

Correlation of Fibroblast Growth Factor 23 with Markers of Inflammation and Endothelial Dysfunction in End-Stage Renal Disease and Type 2 Diabetes Patients on Peritoneal Dialysis

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Abstract

Objective: This study was design to evaluate the correlation between FGF-23 and other cardiovascular risk factors (markers of endothelial dysfunction and inflammation) in ESRD patients with T2DM on PD.

Method: Sixty serum samples collected (four times, at weeks 0, 8, 10, and 18) from 15 patients with Type 2 Diabetes (T2DM) and ESRD on PD were analysed to study the relationship between the levels of FGF-23 and markers of endothelial dysfunction and inflammation. ELISA kits were used to quantitate Endothelin-1 (ET-1), soluble vascular adhesion molecule, Plasminogen Activator Inhibitor-1 (PAI-1), FGF-23, C-reactive protein, Interleukin-6 (IL-6), Tumor Necrosis Factor Alpha (TNF- α) and cluster of differentiation 146 (CD 146). The association between FGF-23, mineral metabolites, and markers of inflammation were analyzed using Spearman correlations. Linear regression models were used to examine the univariate and multivariate adjusted associations between FGF-23 as primary exposure and individual inflammatory markers [including IL-6, PAI-1 and CD146] as the dependent variables. Multivariate analyses were adjusted for factors associated with mineral metabolism such as serum phosphate, Ca \times PO₄ product, and LDL concentration. Two-sided p values < 0.05 were considered statistically significant.

Result: FGF-23 was positively correlated with phosphate (r=0.57; p<0.0001) and Ca \times PO₄ product (r=0.61; p<0.0001). FGF-23 showed the strongest correlation with IL-6 (r=0.32; p<0.05), PAI-1 (r=0.21; p<0.05) and CD 146 (r=0.29; p<0.05). In univariate and multivariate regression analysis, FGF-23 was significantly associated with IL-6, PAI-1, and CD 146. These results were qualitatively unchanged in the model that was further adjusted for Ca \times PO₄ product, serum phosphate, and LDL.

Conclusion: Our results indicate that FGF-23 impacts the cardiovascular health of T2DM patients on PD through mechanisms, which are independent from phosphate levels and linked directly to inflammation and endothelial dysfunction.

Keywords: Fibroblast growth factor-23; End-stage renal disease; Peritoneal dialysis; Inflammatory markers; Type 2 diabetes; Endothelial dysfunction; Mineral metabolites

Introduction

Distorted bone and mineral metabolism are considered as risk factors for cardiovascular mortality in patients with End-Stage Renal Disease (ESRD). Fibroblast Growth Factor-23 (FGF-23), produced in the bone, is a strong predictor of adverse cardiovascular events in ESRD patients [1]. A strong correlation exists between FGF-23 level and mortality in this patient [2-5]. FGF-23 is considered as a unique risk marker for the cardiovascular mortality in Chronic Kidney Disease (CKD) patients [6]. The FGF-23 concentration remains unchanged throughout the day in CKD patients [7]. Increased Soluble Parathyroid Hormone (S-PTH) and FGF-23 levels are associated with the progression of vascular calcification in Peritoneal Dialysis (PD) patients [8].

Gutierrez et al. reported that increased levels of FGF-23 are linked with a noticeable increase in mortality and sickness in CKD patients,

ESRD patients, and kidney transplant recipients [3]. However, the clinical outcome in ESRD patients is also associated with the level of systemic inflammation and endothelial dysfunction. Elevated serum C-reactive protein (CRP) is a strong and more robust predictor of cardiovascular mortality than well-established risk factors such as LDL cholesterol level. The association between elevated CRP and increased mortality is reported in both Hemodialysis (HD) [9,10] and PD [8,11] populations. Several previous experimental, clinical, and interventional studies have documented that endothelial dysfunction; vascular calcification, oxidative stress, and inflammation are central factors in the complex control of mineral metabolism [12]. Patients with diabetes are the most vulnerable due to compromised metabolism [13].

Recent studies demonstrated that vascular disease is an inflammatory condition [14,15]. Cytokines initiate and coordinate inflammatory responses that involve increased hepatic production of acute phase reactants, including CRP, interleukin 6 (IL-6), Tumor Necrosis Factor-alpha (TNF- α), and cluster of differentiation 146 (CD146) [10,16]. Elevated serum levels of pro-inflammatory cytokines

such as CRP and intact Parathyroid Hormone (iPTH) have been linked to increased mortality in dialysis patients [16]. A strong association is observed between inflammation and atherosclerosis in Hemodialysis (HD) patients [3,16]. Inflammatory markers are independent predictors of atherosclerotic plaques in the carotid arteries [9-12,15]. These inflammatory markers are strong predictors of subsequent cardiovascular disease and cardiovascular mortality, both in the general population and among patients on dialysis [14]. Inflammation is believed to be linked to endothelial dysfunction [14].

The aim of our study was to evaluate the correlation between FGF-23 and other more traditional cardiovascular risk factors (CRP, lipids, and markers of endothelial dysfunction and inflammation) in ESRD patients with T2DM on PD.

Methods

Sixty serum samples collected four times at weeks 0, 8, 10, and 18 from 15 adult subjects between 18- and 65-year-old with T2DM on PD whose serum phosphate levels were above 5.5 mmol/L were analyzed. Patients were enrolled from the Amarillo Kidney Specialist, LLC dialysis unit after providing informed consent. The study protocol was approved by the Texas Tech University Health Sciences Center Institutional Review Board. Subjects with Parathyroid Hormone (PTH) levels greater than 1,000 mmol/L, calciphylaxis, or a history of hypercalcemia within the past three months were excluded from the study. Whole blood was collected in a vacutainer (Becton Dickinson, Franklin Lake, NJ). After collection of the blood, it was allowed to clot at room temperature for 30 minutes. The clot was removed by centrifugation at 1,000-2,000 x g for 10 minutes in a refrigerated centrifuge. Serum was transferred to clean polypropylene tubes and 0.5 ml aliquots were stored at -20°C. Serum samples were analyzed for FGF-23, ET-1, PAI-1, soluble Vascular Adhesion Molecule (sVCAM), soluble Intercellular Adhesion Molecule (sICAM), IL-6, Interleukin 1β (IL-1β), TNF-α, CRP, and CD146. Endothelial function was evaluated by measurement of flow mediated dilation by Endo-PAT (Itamar Medical, Franklin, MA). In addition, serum was analyzed for albumin, calcium, phosphate, PTH, and lipids by a commercial laboratory (Quest Laboratory, Amarillo, TX).

Serum FGF-23, sVCAM, sICAM, ET-1, PAI-1, CD146 and pro-inflammatory cytokines

Enzyme-linked Immunosorbent Assays (ELISAs) were used according to manufacturer's instructions (R&D Systems, Minneapolis, MN) to measure the serum levels of FGF-23, sVCAM, sICAM, ET-1, PAI-1, IL-6, IL-1β, TNF-α, and CRP. The direct ELISA method was used to measure CD146 levels [17].

Statistical analysis

ELISA data were analyzed by using 4 parameter logistic equations (Supplementary data). Continuous variables were summarized as mean ± SEM (Table 1). FGF-23, IL-6, CRP, TNF-α, were natural log (ln)-transformed. Associations between FGF-23, mineral metabolites, and markers of inflammation were analyzed using Spearman correlations. Univariate and multivariate regression analyses were conducted to examine the relationships between FGF-23, inflammatory markers (IL-6), markers of endothelial dysfunction (PAI-1, CD146), and mineral metabolites. In these, analyses were limited to the inflammatory markers that were found positive and significantly correlated with FGF-23 by Spearman test.

Socio-demographic	
Sex	8 female, 7 male
Age (Years)	56.6 ± 2.4
BMI	34.0 ± 1.3
Anemia	
Hematocrit (%)	35.49 ± 0.61
HCT CALC (HGBX3)	34.94 ± 0.62
Hemoglobin (g/dL)	11.68 ± 0.20
Iron management	
Iron saturation (%)	34.47 ± 3.51
Iron	79.57 ± 7.78
TIBC (µg/dL)	233.18 ± 9.05
UIBC (µg/dL)	149.67 ± 12.46
Osteodystrophy	
Calcium (mg/dL)	9.11 ± 0.33
Phosphorus (mg/dL)	5.75 ± 0.48
Cal×Phos product	51.56 ± 4.48
Alkaline Phosphatase (mg/dL)	111.13 ± 20.40
Calcium corrected	9.35 ± 0.32
Cal×Phos product corrected	53.11 ± 4.68
Nutrition and Lipids	
Potassium (mg/dL)	4.49 ± 0.13
Albumin (mg/dL)	3.70 ± 0.10
Adequacy of dialysis and renal function	
Creatinine (µmoles/L)	7.44 ± 0.81
Blood Urea Nitrogen	54.73 ± 3.04
Kt/V (K=dialyzer clearance of urea, t=dialysis time and V=volume of distribution of urea in the body)	2.47 ± 0.13
Residual renal function (mL/Min)	6.68 ± 1.36
Duration of dialysis	9 patients (>6 months) 6 patients (<6 Months)
Dialysis regimen	2 patients were on CAPD 13 Patients were on CCPD

Table 1: Baseline characteristics of patients

TIBC: Total Iron Binding Capacity; UIBC: Unsaturated Iron Binding Capacity; CAPD: Continuous Ambulatory Peritoneal Dialysis; CCPD: Continuous Cycling Peritoneal Dialysis

Univariate regression analysis was done in order to check to see if FGF-23 had influenced one of the variables. Using univariate

regression analysis, a model was created by using FGF-23 as a dependent variable and IL-6, PAI-1, and CD146 as single independent variables. The same univariate model was adjusted by adding three more independent variables, i.e. calcium \times phosphorus product (Ca \times PO₄ product), serum phosphate (Phos) and LDL. Multiple regression analysis was performed to analyze whether mineral metabolism significantly affected associations that were found using univariate regression.

Two-sided p values <0.05 were considered to represent statistically significant differences. Data were analyzed using IBM SPSS version 20.0 (SPSS, Chicago, IL).

Variables	FGF-23	ET-1	sICAM	IL6	PAI-1	TNF α	sVCAM	IL-1 β	CD146	CRP
FGF-23	1.00	0.13	-0.17	0.36*	0.21*	0.01	-0.12	-0.12	0.29*	0.14
Calcium (mg/dL)	0.11	-0.26*	0.08	-0.47*	-0.11	0.04	-0.06	-0.12	-0.10	-0.27*
Phosphate (mg/dL)	0.54*	0.49*	-0.11	0.45*	0.11	0.07	0.20*	0.24*	0.22*	0.19
Ca \times PO ₄ product	0.63*	0.41*	-0.09	0.29*	0.07	0.09	0.19	0.20	0.19	0.09
Triglyceride (mg/dL)	-0.23	-0.16	0.48*	0.05	0.34*	0.11	-0.33*	0.31*	0.19	-0.001
LDL (mg/dL)	-0.28*	0.08	0.19	-0.47*	-0.14	-0.15	0.11	0.18	-0.02	-0.17
HDL (mg/dL)	-0.004	0.26*	-0.42*	0.01	-0.16	-0.18	0.53*	-0.05	-0.34*	0.10
Total Cholesterol	-0.35	0.19	0.30	-0.36	0.02	-0.14	0.14	0.35	0.01	-0.06
Intact PTH (pg/mL)	-0.02	0.12	-0.05	0.02	0.01	-0.08	0.02	-0.04	-0.29*	0.03

Table 2: Spearman correlation between markers of mineral metabolism and cardiovascular risks factors; FGF-23: Fibroblast Growth factor-23; ET-1: Endothelin-1; sICAM: soluble Intercellular Adhesion Molecule; IL6: Interleukin 6; PAI-1: Plasminogen Activator Inhibitor-1; TNF α : Tumor Necrosis Factor-alpha; sVCAM: soluble Vascular Adhesion Molecule-1; IL-1 β : Interleukin 1 β ; CD 146: Cluster of Differentiation 146; CRP:C-Reactive Protein; Ca \times PO₄product: Calcium Phosphorus product; LDL: Low-Density Lipoprotein; HDL: High-Density Lipoprotein. * p<0.05 correlations are statistically significant. Serum concentrations of FGF-23, sICAM, PAI-1, sVCAM, and CD146 were measured in ng/mL; Serum concentrations of ET-1, IL-6, TNF α , IL-1 β , and CRP were measured in pg/mL.

FGF-23 also had strong positive and significant correlations with Phos (r=0.54; p<0.05), Ca \times PO₄ product(r=0.63; p<0.05), and LDL(r=0.28; p<0.05).

The multiple regression model with all four predictors produced adjusted R²=0.54, 0.55, 0.57; p<0.001, respectively for IL-6, PAI-1, and CD146 (Table 3). This indicated strong associations between FGF-23 and IL-6, PAI-1, and CD146.

Variable	Univariate		Multivariate adjusted model†	
	Difference (95% CI)	p value	Difference (95% CI)	p value
lnFGF-23				
lnIL-6	0.11	p=0.01	0.54	p<0.001
lnPAI-1	0.02	p=0.01	0.55	p<0.001
lnCD 146	0.06	p=0.05	0.57	p=0.05

Table 3: Univariate and multivariate linear regression analyses between inflammatory markers as dependent variables and fgf-23 as the principal factor

CI: Confidence interval; Ln: Log-normal; FGF-23: Fibroblast Growth Factor-23; IL-6: Interleukin 6; PAI-1: Plasminogen Activator

Results

Socio-demographic characteristics and basic biochemical data of 15 patients are presented in Table 1. As expected, the study participants had elevated Phos levels. FGF-23 showed the significant and strong positive correlation with inflammatory marker IL6 (r=0.36; p<0.05), and with two of the endothelial dysfunction markers: PAI-1 (r=0.21; p<0.05) and CD 146 (r=0.29; p<0.05) (Table 2). In addition, both IL-6 and IL-1 β were positively correlated with levels of Phos, and IL-6 was positively correlated with the CaxPhos product. The same degree of positive correlation was noticed between markers of endothelial dysfunction (ET-1 and CD146) and Phos.

Inhibitor-1; CD 146: Cluster of Differentiation 146. †Model adjusted for Calcium \times phosphorus product: serum phosphate and low-density lipoprotein

The results were qualitatively improved in models that were further adjusted for Ca \times PO₄ product, serum Phos, and LDL (Table 3). Significant interactions were detected between FGF-23 and IL-6 (p<0.001), PAI-1 (p<0.001), and CD146 in the multivariable-adjusted model (Table 3).

Discussion

In CKD and ESRD patients, high levels of FGF-23 are associated with increased morbidity and mortality rates [10,11]. In the past, the major risk factors for morbidity and mortality in dialysis patients were primarily linked to hypercholesterolemia, hypertension, and obesity. More recently, risk factors such as anemia, inflammation, and abnormalities in bone and mineral metabolism were incorporated [3]. These risk factors are considered to be responsible for the higher mortality rates witnessed in CKD and ESRD patients [18]. However, without in-depth knowledge of the pathophysiologic mechanisms, it is difficult to identify efficient targets for therapeutic interventions. Over the past few years, FGF-23 has been recognized as a predictor of a poor prognosis in ESRD patients [18]. In the present study, correlation and association of FGF-23 levels with IL-6, PAI-1, CD 146, and some other inflammatory, endothelial dysfunction factors, and mineral

metabolites in ESRD patients with T2DM were investigated. This association was studied separately from other inflammation-promoting factors such as total cholesterol and triglyceride.

Several studies have confirmed that FGF-23 might be responsible for mortality, Left Ventricular Hypertrophy (LVH), and the progression of CKD, independent of serum phosphate levels [19-21]. Findings in this study further corroborates the observations of previous studies that there is a strong correlation in the levels of FGF-23 and interleukins in ESRD patients undergoing hemodialysis [15] and in CKD patients not yet on dialysis [2].

In our study, a unique population of patients with the highest mortality rate (T2DM patients with ESRD on PD) demonstrated correlations between FGF-23 level and acute phase response markers such as IL-6, and endothelial dysfunction markers, in particular PAI-1 and CD 146.

Faulet al. Recently demonstrated a direct role for elevated FGF-23 in the pathogenesis of LVH and suggested a new mechanism to explain the high rates of LVH in patients with CKD. They reported that FGF-23 directly promoted pathological hypertrophy of isolated cardiomyocytes and that mice developed LVH after intra-ventricular or intravenous injection of FGF-23 [22]. In our study, FGF-23 did not significantly correlate with the endothelial dysfunction markers, ET-1, sICAM, or sVCAM. Similar findings were observed by Almorh et al. in ESRD patients on HD [23]. Concurrently, we found that the markers PAI-1 and CD 146 and $\text{Ca} \times \text{PO}_4$ product were more sensitive than ET-1, sICAM, or sVCAM in the presence of elevated FGF-23. As we found in our study, sICAM and sVCAM were more tightly associated with lipid metabolism than to the mineral imbalance stage, although ET-1 and sVCAM showed some positive correlations with Phos. These interesting finding require further research to understand the relationships between these major players in cardiovascular health.

FGF-23 has been reported to reduce levels of circulatory 1,25-dihydroxycholecalciferol (1,25D) by inhibition of renal 1-hydroxylase and stimulation of 24-hydroxylase, ultimately enhancing the rate of 1,25D decay [24,25]. Since reduced 1,25D levels are associated with higher levels of pro-inflammatory markers such as IL-6 and ET-1 [26], it is possible that FGF-23 mediates increase of some endothelial inflammation and endothelial dysfunction markers, acting through 1,25D. This assumption of indirect action requires further investigation.

Determining the mechanisms involved in cross-sectional interrelationships between FGF-23 and different inflammatory markers area complicated task. It is possible that FGF-23 acts directly to induce inflammation. Mendoza et al. reported that FGF-23 induces signaling in cells that do not express klotho, via activation of FGF receptors [27], which are present in adipose tissue [28]. Thus, it is possible that FGF-23 could induce expression of fat-derived cytokines, such as IL-6, PAI-1, and CD 146 [29,30] through klotho-independent mechanisms. Chitalia et al. Findings were similar to our results in that FGF-23 was associated with increased interleukins [2].

In addition, observations from our study concur with earlier studies that reported an association between levels of serum phosphate and inflammatory markers in ESRD patients [27,28]. We observed that serum phosphate levels are also correlated with the inflammatory and endothelial dysfunction markers ET-1, IL-6, CD146, IL-1 β , and VCAM. These findings underline the critical role played by serum phosphate in the occurrence of inflammation and micro-vascular problems in ESRD patients.

Conclusion

Our results indicate that FGF-23 impacts the cardiovascular health of T2DM patients on PD through mechanisms, which are independent from phosphate levels and linked directly to inflammation and endothelial dysfunction. Further studies are necessary to explain the precise mechanisms through which FGF-23 might impact morbidity and mortality of ESRD patients with T2DM on PD. Studies of FGF-23 pathogenicity may also help in the identification of new therapeutic targets and the development of new therapeutic methods.

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