CD133 and MYCN Amplification Induce Chemo-Resistance and Reduce Average Survival Time in Pediatric Neuroblastoma

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Abstract

Objectives: Neuroblastoma (NB) is the most common pediatric solid tumor derived from the sympathetic nervous system. MYCN gene exists in nearly half of the NB patient and its association with rapid disease progression and poor outcome is controversial. Cancer Stem Cells (CSCs) characterization in NB has been rarely studied. This study is to figure out whether the MYCN gene and CSCs are associated with chemotherapy resistance and survival time in the NB patients.

Methods: Based on unequivocal pathological diagnosis, 50 NB patients are recruited. MYSN amplification is measured before any therapy. The CSCs are derived and their multi-potencies are tested by directed differentiation. Response to chemotherapy and average survival time of these patients are gathered and compared with following groups: CD133+, CD133-, MYCN ≥ 5, MYCN<5, CD133+ plus MYCN ≥ 5, CD133- plus MYCN<5.

Results: CD133+ CSCs differentiate into neuron like cells; CD133+ patients have significant poorer response to chemotherapy comparing with the CD133- patients (P<0.01); CD133+ and MYCN ≥ 5 patients have significantly shorter average survival time than the CD133- and MYCN<5 patients (P<0.01).

Conclusions: CD133+ CSCs produce chemo-resistance. CD133 and MYCN amplification can be used together as a prognostic value for predicting disease outcome.

Keywords: Cancer stem cells; CD133; Chemo-resistance; MYCN; Neuroblastoma

Introduction

Neuroblastoma (NB) is the most common extracranial pediatric solid tumor. NB tumor cells are embryonic neural crest-derived cells, which give rise to the nerve cells of the sympathetic nervous system in normal development [1,2]. It is widely assumed that NBs are embryonic tumors originating from a developmental defect that prevents normal cell differentiation and gets stuck in a continuous replication status [3,4]. It most frequently originates in the adrenal medulla and paraspinal or paravertebral regions where the sympathetic nerve tissue are present from neck to pelvis [5,6].

The prevalence of NBs is about one in 7000 live births; the incidence is about 10.54 cases per 1 million per year in the children younger than 15 years [7]. About 90% are younger than 5 years at diagnosis, 37% are infant, the median age of diagnosis is 19 months [8]. Most NB cases are sporadic, only 1% to 2% patients have family history of NB. The most common symptom of NB is an abdominal mass, and other signs include proptosis and periorbital ecchymosis, abdominal distention, bone pain, pancytopenia, fever, hypertension, anemia, paralysis and watery diarrhea etc [9,10]. The clinical course is heterogeneous ranging from spontaneous regression in neonates to life-threatening metastatic disease in older children [5]. The medical treatments include observation, chemotherapy, surgery, immunotherapy, autologous peripheral blood stem cell transplantation (APBSCT). Since the five-year survival rate was significantly improved in recent five years, high-risk NBs are still remain the most difficult tumor to cure, with only 40% of long-term survival despite intensive multi-modal therapy [6,11,3].

In recent years, accumulating evidence demonstrates that tumor is a hierarchy organization regulated by a minority of cells, called Cancer Stem Cells (CSCs) [12-14]. CSCs constitute less than 1% of tumor cells and have exclusive ability to self-sustain and self-renew to generate heterogeneous lineages of cancer cells to comprise the tumor [15]. The existence of self-renewing and multi-potent CSCs in NBs was partially addressed by Ross's research team [16,17]. Kaplan's research demonstrated that dissociated cells from aggressive NB tumors grew like spheres, differentiated into neurons, and formed NB tumors when orthotopically injected into severe combined immunodeficient/Beige mice [18]. CD133 is a cell-surface glycoprotein harboring five transmembrane domains [19]. Knocking down of the CD133 in NB cells effectively resulted in cell growth retardation [20]. Up-regulated CD133 decreased sensitivity to cytostatics in clinic [21]. Whether CD133+ cells have effects on responses towards chemotherapy and average survival time need to be demonstrated. At the best of our knowledge, there is no data has been published on this point.

MYCN amplification (defined as more than 10 copies per diploid genome) exists in 40% to 50% of high-risk NBs [22,23]. Most unfavorable clinical and pathobiological features are associated, to some degree, with MYCN. However, there is controversial study demonstrated MYCN is not related to the NB progression [24,25]. Whether MYCN amplification is associated with the NB progression needs to be further studied.

The purpose of this study is to demonstrate that whether the MYCN

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amplification and existence of the CD133 positive cells in the NBs are correlated with chemo-resistance and average survival time. We recruit NB patients, collect peripheral blood to measure MYSN amplification levels before any therapy. After surgery, CSCs are derived, and the CD133+ cells are calculated. The correlation of MYSN amplification and CD133+ CSCs with disease severity and overall survival rate is statistically calculated.

Materials and Methods

Patient recruitment

This study was reviewed and approved by The Second hospital of Hebei Medical University Ethics Committee before the study began. Written informed consent was provided to participants recruited into this study. The minimum criterion of a diagnosis of NB must be based on one of the following: an unequivocal pathological diagnosis from tumor tissue by light microscopy determined by the International Neuroblastoma Pathology Classification; the combination of bone marrow aspirate or trephine biopsy containing unequivocal tumor cells and increased levels of urinary catecholamine metabolites [26]. The recruitment started from Jan 2014 and finished in May 2016.

The International Neuroblastoma Staging System (INSS) stratifies neuroblastoma according to anatomical presence at diagnosis: stage I, IIa, IIb, III, IV, and IVS [1,3,4]. The International Neuroblastoma Risk Group Staging System (INRGSS) classifies NB into four pre-treatment groups: very low, low, intermediate and high risk [27]. In this study, we recruited 50 patients from 3 months to 8 years old, NB stages from IIa to IV. The INRGSS classification of these NB patients are from very low to high risk. The characteristics of patients recruited into this study are present in Table 1.

Clinical treatments

In this study, the MYCN amplification was calculated before any therapy. For very low risk NB patients, intensive observation every two months after surgery is carried on in clinic; for low risk, intermediate and high risk NB patients, surgery is performed before or during chemotherapy. For the patients over 18 months old, radiotherapy is used after chemotherapy; Criteria for chemotherapy must reach following clinical conditions before usage: no uncontrolled infection and organ dysfunctions, Alanine Transaminase (ALT) level is less than three-fold higher than the normal level, Neutrophils Absolute Value (NAC) is over 9 x 10^9/L and platelets numbers is over 75 x 10^10/L.

The chemical therapy protocol for low to intermediate risk NB patients is as following: (1.5 mg/m^2)+VP16 (50 mg/m^2)+CTX (1.0g/m^2) for the first cycle, VCR (1.5 mg/m^2)+CBP (550 mg/m^2)+VP16 (160 mg/m^2)+CTX (1.0g/m^2) for the second cycle. These two cycles rotate to use. 9-cis-RA is 150-160 mg/m^2 for 6 months. For high risk NB patients, the chemical therapy protocol is as following: VCR (1.5 mg/m^2)+CDDP (25 mg/m^2)+VP16 (100 mg/m^2)+CTX (1.0g/m^2), IFOS (1.5 mg/m^2)+CBP (550 mg/m^2)+THP (30 mg/m^2). These two cycles rotate to use. 9-cis-RA is 150-160 mg/m^2 for 6 months. For the patients whose body weights is less than 12 kg, the concentrations of chemical drugs are as following: VCR (0.05 mg/kg), CBP (16 mg/kg), ADR (1 mg/kg), CTX (33 mg/kg), VP16 (5 mg/kg), 9-cis-RA (5.33 mg/kg/d), CDDP (0.85 mg/kg), IFOS (50 mg/kg), THP (1 mg/kg).

MYCN amplification

Total RNA is extracted from the NB patient and healthy volunteer peripheral blood. Total RNAs are extracted using RNeasy columns (Qiagen) according to the manufacture instructions. Total RNA (1 g) is reverse transcribed into cDNA (Clontech). SYBR green real-time PCR kit (Applied Biosystems) is used to quantify gene expression levels. Data is analyzed with SDS Relative Quantification Software version 2.2.2 (Applied Biosystems). Ct values were exported into Excel software for data analysis. The MYCN amplification levels are compared between the NB patients and healthy volunteers. MYCN forward primer is: TCTCTCGGACGCGCA. MYCN reverse primer is: TGGCAGAGGAGACC.

Tissue dissection and preparation of single cell suspension

Tumor samples are collected and derived for the CSCs immediately. The tumor tissue is isolated and minced with a scissor, and rinsed with HBSS solution (without Ca^2+ and Mg^2+ (Life Technologies, 14170112) with 20% fetal bovine serum (FBS) (Life Technologies, 10439001) at least three time to get rid of blood. Equal amount of the minced tissue is separated into three 15 ml centrifuge tubes. The tissue pieces were incubated at 37°C for 15 min in 10 ml of dissociation solution: HBSS (without Ca^2+ and Mg^2+) supplemented with 0.015M HEPES (Life Technologies), 5.4 mg/ml D-glucose, 0.25% trypsin (Life Technologies, 15050057), 80 U/ml DNase (Sigma), 0.7 mg/ml hyaluronidase (Sigma), and 2 mg/ml kynurenic acid (Sigma), followed by gentle trituration, and incubation in the same solution at 37°C for another 10 min. Excess solution was removed and the tissue further mechanically dissociated to a single-cell suspension.

Neurosphere cultures

Single cells are plated at a density of 100 cells per well in 6-well ultra-low attachment multilaw polystyrene plates (Falcon, Becton Dickinson, Franklin Lakes, NJ, USA) in a serum-free expansion medium composed of a 1:1 mixture of Dulbecco’s Modified Eagle’s Medium (DMEM) and Ham’s F12 Nutrient Mixture (Life technologies), supplemented with 40 ng/ml epidermal growth factor (R&D Systems), 20 ng/ml basic fibroblastic growth factor (R&D Systems), 1% B27 supplement (Life Technologies), 2ug/ml heparin (Sigma Aldrich) and 100 units/ml penicillin/streptomycin. Cultures are maintained at 37°C in a humid atmosphere with 95% O2 and 5% CO2. 50% of culture medium is changed with fresh medium with fresh growth factors every other day. After 7 days, neurospheres are either passed for FACS analysis or passaged by mechanical dissociation and re-suspended.

CSCs differentiation into neurons

The 6 well plates are coated with 20 mg/ml Poly–L–ornithine (Sigma,
Fluorescence activated cell sorting (FACS)

Cells were mechanically dissociated to obtain a single-cell suspension in a solution containing 2 mm EDTA (Sigma) and 0.5% FBS (Sigma) in PBS. Cells are harvested by centrifugation (500 g, 5 min, 4°C), incubated with anti-mouse CD133 antibody (Chemicon), then incubated with secondary antibody conjugated with phycoerythrine (PE) for 2 hours on ice, washed with PBS twice then filtering through a 70-μm cup filter ( Falcon) and kept on ice until FACS analysis. Cos7 cells are used as a negative control, liver tumor cells (known as high CD133 expression, kindly provided by Dr. Jun Li) are used as a positive control. Cell sorting is performed on a FACS Vantage TS flow cytometer (Becton Dickinson, San Jose, CA, USA).

Immunohistochemistry

The cells are fixed with 4% paraformaldehyde for 10 minutes at room temperature. After being blocked with 5% normal goat serum in PBS, the cells are incubated with primary antibody neuronal class III b-tubulin antibody (Chemicon, MAB1637) overnight at 4°C. After being washed for 3 times with PBS, the cells are incubated with secondary antibody Alexa Fluoro®647-AffiniPure Goat anti-Mouse IgG (Jackson ImmunoResearch, 115-605-205) for 2 hours at room temperature. The labeled slides are dehydrated in alcohol and xylene before being cover-slipped with mounting media.

Statistical analysis

Numerical data is expressed as mean ± standard deviation (SD). Paired Student's t-test (two tailed) is used for statistical analysis using SPSS.v16.0 for windows (SPSS Inc., Chicago, IL, USA). P values less than 0.05 is considered statistically significant and indicated with*; P values less than 0.01 were indicated with**.

Results

CT and pathological images of a particular NB patient

The CT tumor images of a specific NB patient are shown in Figure 1A and 1B. Figure 1A is a CT image before routine chemotherapy; Figure 1B is a CT image after chemotherapy of the same patient. The solid tumor almost disappears after chemotherapy. Most NB patients get partial response towards chemotherapy unfortunately. Figure 1C and 1D are pathological images of the NB tumor stained by H & E staining. On microscopy, the tumor cells are typically small, round and blue, and rosette patterns.

CSCs derived from the NB can form neospheres

Once the tumor tissue is separated into single cells, these single cells are plated on low attachment plates and grow in serum free medium with growth factors. As shown in Figure 1E, it is the third day after plating. Figure 1F shows the seventh day after plating. The neospheres grows bigger. These phenomenon demonstrates that there are CSCs existing inside of the neosphere.

CSCs can differentiate into neuron like cells

As multipotent stem cells, CSCs can differentiate into somatic cells. Whether the CSCs derived from the NB tissue can differentiate into neuron cells determines its identity. As shown in Figure 2A and 2B, once the neurospheres attaches to the bottom of plate, cells crawl out of the cell cluster. When the medium changes into neuron supporting medium for 2 weeks, the neuron like cells appear as shown in Figure 2C. These cells are b-tubulin positive as shown in Figure 2D, demonstrating that these cells are neurons.

FACS results for separating CD133+ cells from the CSCs

Cos7 cells are used as negative control as shown in Figure 3A, liver tumor cells known has high CD133 expression level are used as positive control as shown in Figure 3B. The neurospheres are harvested and stained with CD133 antibody. FACS shows that there are 0.1% CD133 positive cells in the CSCs derived from the NB tissue as shown in Figure 3C. These results demonstrates that there are cancer stem cells existing in the NB tissue. Whether and how these stem cells influence cancer pathophysiology and reaction towards to chemical therapy needs to be further studied (Table 2).

Correlation between CD133+ and MYCN amplification with chemotherapy response and average survival time

CD133+ NB patients have significant less chance to have complete response (P<0.01), very good partial response and partial response (P<0.05) than CD133- patients. However, CD133 expression has no significant influence on average survival time (P>0.05). MYCN amplification over 5 folds has similar effects on chemotherapy response with CD133 expression. However, MYCN amplification over 5 folds can significantly decrease average survival time comparing with MYCN amplification less that 5 folds (P<0.05); CD133+ NB patients who have MYCN amplification over 5 folds has significant shorter average survival time comparing with CD133- and MYCN amplification less than 5 folds (P<0.01) (Table 3 and Figure 4).

<table>
<thead>
<tr>
<th>Samples</th>
<th>Cos7 as the negative control</th>
<th>Liver tumor cells as positive control</th>
<th>NB sample</th>
</tr>
</thead>
<tbody>
<tr>
<td>Percentage of CSCs</td>
<td>0%</td>
<td>15%</td>
<td>0.1%</td>
</tr>
</tbody>
</table>

Table 2: CSCs percentages summary. There is 8.5% of CD133+ cells in liver tumor cells. There is only 0.1% of CD133+ cells in CSCs derived from neuroblastoma.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Patients</th>
<th>Complete response (%)</th>
<th>Very good partial response (%)</th>
<th>Partial response (%)</th>
<th>Average survival (months) (Average ± SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD133+</td>
<td>12</td>
<td>1 (8)</td>
<td>2 (17)</td>
<td>9 (75)</td>
<td>36.3 ± 6.9</td>
</tr>
<tr>
<td>CD133-</td>
<td>38</td>
<td>8 (21)**</td>
<td>19 (50)*</td>
<td>11 (29)*</td>
<td>41.5 ± 9.8</td>
</tr>
<tr>
<td>MYCN ≤ 5</td>
<td>21</td>
<td>0</td>
<td>7 (33)</td>
<td>14 (67)</td>
<td>24.6 ± 4.7</td>
</tr>
<tr>
<td>MYCN &lt; 5</td>
<td>27</td>
<td>9 (33)**</td>
<td>12 (45)</td>
<td>6 (22)*</td>
<td>40.2 ± 6.9*</td>
</tr>
<tr>
<td>CD133+ and MYCN &gt; 5</td>
<td>10</td>
<td>1 (10)</td>
<td>2 (20)</td>
<td>7 (70)</td>
<td>12.4±2.8</td>
</tr>
<tr>
<td>CD133+ and MYCN = 5</td>
<td>25</td>
<td>8(32)**</td>
<td>12(48)**</td>
<td>5(20)**</td>
<td>42.7 ± 8.3**</td>
</tr>
</tbody>
</table>

Table 3: Correlation between CD133 and MYCN amplification. *P<0.05; **P<0.01. CD133+ and/or MYCN ≥ 5 can significantly decrease response to chemical therapy comparing with CD133- and/or MYCN<5 (P<0.05).
Discussion

Neuroblastoma (NB) is the most common pediatric solid tumor derived from the sympathetic nervous system. The clinical course is heterogeneous ranging from spontaneous regression in neonates to life-threatening metastatic disease in older children. MYCN amplification level is recruited into the INRGSS system, and it is used to predict patient survival time and direct clinical treatment protocol. However, it
Figure 2: CSCs can differentiate into neuron like cells. Floating neurospheres can attach to the coated plate bottom and cells can crawl out of the cell cluster as shown in A and B. Under the neuron growth medium, these CSCs can differentiate into neuron like cells as shown in figure C, which can be stained positive by using β-tubulin antibody as shown in figure D. This suggests that these cells are neurons.

Figure 3: CD133 expression results by FACS A: Cos7 cells as negative control; B: liver tumor cells as positive control; C: CSCs derived from neuroblastoma
is not rare to find some NB patients relapse after very good response to chemotherapy. This suggests that there is self-renewing, multi-potent cancer stem cells existing in the NB tumor tissue, and these cells induce chemo-resistance. In this study, we demonstrated that there are CSCs existing in the NB tissue, and these CSCs are CD133 positive and can differentiate into neuron like cells. The ratio of CD133 positive cells are around 0.1%. There are some NB patients are CD133 negative but have shorter average survival time, indicating that the CD133 antibody epitope affinity and neuropheres culture conditions might reduce the selection efficiency. Better culture condition and stable cell separation skills are required in the further studies.

MYCN amplification level has been recognized as a predictor for disease progression. In this study, we demonstrated that CD133 can be used as a response indicator for chemotherapy. The CD133 positive NB patients have significantly worse response comparing with the CD133 negative NB patients. MYCN and CD133 together can be used as indicators for the average survival time of the NB patients. We only recruited 50 patients into this study, bigger study groups are needed in the further studies.

It has been demonstrated that Valproic Acid (VPA), as HDAC inhibitor can increase CD133+ cells and show low sensitivity to cytostatics in NB [22]. The epigenetic effect of the HDAC inhibitor suggests that the viability of the CD133 protein expression is related to the acetylation of histones and methylation status of the CD133 promoter. This means that the CD133 gene promoter might be considered as a target for drug exploration.

In this study, we excluded the NB patients who receive autologous peripheral blood stem cell transplantation (APBSCT) and/or immunotherapy due to uncertainty of these treatments. In our future study, we shall closely observe the clinical effects of these two methods and recruit these patients into our study.

Overall, we demonstrate that CD133+ cells exist in the NB tissue, and these cells are self-renewing and multi-potent stem cells. CD133+ CSCs produce chemo-resistance. CD133 and MYCN amplification can be used together as a prognostic value for predicting disease outcome.

**Figure 4:** Correlation between CD133 and MYCN amplification with chemotherapy responses and average survival time of the NB patients. *P<0.05; **P<0.01. CD133+ and/or MYCN ≥ 5 can significantly decrease response to chemotherapy compared with CD133- and/or MYCN<5 (P<0.05). CD133+ and MYCN ≥ 5 significantly reduce average survival time comparing with other options (P<0.01).


