

Cytotoxic Effects of Aflatoxin B1 Standard in Relation to Aflatoxin Extracts from South African Compound Feeds on Human Lymphocytes

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Abstract

Cytotoxicity testing of aflatoxin (AF) on the viability of cells grown in cultures can be widely used to predict the potential toxic effects of AF in animals. To this end, an in vitro experimental study was conducted to ascertain the toxic effects of AF extracts obtained from compound feeds in South Africa on human lymphocytes in comparison to that of an AFB1 standard. The approach adopted was on the basis of viable cells reducing methyl tetrazolium bromide (MTT) from blue to a purple formazan dye, which was then spectrophotometrically quantified to provide the rate of cytotoxicity.

Data obtained indicated no cytotoxic response in control cells, as the viability of cells without treatment with AF standard or methanolic extracts of AF extracts [negative control] using methanol as the reconstituting solvent, was 99.9% after 24 hrs of incubation. However, cell viability significantly ($p < 0.001$) decreased upon exposure to AF extracts especially for poultry feed. This was influenced by both the dose and duration of exposure, which was much more pronounced when the cells were exposed to AFB1 standard than for all the AF extracts tested. This implies that these feeds on exposure to AF can greatly influence animal health with respect to both the contamination dose and exposure time.

Keywords: Aflatoxins; Cytotoxicity; Lymphocyte; Methyl tetrazolium bromide

Introduction

Aflatoxins [AFs] are a group of mycotoxins produced as secondary metabolites notably by *Aspergillus flavus* and *A. parasiticus*. Other species such as *A. nomius*, *A. bombycis*, *A. ochraceoroseus*, *A. pseudotamarii*, *A. tamarii*, and *A. australis* [1,2] are also recognized producers. There are four main types of aflatoxins; aflatoxin B₁ (AFB₁), AFB₂, AFG₁ and AFG₂, owing to their colour [blue and green, denoted by the letters] and retardation factor (R_c) when viewed under UV light. The most important of these four toxins in terms of prevalence and abundance is AFB₁. Aflatoxin B₁ is also the most potent of all AFs and has been classified as a Class 1 carcinogen by the International Agency for Research on Cancer, IARC. This natural toxin is commonly reported to induce various health effects in humans and animal species; which could be hepatotoxic, mutagenic, genotoxic or carcinogenic [3]. Chronic exposure to low levels of AF may well be a risk factor involved in the etiology of hepatocellular carcinoma and immune system suppression/dysfunction [4,5].

Typically, AFs as metabolites are not chemically similar to those essential metabolites (e.g. polyamine) found in cells. A peculiar problem with respect to their occurrence in compound feeds is the ability of the animal to assimilate high amounts into their system. This may result in decreased animal performance and even death. The presence of the toxin in the animal's system can be further complicated when it is metabolized into animal products such as eggs, meat and milk for human consumption [6]. Therefore, it is of principal importance that compound feeds (feedstuffs or animal feeds that are formulated from various raw materials and fortified with additives) are routinely screened to check for the presence and degree of AFs contamination.

There are several approaches to this, which include direct measurements of the toxins and screening of AFs-producing fungi that gives an indication of the likely AFs presence in a commodity. However, the later approach may only give an indication of the presence of the fungal strain, but not the presence of the toxin. The fact is, some

potential AF producers may not produce the toxin under certain conditions [7]. It might be that some strains may have died before they are screened, leaving the already produced toxins in the compound feed matrix [8]. Biologically, cytotoxicity testing of AFs on human lymphocytes against extracts of compound feeds is another approach to check for the presence of toxin contamination. This approach that has been used to detect toxicity in environmental systems, involves the use of target cells, rather than using animal models [9]. The test is rapid and cytotoxicity of the test material can be quantitatively expressed against that of an untreated control. Nevertheless, limitations are experienced in terms of false positive results. Although there is substantial financial investment in growing cell lines that have to be maintained under special aseptic working conditions [10]. Apparently, due to the high expense, cytotoxicity-testing has been modified making use of cells available to laboratory workers i.e. blood cells, and in particular, lymphocytes. These can be used on a routine basis with ethical considerations and handled under sterile conditions. An advantage of this technique is that they do not require long maintenance periods under sterile conditions, unlike the case for cell cultures, due to shorter maturation times [11].

In this study, isolated and purified human lymphocyte cells were exposed to AFs extracts from compound feeds for investigation, as a biological means for testing the safety of these feeds.

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Materials and Methods

Equipment and reagents

Unless otherwise stated, all chemicals used were of analytical grade. Tetrazolium salt (3-4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), MTT assay kit, histopaque 1077, solvents and dimethylsulphoxide (DMSO) were purchased from Sigma Aldrich [St Louis, USA]. Complete tissue culture medium [RPMI-1640 supplemented with 10% foetal calf serum [FCS] and L-glutamine were purchased from Promega Corporation [Madison, USA]. Penicillin (100 U/ml) and streptomycin (100 µg/ml) were also Sigma, Aldrich. A humidified incubator (Incotherm-Labotec) was used and the temperature set at 37°C. The 96-well microtitre plates were obtained from Corning Cell Wells™ (Corning, USA). Additional materials and equipment included: Hank's Balanced Salt Solution (Adcock Ingram), AFB₁ standard (ARC, South Africa) ELISA microplate Reader [Modello: A2; Rome, Italy], centrifuge (Shalom S. Africa), haemocytometer [Shalom S. Africa] a phase contrast light microscope (Olympus B061, Wirsam Scientific, S. Africa), phytohaemagglutinin-p (PHA-p) (Sigma, Aldrich), penstrep-fungizone (Adcock Ingram), phosphate buffer saline [pH 7.4], trypan blue solution and fluorometer (Modello: A2, Rome, Italy). The PO7/V15/26.01.05 aflaprep kit with immunoaffinity column (Afla prep) was obtained from R-Biopharm Rhones Ltd (AG; Darmstadt, Germany).

Selection of aflatoxin extracts from South African compound feeds

The protocols of Candlish et al. [12], for extracting and quantification of AFs in compound feeds, were adopted with some modifications in obtaining extracts for the present study. The milled sample (12.5 g) and 1 gram NaCl were weighed into a solvent resistant blender jar into which 62.5 ml methanol and distilled water (60:40, v/v) were added and blended for 60 s. The extract was filtered and diluted with distilled H₂O (62.5 ml), which was mixed thoroughly by swirling. Sample extract (25 ml) was passed through a filter paper (Whatman No. 4) and 10 ml of the filtrate obtained (equivalent to 1 g of sample) was passed through an immuno-affinity column at a flow rate of 2-3 ml/min, after which, the immuno-affinity column was washed using 10 ml of phosphate buffered saline (PBS) at a flow rate of 5 ml/min. The analytes were then eluted (1 drop/s) using 1 ml of methanol and collected in an amber vial. Back flushing was employed thrice with the eluent to ensure complete release of AFs into the solution. The extract was dried in a fume cupboard using N₂ gas and stored at 0°C until use for further analysis. Ten AF extracts from compound feeds, mainly for poultry and cattle, were used to perform the cytotoxicity study. Selection of the feeds was based upon most significant levels of AFs analysed in the study.

Cytotoxicity testing

Aflatoxin B₁ standard at concentration levels of 20, 40 and 80 µl/ml as well as AFs containing feed extracts at different concentration levels were tested *in vitro* to evaluate their effects on the viability of human lymphocytes after 24, 48 and 72 h of exposure. A one-way analysis of variance (ANOVA) was performed on the cytotoxicity data using a pairwise multiple comparison procedures (Holm-Sidak method). Cytotoxicity testing was done using venous blood from a healthy male volunteer after the University of Johannesburg's Clinic Ethics Committee granted ethical clearance. Blood was collected by venous puncture, using a 2×15 ml sterile syringe and immediately transferred into a 10 ml heparin tube. The blood was diluted with an equal volume

of RPMI-1640. Ten millilitre of the mixture was layered onto 5 ml of histopaque in a 15 ml polypropylene conical tube and then centrifuged at 1,500 rpm for 30 mins at ambient temperature. The interface layer consisting of mononuclear cells was carefully removed using a sterile pipette and washed twice with Hank's Balanced Salt Solution (Adcock Ingram) by centrifugation at 4°C. About 20 µl of cell suspension, 80 µl of 0.2% trypan blue solution were mixed in an Eppendorf tube and incubated for 2 mins at room temperature. Using a sterile Pasteur pipette, 10 µl of the trypan blue cell suspension mixture was then transferred to a haemocytometer chamber and covered with a cover glass slip. Viable and non-viable cells were counted: coloured (blue) cells were considered dead while uncoloured cells excluding the dye were considered viable. Viability was determined as:

$$\% \text{ Cell viability} = (\text{viable cell counted} / \text{total number of cells}) \times 100$$

The concentration and number of cells were calculated using the formula:

$$\text{Cell/ml} = (n/v) \times (5) \times 10^4$$

Where: n = number of cells counted

$$v = \text{area of big squares counted} \times \text{depth} (0.1)$$

$$\text{DF} = \text{dilution factor} (10 \mu\text{l of blood} : 40 \mu\text{l of Trypan blue}) = 5$$

Mononuclear cells were transferred into a complete culture medium containing 1.5% L-glutamine, 10% foetal calf serum (FCS) and 1% penstrep (penicillin and streptomycin). Cells were seeded into 96 well plate containing methanol (used as a negative control), pure AFB₁ standard (used as a positive control) or AF extract reconstituted in methanol and incubated in a 5% buffered and humidified incubator for 48 hrs at 65°C.

The cytotoxicity assay performed herein is a biological method to assess the quality of compound feeds as well as a confirmatory test for the AF contents in similar feeds obtained via HPLC as reported elsewhere [13]. In this regard, the potential cytotoxic effect of sample extracts on lymphocytes was assessed *in vitro* by 3, 4, 5-dimethylthiazol-2, 5-diphenyl tetrazolium bromide (MTT) assay as described by Meky et al. [14]. This method assesses the ability of cells to convert MTT to formazan crystals. 3-[4, 5-dimethylthiazol-2, 5-diphenyl tetrazolium bromide [50 mg] was dissolved in 10 ml of 0.14 M phosphate buffered saline [PBS] [pH of 7.4] and filtered through a Whatman No.1 filter paper. About 25 µl of 5 mg/ml MTT solution was added into each cell contained in the 96 well plate and gently shaken using a vibrating shaker [Wirsam Scientific, S. Africa]. The contents were incubated for 2 hrs at 37°C in a 5% buffered and humidified incubator. Thereafter, 50 µl DMSO was added to each reaction and further incubated for 4 hrs to solubilize the formazan crystals formed. A microplate reader set at a wavelength of 620 nm was used to measure optical density [OD] values. The percentage of viable cells obtained after the assay was calculated as follows:

$$\% \text{ Cell viability} = [\text{Mean OD values of treated cells} / \text{Mean OD values of Control}] \times 100\%$$

Statistical analysis of data

A linear regression analysis was done on SigmaPlot 10.0 for Windows [15]. Data were then graphically represented. Among treatment groups, mean values were estimated to be different if the level of probability (p) was <0.05.

Results

It was observed, as data obtained showed that the viability of cells without any treatment with AFB₁ standard or extracts, was 99.9%, 99.7% and 99.7% after 24, 48 and 72 h, respectively, of incubation. The viability of cells after 24 hrs exposure was strongly influenced by the concentration of AFB₁ standard and sample extracts. This showed a higher reduction in cell viability when compared to the control or those of extracts with lower AFs contents as seen in Figure 1. Cell viability decreased significantly ($p < 0.001$) over time due to continued exposure and increased AF dosage under the same conditions. The AFB₁ standard (80 µl/ml) used as a point of reference exhibited the greatest cytotoxic effect in causing cell mortality ([73% cell viability recorded after 24 hrs of exposure), which increased over time (59% cell viability recorded after 72 hrs of exposure). The AFs extract concentrations from compound feeds, as determined by HPLC results which have already been submitted in another study [13] shows a relative correlation in respect to the determined cytotoxicity seen in this study, hence the relative cell percentage viabilities seen in Figure 1 and Tables 1-3.

Discussion

In this study, the cytotoxic effect of feed extracts containing AFs on cell viability of human lymphocytes cells in comparison to that of standard AFB₁ was performed *in vitro*. *In vitro* cytotoxicity testing initially described by Jelinek [16] on chick embryos toxicity screening test was against toxic potentials of different chemicals but not on lymphocytes against AFs. Reports of Henry and Wyatt [17] and Sehata et al. [18] also established the toxicity of different chemicals which include anti-tumour drugs, antipyretics, antibiotics and ergot alkaloids on chick embryo and rat brain but not on lymphocytes against AFs (Figures 2 and 3).

The toxico-pathological potential of AFs, both for the AFB₁ standard and AFs extracts from different compound feeds, to the lymphocytes was relatively high. This was observed as increased cytotoxicity [measured as decreased viability] as compared to the very low cytotoxicity posed to cells that received no treatment with AF standard or extract. A further decrease was observed over time as exposure dose increased. In

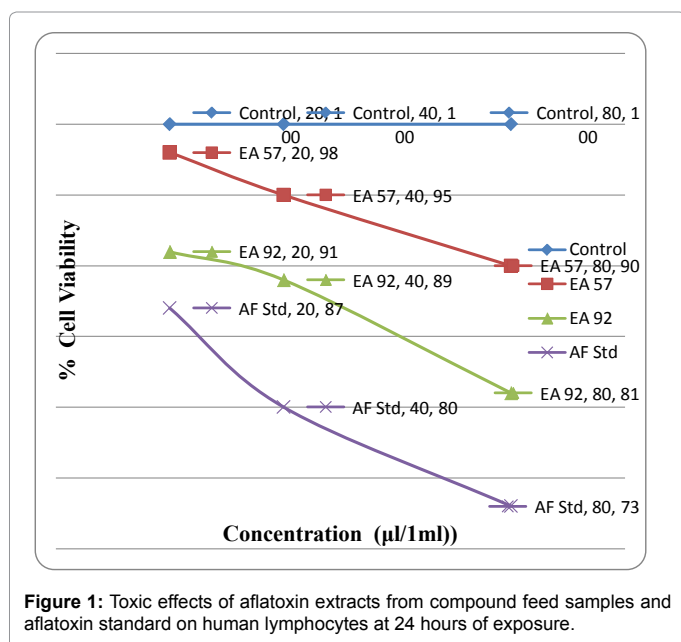


Figure 1: Toxic effects of aflatoxin extracts from compound feed samples and aflatoxin standard on human lymphocytes at 24 hours of exposure.

Sample Codes	Compound feed types	% Cell Viability ± SD		
		20 µl/ml	40 µl/ml	80 µl/ml
EA 92	Poultry	91 ± 2.1	89 ± 0.9	81 ± 5.0
EA 57	Cattle	98 ± 0.7	95 ± 1.3	90 ± 2.7
EA 83	Cattle	96 ± 1.1	90 ± 3.3	87 ± 1.5
EA 26	Poultry	97 ± 1.1	91 ± 4.0	89 ± 2.0
EA 91	Cattle	98 ± 3.4	90 ± 2.8	86 ± 2.9
EA 70	Poultry	98 ± 0.3	93 ± 2.2	91 ± 0.5
EA 88	Poultry	94 ± 2.1	91 ± 3.0	89 ± 3.9
EA 78	Cattle	99 ± 5.3	96 ± 2.7	90 ± 1.1
EA 21	Poultry	95 ± 2.1	91 ± 4.2	89 ± 0.9
EA 34	Poultry	97 ± 4.4	92 ± 3.0	85 ± 2.7
AF B ₁		87 ± 2.1	80 ± 0.9	73 ± 1.3

Appendix II

Table 1: Toxicity on Human Lymphocytes after 24 hrs.

EA=Sample Code, AF Std=Aflatoxin standard, SD=Stanard Deviation

Sample Codes	Compound feed types	% Cell Viability ± SD		
		20 µl/ml	40 µl/ml	80 µl/ml
EA 92	Poultry	86 ± 0.7	81 ± 1.1	77 ± 2.0
EA 57	Cattle	94 ± 2.1	89 ± 2.8	86 ± 2.8
EA 83	Cattle	94 ± 1.3	89 ± 0.9	82 ± 1.2
EA 26	Poultry	89 ± 1.1	83 ± 2.0	79 ± 1.9
EA 91	Cattle	92 ± 0.9	87 ± 2.3	81 ± 3.5
EA 70	Poultry	95 ± 3.3	88 ± 4.8	83 ± 3.0
EA 88	Poultry	90 ± 5.3	84 ± 1.1	80 ± 1.1
EA 78	Cattle	87 ± 0.9	82 ± 2.5	79 ± 2.7
EA 21	Poultry	91 ± 2.7	87 ± 3.1	81 ± 3.2
EA 34	Poultry	90 ± 1.5	85 ± 2.5	79 ± 2.9
AF Std		79 ± 1.3	73 ± 2.0	69 ± 2.2

Table 2. % Cell viability of human lymphocytes induced by AF extracts in feeds after 48 hrs of exposure.

EA=Sample Code, AF Std=Aflatoxin standard, SD=Stanard Deviation

Sample	Compound feed types	% Cell Viability ± SD		
		20 µl/ml	40 µl/ml	80 µl/ml
EA 92	Poultry	80 ± 3.7	76 ± 0.7	70 ± 2.5
EA 57	Cattle	90 ± 2.2	83 ± 1.9	79 ± 4.1
EA 83	Cattle	89 ± 4.1	81 ± 2.0	77 ± 2.8
EA 26	Poultry	82 ± 2.5	79 ± 2.0	72 ± 3.3
EA 91	Cattle	88 ± 2.1	81 ± 2.5	75 ± 2.1
EA 70	Poultry	87 ± 3.0	83 ± 4.1	78 ± 3.5
EA 88	Poultry	87 ± 2.8	80 ± 3.6	73 ± 0.8
EA 78	Cattle	84 ± 2.2	79 ± 0.9	76 ± 2.4
EA 21	Poultry	89 ± 3.0	84 ± 2.1	77 ± 1.2
EA 34	Poultry	85 ± 2.7	79 ± 2.0	71 ± 3.7
AF B ₁		71 ± 0.9	66 ± 0.6	59 ± 0.5

Table 3: Toxicity on Human Lymphocytes after 72 hrs.

EA=Sample Code, AF Std=Aflatoxin standard, SD=Standard deviation

overall, AFB₁ standard revealed the highest toxicity as expressed by the lowest percentage lymphocyte viability when compared to that for feed extracts which also exhibited some but lower cytotoxicity tendencies. This agrees with the reports of Büngrer et al. [19] and Meky et al. [14] in which human cell mortality increased with increasing AFs levels. Our study further reveals the implication of duration in exposure. Given the high dietary intake of AFs which led to many outbreaks of aflatoxicosis in animals reported in South Africa [2,20,21] and other

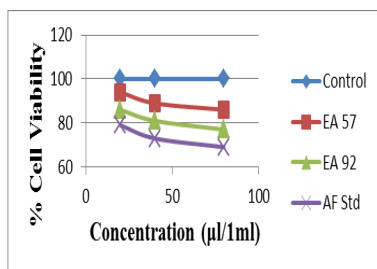


Figure 2: Toxic effects of aflatoxin extracts from compound feed samples and aflatoxin standard on human lymphocytes at 48 hours of exposure.

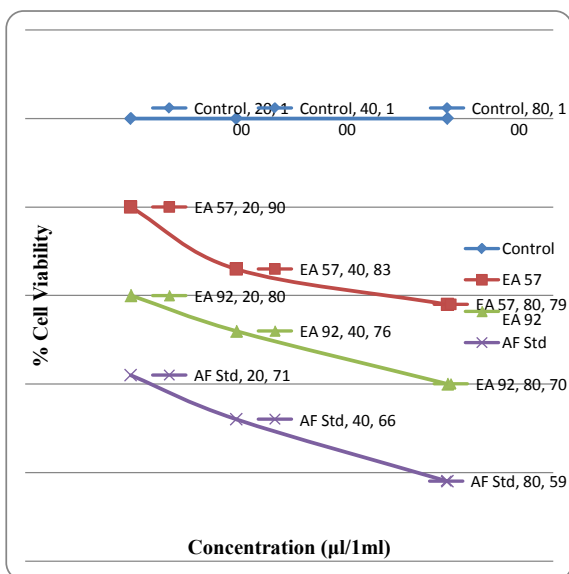


Figure 3: Toxic effects of aflatoxin extracts from compound feed samples and aflatoxin standard on human lymphocytes at 72 hours of exposure.

parts of the world [1,22]. This possible cause might be linked to the ability of this toxin causing immunosuppression as an initial step of immunomodulation due to loss of cells. This immunosuppressive effect can render the animals susceptible to secondary infections; predisposing chickens to candidiasis, Marek's disease, coccidiosis and salmonellosis [23], pigs to erysipelas and salmonellosis and cattle to fascioliasis, intramammary and clostridial infections [24].

Generally, aflatoxins, mainly AFB₁, are metabolized by the enzyme cytochrome P-450 enzyme [CYP450] to a toxic AFB₁-epoxide that binds to cellular DNA, RNA and protein forming an adduct that possibly results in the carcinogenic effect of AFB₁ in both animals and humans [3]. Another mechanism of action of AFs involves the inhibition of DNA synthesis [24], which may result in aflatoxicosis in animals with vague initial presentations [25]. It is known, that T lymphocytes play a pivotal role in the immune system, being responsible for immune responses, acting as a natural defence mechanism against host invasion of diseases. This may lead to a gradual but harmful onset of a disease condition. Some signs and symptoms due to such secondary infections maybe lethargy and anorexia. Sometimes expressions like haematochezia, melena and haematemesis [26] are present as disease condition, and widespread petechiae and ecchymoses [27] may occur. Some animals may present with peripheral oedema or ascites, polyuria and polydipsia [28] in the progressive stages of the disease. Though

fatality rate due to aflatoxicosis is high, its signs are also consistent with liver failures that are specific for AFs aetiology. However, the toxicity of the AFs extracts is lower than that of AF standard. One of the reasons may be due to the chemical compositions of the feed samples. Another reason could also be possibility of breakdown into residues. Animal feed consists of complex adjuncts and matrice which interfere with extraction and purification of the toxins to obtain up to 99.5% purity during laboratory analysis. It was therefore not unexpected that the pure aflatoxin standard would exert more damage to the lymphocytes. This does not in any sense downplay the cytotoxic potential of the crude hepatotoxin *in vivo*, especially over a period of time. Therefore, continued ingestion of the contaminated feeds may be risky to the animals [29-32].

Conclusion

The analysis of compound feeds from South Africa for AFs presence and contamination, using the MTT assay pre-analysis screening, showed feed samples with residual toxicity. The toxicity of AFB₁ standard on human lymphocytes was compared to that of compound feed fraction extracts and adequate comparability of the results were observed. It was not, however, possible to deduce the biochemical mode of action of the AFB₁ standard and AFs extract based on the observed reduction in cell viability potentiated by either the standard or extracts. Nevertheless, cytotoxicity results obtained showed a high through-put, which confirms the sensitivity of the test performed to an extent in the detection of AFs contained in compound feeds. This makes it a useful biological screening tool to ascertain the quality of compound feeds.

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References

- Akinrinmade JF, Akinrinde AS (2012) Aflatoxin status of some commercial dry dog foods in Ibadan, Nigeria. *African Journal of Biotechnology* 11: 11463-11467.
- Arnot LF, Duncan NM, Coetzer H, Botha CJ (2012) An outbreak of canine aflatoxicosis in Gauteng Province, South Africa. *Journal of South Africa Veterinary Association*, South Africa.
- Ayub MY, Sachan DS (1997) Dietary factors affecting aflatoxin Bi carcinogenicity. *Malaysian Journal of Nutrition* 3: 161-179.
- Jiang Y, Jolly PE, Preko P, Wang JS, Ellis WO, et al. (2008) Aflatoxin-related immune dysfunction in health and in human immunodeficiency virus disease. *Clin Dev Immunol*.
- Wu F, Narrod C, Tiongco M, Liu Y (2011) The Health Economics of Aflatoxin: Global Burden of disease: Afla control, Working Paper 4.
- Veldman V, Meijst J, Borgreave J, Heers-Van der tol J (1992) Carry over of aflatoxin from cow's food to milk. *Animal Production* 55: 163-168.
- Frisvad JC, Thrán U, Samson RA, Pitt J I (2006) Important mycotoxins and the fungi which produce them. *Adv Exp Med Biol* 571: 3-31.
- Vosough M, Bayat M, Salemi A (2010) Matrix-free analysis of aflatoxins in pistachio nuts using parallel factor modeling of liquid chromatography diodearray detection data. *Analytical chimica acta* 663: 11-18.
- Braydich Stolle L, Hussain S, Schlager JJ, Hofmann M (2005) In vitro Cytotoxicity of Nanoparticles in mammalian Germline Stem Cells. *Toxicological Sciences* 88: 412-419.
- Pence VC (2011) Evaluating costs for the in vitro propagation and preservation of endangered plants. *In Vitro Cellular and Developmental Biology-Plant* 47: 176-197.

11. Harry JG, Billingsley M, Bruinink A, Campbell IL, Classen W, et al. (1998) Reviews in Environmental Health: Environmental Health Perspectives.
12. Candlish AG, Haynes CA, Stimson WH (1998) Detection and determination of aflatoxins using affinity chromatography. *International Journal of Food Science and Technology* 23: 479-485.
13. Iheanacho HE, Dutton MF, Steenkamp PA, Steenkamp L, Makun HA (2014) Real time PCR of Nor-1 (aflD) gene of aflatoxin producing fungi and its correlative quantization to aflatoxin levels in South African compound feeds. *Journal of Microbiological Methods* 97: 63-67.
14. Meky FA, Hardie LJ, Evans SW, WildC P (2001) Deoxynivalenol-induced immunomodulation of human lymphocyte proliferation and cytokine production. *Food Chemical Toxicology* 39: 827-836.
15. Systat INC (2006) SigmaPlot 10.0 for Windows Systat Software Inc, Point Richmond, CA, 94804-2028, USA.
16. Jelinek R (1977) The Chick Embryotoxicity screening test (CHEST). *Methods in prenatal toxicology*, Georg Thieme Publishers Stuttgart. Pp. 381-386.
17. Henry MH, Wyatt RD (2001) The Toxicity of Fumonisin B1, B2, and B3, Individually and in combination, in chicken embryos. *Poultry Science* 80: 401-407.
18. Sehata S, Kiyosawa N, Makino T, Atsumi F, Ito k, et al. (2004) Morphological and microarray analysis of T-2 toxin-induced rat fetal brain lesion. *Food and Chemical Toxicology* 42: 1727-1736.
19. Bünger J, Westphal G, önnich A, Hinnendahl B, Hallier E, et al. (2004) Cytotoxicity of occupationally and environmentally relevant mycotoxins. *Toxicology* 202: 199-211.
20. Otto H (2011) Tainted food kills 16 dogs. *News24*.
21. Van Halderen A, Garthwaite I, Wessels JC (2000) Outbreak of bovine aflatoxicosis in South Africa. *World Mycotoxin Journal* 4: 87-100.
22. Thomas WK, Bill DR, Gerald NW, John DG (2010) Aflatoxin: A 50-Year odyssey of mechanistic and translational Toxicology. *Oxford Journals of Toxicological Sciences* 120: 28-48.
23. Robens JF, Richard JL (1992) Aflatoxins in Animal and Human Health. In: WARE, G. (ed.) *Reviews of Environmental Contamination and Toxicology*. Springer US.
24. El khoury A, Atoui A, Rizk T, Lteif R, Kallassy M, et al. (2011) Differentiation between *Aspergillus flavus* and *Aspergillus parasiticus* from pure culture and aflatoxin-contaminated grapes using PCR-RFLP analysis of aflR-aflJ intergenic spacer. *Journal of Food Science* 76: 247-253.
25. Dixon JB, Kannevischer I, Tenorio arvide MG, Barrientos velazquez AL (2008) Aflatoxin sequestration in animal feeds by quality-labeled smectite clays: An introductory plan. *Applied Clay Science* 40: 201-208.
26. Tedesco D, Steidler S, Galletti S (2004) Efficacy of silymarin-phospholipid complex in reducing the toxicity of aflatoxin B1 in broiler chicks. *Poultry Science* 83: 1839-1843.
27. Galvano F, Piva A, Ritieni A (2001) Dietary strategies to counteract the effects of mycotoxins a review. *Journal of food Protection* 64: 120-131.
28. Wilson D (2010) *Clinical Veterinary Advisor: The Horse*, Elsevier Health Sciences.
29. Smith JE, Moss MO (1985) Structure and formation of mycotoxins. In: *Mycotoxins. Formation, analysis and significance*. John Willey and Sons, Toronto, Canada, 31-49.
30. Ito Y, Peterson SW, Wicklow DT, Goto T (2001) *Aspergillus pseudotamarii* a new aflatoxin producing species in *Aspergillus* section Flavi. *Journal of Mycological Research* 105: 233-239.
31. Varga J, Frisvad JC, Samson RA (2009) A reappraisal of fungi producing aflatoxins. *World Mycotoxin Journal* 2: 263-277.
32. Wu F (2010) The global burden of disease caused by foodborne aflatoxin WHO Commissioned Report, Food.