

Conformation of the Antigenic Peptide Tyrosinase (192-200) in Water and DMSO-d₆ by NMR Spectroscopy and MD Simulations

Evans C Coutinho^{1*}, Mishra NB¹, Deep Bhattacharya¹, Sudha Srivastava², Mamata Joshi² and Mushtaque Shaikh¹

¹Department of Pharmaceutical Chemistry, Bombay College of Pharmacy, Kalina, Santacruz [E], India

²National Facility for High Field NMR, Tata Institute of Fundamental Research, Homi Bhabha Road, Navy Nagar, Colaba, India

Abstract

Melanomas represent the most resistant form of cancer. If detected early, a cure is possible with the conventional therapies, while metastatic forms are almost resistant to conventional therapies. A new area i.e. immune based therapy shows promise of cytolytic activity against tumor cells. Production of antibodies against the antigenic epitopes present in the melanoma is the main basis of this immune based therapy. Production of T cell response against the tumor cells has been seen by synthetic antigenic epitopes. The antigenic nonameric peptide epitope of tyrosinase (192-200) with the sequence S¹EIWR⁵DIDF⁹ causes significant induction of the T cell response in melanoma patients. In realm of the significance of this peptide, a solution state NMR structure of 192-200 amino acid section of tyrosinase has been investigated, to assimilate the relationship between the antigenicity and the conformation of the peptide. NMR studies were carried out in H₂O:D₂O (95:5) and DMSO-d₆ solvents. Molecular dynamics simulations with NMR constraints were carried out using the GROMACS v .4.6.5 package. The results suggest the prevalence of a β-sheet structure both in H₂O as well as in DMSO-d₆.

Abbreviations: DMSO-d₆: Dimethyl Sulfoxide-D₆; TAA: Tumor Associated Antigen; MAA: Melanoma Associated Antigens; CTLs: Cytolytic T Cell Lymphocytes; TCR: T Cell Receptor; APC: Antigenic Presenting Cell; HLA: Human Leukocyte Antigens; TRP: Tyrosinase-Related Protein; TSP: 3-(Trimethylsilyl)Propanoic Acid, TOCSY: Total Correlation Spectroscopy, ROESY: Rotating Frame Overhauser Effect Spectroscopy; Roe: Rotating Frame Overhauser Effect; RMSD: Root Mean Square Deviation, OPLS: Optimized Potentials For Liquid Simulations, PDB: Protein Data Bank; Ppb: Parts Per Billion

Keywords: Nuclear magnetic resonance; Tyrosinase (192-200); Molecular dynamics simulations; Antigenic peptide; GROMACS

Introduction

Melanoma is a malignant tumor of the melanocytes which are pigment producing cells in the skin [1]. It is the major cause of deaths related to skin cancers. Although melanoma is completely curable by classical therapies such as surgery at an early stage, metastatic melanoma is many a times resistant to classical therapies [2]. Thus, there is an immediate need for new therapeutic approaches. One of the most investigated area is the use of immune based therapies as adjuvant treatment for melanoma, which goads the immune system to produce antibodies against the tumor cells [3,4]. These therapies are able to eliminate the tumor with very fewer side effects and establish an immunologic memory, providing long-term protection against tumor recurrence or metastasis [5,6]. In tumor rejection the important requirement is the ability of the immune system to recognize TAA, and to elicit an immune response against the tumor cells [7,8]. These antigens range from the 3D structures of small molecules that are identified by antibodies, to amino acid sequences which are recognized by CTLs. Induction of T cell mediated immune response is activated by the following two steps. In the first step, TAA derived peptides are thought to bind to empty MHC class-I molecules on dendritic cells, which then activate T cells by migration to the draining lymph nodes. In the case where the lymphocyte only receives signal 1, there is no activation of T cells, resulting in the tumor escaping the attack and a complete failure to stop tumor growth [9]. The barrier to the loss of signal can be overcome by the wide varieties of MAA with a

great number of epitopes that are recognized by CTLs. One class of MAA which is identified contains tyrosinase, melan A, gp 100, TRP-1 and TRP-2. The first melanocytes differentiating antigen recognized was tyrosinase [8]. This melanosomal membrane protein consists of 529 amino acids, previously known to be required for the melanin synthesis [10]. An antigenic peptide tyrosinase (192-200) with the sequence S¹EIWR⁵DIDF⁹ can be recognized by CTL on the HLA-B44 molecule. According to the recently inferred consensus motif for the anchor residues binding to the HLA-B44, the predominant position for Glu is position 2, Tyr or Phe at positions 9 or 10 and hydrophobic residues at position 3 [11]. The tyrosinase (192-200) nonapeptide fits the consensus very well with Glu at position 2, Phe at position 9 and Ile at position 3 [12]. Among the cells expressing tyrosinase endogenous peptide subtypes of HLA-B44, the HLA-B*4403 subtype is lysed by the anti-MZ2-C (an antigen produced by the melanoma patients) CTL clone, whereas those of the B*4402 subtype undergo very insignificant lysis. They differ by a single Asp (B*4402) to Leu (B*4403) substitution in position 156 of the α2-domain [13]. This restricted lysis by selective subtype could be explained either by the binding of the peptides to only one subtype or by differences in the bound peptide structure induced by differences in the groove of the two subtypes. It is observed that the tyrosinase nonapeptide binds to both subtypes. This is consistent with the finding that the consensus motif are similar for both subtypes B*4402 and B*4403, since it is based on anchoring residues. It appears, therefore, that different conformations of the peptide in the B*4402 and B*4403

***Corresponding author:** Evans C Coutinho, Department of Pharmaceutical Chemistry, Bombay College of Pharmacy, Kalina, Santacruz [E], Mumbai 400098, India, E-mail: evans@bcp.edu.in

Received June 19, 2015; **Accepted** July 13, 2015; **Published** July 16, 2015

Citation: Coutinho EC, Mishra NB, Bhattacharya D, Srivastava S, Joshi M, Shaikh M et al. (2015) Conformation of the Antigenic Peptide Tyrosinase (192-200) in Water and DMSO-d₆ by NMR Spectroscopy and MD Simulations. Pharm Anal Acta 6: 392. doi:10.4172/21532435.1000392

Copyright: © 2015 Coutinho EC, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

grooves are responsible for the differences in the CTL recognition sites [11]. The T cell epitope of tyrosinase (192-200), has shown promising results in clinical trials with fewer side effects and with significant T cell response. In view of this we have investigated the solution NMR structure of tyrosinase (192-200) by NMR and molecular dynamics simulations. We have investigated its conformation in two solvents i.e. in H₂O:D₂O and DMSO-d₆. Water provides the environment similar to the body fluids. Based on some observations that the liquid phase in the cytoplasm and the intersynaptic space in receptor binding sites have a viscosity higher than bulk water, it has been suggested that DMSO with a viscosity approximately 2.5 times that of water or other high viscosity solvents may provide a more realistic environment for the study [14,15]. Presence of any strong intramolecular hydrogen bonds in the peptides can also be studied as these stronger bonds cannot be disrupted by interaction with DMSO-d₆ [16].

Material and Methods

NMR experiments

The peptide, S'EIWR²DIDF⁹ was purchased from Bachem, USA. An amount equal to 1 mg of the peptide was dissolved in 0.6 ml of a mixture of H₂O (95%) and D₂O (5%). For the study in DMSO-d₆, a similar amount of the peptide was dissolved in 0.6 ml of DMSO-d₆ purged in a nitrogen atmosphere to exclude moisture.

The NMR experiments were carried out on 600 MHz and 800 MHz Bruker Avance NMR spectrometers, as per the requirement. The length of the 90° pulse was calibrated at the beginning of the experiments and the temperature was controlled throughout the experiment to ± 0.1 K. 1D proton spectra for the sample in water [H₂O: D₂O (95: 5)] were acquired with a spectral width of 8012 Hz, 512 scans, and digitized with 16K data points. Solvent suppression for the sample in water was accomplished with the 3-9-19 WATERGATE pulse sequence. All experiments were recorded with the conventional pulse sequences. The temperature coefficients were measured by recording ¹H spectra in the range 290 to 330K. The spin system assignment for amino acids was carried out using 2D TOCSY [17] with mixing time of 150 ms and sequential assignment carried out by 2D-ROESY [18] with mixing time of 300 ms. All experiments were run at 300K. For 2D experiments, 512 data points were collected in the time domain. At the beginning of each experiment, 16 dummy scans were collected to allow the system to reach thermal equilibrium. 2D spectra except TOCSY were acquired with a sweep width of 14000 Hz. Typically, 512 FIDs of 2048 complex points were collected. The TOCSY experiment was carried out using spin lock pulse of 100 ms with 512 experiments at 300K and a spectral width of 8012 Hz for 4096 complex data points. The data were multiplied with $\pi/4$ and $\pi/8$ sine bell window functions along the t_1 and t_2 axes prior to Fourier transformation. All NMR data were processed using Topspin 3.2 software [19]. In H₂O: D₂O the ¹H chemical shift were calibrated with the internal standard TSP.

For the sample in DMSO-d₆, 1D proton spectra were recorded with a sweep width of 13586 Hz and 16K data points. A total of 1024 scans were accumulated, and an acquisition time of 2.05 s was used for the peptide in DMSO-d₆. 16 dummy scans were taken prior to the start of the experiment for thermal equilibrium. All 2D spectral widths were 13586 Hz. The data size in the time domain was 512 points in t_1 and 2048 points in t_2 domains. For each t_1 value, 96 transients were accumulated. The data were multiplied with $\pi/4$ and $\pi/8$ sine bell window functions along the t_1 and t_2 axes prior to Fourier transformation. The observed

512*2k complex data matrices were zero-filled to 2K*2K points (ROESY and TOCSY). DMSO-d₆ was considered as its own internal standard as it distinctly shows a peak at 2.5 ppm.

Measurement of inter-proton distances

ROESY is useful for determining signals arising from protons that are close to each other in space even though they are not covalently bonded. ROESY also detects chemical and conformational exchange among NH protons. The rOe is the result of modulation of the dipole-dipole coupling between different nuclear spins by Brownian motion of the molecules in solution. Dipole-dipole interactions are electrostatic interactions present between permanent dipoles in molecules. The rOe intensity varies proportionately with the distance between nuclei separated by a distance less than 5.0 Å [20,21]. The rOe's were converted to distances using the relationship of the rOe intensity to the inverse sixth power of the distance between the protons, as expressed in the following equation.

$$r_{ij} = r_{kl} (I_{kl}/I_{ij})^{1/6}$$

where, r_{ij} and I_{ij} are the distance and integrated rOe volume between two proton i and j and r_{kl} and I_{kl} are the same quantities for the pair of reference protons [22]. The inter-nuclear distance between the β -H₁ and β -H₂ protons of phenylalanine, 1.732 Å was taken as the reference distance because these protons are non-degenerate and the inter-nuclear distance is invariant with peptide conformations. The distances were cast as restraints using a ± 0.3 Å limit to define the upper and lower bounds on the distances.

Dihedral restraints

The dihedral angle (φ) was calculated from the ³J_{NH α coupling constant from the modified Karplus equation, and these were cast as restraints allowing a range of ± 10° on the calculated value [23,24] in the MD simulations.}

$${}^3J_{\text{NH}\alpha} = A + B \cos \varphi + C \cos 2\varphi$$

MD simulations

MD simulations were carried out using GROMACS simulation software package v.4.6.5 [25-28]. The partial charges and potentials for all atoms were assigned using the OPLS-AA 2005 force field [29]. A 3D structure for the sequence was built and saved as PDB format using the Schrödinger module Maestro and the structure was centered in a solvated box containing 26310 TIP3P water molecules [30] or 8027 molecules of DMSO-d₆ with the GROMACS software package. The system was neutralized using three Na⁺ and one Cl⁻ as counter ions. Further the system was energy minimized using the steepest descent algorithm [31] for of 1,000,000 steps with a time steps of 2 ps by constraining the C-H bonds using the LINCS algorithm. The equilibration in MD was carried out in two steps. The first step (100 ps) was conducted under an NVT (constant number of particles, volume, and temperature) ensemble and the subsequently the system was equilibrated under NPT (constant number of particles, pressure, and temperature) conditions for 100 ps. The production stage with NMR restraints, was performed for 2500 ps.

Results and Discussion

NMR analysis

Sequence specific resonance assignment: Sequential assignments were made according to the methods discussed by Wüthrich et al [32]. In sequential assignment the connectivities between ¹H spin systems

of sequentially neighboring amino acids were established. The amino acids were first classified on the basis of their side chain spin system and identified [32]. The Phe-9 residue was identified from the rOe's arising from H β proton to its aromatic rings and the absence of the H γ resonance in the 2D-TOCSY spectrum. Three AMX systems (one Ser and two Asp residues) were identified. Identification of all three H α -H β connectivities was straight forward from the TOCSY spectrum. The individual Asp residues were distinguished on the basis of their sequential rOe's to their neighboring residues. Ile-3 and Ile-7 were identified on the basis of NH-H α -H β -H γ -H δ connectivities and individual resonances identified by the sequential rOe's to their neighboring residues. Arg-5 was identified by the presence of NH-H γ connectivity, where the H γ resonance is comparatively upfield of H β . Trp-4 was assigned by the presence of rOe from the aromatic ring to its H β resonance which is easily observed in the 2D-ROESY spectrum. Glu-2 was identified on the basis of the upfield shift for its H β resonance, and was confirmed by the rOe to its neighboring residue. Ser-1 was identified by the rOe to Glu-2. After assigning respective resonances for the specific amino acid residues, sequence specific resonance assignment was done by rOe's of 2D ROESY. The cross-peaks observed in the 2D-ROESY spectrum helped in complete sequential assignment. For the peptide in water, the sequence specific resonance assignment was made by walking from the N terminal to the C terminal using the rOe's (H α_i to NH $_{i+1}$). Absence of long range rOe's are indicative of a β -sheet structure in both solvents [33,34]. The chemical shifts for the peptide is given in Table 1 (for H₂O:D₂O) and in Table 2 (for DMSO-d₆).

Chemical shift index (CSI)

The CSI is a graph based technique which assigns every assigned chemical shift value to a three state (-1, 0, +1) index. The method makes the use of fact that α H chemical shifts of amino acid residues in helices tends to be shifted upfield (i.e. towards the right side of a NMR spectrum) relative to their random coil values and downfield (i.e. towards the left side of a NMR spectrum) in beta strands. The CSI values of H α resonances are used to draw inferences about the secondary structure of the peptide using the rules laid down by Wishart et al [35]. Helical regions and stretches containing turns are usually characterized by a continuous stretch of negative deviations ('-1') from the random coil values for the α H chemical shifts and a positive deviation ('+1') in the case of ¹³C α chemical shifts. β -turns are characterized by the presence of a continuous stretch of 0 (without any deviation). A β -strand shows exactly an opposite picture [35-36]. The CSI of the peptide SEIWRDIDF in water and DMSO-d₆ are given in Figure 3. The CSI shows that peptide adopts a predominantly β sheet structure in both the solvents.

Temperature coefficients of amide proton resonances

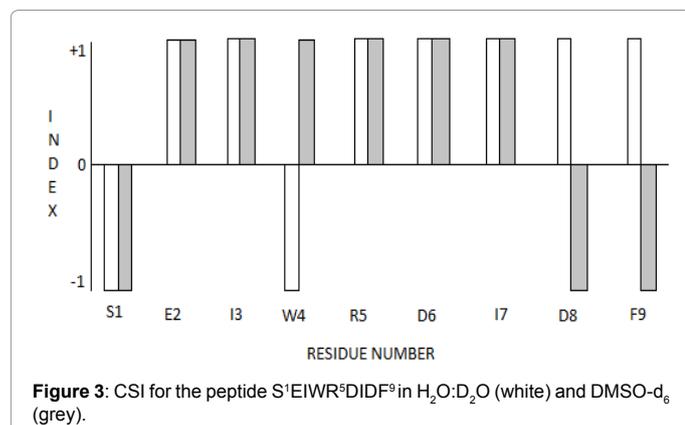
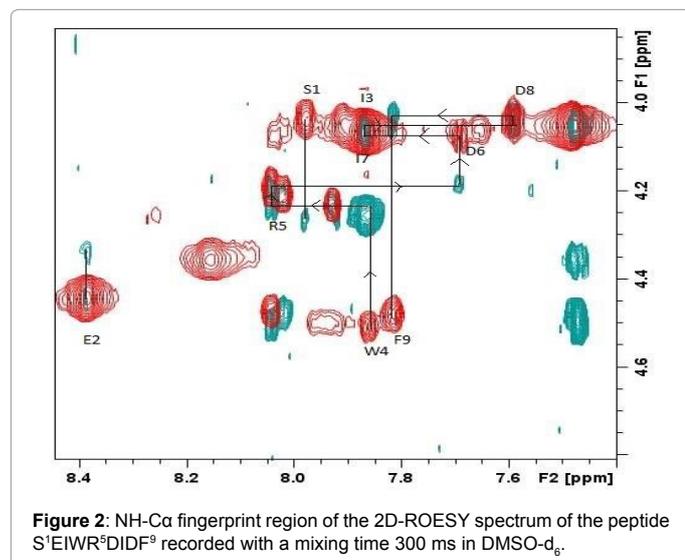
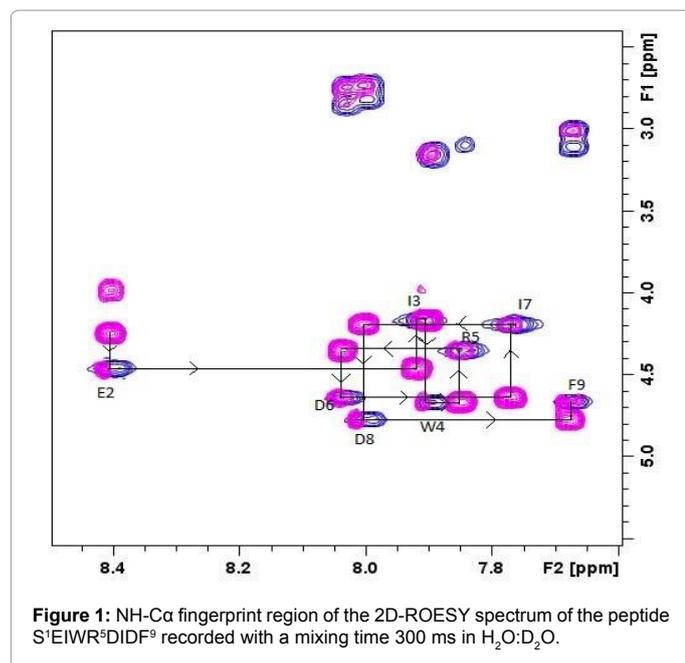
Temperature coefficients of amide proton embarks upon the presence of intra-molecular hydrogen bonding as chemical shifts are temperature dependent [37]. Temperature coefficient values of the amide proton has been extensively used to predict hydrogen bonding. Small values of the temperature coefficients (< 3.0 ppb/K) of amide protons are usually indicative of intramolecular hydrogen bonds [38]. The temperature dependent of amide proton chemical shift was determined in the temperature range 290 to 330 K. The temperature

Residue	NH	H α	H β	Others	³ J _{NHα}	Torsion angles(ϕ)	Temp. coefficient of NH resonances
S1	-	4.2	3.9				
E2	8.39	4.44	2.33	H γ 1.92, 1.84	7.97	-159	-4.8
I3	7.91	4.24	1.69	H γ 1.32, 1.01 H δ 0.73	8.5	-164	-6.2
W4	7.88	4.65	2.33	Ring N-H 9.70	8.2	-161	-4.3
R5	7.84	4.49	1.74, 1.54	H γ 1.44 H δ 3.08 NH 6.96	8.2	-161	-5.3
D6	8.02	4.87	2.88, 2.72		7.68	-156	-5.3
I7	7.76	4.18	1.76	H γ 1.34, 1.0 H δ 0.71	8.57	-165	-6.3
D8	7.99	4.86	2.80, 2.72		8.27	-162	-4.9
F9	7.66	4.77	3.09, 2.98	3,5H 7.21 2,6H 7.10	8.57	-165	-4.3

Table 1: ¹H chemical shifts (in ppm), ³J_{NH α} (in Hz), torsion angles (in degrees), temperature coefficients of NH resonances (ppb/K) of the peptide S'EIWR⁵DIDF⁹ in H₂O:D₂O.

Residue	NH	H α	H β	Others	Temp. coefficient of NH resonances
S1	7.98	4.04	3.82		-5.0
E2	8.39	4.41	2.62	H γ 1.78, 1.84	-6.9
I3	7.87	4.16	1.64	H γ 1.24, 0.94 H δ 0.63	-4.5
W4	7.86	4.91	2.33	NH 9.78	-4.1
R5	8.04	4.54	2.54, 2.39	H γ 1.65 H δ 4.1	-5.0
D6	8.02	4.77	2.88, 2.72		-5.2
I7	7.87	4.06	1.70	H γ 1.32, 1.0 H δ 0.71	-5.2
D8	7.68	4.08	2.42, 2.29		-4.9
F9	7.81	4.48	3.05, 2.90	3,5-H 7.47 2,6-H 7.23	-3.9

Table 2: ¹H chemical shifts (in ppm), temperature coefficients of NH resonances (ppb/K) of the peptide S'EIWR⁵DIDF⁹ in DMSO-d₆.



coefficient of NH chemical shifts of the peptide in water are given in Table 1. The values fall in the range of -3.2 to -6.3 ppb/K. In all cases the chemical shifts varied linearly with temperature. These values indicates that the amide resonances are all exposed to the solvent and suggests a β sheet structure for the peptide in the two solvents [33].

$^3J_{\text{NH}\alpha}$ coupling constants

The $^3J_{\text{NH}\alpha}$ coupling constant is a function of the dihedral angle ϕ and is used to identify the conformation of the peptide [32]. The dihedral angles ϕ were calculated from the $^3J_{\text{NH}\alpha}$ values using the Karplus equation [23,39] as explained earlier. The ϕ values so obtained for the peptide in water are given in Tables 1. The $^3J_{\text{NH}\alpha}$ coupling constants and the ϕ values extracted from them are characteristic of a predominantly extended backbone conformation [39].

Calculation of 3D structure using NMR constraints

Restrained molecular dynamics simulations was performed using GROMACS. The statistical analysis of the simulation are given in Table 3. For the peptide in water, of the 100 structures sampled from the MD trajectory, 5 structures with the lowest global deviations from the input restraints were selected. A superposition of the backbone atoms of the 5 conformers is shown in Figure 4. The NMR structure with the lowest violations of imposed restraints is depicted in Figure 5. The backbone torsion angles (ϕ , ψ) for S¹EIWR³DIDF⁹ averaged over the entire MD trajectory are given in Table 4. The Ramachandran plots are given in Figure 4 (H₂O:D₂O) and Figure 5 (DMSO- d_6). The mean global RMSD was found to be 0.30 Å.

For DMSO- d_6 , of the 100 structures, 6 structures with the lowest global deviations from the input restraints were selected. A superposition of the backbone atoms of the peptide in the 6 conformations is shown in Figure 6. The NMR structure with the lowest violations of the imposed restraints is depicted in Figure 7. The mean global RMSD was found to be 0.28 Å.

Conclusions

The antigenic epitope tyrosinase (192-200) has shown wide applicability in the treatment of the melanoma associated cancers. It is significantly responsible for T cell response against tumor cells which is predominantly dependent upon the conformational structure of the peptide. In this study we have underlined the NMR structure of the antigenic epitope in two media i.e. H₂O:D₂O (95:5) and DMSO- d_6 . From the NMR parameters i.e. $^3J_{\text{NH}\alpha}$ coupling constants, temperature coefficients of NH chemical shifts and CSI values it was deduced that the peptide in both solvents exhibits an extended β sheet conformation. From this it may be concluded that the peptide in a β sheet conformation perhaps binds to TCR to elicit the biological response. This could open new areas of research for optimizing synthetic peptidomimetics specifically targeting TCR thus aiding the rational design of peptides that can bind to T-cells.

Conflict of Interest

The authors confirm that this article has no conflict of interest.

Acknowledgment

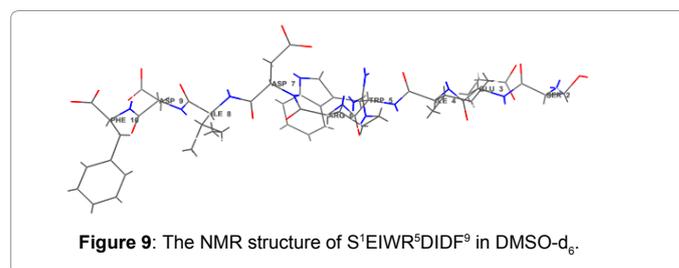
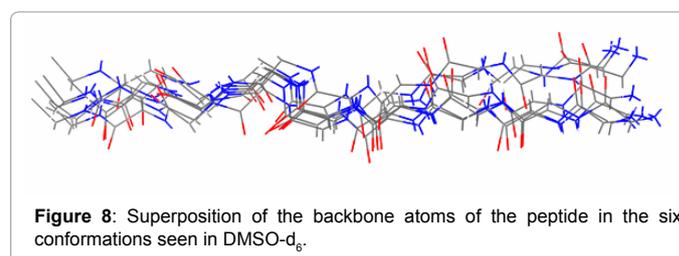
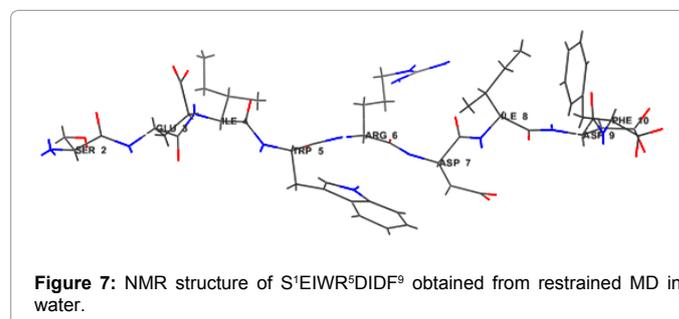
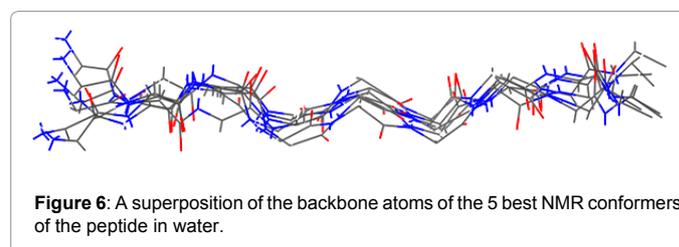
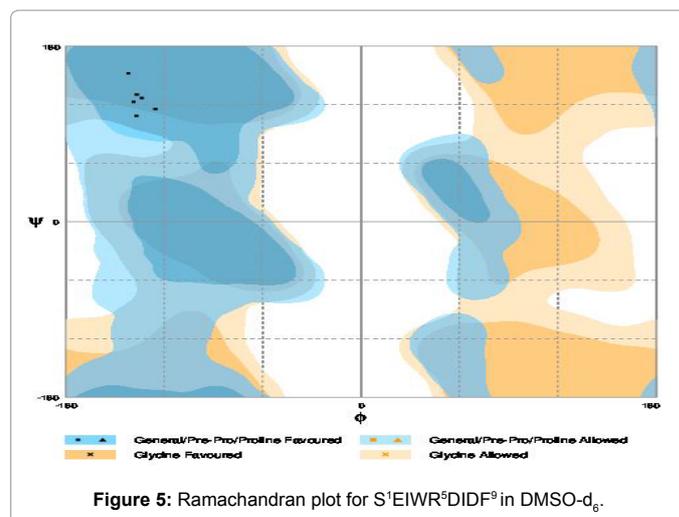
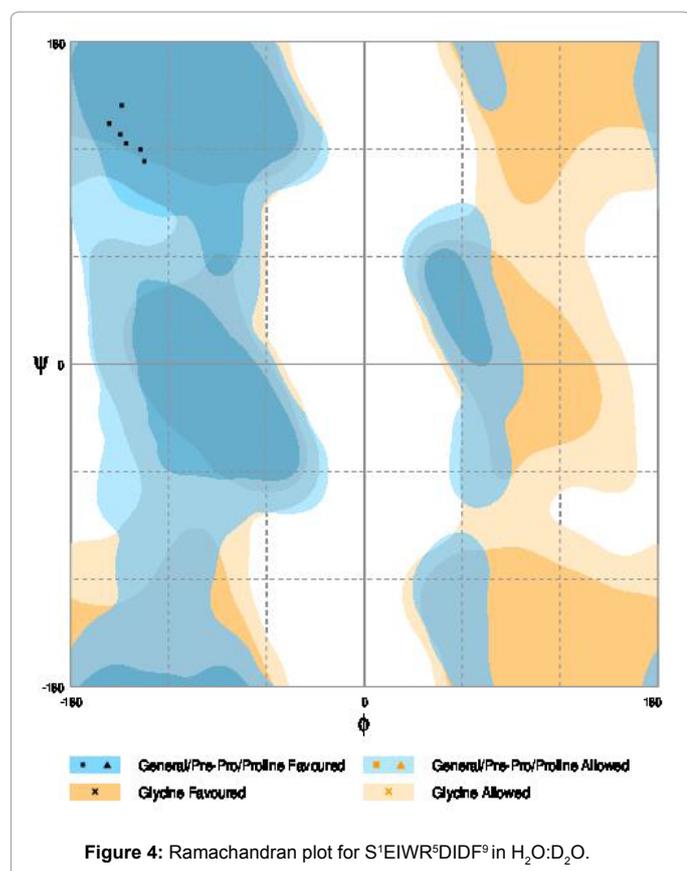
We acknowledge the NMR facilities provided by the National Facility for High Field NMR located at Tata Institute of Fundamental Research, Mumbai and the

Distance restraints			
All	H ₂ O	52	
	DMSO-d ₆	27	
Intra residue	H ₂ O	44	
	DMSO-d ₆	22	
Sequential	H ₂ O	8	
	DMSO-d ₆	5	
NOE violations (>0.3 Å)	H ₂ O	3	
	DMSO-d ₆	5	
RMSD (Å)			
Maximum	H ₂ O	0.61	
	DMSO-d ₆	0.48	
Minimum	H ₂ O	0.08	
	DMSO-d ₆	0.06	

Table 3: Structural statistics for the tyrosinase (192-200) in H₂O:D₂O and DMSO-d₆

Amino acid residues	H ₂ O:D ₂ O		DMSO-d ₆	
	φ	ψ	Φ	ψ
S ¹	-	143	-	129
E ²	-137	130	-142	152
I ³	-135	123	-134	126
W ⁴	-149	144	-139	122
R ⁵	-156	134	-137	130
D ⁶	-146	119	-125	115
I ⁷	-149	128	-137	118
D ⁸	-148	133	-144	136
F ⁹	-135	-	-131	-

Table 4: The backbone torsion angles (φ, ψ) for S¹EIWR⁵DIDF⁹ averaged over the entire MD trajectory.



computational and infrastructural facilities provided by Department of Science and Technology, New Delhi through their SERB program (SR/FST/LSI-163/2003; SR/SO/HS-0117/2012) and Council of Scientific and Industrial Research, New Delhi (01/2399/10/EMRII).

References

1. Kwon BS1 (1993) Pigmentation genes: the tyrosinase gene family and the pmel 17 gene family.
2. MacKie RM (1995) Melanoma prevention and early detection. *Br Med Bull* 51: 570-583.
3. Ohman Forslund K, Nordqvist K (2001) The melanoma antigen genes--any clues to their functions in normal tissues? *Exp Cell Res* 265: 185-194.
4. Morton DL, Foshag LJ, Hoon DS, Nizze JA, Famatiga E, et al. (1992) Prolongation of survival in metastatic melanoma after active specific immunotherapy with a new polyvalent melanoma vaccine. *Ann Surg* 216: 463-482.
5. Jimenez M, Nieto J, Rico M, Santoro J, Herranz J (1986) A study of the NH NMR signals of Gly-Gly-X-Ala tetrapeptides in H₂O at low temperature. *Journal of Molecular Structure* 143: 435-438.
6. Bancheureau J, Palucka AK, Dhodapkar M, Burkeholder S, Taquet N, et al. (2001) Immune and clinical responses in patients with metastatic melanoma to CD34(+) progenitor-derived dendritic cell vaccine. *Cancer Res* 61: 6451-6458.
7. Van den Eynde B, Lethe B, Van Pel A, Boon T (1992) Tumor Rejection Antigens and Immune Surveillance. *Modern Trends in Human Leukemia IX* pp: 279-285.
8. Kirkin AF, Dzhandzhugazyan K, Zeuthen J (1998) Melanoma-associated antigens recognized by cytotoxic T lymphocytes. *APMIS* 106: 665-679.
9. Scheibenbogen C, Schadendorf D, Bechrakis NE, Nagorsen D, Hofmann U, et al. (2003) Effects of granulocyte-macrophage colony-stimulating factor and foreign helper protein as immunologic adjuvants on the T-cell response to vaccination with tyrosinase peptides. *Int J Cancer* 104: 188-194.
10. Bouchard B, Fuller BB, Vijayasaradhi S, Houghton AN (1989) Induction of pigmentation in mouse fibroblasts by expression of human tyrosinase cDNA. *J Exp Med* 169: 2029-2042.
11. Fleischhauer K, Avila D, Vilbois F, Traversari C, Bordignon C (1994) Characterization of natural peptide ligands for HLA-B*4402 and -B*4403: implications for peptide involvement in allorecognition of a single amino acid change in the HLA-B44 heavy chain. *Tissue Antigens* 44: 311-317.
12. Brichard VG, Herman J, Van Pel A, Wildmann C, Gaugler B, et al. (1996) A tyrosinase nonapeptide presented by HLA-B44 is recognized on a human melanoma by autologous cytolytic T lymphocytes. *Eur J Immunol* 26: 224-230.
13. Kawakami Y, Eliyahu S, Delgado CH, Robbins PF, Sakaguchi K, et al. (1994) Identification of a human melanoma antigen recognized by tumor-infiltrating lymphocytes associated with in vivo tumor rejection. *Proceedings of the National Academy of Sciences of the United States of America* 91: 6458-6462.
14. Amodeo P, Motta A, Picone D, Saviano G, Tancredi T, et al. (1991) Viscosity as a conformational sieve. NOE of linear peptides in cryoprotective mixtures. *Journal of Magnetic Resonance* 95: 201-207.
15. Shen CL, Murphy RM (1995) Solvent effects on self-assembly of beta-amyloid peptide. *Biophys J* 69: 640-651.
16. Jackson M1, Mantsch HH (1991) Beware of proteins in DMSO. *Biochim Biophys Acta* 1078: 231-235.
17. Bax A, Davis DG (1985) MLEV-17-based two-dimensional homonuclear magnetization transfer spectroscopy. *Journal of Magnetic Resonance* 65: 355-360.
18. Bax A, Davis DG (1985) Practical aspects of two-dimensional transverse NOE spectroscopy. *Journal of Magnetic Resonance* 63: 207-213.
19. Donaldson AF. TopSPIN User Manual.
20. Van de Ven F, Blommers M, Schouten R, Hilbers C (1991) Calculation of interproton distances from NOE intensities. A relaxation matrix approach without requirement of a molecular model. *Journal of Magnetic Resonance* 94: 140-151.
21. James TL, Young GB, Broido MS, Keepers JW, Jamin N, et al. (1985) Quantitative internuclear distances via two-dimensional nuclear magnetic resonance spectra: A test case and a DNA octamer duplex. *Journal of Biosciences* 8: 553-562.
22. Jackman LM, Sternhell S, DHR Barton, W Doering (2013) Application of Nuclear Magnetic Resonance Spectroscopy in Organic Chemistry (2nd edn) International Series in Organic Chemistry, Elsevier.
23. Karplus M (1963) Vicinal proton coupling in nuclear magnetic resonance. *Journal of the American Chemical Society* 85: 2870-2871.
24. Malde AK, Srivastava SS, Coutinho EC (2007) Understanding interactions of gastric inhibitory polypeptide (GIP) with its G-protein coupled receptor through NMR and molecular modeling. *J Pept Sci* 13: 287-300.
25. Hess B, Kutzner C, Van Der Spoel D, Lindahl E (2008) GROMACS 4: algorithms for highly efficient, load-balanced, and scalable molecular simulation. *J Chem Theory Comput* 4: 435-447.
26. Van Der Spoel D, Lindahl E, Hess B, Groenhof G, Mark AE, et al. (2005) GROMACS: fast, flexible, and free. *J Comput Chem* 26: 1701-1718.
27. Lindahl E, Hess B, Van Der Spoel D (2001) GROMACS 3.0: a package for molecular simulation and trajectory analysis. *Molecular modeling annual* 7: 306-317.
28. Berendsen HJ, van der Spoel D, van Drunen R (1995) GROMACS: A message-passing parallel molecular dynamics implementation. *Computer Physics Communications* 91: 43-56.
29. Jorgensen WL (1998) OPLS force fields. *Encyclopedia of computational chemistry*.
30. Price DJ, Brooks CL 3rd (2004) A modified TIP3P water potential for simulation with Ewald summation. *J Chem Phys* 121: 10096-10103.
31. Luenberger DG (1973) Introduction to linear and nonlinear programming, Addison-Wesley Reading, MA.
32. Wüthrich K (1986) NMR of proteins and nucleic acids: Wiley New York.
33. Desai P, Coutinho E, Srivastava S, Haq W, Katti S (2002) Conformation of an immunoreactive undecapeptide fragment (10-20) of Asp f 1 by NMR and molecular modeling. *Letters in Peptide Science* 9: 21-34.
34. Bhattacharya D, Mishra N, Coutinho E, Srivastava S, Pissurlenkar R (2015) Conformational Study on Pheromontropin neuropeptide using NMR and Molecular Dynamics. *Pharm Anal Acta* 6: 2.
35. Wishart DS, Sykes BD, Richards FM (1992) The chemical shift index: a fast and simple method for the assignment of protein secondary structure through NMR spectroscopy. *Biochemistry* 31: 1647-1651.
36. Tikhele SH, Pissurlenkar RR, Srivastava S, Saran A, Coutinho EC (2010) Mapping interactions of gastric inhibitory polypeptide with GIPR N-terminus using NMR and molecular dynamics simulations. *Journal of peptide science : an official publication of the European Peptide Society* 16: 383-391.
37. Ohnishi M, Urry DW (1969) Temperature dependence of amide proton chemical shifts: the secondary structures of gramicidin S and valinomycin. *Biochem Biophys Res Commun* 36:194-202.
38. Tripathi J, Desai P, Srivastava S, Coutinho E (2009) Conformational studies of nonapeptide using NMR. *Indian journal of chemistry Section B, Organic including medicinal* 48: 103.
39. Bystrov VF (1976) Spin-spin coupling and the conformational states of peptide systems. *Progress in Nuclear Magnetic Resonance Spectroscopy* 10: 41-82.