

Cool Water Off-flavor Algae and Water Quality in Four Arkansas Commercial Catfish Farms

Lin X* and Peter P

Aquaculture/Fisheries Center, University of Arkansas at Pine Bluff, USA

Abstract

Catfish pond culture during the cool weather period of late fall, winter and early spring have experienced problems related to algal populations. Geosmin tainting by off-flavor cyanobacteria is more abundant at this time. Toxic algae episodes also appear to be more common during cool weather periods, including the devastating new golden algae (*Pyrrnesium parvum*) HAB, in adjacent Texas and believed to have occurred in Arkansas. Water and phytoplankton samples were collected weekly at 0800-1000 from November to April for three consecutive years from 12 ponds at 4 commercial catfish farms located in the southeastern Arkansas. *Oscillatoria chalybea* was found from November-January at low levels and *Pseudanabaena limnetica* occurred from January-April. Results from multiple regression show that the values of the expected number of *Anabaena circinalis* is significantly associated with dissolved oxygen, pH, total ammonia nitrogen, nitrite nitrogen, green algae, Chlorophyll a and Pheophytin a and the expected number of *Oscillatoria chalybea* is associated with total ammonia nitrogen, un-ionized ammonia, Pheophytin a cyanobacteria, and diatoms, whereas the expected number of *Pseudanabaena limnetica* is linked to temperature, total ammonia nitrogen and un-ionized ammonia, respectively.

Keywords: Cyanobacteria; Pheophytina; Arkansas; Circinalis; Phytoplankton

Introduction

Water quality during the warm catfish growing season has attracted a great deal of attention, and less so during the cool season when water quality is judged to be better due to reduced feeding (extensive management) and lower temperatures. However, winter mortalities suggest water quality issues exist, and off-flavor has been found. One of the few studies to have focused on phytoplankton and water quality during the over wintering season for catfish is that of Tucker and van Der Ploeg [1]. They evaluated phytoplankton and water quality on a monthly basis for an annual cycle in 10 commercial ponds on a central Mississippi catfish farm. They noted light was the controlling factor in phytoplankton abundance, although some ponds maintained high algal biomass through the winter. They also stated that poorest water quality occurs during the summer. Their study however did not evaluate pH or UIA, which is the most toxic compound commonly, encountered in catfish ponds. In addition, their study was of one cool weather season and consisted of once monthly sampling. Tucker and Hargreaves [2] have also speculated on the role of sediment overwintering of cyanobacteria in later dominance. Little is known on the stages leading up to massive appearance of cyanobacteria in early summer.

The algal community is affected by manmade nutrient input and herbicides, as well as natural processes. Natural processes include: climate (temperature and sunlight), water movement and turnovers, presence of competing Macrophytes and abundance of plankton-consuming macro and microorganisms. Feeding is often practiced during the warmer interludes during cool weather season, however the impact on the algal population and water quality is not known. Plankton/water quality research has focused on the growing season for fish. This is appropriate as this period is the growth period, which may be influenced by the algae and, directly and indirectly, by water quality. However, survival and health of the overwintered population in ponds is important to the success of the growing season. If the fish are in poor health going to the spring warmup period, disease can be expected, and mortalities during cool season can be substantial. Research is needed on plankton community and water quality levels during the cool weather (fall-winter-spring) season, as influenced by climatic, management and limnological conditions. Plankton populations/communities and

dynamics, including those responsible for off-flavours and toxins, need to be known for catfish pond systems. Water quality levels and dynamics research is also needed to assess stress and impacts on the cultured species. Finally, examining the substrate and water samples for cyanobacteria spores and development may provide clues to the progression of cyanobacteria dominance and enable proactive controls. The objectives of this study are: 1) to compare effects of months on water quality parameters and phytoplankton; 2) to study the relationship between water quality parameters and phytoplankton in cool season; 3) to assess relationship between cool water off-flavour algae and water quality parameter and phytoplankton. In addition, this study seeks to expand the Tucker and van Der Ploeg study by adding pH, UIA and plankton of Arkansas commercial catfish ponds.

Methods

Water and phytoplankton samples were collected from 12 production ponds at 4 commercial catfish farms located in the Mississippi River Alluvial Plain of southeastern Arkansas, and within a 16 km radius of Dumas, AR. The mean total alkalinity and hardness of commercial catfish ponds from this area ranged from 125 to 275 mg/L and 110 to 250 mg/L as CaCO₃ [3]. Commercial catfish farms were sampled weekly at 0800-1000 from November to May for three consecutive years. A water column sampler was submerged to 0.9 m at approximately the midpoint along one side of each pond and approximately 1.5 m from the shoreline. A single morning sampling was chosen to reduce sample time intervals between ponds, to coincide with low wind levels and reduce phytoplankton patchiness, and when dissolved oxygen levels were below saturation. Water samples were

*Corresponding author: Lin X, Aquaculture/Fisheries Center, Mail Slot 4912, University of Arkansas at Pine Bluff, 1200 N. University Drive, Pine Bluff, AR 71601, USA, Tel: 870-575-8157; Fax: 870-575-4637; E-mail: xiel@uapb.edu

Received November 03, 2015; Accepted December 10, 2015; Published January 01, 2016

Citation: Lin X, Peter P (2016) Cool Water Off-flavor Algae and Water Quality in Four Arkansas Commercial Catfish Farms. J Fisheries Livest Prod 4: 158. doi:10.4172/2332-2608.1000158

Copyright: © 2016 Lin X, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

returned to the laboratory in 300 ml opaque Nalgene bottles filled to overflowing in an insulated ice chest, and total dissolved solids (TDS) (mg/L), total ammonia (TAN) (mg/L), dissolved oxygen (DO), and water temperature (Temp) were obtained from a multiprobe water quality instrument. TAN and nitrite nitrogen (NO₂-N) were determined with Hach colorimeters un-ionized ammonia (UIA) were later calculated using the formula of Emerson et al. [4]. Chlorophyll a (Chla) corrected for Pheophytin a (Pheo) were determined following APHA [5]. Phytoplankton samples were taken from the column samples and fixed with 10% formalin. Phytoplankters were identified and counted by natural units from unconcentrated samples to the genus level and species level with cyanobacteria following Cocke [6].

Pearson correlation analysis was used to identify and quantify the association between water quality parameters and phytoplankton concentrations and compositions. The influence of month on the water quality and phytoplankton parameters was assessed using one-way ANOVA and multiple linear regression to exam the effect of month, water quality parameters and phytoplankton on cool water off-flavor algae *Anabaena circinalis*, *Oscillatoria chalybea*, and *Pseudanabaena limnetica*, respectively. Cool water off-flavor algae were log 10 transformed prior to analyses. Significance level was set at P < 0.05. All analyses were carried out in SAS, version 9.2 (SAS Institute Inc., Cary, North Carolina, USA).

Results and Discussion

Monthly Variations of water quality and phytoplankton parameters during the three-year combined samples were shown in (Tables 1 and 2). Ranges for water quality and phytoplankton parameters varied considerably over the cool season: DO, 4.09-15.78 mg/L; pH, 7.05-8.60; TAN, 0.01-4.42 mg/L; TDS, 290-1382 mg/L; UIA, 0.0002-0.15 mg/L; NO₂-N, 0-0.68; Chla, 0.02-1.832 mg/L; Pheo, 0.001-0.773 mg/L; cyanobacteria (BG), 0-241,560 units/mL; diatoms (D), 0-24,730 units/mL; Euglenoid algae (E), 0-24,430 units/mL; green algae (G), 0-21,130 units/mL.

From Pearson correlation analyses among all 13 parameters and cool seasons, a strong negative correlation (-0.74) was found between DO and Temp. Also, strong positive correlations (0.63 and 0.73) were also observed between UIA and TAN, and BG and Chla (Table 3).

Numbers of *Anabaena circinalis* ranged from 0 to 30,000 units/mL (5/17/07), *Oscillatoria chalybea* from 0 to 200 units/mL, and *Pseudanabaena limnetica* from 0 to 8000 units/mL (4/4/07). *Oscillatoria chalybea* filaments were observed in a shortened condition and in association with thickened one or two cell potential "spores" on 2/27 and 10/19. The prevalence of *Anabaena circinalis* was greater than *Oscillatoria chalybea* and *Pseudanabaena limnetica* during

Water Quality Parameters							
Month	DO (mg/L)	Temp (°C)	pH	TAN (mg/L)	TDS (mg/L)	UIA (mg/L)	NO ₂ -N (mg/L)
November	10.48 (0.37)bc	12.16 (0.69)c	7.71 (0.06)bcd	0.96 (0.19)abc	574.44 (56.89)ab	0.01 (0.004)abc	0.01 (0.02)a
December	10.12 (0.27)c	12.39 (0.50)c	7.67 (0.04)cd	1.21 (0.14)ab	647.47 (41.39)a	0.01 (0.003)abc	0.09 (0.01)a
January	11.37 (0.21)a	7.87 (0.39)e	7.58 (0.03)d	1.37 (0.11)a	601.76 (32.54)ab	0.01 (0.001)c	0.04 (0.01)b
February	11.19 (0.17)ab	9.48 (0.30)d	7.84 (0.04)a	0.88 (0.09)c	553.05 (25.16)ab	0.01 (0.002)bc	0.02 (0.01)b
March	8.32 (0.19)d	16.89 (0.36)b	7.80 (0.03)ab	0.75 (0.10)c	545.97 (29.49)b	0.02 (0.002)a	0.04 (0.01)b
April	6.95 (0.28)e	18.83 (0.51)a	7.70 (0.04)bc	0.90 (0.15)bc	587.22 (42.67)ab	0.02(0.003)ab	0.05 (0.01)b

Table 1: Mean (SE) monthly water quality parameters in 12 commercial catfish ponds. Means in columns with common letters are not significantly different; P>0.05. Temp=Water temperature; TAN=Total ammonia nitrogen; TDS=Total dissolved solid; UIA=Un-ionized ammonia; NO₂-N=nitrite-nitrogen.

Phytoplankton						
Month	Chla (mg/L)	Pheo (mg/L)	BG (units/mL)	D (units/mL)	E (units/mL)	G (units/mL)
November	0.25 (0.05)ab	0.058 (0.014)a	3616 (6138)c	4396 (1273)ab	548 (506)ab	1483 (579)a
December	0.28 (0.04)ab	0.067 (0.01)a	11699 (4466)bc	3657 (926)ab	1621 (368)a	1271 (421)a
January	0.24 (0.03)b	0.062 (0.008)a	10514 (3543)bc	2771 (735)b	507 (292)b	1054 (334)a
February	0.23 (0.02)b	0.056 (0.007)a	10057 (2715)c	3471 (563)b	702 (224)b	962 (256)a
March	0.34 (0.03)a	0.070 (0.007)a	19761 (3182)ab	5401 (659)a	114 (263)b	493 (300)a
April	0.34 (0.04)ab	0.061 (0.011)a	25717 (4604)a	3546 (954)ab	237 (379)b	995 (443)a

Table 2: Mean (SE) monthly phytoplankton parameters in 12 commercial catfish ponds. Means in columns with common letters are not significantly different; P > 0.05. Chla = Chlorophyll a; Pheo = Pheophytin a; BG = Blue green algae; D = Diatom; E = Euglenoid algae; G = Green algae.

	Temp	DO	pH	TAN	TDS	UIA	NO ₂	Chla	Pheo	BG	D	E	G	Total P.
Temp	1													
DO	-0.74*	1												
pH	-0.06	0.40*	1											
TAN	-0.41*	0.06	-0.35	1										
TDS	-0.26*	0.11	-0.51	0.25	1									
UIA	0.02	-0.02	0.11*	0.63*	-0.03	1								
NO ₂	-0.12	-0.14*	-0.15	0.33*	0.19	0.44*	1							
Chla	0.14*	0.01	0.32*	0.14	0.05	0.26*	0.28	1						
Pheo	0.14*	-0.15	0.44	-0.24	-0.39	0.04	0.07	0.48*	1					
BG	0.32*	-0.27	0.06	0.22*	-0.04	0.21*	0.23*	0.73*	0.09*	1				
D	0.17*	0.01	0.12	-0.39	-0.08	0.1	-0.18*	0.17	-0.01	-0.23	1			
E	-0.18	0.22*	0.1	-0.14	0.09	-0.07	-0.02	-0.04	0.06	-0.21*	0.18	1		
G	-0.26*	-0.08	-0.23	0.51	-0.17*	-0.02	0.06	-0.33*	-0.11	-0.20*	-0.27*	0.03	1	
Total P	-0.17*	-0.23*	0.25*	-0.11	0.01	0.05	-0.09	0.44*	0.25*	0.79*	0.2	0.12	0.2	1

Table 3: Correlations coefficients (R) among water quality parameters and plankton. Values ≥0.60 in bold *p <= 0.05. Total. P = Total phytoplankton.

	<i>Anabaena circinalis</i>	<i>Oscillatoria chalybea</i>	<i>Pseudanabaena limnetica</i>
Month	Mean (count/mL)	Mean (count/mL)	Mean (count/mL)
November	50 (100)	150 (100)	0 (0)
December	1225 (1812)	50 (93)	0 (0)
January	462 (626)	29 (72)	57 (185)
February	814 (975)	0 (0)	15 (53)
March	3550 (2863)	0 (0)	38 (109)
April	3942 (6265)	0 (0)	1143 (3023)

Table 4: Mean (SD) of cool water off-flavor algae in four Arkansas commercial catfish farms.

December-April (Table 4). Results from multiple regressions (Table 5) shows that the values of the expected number of *Anabaena circinalis* is significantly different between month and is significantly related to DO, pH, TDS, UIA, Chla and Pheo. The expected number of *Oscillatoria chalybea* is statistically associated with DO, pH, TAN, Chla, Pheo, BD, and D, whereas the expected number of *Pseudanabaena limnetica* is linked to DO, TAN and Chla. The deviance goodness-of-fit test indicated that the multiple regression models fit the data reasonably well. The mean number of *Anabaena circinalis*, *Oscillatoria chalybea* and *Pseudanabaena limnetica* changed dramatically during the cool season. The expected number of *Anabaena circinalis* in November differed significantly from that in April. The expected number of *Oscillatoria chalybea* in November differed significantly from the rest of the cool season. The expected number of *Pseudanabaena limnetica* was significantly different among season, with lower values found in November-March. The coefficients (0.34) for DO indicates that, if every unit increases in DO while holding other variables constant, the mean number of *Anabaena circinalis* would increase by $\exp(0.34) = 1$. While TAN is positively associated with *Oscillatoria chalybea* and negatively associated with *Anabaena circinalis* and *Pseudanabaena limnetica*, *Oscillatoria chalybea* and *Pseudanabaena limnetica* are directly related to UIA. The coefficient of 76.76 for UIA indicates that the magnitude of the effect from UIA on *Pseudanabaena limnetica* is quite large. The coefficient 0.21 for G suggests that, if every unit increases in G, mean number of *Anabaena circinalis* would increase by $\exp(0.21) = 1$.

Water temperature during the cool season directly affected DO by increased solubility at lower temperatures. Few low DO values were observed, and generally ranged from about 6-12 mg/L. DO was higher earlier in January followed by a slight decline in April. Levels of pH were lower early in the cool season, followed by a slight increase in February and March, then a decrease again in the April. The DO and pH values measured in this study were similar to those reported in the study of commercial channel catfish pond effluents in central and west-central Alabama by Schwartz and Boyd [7]. The optimum range of DO and pH for fish and aquatic life is > 4-5 mg/L and 6.0-9.0 [8], respectively. The critical parameter UIA remained relatively low and constant in all ponds during the cool season, reflecting pH values around 7 and the low temperatures. Tucker and Van Der Pleog [1] reported that chlorophyll a can reach concentrations of 450-605 $\mu\text{g}\cdot\text{L}^{-1}$ in commercial catfish ponds in Mississippi during the summer months. Only 15% of Chla measurements were above 450 mg/L during cool season.

The high correlation between Temp and DO (-0.74), and BG and Chla (0.73) observed in this study was also observed in commercial baitfish ponds (-0.70 and 0.67) [9]. The correlations between Total Phytoplankton (Total P.) and BG, BG and Chla in this study (0.79 and 0.73) was greater than 0.656 and 0.226 reported by Zimba et al. (2001). However, Total P. was less correlated with Cha (0.44) in our study than 0.711 reported by Zimba et al. [10].

Phytoplankton composition was fairly equitable in November (36.0%, 43.8% and 14.8%) BG, D, and G, respectively. However, BG dominated (64.1%, 70.8%, 66.2%, 76.7% and 84.3%) in Dec., Jan., Feb., Mar., and April, respectively. Only during December did E exceed G, and D exceeded G in all months. Tucker and van Der Pleog [1] found BG dominated all year long in three of ten ponds, and from July to September in other seven ponds, and E and small G were more abundant in spring [1]. In this study, the four instances of geosmin/earthy off-flavor odor were noted when Total P. (25,769 units/mL and

Category	Coeff.	SE	t-value	p-value
<i>Anabaena circinalis</i>				
Intercept	32.08	9.13	3.51	0.0009
Month				
December	0.81	0.68	1.19	0.2398
January	0.36	0.66	0.54	0.5932
February	0.59	0.62	0.96	0.3426
March	1.18	0.77	1.52	0.133
April	2.21	0.97	2.28	0.0266
DO	0.34	0.16	2.13	0.036
pH	-4.3	1.3	-3.31	0.0016
NO ₂	-14.17	6.61	-2.14	0.0364
TAN	-0.63	0.22	-2.88	0.0056
G	0.21	0.092	2.24	0.029
Chla	0.0028	0.0011	2.73	0.0044
Phae	-0.013	0.0035	-3.62	0.0006
Adjusted R ²	0.47			
<i>Oscillatoria chalybea</i>				
Intercept	1.29	0.29	4.51	<.0001
Month				
December	-1.44	0.32	-4.46	<.0001
January	-1.75	0.31	-5.69	<.0001
February	-1.69	0.3	-5.65	<.0001
March	-1.77	0.3	-5.85	<.0001
April	-1.97	0.34	-5.84	<.0001
TAN	0.76	0.16	4.79	<.0001
UIA	-59.97	15.63	-3.84	0.0003
Phae	0.006	0.0014	4.27	<.0001
BG	0.000016	0.000006	2.86	0.0059
D	-0.14	0.044	-3.1	0.0029
Adjusted R ²	0.51			
<i>Pseudanabaena limnetica</i>				
Intercept	3.68	1.046	3.52	0.0008
Month				
December	0.34	0.44	0.77	0.44
January	0.74	0.41	1.82	0.073
February	-0.39	0.44	-0.88	0.38
March	1.74	0.55	3.17	0.0024
April	2.86	0.71	4.01	0.0002
Temp	-0.32	0.082	-3.95	0.0002
TAN	-1.01	0.19	-5.12	<.0001
UIA	76.76	14.29	5.37	<.0001
Adjusted R ²	0.31			

Table 5: Relative importance of water quality or months for the number of *Anabaena circinalis*, *Oscillatoria chalybea* and *Pseudanabaena limnetica*, respectively. Reference was November.

30,495 units/mL) and BG percentages (76.7% and 84.3%) were highest in March and April, respectively.

Total alkalinity and hardness in this study were similar to ponds at Stoneville in the study by Hairiyadi et al. [11]. Hairiyadi et al. [11] reported BG was the prominent component of the phytoplankton at both locations (ponds at Auburn, Alabama and ponds at Stoneville, Mississippi). However, BG community composition differed between Auburn and Stoneville ponds. *Anabaena circinalis* and *Oscillatoria chalybea* both occurred at ponds at Stoneville and did not occur at ponds at Auburn. The major differences in water quality were higher pH (8.57 vs 8.32 at afternoon and 8.13 vs 7.32 at dawn), total alkalinity (244.2 vs 35.2 mg/L as CaCO₃), total hardness (251.8 vs 28.2 mg/L as CaCO₃), and dissolved inorganic phosphorus (0.107 vs 0.012 mg/L) at Stoneville and greater TAN (0.47 vs 0.12 mg/L) and nitrite-N concentration (0.016 vs 0.004 mg/L) at Auburn.

Geosmin and 2-methylisoborneol (MIB) are the most common causes of off-flavour in catfish [12,13]. These compounds are produced by species of cyanobacteria (blue-green algae) found in catfish aquaculture ponds [14]. van Der Ploeg et al. [15] found that the presence of geosmin is the main reason of off flavour in soft-water ponds (total alkalinity and hardness 15-30 mg/L as CaCO₃) in east Alabama and MIB in hard-water ponds (total alkalinity and hardness 150-300 mg/L as CaCO₃) west Mississippi. *Oscillatoria chalybea* was identified responsible for producing 2-methylisoborneol in Mississippi catfish ponds [16,17], whereas, *Anabaena circinalis* and *Pseudanabaena limnetica* were observed in commercial catfish ponds in the Mississippi-Alabama Blackland Prairie region [18]. Geosmin production by *Anabaena circinalis* has been identified in previous studies [19,20] and MIB production by *Pseudanabaena limnetica* in a drinking water reservoir in southern California [21]. No correlation was found between geosmin production and increasing light intensity or temperature [22]. However, Hurlburt et al. [23] reported that weak and positive correlation was found between geosmin concentration and air and soil temperatures at a Louisiana commercial catfish farm. Bowmer et al. [24] showed that, for *Anabaena circinalis*, the ratio of geosmin to biomass correlated with the change of light intensity. Moreover, geosmin concentration was strongly correlated with increasing ammonium-N nitrogen concentrations [25].

Conclusions

Thus, although water quality was acceptable, off flavor-causing algae *Anabaena circinalis* reached levels likely to cause geosmin-tainting during December, March and April. *Oscillatoria chalybea* was found from Nov-Jan at low levels and *Pseudanabaena limnetica* occurred from Jan-April. More research is needed to be conducted to confirm the findings.

Acknowledgements

Student assistants were Malissa Miller and Coco Lihono. USDA/CSREES Project ARX 05024 funded the three-year project.

References

1. Tucker CS, vander PM (1993) Seasonal changes in water quality in commercial channel catfish ponds in Mississippi. Journal of World Aquaculture Society 24: 473-481.
2. Tucker CS, Hargreaves JA (2004) Biology and culture of channel catfish. Elsevier, Amsterdam.
3. Farrelly JC (2014) Water quality and fish production in catfish pond production systems: tradition, intensively-aerated, and split ponds in Arkansas. University of Arkansas at Pine Bluff. Master Thesis.
4. Emerson K, Russo RC, Lund RE, Thurston RV (1975) Aqueous ammonia equilibrium calculations: effect of pH and temperature. Journal of the Fisheries Research Board of Canada 32: 2379-2383.
5. APHA (American Public Health Association) (1999) Standard methods for the examination of water and waste water (20th edn.). American Public Health Association, Washington, DC.
6. Cocke EC (1967) The myxophyceae of North Carolina. Edwards Brothers, Ann Arbor, MI.
7. Schwartz MF, Boyd CE (1994) Channel catfish pond effluents. The Progressive Fish-Culturist 56: 273-281.
8. Buttner JK, Soderberg RW, Terlizzi DE (1993) An Introduction to water chemistry in freshwater aquaculture. Northeastern Regional Aquaculture Center Fact Sheet.
9. Xie L, Perschbacher PW (2011) Water quality and zooplankton correlations from cool season sampling on four commercial baitfish farms. America Aquaculture 2011.
10. Zimba PV, Grimm CC, Dionigi CP, Weirich CR (2001) Phytoplankton community structure, biomass, and off flavor: pond size relationships in Louisiana catfish ponds. Journal of the World Aquaculture Society 32: 96-104.
11. Hariyadi S, Tucker CS, Steeby JA, Ploeg VM, Boyd CE (1994) Environmental conditions and channel catfish *Ictalurus punctatus* production under similar pond management regimes in Alabama and Mississippi. Journal of World Aquaculture Society 25: 236-249.
12. Lovell RT, Lelana IY, Boyd CE, Armstrong MS (1986) Geosmin and Musty-muddy flavors in Pond-raised channel catfish. Transactions of the American Fisheries Society 115: 485-489.
13. Martin JF, McCoy CP, Greenleaf W, Bennett L (1987) Analysis of 2-methylisoborneol in water, mud, and channel catfish (*Ictalurus punctatus*) from commercial culture ponds in Mississippi. Canadian Journal of Fisheries and Aquatic Sciences 44: 909-912.
14. Paerl HW, Tucker CS (1995) Ecology of blue-green algae in aquaculture ponds. Journal of World Aquaculture Society 26: 109-131.
15. Vander PM, Tucker CS, Boyd CE (1992) Geosmin and 2-methylisoborneol production by cyanobacteria in fish ponds in the southeastern United States. Water Science and Technology 25: 283-290.
16. Martin JF, Izaguirre G, Waterstrat P (1991) A planktonic *Oscillatoria* species from Mississippi catfish ponds that produces the off-flavor compound 2-methylisoborneol. Water Research 25: 1447-1451.
17. Vander PM, Dennis ME, de Regt MQ (1995) Biology of *Oscillatoria* cf. *chalybea*, a 2-methylisoborneol producing blue-green alga of Mississippi catfish ponds. Water Science and Technology 31: 173-180.
18. Schrader KK, Dennis ME (2005) Cyanobacteria and earthy/musty compounds found in commercial catfish (*Ictalurus punctatus*) ponds in the Mississippi Delta and Mississippi-Alabama Black land Prairie. Water Research 39: 2807-2814.
19. Henley DE (1970) Odorous metabolite and other selected studies of cyanophyta. Ph.D.thesis, North Texas State University, USA.
20. Rosen BH, MacLeod BW, Simpson MR (1992) Accumulation and release of geosmin during the growth phases of *Anabaena circinalis* (Kutz.) Rabenhorst. Water Science and Technology 25: 185-190.
21. Izaguirre G, Taylor WD (1998) A *Pseudanabaena* species from Castaic Lake, California, that produces 2-methylisoborneol. Water Research 32: 1673-1677.
22. Saadoun IM, Schrader KK, Blevins WT (2001) Environmental and nutritional factors affecting geosmin synthesis by *Anabaena* sp. Water Research 35: 1209-1218.
23. Hurlburt BK, Brashear SS, Lloyd SW, Grimm CC, Thomson JL, et al. (2009) Impact of weather on off flavor episodes at a Louisiana commercial catfish farm. Aquaculture Research 40: 566-574.
24. Bowmer K, Padovan A, Oliver RL, Korth W, Ganf GG (1992) Physiology of geosmin production by *Anabaena circinalis* isolated from the Murrumbidgee River, Australia. Water Science and Technology 25: 259-267.
25. Schrader KK, de Regt MQ, Tidwell PD, Tucker CS, Duke SO (1998) Compounds with selective toxicity towards the off-flavor metabolite-producing Cyanobacterium *Oscillatoria* cf. *chalybea* Aquaculture 163: 85-99.