-differentiated tripolar cells, and an undifferentiated amelanotic bipolar sub-population [9]. This detected proliferation of melanocytic precursors preceded the characteristic perifollicular pigmentation, demonstrating the relevance of follicles during vitiligo repigmentation (Figure 5).

Such stimulation of melanocyte reservoir in the bulge areas was described before by Starrico who demonstrated amelanotic melanocytes in the ORS of hair follicles and proved that after phototherapy or mechanical removal of skin epidermis those immature cells start to proliferate, migrate and be stimulated [23-25].

Hair Follicle and Vitiligo Regression

The famous repigmentation pattern in vitiligo usually begins in the perifollicular area and treatment failure is expected in patches with leukotrichia. The presence of white hairs within vitiligo patches used to be a sign of recalcitrance and a poor prognostic signal [26,27].

The difficulty in pigmenting non-hairy glabrous skin such as on palms is well known, because of the absence of a melanocyte reservoir due to lack of hair follicles. In addition to hair bearing skin, with totally depigmented terminal hairs, which is less likely to respond due to the destruction of the melanocyte reservoir [28].

Again, this hypothesis is not a sharp indication for treatment failure as some vitiligo cases responded to phototherapy (Figure 6) in spite of the presence of poliosis, although others may fail to repigment.

Surgical Procedures for Vitiligo

The introduction of surgical procedures was mandated in such difficult to treat cases of vitiligo with poliosis. Hair grafting was described to treat resistant areas aiming to provide new generation of melanocytes and precursors. Only the upper two thirds of the follicle was used and grafted when glaborous skin is targeted to avoid unwanted growth of coarse terminal hair. Successful surface repigmentation was attained after hair grafting [27].

Refilling of the melanocyte reservoir of the hair follicle by retrograde migration was described during treatment of patients with stable vitiligo lesions with depigmented hairs and expected depletion of the melanocyte reservoir in the hair follicle using transplantation of autologous cultured melanocytes in fibrin suspension [29].
in the outer root sheath corresponded with the hypothesis that c-Kit surface epidermal c-kit staining was not obtained yet staining of cells (MSCs) residing possibly also in hair follicles [30]. Although stable patches of vitiligo through stimulation of Melanocyte Stem Cells (MSCs) [15,21,30]. Subsequently, repigmentation of leukotrícia in vitiligo patches after dermabrasion maybe expected too and should be examined.

Epilation of white hairs was advised in a study to accelerate the hair cycle, and new black hairs may emerge in new anagen phase [27].

Figure 6: In spite of the loss of follicular color (leukotrícia), successful patchy repigmentation of vitiligo was obtained in this case after phototherapy.

Figure 7: Child with vitiligo patch on forehead and scalp with manifested leukotrícia dyed with Henna as camouflage (Left Panel). After Epithelial grafting majority of hair follicles successfully repigmented following surface skin repigmentation and persisted for several years of follow up (Right Panel).

Hair Repigmentation

Interestingly, epithelial grafting on dermabraded vitiligious skin with poliosis successfully repigmented the recalcitrant surface skin and also repigmented the white hair follicles afterward which persisted for several years (Figure 7). The duration required for hair follicles to repigment was not constant for all cases as was the surface skin but varied from 4 to 11 months after the procedure and this was claimed to different anagen durations in different sites of the body [23]. Retrograde migration theory was not the explanation here rather a stimulation of melanocyte reservoir in the bulge areas after dermabrasion and phototherapy following, in accordance to old previous reports demonstrating that mechanical removal of skin epidermis can stimulate those immature cells to start to proliferate and migrate [23-25].

Cells positively stained for c-kit were demonstrated in the outer root sheath of hair follicles after dermabraded vitiligious skin in another study where dermabraded proved to help repigmenting stable patches of vitiligo through stimulation of Melanocyte Stem Cells (MSCs) residing possibly also in hair follicles [30]. Although surface epidermal c-kit staining was not obtained yet staining of cells in the outer root sheath corresponded with the hypothesis that c-Kit is required mainly for hair supply rather than epidermal supply of MSCs [15,21,30]. Subsequently, repigmentation of leukotrícia in vitiligo patches after dermabrasion maybe expected too and should be examined.

The destructive process during catagen stage or during vitiligo immune-cytotoxic activity affects only mature functioning cells, while other immature cells should escape this slaughter.

Stimulation of these follicular melanocytic precursors can lead to their maturation, differentiation and migration to supply new anagen follicles and surface epidermis with new functioning melanocytic population.

References


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