Research Article Open Access

Correlation between bla_{SHV} Gene and Biofilm Formation among Beta Lactamase Producing Uropathogenic Isolates from Patients in Erbil City in Iraq

Hosan Yousif Hassan^{1*}, Safaa Toma Hanna Aka¹ and Salah Tofik Jalal²

- ¹Department of Pharmacognosy, College of Pharmacy, Hawler Medical University, Erbil City, Kurdistan Region, Iraq
- ²Department of Medical Microbiology, College of Health Sciences, Hawler Medical University, Erbil City, Kurdistan Region, Iraq

Abstract

Background and objective: Bacterial resistance has been considered as one of the most serious threats in infectious disease, especially in development countries. The ability of biofilm formation is often associated with antibiotic resistance. The aim of the study is to detect the correlation between the ability of biofilm formation and *SHV* gene, in addition to investigate the prevalence of *SHV* gene among the beta-lactamase producing strains.

Methods: This cross-sectional study was carried out during the period September to December 2017 at Rizgary Teaching Hospital in Erbil. A total of 31 beta-lactamase producing uropathogens were collected, identification and antibiotic sensitivity test was done using VITEK-2 automated system. Biofilm formation was determined by 96 well flat bottom microtiter plates. Molecular detection was performed for one of the ESBL genes, the *SHV* gene.

Results: The prevalence of SHV gene among the beta-lactamase producing isolates was 29%. The findings showed a significant association between the presence of SHV gene and the bacterial species (p=0.001). In addition 44.4% of SHV positive isolates were K. pneumoniae pathogens with the ability of moderate or strong biofilm formation which showed to have a significant association (p=0.044). From the beta-lactamase producing strains 90.3% appeared to be multi-drug resistant isolates.

Conclusion: The SHV gene was significantly more present in K. pneumoniae in compare to E. coli strains. However, most of SHV positive isolates were K. pneumoniae pathogens with ability of moderate or strong biofilm formation.

Keywords: Biofilm; SHV gene; Uropathogens; ESBL

Introduction

Bacterial resistance has been considered as one of the most serious threats in infectious disease, especially in development countries. However a geographical difference has been observed in the antibiotic sensitivity profile of uropathogens. A study from India reported that most of the uropathogenic *E. coli* strains were multi-drug resistant [1].

The World Health Organization (WHO) has classified resistant bacteria according to critically of need for new antibiotics into three groups; critical, high and medium priority. Enterobacteriaceae extended spectrum β -lactamase (ESBLs) enzymes producing bacteria are classified within the critical priority. ESBLs have the ability to hydrolyze broad spectrum of β lactams (third-generation cephalosporins) and monobactams, but do not affect carbapenems or cephamycin [2-4]. Based on their primary structure ESBLs are classified into four classes A, B, C and D enzymes. The class A ESBLs include cefotaximase (*CTX-M*), temoneira (*TEM*) and sulfhydryl variable (SHV) [5]. Additionally it is reported that the most common site for ESBL production are UTIs [6]. Biofilm related infections are often associated with antibiotic resistance [7].

Biofilms are significant colonizers of medical devices for instance urinary, arterial and venous catheters [8]. The main cause of catheter-associated UTI is due to the biofilm formation on the catheter surface [9]. Studies reported that about 80% of all infectious diseases are biofilm related and are also responsible for more than 60% of nosocomial infections. Empirical or antibiotic treatment based on antibiotic sensitivity test is usually inefficient towards infection caused by biofilm producing isolates [10]. Many of the recurrent UTIs are assumed to be caused by biofilm producing uropathogenic isolates [11].

Therefore the aim of the study is to detect the correlation between the ability of biofilm formation and SHV gene, in addition to investigate

the prevalence of SHV gene among ESBL-isolates. Furthermore we observed the antibiotic sensitivity profile of beta-lactamase producing strains.

Materials and Methods

Study design and specimen collection

This cross-sectional study was carried out during the period September to December 2017. A total of 31 beta-lactamase producing uropathogens of *E. coli* and *K. pneumoniae* were collected from patients attending Urology Department at Rizgary Teaching Hospital in Erbil city/ Iraq. The identification and antibiotic sensitivity test was done using VITEK-2 automated system with the ESBL test panel, quality control strains (*E. coli* ATCC 25922).

Biofilm assay

Biofilm formation was determined by the 96 well tissue culture plate (TCP) assay, as described in previous studies [12]. Biofilms were stained with 0.1% crystal violet solution for 15 minutes and the plates were washed in distilled water and dried at room temperature. The optical density (OD) of the biofilms was measured at the wavelength

*Corresponding author: Hosan Yousif Hassan, Department of Pharmacognosy, College of Pharmacy, Hawler Medical University, Erbil city, Kurdistan Region, Iraq, Tel: 004917658401906; E-mail: hozan_aspindara@yahoo.de

Received May 27, 2019; Accepted June 14, 2019; Published June 21, 2019

Citation: Hassan HY, Aka STH, Jalal ST (2019) Correlation between *bla_{SHV}* Gene and Biofilm Formation among Beta Lactamase Producing Uropathogenic Isolates from Patients in Erbil City in Iraq. Mol Biol 8: 231.

Copyright: © 2019 Hassan HY, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

630 nm using an ELISA reader. Briefly the isolates were classified into four different categories according to their biofilm-forming ability depending on their 2 ODc (the average OD of the control negative at threefold its standard deviation) of bacteria. The categories were classified as follows, not able to form a biofilm (OD \leq ODc), weakly able to form a biofilm (ODc<OD \leq 2xODc), moderately able to form a biofilm (2xODc < OD \leq 4xODc), strongly able to form a biofilm (4xODc<OD).

Molecular detection of bla_{SHV} gene

All isolates which were tested as ESBL positive by VITEK-2 system were screened for detection of $bla_{\rm SHV}$ Each bacterial isolate was sub cultured on MacConkey agar for 24 hrs at 37°C. From the plates several single colonies were picked and suspended in 500 mL sterile distal water, and then the bacteria were lysed by heating at 95 °C for 10 minutes. The cell debris were removed by centrifugation at 16000 rpm for 2 minutes. PCR amplification were performed in a final volume of 25 μ l containing 12.5 μ l Master mix, 2.5 μ l of each primer forward and backwards, 5 μ l DNA template and 2.5 μ l of nuclease free water. Primers have been used in previous studies, bla_{SHV} forward primer 5'TTAACTCCCTGTTAGCCA 3' and reverse 5'GATTTGCTGATTTCGCCC 3' have been used with a product size 768 base pairs [13].

The cycling parameters were as followed, initial denaturation at 94 $^{\circ}$ C for 3 min followed by 30 cycles. Each denaturation at 94 $^{\circ}$ C for 30 seconds, annealing at 50 $^{\circ}$ C for 30 seconds, amplification at 72 $^{\circ}$ C for 2 minutes and final extension for 10 min [14]. Finally the amplified products were applied to electrophoresis with 1.2% agarose gel in 1XTBE buffer.

Ethical approval

Approval number 181017/78 was obtained from the Ethical committee at College of Pharmacy, Hawler Medical University. In addition, during data collection all of the attended patients were asked for verbal consents.

Statistical analysis

Data analysis was performed using the statistical package for social sciences (SPSS) version 23. The Chi square and Fischer's exact test was used to evaluate any association between two variables a *p* value equal to or less than 0.05 was considered statistically significant.

Results

The antibiotic sensitivity profile of ESBL producing isolates

The antibiotic sensitivity profile of the ESBL isolates indicate that about 90.3% (n=28) were multi-drug resistant (MDR) bacteria, with resistance to more than three classes of antibiotics.

Resistance towards cephalosporin antibiotic class and penicillin class was 100%. Furthermore, the resistance for ciprofloxacin, trimethoprim and gentamycin was moderate with each of 58% (n=18), 58% (n=18) and 64.5% (n=20). In addition nitrofurantoin resistance or intermediate profile was observed in 19.3% (n=6) of the isolates. However, all bacterial strains showed to be sensitive to carbapenem class antibiotics.

The prevalence of ${\it bla}_{\it SHV}$ gene and the distribution according to bacterial species

From the total 31 phenotypically confirmed ESBL isolates, 29% (n=9) possessed the resistant *SHV* gene. The gene was present in all *K. pneumoniae* isolates 100% (n=5), while only 15.4% (n=4) of *E. coli*

strains harbored the gene. A significant difference has been observed between the presence of SHV gene and the bacterial species (p=0.001) as described in Table 1.

Correlation between bla_{SHV} gene and biofilm production

About 44.4% (n=4) of the SHV positive isolates were moderate or strong biofilm producers, while only 4.5% (n=1) from the SHV negative isolates showed moderate or strong biofilm producers. A significant association with p = 0.044 is present between the presence of SHV gene and the strength of biofilm formation as described in Table 2. Figure 1 showing the PCR results for 31 samples of this study, bands of expected size (768 bp) were seen in positive samples.

Discussion

The emergence of MDR bacteria have changed simple infections into complicated ones. The antibiotic resistance profile of beta-lactamase producing bacteria of the present study was examined, and 90.3% appeared to be MDR isolates. Studies described antibiotic resistance and its significant association with MDR [15,16]. Furthermore, the pathogens were 100% susceptible to last resort antibiotic carbapenems, a finding which is in agreement with other investigations [15,17]. However, in contrast, a recent investigation from Egypt reported 20.4% resistance towards class carbapenems antibiotic [18]. From the current study, resistance of ciprofloxacin was 58% and nitrofurantoin 19.3%. These findings are higher than those reported from investigations in Europe and America which ranged for ciprofloxacin between 4%-39.8% with a susceptibility rate for nitrofurantoin above 99.4% [17,19-22]. Furthermore, studies from India and the Middle East countries described a moderate resistance for ciprofloxacin ranging from 45%-55% and nitrofurantoin 7.5% [14,16,23].

| Bacterial species | SHV gene | | | | |
|-------------------|------------------|------------------|---------------|---------|--|
| | Negative No. (%) | Positive No. (%) | Total No. (%) | p-value | |
| E. coli | 22 (84.6) | 4 (15.4) | 26 (100) | | |
| K. pneumoniae | 0 (0) | 5 (100) | 5 (100) | 0.001 | |
| Total | 22 (71) | 9 (29) | 31 (100) | | |

Table 1: The association between SHV gene and the bacterial species.

| | SHV gene | | | | |
|--------------|------------------|------------------|---------------|---------|--|
| Biofilm | Negative No. (%) | Positive No. (%) | Total No. (%) | p-value | |
| Non-adherent | 8 (36.4) | 1 (11.1) | 9 (29) | | |
| Weak | 13 (59.1) | 4 (44.4) | 17 (54.8) | | |
| Moderate | 1 (4.5) | 3 (33.3) | 4 (12.9) | | |
| Strong | 0 (0) | 1 (11.1) | 1 (3.2) | 0.044 | |
| Total | 22 (100) | 9 (100) | 31 (100) |] | |

Table 2: The association between the strength of biofilm formation and SHV gene.



Figure 1: Amplification of bla_{SHV} gene from 8 samples of both *E. coli* and *K. pneumoniae*. Agarose gel electrophoresis of PCR amplicons for the target gene. M: 100-1000 bp DNA molecular weight marker, lanes 1-4 are *K. pneumoniae* isolates and lanes 5-8 are *E. coli*.

The prevalence of *SHV* gene among the ESBL producing *E. coli* and *K. pneumoniae* isolates in the study was 29%, which is higher in compare to European countries like Norway with 5% [24,25], the United Kingdom 2.4% (25), Portugal 23.3% [26]. On the other hand, the rate is higher in Kenya 33% [27], India 57.5% [28], Thailand 87.4% [29]. Furthermore, studies from neighboring countries like Iran which included only *K. pneumoniae* samples reported a prevalence of 69.44% [30]. In Iraq, investigations reported a relatively high percentage of *SHV* gene among *K. pneumoniae* which ranged between 55%-87.7% [14,31].

The resistance gene was observed in 100% of K. pneumoniae isolates in compare to *E. coli* in which only 15.4% harbor the gene. The findings showed a significant association between the presence of SHV gene and the bacterial species (p=0.001) which is in agreement with other investigations [14,32,33]. The SHV gene was screened mainly among K. pneumoniae, a possible explanation might be that the SHV-1 was categorized as a plasmid-mediated β-lactamase. It is believed that K. pneumoniae isolates encode the beta-lactamase enzyme on their chromosomes [14,34]. Furthermore, the present investigation stated that 44.4% of SHV positive isolates were K. pneumoniae pathogens with the ability of moderate or strong biofilm formation which showed to have a significant association (p=0.044). A possible explanation might be that for K. pneumoniae, the presence of SHV gene and strength of biofilm production are significantly correlated to each other. A limitation of the study is that from the three genes SHV, CTX, and TEM of ESBL, PCR was performed only for SHV gene.

Conclusion

The *SHV* gene was significantly more present in *K. pneumoniae* isolates in compare to *E. coli*. However, the present investigation stated that most of *SHV* positive isolates were *K. pneumoniae* pathogens with ability of moderate or strong biofilm formation.

References

- Niranjan V, Malini A (2014) Antimicrobial resistance pattern in Escherichia coli causing urinary tract infection among inpatients. Indian J Med Res 139: 945-049.
- 2. Pfaller MA, Segreti J (2006) Overview of the epidemiological profileand laboratory detection of extended-spectrum β -Lactamases. Clin Infect Dis. 42 Suppl 4: S153-S163.
- Freire-Moran L, Aronsson B, Manz C, Gyssens IC, So AD, et al. (2011) Critical shortage of new antibiotics in development against multidrug-resistant bacteria-Time to react is now. Drug Resist Updat 14: 118-124.
- 4. World Health Organization (2018) WHO Publishes List of Bacteria for Which New Antibiotics Are Urgently Needed. Geneva.
- Seyedjavadi SS, Goudarzi M, Sabzehali F (2016) Relation between blaTEM, blaSHV and blaCTX-M genes and acute urinary tract infections. J Acute Dis 5: 71-76.
- Tansarli GS, Poulikakos P, Kapaskelis A, Falagas ME (2014) Proportion
 of extended-spectrum β-lactamase (ESBL)-producing isolates among
 Enterobacteriaceae in Africa: evaluation of the evidence-systematic review. J
 Antimicrob Chemother 69: 1177-1184.
- Fux CA, Costerton JW, Stewart PS, Stoodley P (2005) Survival strategies of infectious biofilms. Trends Microbiol 13: 34-40.
- Davies D (2003) Understanding biofilm resistance to antibacterial agents. Nat Rev Drug Discov 2: 114-122.
- Svensson L, Poljakovic M, Demirel I, Sahlberg C, Persson K (2018) Host-Derived Nitric Oxide and Its Antibacterial Effects in the Urinary Tract. Adv Microb Physiol 73: 1-62.
- 10. Di Domenico EG, Toma L, Provot C, Ascenzioni F, Sperduti I, et al. (2016) Development of an in vitro assay, based on the biofilm ring test®, for rapid profiling of biofilm-growing bacteria. Front Microbiol 7: 1429.
- 11. Rijavec M, Müller-Premru M, Zakotnik B, Žgur-Bertok D (2008) Virulence factors

- and biofilm production among Escherichia coli strains causing bacteraemia of urinary tract origin. J Med Microbiol 57: 1329-1334.
- Stepanović S, Vuković D, Dakić I, Savić B, Švabić-Vlahović M (2000) A modified microtiter-plate test for quantification of staphylococcal biofilm formation. J Microbiol Methods 40: 175-179.
- Briñas L, Zarazaga M, Sáenz Y, Ruiz-Larrea F, Torres C (2002) β-Lactamases in ampicillin-resistant Escherichia coli isolates from foods, humans, and healthy animals. Antimicrob Agents Chemother 46: 3156-3163.
- Haji SH, Jalal ST, Omer SA, Mawlood AH (2018) Molecular detection of SHV-Type ESBL in E. coli and K.pneumoniae and their antimicrobial resistance profile. Zanco J Med Sci 22: 262-272.
- Al-Mayahie SM (2013) Phenotypic and genotypic comparison of ESBL production by vaginal Escherichia coli isolates from pregnant and non-pregnant women. Ann Clin Microbiol Antimicrob 12: 7.
- Rajabi Z, Dallal MS, Sabbaghi A, Molla Aghamirzaeie H, Lari AR, et al. (2013)
 Prevalence of AmpC and SHV β-lactamases in clinical isolates of Escherichia coli from Tehran Hospitals. Jundishapur J Microbiol 6: 176-180.
- Patel AH, Bhavsar RH, Trivedi P, Mehta SR (2015) Urinary tract infection in children: Clinical profile, bacteriology and antibiotic sensitivity pattern. GCSMC J Med Sci 4: 75-81.
- Mahgoub FM, El-Gamal SAE-H (2018) Microbiological Profile of Urinary Tract Infections with special Reference to Antibiotic Susceptibility Pattern of Escherichia coli Isolates. Int J Curr Microbiol App Sci 7: 911-920.
- Coque TM, Baquero F, Canton R (2008) Increasing prevalence of ESBLproducing Enterobacteriaceae in Europe. Euro Surveill 13: 19044.
- Kamenski G, Wagner G, Zehetmayer S, Fink W, Spiegel W, et al (2012)
 Antibacterial resistances in uncomplicated urinary tract infections in women:
 ECO·SENS II data from primary health care in Austria. BMC Infect Dis 12: 222.
- Kresken M, Körber-Irrgang B, Biedenbach DJ, Batista N, Besard V, et al. (2016) Comparative in vitro activity of oral antimicrobial agents against Enterobacteriaceae from patients with community-acquired urinary tract infections in three European countries. Clin Microbiol Infect 22: 63e1-63e5.
- 22. Frazee BW, Trivedi T, Montgomery M, Petrovic DF, Yamaji R, et al. (2018) Emergency Department Urinary Tract Infections Caused by Extended-Spectrum β-Lactamase-Producing Enterobacteriaceae: Many Patients Have No Identifiable Risk Factor and Discordant Empiric Therapy Is Common. Ann Emerg Med 72: 449-456.
- Tadepalli S, Prudhivi S, Babu Myneni R, Rao S (2016) Biofilm formation in uropathogenic Escherichia coli isolates and its association with extended spectrum betalactamase production and drug resistance. Saudi J Pathol Microbiol 1: 260-264.
- Søraas A, Sundsfjord A, Sandven I, Brunborg C, Jenum Pal A (2013) Risk factors for community-acquired urinary tract infections caused by ESBLproducing enterobacteriaceae-a case-control study in a low prevalence country. PLoS One 8: e69581.
- Reid R, Baho S, Samarasinghe S (2017) Genotypic Identification of ESBL Producing Urinary Tract Infections In Leicestershire Area, UK. ASM Microbe 2017.
- Fernandes R, Amador P, Oliveira C, Prudêncio C (2014) Molecular characterization of ESBL-producing Enterobacteriaceae in northern Portugal. ScientificWorldJournal 2014: 782897.
- 27. Maina D, Revathi G, Whitelaw AC (2017) Molecular characterization of multidrug-resistant Klebsiella pneumoniae and Escherichia coli harbouring extended spectrum beta-lactamases and carbapenemases genes at a tertiary hospital, Kenya. Microbiologia Medica 32.
- 28. Sharma M, Pathak S, Srivastava P (2013) Prevalence and antibiogram of Extended Spectrum β-Lactamase (ESBL) producing Gram negative bacilli and further molecular characterization of ESBL producing Escherichia coli and Klebsiella spp. J Clin Diagn Res 7: 2173-2177.
- 29. Kiratisin P, Apisarnthanarak A, Laesripa C, Saifon P (2008) Molecular characterization and epidemiology of extended-spectrum-β-lactamase-producing Escherichia coli and Klebsiella pneumoniae isolates causing health care-associated infection in Thailand, where the CTX-M family is endemic. Antimicrob Agents Chemother 52: 2818-2824.
- Izadi N, Naderi Nasab M, Harifi Mood E, Meshkat Z (2014) Prevalence of TEM and SHV Genes in Clinical Isolates of Klebsiella Pneumonia From Mashhad, North-East Iran. Iran J Pathol 9:199-205.

Citation: Hassan HY, Aka STH, Jalal ST (2019) Correlation between bla_{SHV} Gene and Biofilm Formation among Beta Lactamase Producing Uropathogenic Isolates from Patients in Erbil City in Iraq. Mol Biol 8: 231.

Page 4 of 4

- 31. Khalid HM, Yousif SY, Jubrael JM (2017) Bacteriological and molecular characterization of extended spectrum b-lactamases in clinical isolates of klebsiella pneumoniae isolated from kurdistan region, Iraq. J Univ Zakho 1: 158-163.
- 32. Bali EB, Accedil L, Sultan N (2010) Phenotypic and molecular characterization of SHV, TEM, CTX-M and extended-spectrum-lactamase produced by Escherichia coli, Acinobacter baumannii and Klebsiella isolates in a Turkish hospital. Afr J Microbiol Res 4: 650-654.
- 33. Hassan MI, Alkharsah KR, Alzahrani AJ, Obeid OE, Khamis AH, et al. (2013) Detection of extended spectrum beta-lactamases-producing isolates and effect of AmpC overlapping. J Infect Dev Ctries 7: 618-629.
- 34. Paterson DL, Ko W-C, Von Gottberg A, Casellas JM, Mulazimoglu L, et al. (2001) Outcome of cephalosporin treatment for serious infections due to apparently susceptible organisms producing extended-spectrum β-lactamases: implications for the clinical microbiology laboratory. J Clin Microbiol 39: 2206-2212.

Mol Biol, an open access journal ISSN: 2168-9547