

Biological Wonders of Osmolytes: The Need to Know More

Eva Judy and Nand Kishore*

Department of Chemistry, Indian Institute of Technology-Bombay, Powai, Mumbai, India

Abstract

Nature has selected osmolytes to protect intracellular macromolecules against denaturing stress conditions. These molecules are accumulated in the intracellular environment at considerably high concentrations. In general, osmolytes are known to stabilize proteins. However, under certain conditions, their destabilizing properties have also been pointed out. A careful qualitative and quantitative understanding of the mechanism of action of osmolytes with proteins from native to different stages of aggregation/fibrillation is extremely important in rational drug design. This review highlights the importance of naturally occurring osmolytes in protein folding, stabilization, and prevention of fibrillation/aggregation related diseases among others. Continued efforts are required to get quantitative insights into osmolyte-protein interactions along with experimental evidences for the much claimed preferential exclusion/preferential hydration phenomenon of osmolyte action. Mechanistic insights into the disease associated roles of osmolytes needs special attention.

Keywords: Osmolytes; Protein stability; Fibrillation/aggregation; Preferential hydration; Rational drug design

Introduction

Osmolytes have been fascinating to biochemists as these molecules are small solutes which manage cell volume regulation under water stress conditions. Such conditions arise due to extremes of temperature, pressure, alterations in extracellular osmotic conditions and even urea which is a protein denaturing osmolyte [1-4]. There are three major classes of organic compounds which are considered as cellular osmolytes (Figure 1) [2]. These can be further categorized into (i) osmolytes that stabilize proteins raising free energy of both the native and denatured states: trimethylamine N-oxide (TMAO), sarcosine, sorbitol, sucrose, and trehalose, (ii) osmolytes that only moderately change protein stability: glycine betaine, proline, and glycerol, (iii) denaturing osmolytes: urea, and (iv) counteracting osmolytes: mixture of urea and TMAO. Modulation of activity of molecular chaperons (heat shock proteins) due to promotion of local refolding in protein molecules has also been observed [5] which points out to linkage of chemical chaperons (osmolytes) and molecular chaperones in *in vivo* regulation of protein folding.

Significant efforts have been dedicated in understanding solution thermodynamics of protein-osmolyte mixtures [6-15]. Most of the experimental work which has led to the calculation of preferential interaction parameters is based on density measurements or thermal transition temperature of the protein by using differential scanning calorimetry or spectroscopic methods [16,17]. These experimentally determined preferential interaction parameters have been used to address the mode of action of osmolytes on proteins. Based on the free energy of transfer of protein backbone models from water to aqueous osmolyte solutions, the cause of stabilization/destabilization of the unfolded state has been suggested [6,9,10]. Molecular mechanism for osmolyte induced protein stability has been discussed extensively in literature [11,18], though their mode of action in their different roles is still not completely understood.

This mini review focuses on the mode of action of osmolytes on proteins, disease associate roles, and DNA associated effects. The unanswered questions and need for further experimental evidences on action of osmolytes in their different roles has been discussed.

Known Mode of Action on Proteins

Osmolytes are known to alter the chemical potential of proteins

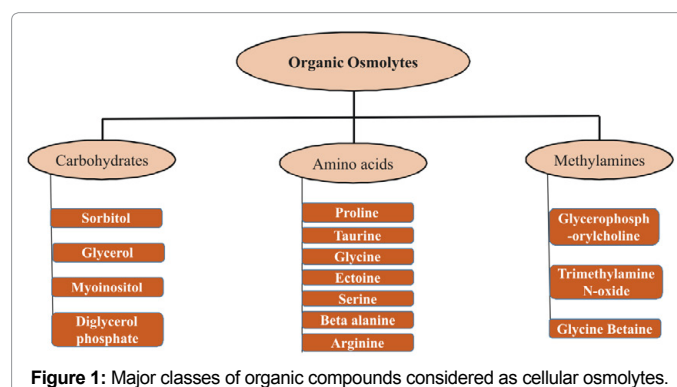


Figure 1: Major classes of organic compounds considered as cellular osmolytes.

in the native and the unfolded states to different extents [19-22]. The difference arises due to competing protein-water and protein-osmolyte interactions [23]. One of the widely used models to explain osmolyte driven protein folding and stabilisation is based on determining free energies of interaction between osmolytes and building blocks of proteins such as amino acids or peptides [24]. This model is based on the principle that the osmolytes undergo unfavourable interactions with the peptide backbone in the unfolded state of the protein which leads to strengthening of its secondary structure (an intermediate structure in the folding pathway) before attaining the folded conformation [7,25].

Gibbs free energy implications of action of osmolytes have been considered in understanding mechanism of stabilization of proteins. It is believed that preferential exclusion of osmolytes increases the standard Gibbs free energy change accompanying unfolding of the protein. It is suggested that stabilization of proteins by protecting osmolytes is not due to stabilization of the native state, rather it

*Corresponding author: Nand Kishore, Department of Chemistry, Indian Institute of Technology-Bombay, Powai, Mumbai, India, Tel: 91-22-2576-7157; E-mail: nandk@chem.iitb.ac.in

Received: September 15, 2016; Accepted: December 12, 2016; Published December 15, 2016

Citation: Judy E, Kishore N (2016) Biological Wonders of Osmolytes: The Need to Know More. Biochem Anal Biochem 5: 304. doi: 10.4172/2161-1009.1000304

Copyright: © 2016 Judy E, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

arises principally from the destabilization of the unfolded state of the protein by raising its Gibbs free energy than that of the native state [26,27]. Osmolytes shift the equilibrium towards the native conformation of protein by increasing the free energy of the unfolded state. It is hypothesised that the osmolyte sequester the water molecules from the protein surroundings thus creating a hydrophobic environment which causes the protein to fold more compactly, thus driving the equilibrium to functionally active conformation [28,29].

Extensive experimental observations on osmolytes-protein interactions have been explained in terms of preferential hydration/preferential interaction phenomenon by Timasheff, Shellman and other researchers [8,16,30-33]. The ability of organic osmolytes to stabilize and protect intracellular proteins under denaturing environmental stresses has been demonstrated by different experiments [2,34,35]. Osmolytes exert a dramatic influence on protein folding process without making new or breaking existing covalent bonds. Protein stabilization is ability of the osmolytes to push the native (N) = denatured (D) equilibrium towards left. An increase in the thermal unfolding transition temperature up to 22°C has been reported for ribonuclease A in the presence of 8.2 M sarcosine which translates to 45000 fold increase in stability of the native form of the protein over that in the absence of the osmolyte. Similarly, the osmolytes led to an increase in the thermal unfolding temperature of lysozyme up to 23°C [36]. Exceptional increase in the thermal stability of lysozyme up to 26.4°C and myoglobin up to 31.8°C was obtained in the presence of hydroxyproline [17].

The increase in the thermal stability of proteins or refolding of unfolded polypeptides by osmolytes also permits the latter to be classified as “chemical chaperones”. Regulation of molecular chaperones (Heat Shock Proteins or HSP’s) *in vitro* and in cells by chemical chaperones probably by promoting local refolding within chaperone protein molecules under combined salt and heat stresses is well documented in literature [5]. This suggests a link between the chemical and molecular chaperones in supporting protein folding in cells. [28]

Disease Associated Roles of Osmolytes

Role in immunological processes

Several key immunological processes are regulated by osmolytes. Specific examples include immunoglobulin assembly and folding, immune cell proliferation, immune cell function regulation, inflammatory response and also protection against photo-immunosuppression [37]. Therefore osmolytes based therapeutic strategies in the treatment of several immunological disorders needs a special attention.

Role in cancer

Specific inhibitory effect of a 240s plasma exposure in the presence of osmolytes against T98G brain cancer cells has been observed without affecting HEK normal cells [38]. The use of non-thermal plasmas has increased recently in the treatment of living tissues [39-43]. It is observed that sucrose, glycerol and TMAO exhibit inhibitory effect on T98G brain cells only. The importance of work can further be realised if we understand the mechanism of action of osmolytes on these cell lines. For example, it will be important to understand whether the properties of cell lines are changed due to interaction with osmolytes or plasma leads to alteration in the properties of osmolytes.

Role in kidney related diseases

The role of organic osmolytes (such as glycine betaine, myoinositol,

sorbitol, and glycerophosphoryl) in human and other mammalian kidneys has also been reported [44]. The concentration of osmolytes was observed to be different in inner medulla and cortex tissue samples [44]. Myoinositol, sorbitol, and glycine betaine have been found to be components of human urine [45,46]. These results strongly point out physiological importance of organic osmolytes in humans and need further investigation in understanding a general mammalian osmoprotectant strategy.

Role in cardiovascular risk factors and urinary excretion

Deficiency of the osmolyte betaine is a possible cardiovascular risk factor [47], and its urinary excretion is increased in diabetes. It is reported that almost 30% of patients affected with diabetes have unusually higher levels of urinary betaine excretion [48]. The correlation of the osmolyte deficiency with fibrates treatment can help in designing proper dietary intake supplements for patients. The contribution of betaine to transmethylation of homocysteine to methionine is reported to be important since BHMT1 pathway is a major route for the elimination of monocysteine which has a role in the development of cardiovascular disease [49]. Thus betaine is an important nutrient in the prevention of chronic disease [50]. Natural osmolyte trimethylamine *N*-Oxide (TMAO) has been shown to correct assembly defects of mutant branched-chain α -ketoacid decarboxylase in maple syrup urine disease [51]. TMAO has also been recently described as risk factor for cardiovascular disease and the mechanism of its accumulation in hemodialysis patients has been discussed [52]. Glycine betaine and proline betaine are present as osmoprotectants in urine. They act as potents for inhibiting growth of bacteria and thus in the treatment of urinary tract infections [46].

Role in lungs and skin related issues

Several antimicrobial substances which kill constantly deposited bacteria in the lungs are present in the thin layer of airway surface liquid. The role of osmolyte xylitol in enhanced killing of such bacteria and hence in prevention of onset of bacterial infection in cystic fibrosis has been hypothesized [53]. It is suggested that delivery of xylitol to airway surface may lead to enhancement of innate bacterial defence system. The effect of glycerol and urea have been explored on permeability of excised skin membranes [54]. It was observed that these two osmolytes (glycerol and urea) penetrate the skin membrane and retain skin permeability characteristics even at low water activity. Being chemical chaperone, the increased uptake of osmolytes by uv-irradiated keratinocytes was correlated with their defence strategy against detrimental effects of such irradiations [55]. The role of taurine in prevention of surfactant induced dry and scaly skin by modulation of proinflammatory response and stimulation of epidermal lipid synthesis has also been reported [56]. *Staphylococcus aureus* (*S. aureus*) is a major cause of skin and soft tissue infections. Osmolyte transport in *Staphylococcus aureus* and its role in pathogenesis have recently been described [57].

Role in prevention of aggregation/fibrillation of protein

Osmolytes have also found important role in the prevention of fibrillation/aggregation of proteins. They can be utilized as therapeutic targets for diseases mainly related to protein misfolding. Protein fibrillation is responsible for several amyloidogenic disorders including diseases such as Alzheimer’s, Parkinson’s, mad cow, diabetes type II, cystic fibrosis, and dialysis related amyloidosis [58,59]. Though there have been several studies describing the effects of osmolytes on fibrillation/aggregation of proteins [60-66], quantitative understanding in terms of energetics of interaction has only recently been addressed

[67,68]. Trehalose has been currently used for the treatment of Huntington's disease in transgenic animal mice [69]. Such studies allow identification of functional groups on potential inhibitors of fibrillation/aggregation and hence in deriving guidelines for novel drug synthesis.

Role in intrinsically disordered proteins (IDPs)

Many domains or regions in proteins are intrinsically disordered (ID) under native conditions. Such ID proteins or IDPs are found disproportionately in cell signalling proteins and transcription factors. These regions in signalling proteins tend to promote molecular recognition by binding specific protein partners [70,71]. The effect of osmolytes on IDPs is known to be opposite to that of globular proteins. For instance, several types of osmolytes induce aggregation/fibrillation in intrinsically disordered protein α -synuclein which is associated with Parkinson's disease [72-74]. Other intrinsically disordered proteins which undergo aggregation/fibrillation by some osmolytes include tau protein [75], the prion protein [76], Alzheimer's amyloid β -peptides [77,78] and glucagon hormone peptide [64].

Osmolytes and DNA

The destabilization of DNA by osmolytes has also been observed [79] and role in modulating protein-DNA interactions has been discussed [80]. For example, mitigation of the binding of ERG1 to DNA in a differential manner has been reported. The mechanism proposed involved interaction of osmolytes with the DB domain of ERG1 instead of its conjugate DNA. The negative modulation of ERG1-DNA interaction can have important therapeutic implications. The role of osmolytes in the regulation of biological activity of transcription factors needs to be seriously examined. The effect was observed to be concentration and/or solvent condition dependent. Potential of osmolytes in trapping DNA-protein binding reactions with natural osmolytes has been reported [81]. The reason for this trapping has been assigned to slowing down of the rate of dissociation of the complex of the nucleic acid with the protein.

Even though we understand the effect of osmolytes on the overall conformation of protein, the mechanism of the osmolytic effect has mostly been attributed to preferential hydration or preferential exclusion phenomenon [82]. In general, contrasting theories of direct interaction mechanism [83-86] and indirect mechanism [82,87-90] have been proposed. Synergy in osmolyte mixtures has potential applications in medicinal and agricultural fields. It is important to understand whether the counteraction of chemical denaturing stress by osmolytes is due to synergy between additive molecules or due to direct interaction with the protein. Thus the counteraction mechanism needs extensive experimental and theoretical proof [91]. Experimental proofs for these direct or indirect mechanism are lacking and there is a need to focus more in this direction.

Conclusion and Future Perspectives

The important role of osmolytes not only in the counteraction of stress conditions for proteins, but the disease associated roles require a thorough understanding of the related mode of action. It will be important to know if the known preferential exclusion phenomenon is able to explain all the observed effects or the mode of action changes depending upon the role of the osmolyte. There is a still lot more to be done to understand the effect of osmolytes on protein conformation, fibrillation and many other processes specifically quantitatively. This requires extensive experimental approaches which can help in establishing whether the mechanism of action of osmolytes on proteins in the native, denatured, and fibrillar/aggregated state is direct,

indirect, or a combination of the two processes. Establishing nature of interactions with the protein at different stages from nucleation to fibrillation can possibly provide more information on the mechanism of action of osmolytes under such conditions.

The mode of action of osmolytes on DNA has been not been addressed to the extent as it has been done for the proteins. The questions which still need to be completely understood are as to whether the protein models with respect to osmolytes can also be extended to nucleic acids or not. Further experimental investigations on osmolytes-nucleic acids interactions, especially addressing the energetics of interactions, can perhaps throw more light on the commonality shared by proteins and nucleic acids with respect to the mode of action of osmolytes on these biological macromolecules.

A thorough understanding of these mechanisms can lead to development of osmolytes as effective therapeutic molecules and hence rational drug design for the prevention and cure of diseases which result due to protein misfolding/fibrillation/aggregation among other factors. Thus extensive efforts are needed in complete understanding of these biological wonders of osmolytes.

References

1. Yancey H, Somero GN (1979) Counteraction of urea destabilization of protein structure by methylamine osmoregulatory compounds of elasmobranch Fishes. *Biochem J* 183: 317-323.
2. Yancey PH, Clark ME, Hand SC, Bowler RD, Somero GN (1982) Living with water stress: evolution of osmolyte systems. *Science* 217: 1214-1222.
3. Somero GN (1986) Protons, osmolytes, and fitness of internal milieu for protein function. *Am J Physiol* 251: 197-213.
4. Yancey PH (2001) Water stress, osmolytes and proteins. *Amer Zool* 41: 699-709.
5. Eliahu N, Rosenthal D, Golubinioff P (2001) Chemical chaperones regulate molecular chaperones in vitro and in cells under combined salt and heat stresses. *J Biol Chem* 276: 39586-39591.
6. Auton M, Bolen WD (2005) Predicting the energetics of osmolyte-induced protein folding/unfolding. *Proc Natl Acad Sci USA* 102: 15065
7. Bolen DW, Baskakov IV (2001) The osmophobic effect: natural selection of a thermodynamic force in protein folding. *J Mol Biol* 310: 955-963.
8. Lee JC, Timasheff SN (1981) The stabilization of proteins by sucrose. *J Biol Chem* 56: 7193-7201.
9. Shellman JA (2002) Fifty years of solvent denaturation. *Biophys Chem* 96: 91-101.
10. Auton M, Bolen DW (2004) Additive transfer free energies of the peptide backbone unit that are independent of the model compound and the choice of concentration scale. *Biochemistry* 43: 1329-1342.
11. Liu Y, Bolen DW (1995) The peptide backbone plays a dominant role in protein stabilization by naturally occurring osmolytes. *Biochemistry* 34: 12884-12891.
12. Timasheff SN (1992) Water as ligand: preferential binding and exclusion of denaturants in protein unfolding. *Biochemistry* 31: 9857-9864.
13. Makhatadze GI, Privalov PL (1992) Protein interactions with urea and guanidinium chloride. A calorimetric study. *J Mol Biol* 226: 491-505.
14. Felitsky DJ, Cannon JG, Capp MW, Hong J, Wynsberghe AWW, et al. (2004) The exclusion of glycine betaine from anionic biopolymer surface: why glycine betaine is an effective osmoprotectant but also a compatible solute. *Biochemistry* 43: 14732-14743.
15. Fonin AV, Uversky VN, Kuznetsova IM, Turoverov KK (2016) Protein folding and stability in the presence of osmolytes. *Biofizika* 61: 222-230.
16. Timasheff SN (1993) The control of protein stability and association by weak interactions with water: how do solvents affect these processes?. *Annual Rev Biophys Biomol Str* 22: 67-97.
17. Kar K, Kishore N (2007) Enhancement of thermal stability and inhibition of protein aggregation by osmolytic effect of hydroxyproline. *Biopolymers* 87: 339-351.

18. Street TO, Bolen DW, Rose GD (2006) A molecular mechanism for osmolyte-induced protein stability. *Proc Natl Acad Sci* 38: 13997-14002.
19. Annunziata O, Asherie N, Lomakin A, Pande J, Ogun O, et al. (2002) Effect of polyethylene glycol on the liquid-liquid phase transition in aqueous protein solutions. *Proc Natl Acad Sci* 99: 14165.
20. Rodríguez-Ropero F, Van Der Vegt NFA (2015) On the urea induced hydrophobic collapse of a water soluble polymer. *Phys Chem Chem Phys* 17: 8491-8498.
21. Knowles DB, Shkel IA, Phan NM, Sterneke M, Lingeman E, et al. (2015) Chemical interactions of polyethylene glycols (PEGs) and glycerol with protein functional groups: Applications to effects of PEG and glycerol on protein processes. *Biochemistry* 54: 3528-3542.
22. Schneck E, Horinek D, Netz RR (2013) Insight into the molecular mechanisms of protein stabilizing osmolytes from global force-field variations. *J Phys Chem B* 117: 8310-8321.
23. Ben-Naim A (2013) *Statistical thermodynamics for chemists and biochemists*, First Edition 1992, Springer.
24. Auton M, Bolen DW (2007) Application of the transfer model to understand how naturally occurring osmolytes affect protein stability. *Methods Enzymol* 428: 397-418.
25. Holthauzen LM, Rösger J, Bolen DW (2010) Hydrogen bonding progressively strengthens upon transfer of the protein urea-denatured state to water and protecting osmolytes. *Biochemistry* 49: 1310-1318.
26. Qu Y, Bolen CL, Bolen DW (1998) Osmolyte-driven contraction of a random coil protein. *Proc Natl Acad Sci* 95: 9268-9273.
27. Wang A, Bolen DW (1997) A naturally occurring protective system in urea-rich cells: Mechanism of osmolyte protection of proteins against urea denaturation. *Biochemistry* 36: 9101-9108.
28. Khan SH, Ahmad N, Ahmad F, Kumar R (2010) Naturally occurring organic osmolytes: From cell physiology to disease prevention. *IUBMB Life* 62: 891-895.
29. Burg MB (1996) Coordinate regulation of organic osmolytes in renal cells. *Kidney Int* 49: 1684-1685.
30. Bhat R, Timasheff SN (1992) Steric exclusion is the principal source of the preferential hydration of proteins in the presence of polyethylene glycols. *Protein Sci* 1: 1133-1143.
31. Shellman JA (2003) Protein stability in mixed solvents: A balance of contact interaction and excluded volume. *Biophys J* 85: 108-125.
32. Schellman JA (1987) Selective binding and solvent denaturation. *Biopolymers* 26: 549-559.
33. Shellman JA (1997) Temperature, stability, and the hydrophobic interaction. *Biophys J* 73: 2960-2964.
34. Clark ME (1985) The osmotic role of amino acids: discovery and function. In *transport processes, ion and osmoregulation*. 412-423.
35. Somero GN (1986) Protons, osmolytes, and fitness of internal milieu for protein function. *Am J Physiol* 251: 197-213.
36. Santoro MM, Liu Y, Khan SMA, Hou LX, Bolen DW (1992) Increased thermal stability of proteins in the presence of naturally occurring osmolytes. *Biochemistry* 31: 5278-5283.
37. Tarun K, Manisha Y, Laishram RS (2016) Role of osmolytes in regulating immune system. *Curr Pharm Des* 22: 3050-3057.
38. Kaushik NK, Attri P, Kaushik N, Choi EH (2013) A preliminary study of the effect of DBD plasma and osmolytes on T98G brain cancer and HEK non-malignant cells. *Molecules* 18: 4917-4928.
39. Chutsirimongkol C, Boonyawan D, Polnikorn N, Techawatthanawisan W, Kundilokchai T (2011) Non-Thermal Plasma for Acne Treatment and Aesthetic Skin Improvement. *J Eur Acad Derma Venere* 25: 1-11.
40. Vandamme M, Robert E, Lerondel S, Sarron V, Ries D, et al. (2012) ROS implication in a new antitumor strategy based on non-thermal plasma. *Int J Cancer* 130: 2185-2194.
41. Noriega E, Sharma G, Laca A, Diaz M, Cong MG (2011) Cold atmospheric gas plasma disinfection of chicken meat and chicken skin contaminated with *Listeria innocua*. *Food Microbiol* 28: 1293-1300.
42. Hirst AM, Frame FM, Arya M, Maitland NJ, O'Connell D (2016) Low temperature plasmas as emerging cancer therapeutics: the state of play and thoughts for the future *Tumour Biol*. 37: 7021-7031.
43. Attri P, Venkatesu P, Kaushika N, Choi EH (2012) TMAO and sorbitol attenuate the deleterious action of atmospheric pressure non-thermal jet plasma on a-chymotrypsin. *RSCAdv* 2: 7146-7155.
44. Sizeland PCB, Chambers ST, Lever M, Bason LM, Robson RA (1993) Organic osmolytes in human and other mammalian kidneys. *Kidney International* 43: 448-453.
45. Lentner C (1981) *Geigy Scientific Tables* (8th edn), New Jersey, Ciba-Geigy 1: 84.
46. Chambers ST, Kunin CM (1987) Isolation of glycine betaine and proline betaine from human urine. Assessment of their role as osmoprotective agents for bacteria and the kidney. *J Clin Invest* 79:731-737.
47. Lever M, McEntyre CJ, George PM, Slow S, Elmslie JL, et al. (2010) Extreme urinary betaine losses in type 2 diabetes combined with bezafibrate treatment are associated with losses of dimethylglycine and choline but not with increased losses of other osmolytes. *Cardiovasc Drugs Ther* 28: 459-468.
48. Lever M, Slow S (2010) The clinical significance of betaine, an osmolyte with a key role in methyl group metabolism. *Clin Biochem* 43: 732-744.
49. Kepmson SA, Zhou Y, Danbolt NC (2014) The betaine/GABA transporter and betaine: roles in brain, kidney, and liver. *Front Physiol* 5: 159.
50. Craig SA (2004) Betaine in human nutrition. *Am J Clin Nutr* 80: 539-549.
51. Song JL, Chuang DT (2001) Natural osmolyte trimethylamine N-oxide corrects assembly defects of mutant branched-chain α -ketoacid decarboxylase in maple syrup urine disease. *J Biol Chem* 276: 40241-40246.
52. Hai X, Landeras V, Dobre MA, DeOreo P, Meyer tw, et al. (2015) Mechanism of prominent Trimethylamine OXide (TMAO) accumulation in hemodialysis patients. *PLoS ONE* 10: 0143731.
53. Zabner J, Seiler MP, Launspach JL, Karp PH, Kearney WR, et al. (2000) The osmolyte xylitol reduces the salt concentration of airway surface liquid and may enhance bacterial killing. *Proc Natl Acad Sci USA* 97: 11614-11619.
54. Björklund S, Engblom J, Thuresson K, Sparr E (2013) Glycerol and urea can be used to increase skin permeability in reduced hydration conditions. *Eur J Pharm Sci* 50: 638-645.
55. Warskulat U, Reinen A, Grether-Beck S, Krutmann WJ, Haussinger D (2004) the osmolyte strategy of Normal Human Keratinocytes in Maintaining Cell Homeostasis. *J Invest Dermatol* 123: 516-521.
56. Anderheggen B, Jassoy C, Waldmann-Laue M, Forster T, Wadle A, et al. (2006) Taurine improves epidermal barrier properties stressed by surfactants. A role for osmolytes in barrier homeostasis. *J Cosmetic Sci* 57: 1-10.
57. Schwan WR, Wetzel KJ (2016) Osmolyte transport in *Staphylococcus aureus* and the role in pathogenesis. *World J Clin Infect Dis* 6:22-27.
58. Chiti F, Dobson CM (2006) Protein misfolding, functional amyloid, and human disease. *Annu Rev Biochem* 75: 333-366.
59. Herczenik E, Gebbink MFBG (2008) Molecular and cellular aspects of protein misfolding and disease. *FASEB J* 22: 2115-2133.
60. Kendrick BS, Carpenter JF, Cleland JL, Randolph TW (1998) A transient expansion of the native state precedes aggregation of recombinant human interferon-c. *Proc Natl Acad Sci USA* 95: 14142-14146.
61. Bhavsar RD, Prasad S, Roy I (2013) Effect of osmolytes on the fibrillation of HypF N. *Biochimie* 95: 2190-2193.
62. Olsen SN, Ramlov H, Westh P (2007) Effects of osmolytes on hexokinase kinetics combined with macromolecular crowding: Test of the osmolyte compatibility hypothesis towards crowded systems. *Comp Biochem Phys A: Mol Integr Physiol* 148: 339-345.
63. Wawera J, Krakowiaka J, Szocinskib M, Lustigc Z, Olszewskid M, et al. (2014) Inhibition of amyloid fibril formation of hen egg white lysozyme by trimethylamine N-oxide at low pH. *Int J Biol Macromol* 70: 214-221.
64. Macchi F, Eisenkolb M, Kiefer H, Otzen DE (2012) The effect of osmolytes on protein fibrillation. *Int J Mol Sci* 13: 3801-3819.
65. Arora A, Hab C, Park CB (2004) Inhibition of insulin amyloid formation by small stress molecules. *FEBS Lett* 564: 121-125.

66. Gao M, Estel K, Seeliger J, Friedrich RP, Dogan S, et al. (2015) Modulation of human IAPP fibrillation: cosolutes, crowders and chaperones. *Phys Chem Chem Phys* 17: 8338-8348.
67. Choudhary S, Kishore N, Hosur RV (2016) Inhibition of insulin fibrillation by osmolytes: Mechanistic Insights. *Scientific Reports* 5: 17599.
68. Choudhary S, Kishore N (2014) Addressing mechanism of fibrillization/ aggregation and its prevention in presence of osmolytes: spectroscopic and calorimetric approach. *PLoS One* 9: 104600.
69. Tanaka M, Machida Y, Niu S, Ikeda T, Jana NR, et al. (2004) Trehalose alleviates polyglutamine-mediated pathology in a mouse model of Huntington disease. *Nat Med* 10: 148-154.
70. Dyson HJ, Wright PE (2005) Intrinsically unstructured proteins and their functions. *Nat Rev* 6: 197-208.
71. Liu J, Perumal NB, Oldfield CJ, Su EW, Uversky VN, et al. (2006) Intrinsic disorder in transcription factors. *Biochemistry* 45: 6873-6888.
72. Uversky VN, Li J, Fink AL (2001) Trimethylamine-N-oxide-induced folding of α -synuclein *FEBS Letters* 509: 31-35.
73. Munishkina LA, Henriques J, Uversky vn, fink al (2004) role of protein-water interactions and electrostatics in α -Synuclein Fibril Formation. *Biochemistry* 43: 3289-3300.
74. Munishkina LA, Cooper EM, Uversky VN, Fink AL (2004) The effect of macromolecular crowding on protein aggregation and amyloid fibril formation. *J Mol Recognit* 17: 456-464.
75. Scaramozzino F, Peterson DW, Farmer P, Gerig JT, Graves DJ, et al. (2006) TMAO promotes fibrillization and microtubule assembly activity in the C-terminal repeat region of Tau. *Biochemistry* 45: 3684-3369.
76. Nandi PK, Bera A, Sizaret PY (2006) Osmolyte trimethylamine N-oxide converts recombinant α -helical prion protein to its soluble β -structured form at high temperature. *J Mol Biol* 206: 810-820.
77. Fung J, Darabie AA, McLaurin J (2005) Contribution of simple saccharides to the stabilization of amyloid structure. *Biochem Biophys Res Commun* 328:1067-1072.
78. Kim HY, Kim Y, Han G, Kim DJ (2010) Regulation of in vitro A β 1-40 aggregation mediated by small molecules. *J Alzheimers Dis* 22: 73-85.
79. Singh LR, Poddar NK, Dar TA, Kumar R, Ahmad F (2011) Protein and DNA destabilization by osmolytes: The other side of the coin. *Life Sciences* 88: 117-125.
80. Mikles DC, Bhat V, Schuchardt BJ, McDonald CB, Farooq A (2015) Effect of osmolytes on the binding of EGR1 transcription factor to DNA. *Biopolymers* 103: 74-87.
81. Sidorova NY, Muradymov S, Rau DC (2005) Trapping DNA-protein binding reactions With neutral osmolytes for the analysis by gel mobility shift and self-cleavage assays. *Nucleic Acids Res* 33: 5145-5155.
82. Timasheff SN (1992) A physicochemical basis for the selection of osmolytes by nature. In: Somero (edn) *Water and Life*: Springer-Verlag, New York, 70-84.
83. Hu CY, Kokubo H, Lynch GC, Bolen DW, Pettitt BM (2010) Backbone additivity in the transfer model of protein solvation. *Protein Sci* 19: 1011-1022.
84. Stumpe MC, Grubmuller H (2007) Interaction of urea with amino acids: Implications for urea-induced protein denaturation. *J Am Chem Soc* 129: 16126-16131.
85. Hua L, Zhou R, Thirumalai D, Berne BJ (2008) Urea denaturation by stronger dispersion interactions with proteins than water implies a 2-stage unfolding. *Proc Natl Acad Sci USA* 105: 16928-16933.
86. Auton M, Bolen DW, Rösgen J (2008) Structural thermodynamics of protein preferential solvation: osmolyte solvation of proteins, aminoacids, and peptides. *Proteins* 73: 802-813.
87. Bennion BJ, Daggett V (2004) Counteraction of urea-induced protein denaturation by trimethylamine N-oxide: A chemical chaperone at atomic resolution. *Proc Natl Acad Sci USA* 101: 6433-6438.
88. Wei H, Fan Y, Gao YQ (2010) Effects of urea, tetramethyl urea, and trimethylamine N-oxide on aqueous solution structure and solvation of protein backbones: a molecular dynamics simulation study. *J Phys Chem B* 114: 557-568.
89. Paul S, Patey GN (2008) Hydrophobic interactions in urea-trimethylamine-N-oxide solutions. *J Phys Chem B* 112: 11106-11111.
90. Canchi DR, Paschek D, Garcia AE (2010) An equilibrium study of protein denaturation by urea. *J Am Chem Soc* 132: 2338-2344
91. Kumar N, Kishore N (2013) Synergistic behavior of glycine betaine - urea mixture: A molecular dynamics study. *J Chem Phys* 139: 115104.