

Cytotoxicity, Antioxidants and Antimicrobial Activities of Lipids Extracted from Some Marine Algae

El Baz FK¹, El-Baroty GS², Ibrahim AE¹ and Abd El Baky HH^{1*}

¹Plant Biochemistry Department, National Research Centre, Dokki, Cairo, Egypt

²Biochemistry Department, Faculty of Agriculture, Cairo University, Cairo, Egypt

Abstract

The lipids of five marine algae species, i.e., two from the Red sea, Rhodophyta (*Laurencia popillose*, *Galaxoura cylindria*), and three from the Mediterranean sea (Chlorophyta, *Ulva fasciata*; and Phaeophyta, *Dilophys fasciola* and *Taonia atomaria*) were evaluated as anticancer, antiviral, antimicrobial and antioxidant activities. Total lipid content was varied significantly, and the values ranged from 0.66 to 2.20%. The highest lipid content was found in *U. fasciata* (2.2% dw). Among fatty acids of all algae species, palmitic was dominating fatty acid and C14:0, C17:0, C18:0, C18:1, C20:4 were presented in significant levels. The polyunsaturated fatty acids C18:2, C22:5 and C20:3 were identified in most algal species.

The biological activities of algal crude lipids were assessed in vitro. The crude lipids at a concentration of 10 µg/ml inhibited HSV-1 virus growth (*in vitro*) and % inhibition ranged from 12.5 to 74.4%. While, total lipids at concentration of 20 µg/ml induced toxic effect in host cells. The algal lipids exhibited a potent inhibitory effect on both breast and liver human cancer cell lines with IC₅₀ values ranged from 0.34 to 7.11 µg/ml. All algal lipids induced remarkable antimicrobial activity *Aspergillus niger* and *Candida albicans*. Marine algal lipids exhibited moderate scavenging activity toward DPPH. radical, and high activity was found in lipids of fractionD. fasciol.

Keywords: Marine macroalgae; Algal lipids; Anticancer; Antiviral; Antibacterial; HSV-1; MCF7 Cell; HEPGcell

Introduction

Algae represent valuable sources of a wide spectrum of complex lipids with different potential applications in food, cosmetic and pharmaceutical industries [1-3]. Lipids of marine macroalgae possess antibacterial, antiviral, antitumor, antiinflammatory, antiproliferative and antioxidant activity [4-6]. Algal lipids contained high concentrations of polyunsaturated fatty acids (PUFA) and vitamins as well as bioactive molecules, e.g. phenolic and terpenes compounds. Marine algae are known to be a good source of healthy food. PUFA prevent atherosclerosis development, and they reduce a frequency of heart and blood vessel diseases [4]. Bhaskar [7] found that the lipids of brown algae.

Sargassum marginatum possessed an inhibition effects against human promyelocytic leukemia. Lipid extracts of *Ginsens marc* showed potent inhibitory activity on human hepatoma (HepG2) and breast (MCF7) cancer cell proliferation in vitro [8]. The aim of current work was to study lipid potential of some Egyptian marine algae as antiviral, anticancer, antimicrobial and antioxidant agents.

Materials and Methods

Collection of algal samples

Samples of *Laurencia popillose* and *Galaxoura cylindria* were collected from Red Sea (Faied and Ein Al-Sokhna in Suez both in April 2007). *Ulva fasciata* and *Taonia atomaria* were collected from the Mediterranean Sea from Abu-Qir near Alexandria in 2007 and *Dilophys fasciola* from Marsa Matrouh in 2005. All algal sample were washed several times with tap water and then left in air for dryness. Samples were grinded and stored in glass containers at room temperature for further experiments.

Identification of algae species

After preparation of herbarium specimens of the algae species,

they were identified by Dr. Rauhaya Abdul-Latif, Professor of Botany Department of Botany, Faculty of Science, El-Azhar University, to whom the authors are very indebted.

Extraction and determination of total lipids

Total lipids of marine algae (100 g) were extracted according to the method described by Roughan and Bratt [9].

Identification of algal lipid fatty acids

The algal lipids were subjected to direct transmethylation in 1.5% sulfuric acid: methanol at 95°C for 2 h [10]. Fatty acid methyl esters were analyzed by gas chromatography (Perkin Elmer Auto system XL) equipped with a flame ionization detector and fused silica capillary column (DB-5 (American) 60 m×0.32 mm, i.d.) with a film thickness of 0.25/25 µm. The column temperature was initially set up at 150°C and gradually increased at rate of 3°C/min up to 250°C. The injector and detector temperatures were 230 and 250°C respectively. The helium was used as a carrier gas at 1 ml/min. The split ratio was 1/100. Fatty acids were identified by comparison between retention times of samples and with those of methyl fatty acid standard mixture (Sigma, >99% purity by GLC).

***Corresponding author:** Abd El Baky HH, Plant Biochemistry Department, National Research Centre, Dokki, Cairo, Egypt, Tel: +20 2 33371211; E-mail: abdelbakyh@hotmail.com

Received October 19, 2014; **Accepted** October 31, 2014; **Published** November 03, 2014

Citation: El Baz FK, El-Baroty GS, Ibrahim AE, Abd El Baky HH (2014) Cytotoxicity, Antioxidants and Antimicrobial Activities of Lipids Extracted from Some Marine Algae. J Aquac Res Development 5: 284. doi:10.4172/2155-9546.1000284

Copyright: © 2014 El Baz FK, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Biological evaluation of algal lipids

Preparation of algal extract for biological bioassays: The algal lipids (100 mg) of lipids were dissolved in 10 ml of DMSO in water (90:1, v/v) and kept at 4°C until used, appropriate dilution of solution were used in each assay.

Antiviral screening of algal lipids: Algal lipids were evaluated as antiviral against herpes simplex virus type 1 (HSV-1). The virus was obtained from Virology Laboratory, Water Pollution Research Department National Research center (NRC), Egypt. The virus was propagated in virocell cultures. Inhibition % of virus was calculated as plaque reduction as a result of being subjected to a given extracts [11].

Antitumor activity of algal lipids: Potential antitumor activity of algal lipids was evaluated using the method of Skehan [12]. Human HEPG2 (liver carcinoma cell lines) and MCF7 (breast carcinoma cell lines) cells were plated in 96 multiwell plate for 24 h before treatment with the algal lipids to allow attachment of cells to the wells of the plate. Lipids and antitumor reference drug (Novantron) were added at serial concentrations to cell monolayer. After incubation for 48 h at 37°C the cytotoxicity was determined spectrophotometrically by measuring the developed color at 570 nm by ELISA reader (Tecan Sunrise absorbance reader (No. 3008746), software Magllan V.4 was used).

Antimicrobial activity of algal lipids: The antimicrobial activities of algal lipids were determined by conventional agar diffusion assay [13] using one gram positive (*Bacillus subtilis* NRRL B- 94) one gram negative (*Escherichia coli* NRRL B- 3703) bacteria, fungi (*Aspergillus niger* NRRL 313), and yeast (*Candida albicans* NRRL 477). The microbial growth inhibition zone was measured after incubation at 30°C by the appearance of clear microbial free inhibition zone, beginning within 24 h for yeast, 24-48 h for bacteria and 72-96 h for fungus.

Antioxidant activity of macro algal lipids using DPPH free radical-scavenging assay

Quantitative measurement of radical scavenging properties of algal lipids was determined according to Ye [14] method.

Ethanol DPPH solution (0.1 mM) was prepared, to give the initial absorbance value of 0.993 at 517 nm. The different concentrations of lipid samples (in 0.1 ml) of each sample (with appropriate dilution if necessary) was added to 3.0 ml of ethanolic DPPH solution. After incubation for 30 min in the dark, the absorbance was measured at 517 nm. The percentage of DPPH which was scavenged was calculated using the following formula: Scavenging (%)=[1-(A sample-A blank/A control)]×100%.

Statistical analysis

Data were statistically analyzed through analysis of variance (Anova) and Duncans test at 0.01 probability level [15].

Results and Discussion

Lipid content of marine algae

Results illustrated in Table 1 showed that the total lipid content (TL) of five macroalgae species (*Ulva fasciata*, *Dilophys fasciola*, *Taonia atomaria*, *Laurencia popillose* and *Galaxoura cylindriea*) were ranged from 0.09 to 2.2% (D.W). Total lipids showed significant differences amongst all algae species. The maximum lipid content was found in *U. fasciata* (2.2%) followed by *D. fasciola* (1.1%) then, *L. popillose* (0.81%) and *T. atomaria* (0.66%). Whereas the minimum value was found in *G. cylindriea* (0.090%). These results are in agreement with those obtained

Total lipids (% dry weight basis)	Macroalgae Species
2.35 ^d	<i>U. fasciata</i>
1.11 ^a	<i>D. fasciola</i>
0.09 ^a	<i>G. cylindriea</i>
0.81 ^c	<i>L. papillose</i>
0.66 ^b	<i>T. atomaria</i>
0.09	LSD

The values represent the mean of three replicate (n=3) Values in the same column with different superscript letters were significantly different (P<0.05)

Table 1: Total lipid contents of some marine alga.

by [16], they found that the twelve species of seaweeds which includes from Chlorophyceae, species from Phaeophyceae (Brown algae) and species from Rhodophyceae (Red algae) had lipid values ranged from 1.33 to 4.6%. In another study carried out by [17], they reported that the lipid contents of Rhodophyta species, Chlorophyta species and Phaeophyta species were ranged from 1.33 to 4.6% based on D.W. On the other hand [18] found that the macroalgal total lipid content was dependent on species specific and genetic origin. Also, accumulation of lipids in macroalgal species was dependent on climate and geography development [16].

Fatty acid composition of macroalgal lipids: Fatty acid composition of marine macroalgae under investigation is illustrated in Table 2. *L. papillose* had the highest palmitic acid (C16:0) content (59.35%) followed by *G. cylindriea* (49.4%), *T. atomaria* (36.27%) and *D. fasciola* (32.58%). While, *U. fasciata* had the minimum C16:0 content (19.2%).

It is very interesting that, all macroalgae species under investigation had an odd fatty acid margaric acid (C17:0). *U. fasciata* had the highest margaric acid content (16.03%) followed by *L. papillose* (8.1%). Moreover, all marine macroalgal had high content of long chain polyunsaturated fatty acids such as eicosatrienoic (C20:3) and docosapentaenoic (C22:5). However, all macroalgal species contained a low amounts of docosapentaenoic (C22:5) except *T. atomaria* (11.89%). Hence, the predominant fatty acid is dependent on the marine algae species. The highest content of eicosatrienoic (C20:3) was detected in *U. fasciata* (54.41%) and oleic acid (C18:1) was abundant in *D. fasciola* (37.49%) (Table 3).

These results are in agreement with those obtained by Shehnaz [19], who found that the major fatty acid of *U. fasciata* was palmitic acid (16:0) (50%) followed by oleic acid (18:1) (12.5%), margaric acid (17:0) (9-10%) and stearic acid (18:0) (4%). Orhan [20] found that palmitic and oleic acids were the most abundant fatty acids in several of marine algae. Among all lipid extracts analyzed, *H. scoparia* contained capric acid, however myristic acid was found in *C. fracta* and *V. sessilis*, which present in freshwater origin. Fatty acid composition of *H. incrassata* lipids were found to be rich of C16:0, C18:1, C 20:1, C20:3, C24:0 and (Lignoceric) C24:1 (Nervonic) fatty acid [21].

Biological evaluation of marine macroalgal lipids

Antiviral activity for marine algal crude lipid extracts against HSV-1: Table 4 showed the antiviral potential of marine algal lipids against HSV-1. The maximum inhibition of HSV-1 was recorded for the lipid extract of *T. atomaria* (74.4%) followed by *D. fasciola* (72%) and *G. cylindriea* (72%) at a concentration of 10 µg/ml. The complete inhibition effect was observed in *T. atomaria*, *D. fasciola* and *G. cylindriea* lipids at the concentration of 20 µg/ml.

	Algal strain				
	Mediterranean sea			Red sea	
	<i>U. fasciata</i>	<i>T. atomaria</i>	<i>D. fasciola</i>	<i>L. papillose</i>	<i>G. cylindriea p</i>
Myristic acid (C _{14:0})	-	2.84	11.04	10.33	11.29
Palmitic acid C _{16:0} *	19.20	36.27	32.58	59.35	49.40
palmitoleic acid C _{16:1}	-	0.74	4.06	5.82	8.39
Margaric acid (C _{17:0})	16.03	0.65	5.72	8.06	0.77
Stearic (C _{18:0})	5.39	14.22	2.07	2.13	4.56
Oleic acid (C _{18:1})	3.41	11.50	37.49	5.11	7.41
Linoleic (C _{18:2})	-	21.34	-	9.48	9.01
Eicosatrienoic (C _{20:3})	54.41	0.55	5.31	-	4.99
Docosapentaenoic (C _{22:5})	1.56	11.89	1.69	0.25	1.92
Saturated FAs %	40.62	53.98	51.41	79.87	66.52
MUFAs %	3.41	12.24	41.55	10.93	15.8
PUFAs %	55.97	33.78	7.00	9.73	15.92

FA, fatty acid; MUFAs, monounsaturated fatty acids; PUFAs, Polyunsaturated fatty acids

Table 2: Fatty acid composition of some marine algal lipids.

Algal Species	Viral inhibition (%)	
	Concentration (µg/ml)	
	10	20
<i>U. fasciata</i>	53.12	68.75
<i>T. atomaria</i>	74.40	T
<i>L. papillose</i>	12.50	28.12
<i>G. cylindriea</i>	72.00	T
<i>D. fasciola</i>	72.00	T

T: Toxicity=complete inhibition

Table 3: Antiviral activity of crude lipids extracted from some marine algae against HSV-1.

Algal lipid concentration (µg/ml)	Inhibition %					Reference control
	Algal Species					
	<i>U. fasciata</i>	<i>T. atomaria</i>	<i>D. fasciola</i>	<i>L. papillose</i>	<i>G. cylindriea</i>	Novantron
1.0	28.3 ^a	77.24 ^a	59.12 ^a	17.21 ^a	22.18 ^a	42.734
2.5	48.6 ^a	77.21 ^a	62.96 ^a	47.27 ^a	52.17 ^a	52.81
5.0	52.15 ^a	80.24 ^a	73.9 ^b	65.66 ^b	65.73 ^a	52.81
10.0	51.62 ^b	82.34 ^a	74.84 ^b	74.01 ^c	69.01 ^b	52.81
IC ₅₀	7.18	0.34	2.55	2.95	1.61	1.40
LSD	13.12	7.81	9.49	11.90	23.02	

The mean (n=3) difference was significant at $P \leq 0.05$

Table 4: Antitumor activity of some marine algal crude lipid extract against MCF7 after 48 h incubation period.

These results are similar to that obtained by Chirasuwan [22], they found that lipid extracts of *S. platensis* possessed antiviral activity against HSV-1 with an IC₅₀ value of 25.1 µg/ml. On the other hand, the 1- docosanol (C_{22:0}) was found to inhibit viral replication by interfering with the early intracellular events surrounding viral entry into target cells. It is possible that interaction between the highly lipophilic compound and components of target cell membranes renders such target cells less susceptible to viral fusion and/or entry.

If this mechanism proves to be correct, 1-docosanol (C_{22:0}) may provide a broad spectrum activity against many different viruses, especially those with lipid-containing envelopes [23]. Pope [24] reported the C_{22:0}, n-docosanol exhibited in vitro antiviral activity against several lipid enveloped viruses including herpes simplex viruses 1 and 2 by a mechanism that interferes with normal viral entry into target cells.

Antitumor activity of marine algal lipids extract against human breast carcinoma (MCF-7) and hepato carcinoma cells (HEPG2): Antitumor potential of macroalgal lipids was evaluated against breast

carcinoma cell line (MCF-7) and hepato carcinoma cells (HepG2) at different concentrations of algal lipids (1, 2.5, 5 and 10 µg/ml) and tumor cells viability after 48 h incubation and the results are presented in Tables 5 and 6. The inhibition % of algal crude lipids on MCF-7 and HepG2 cells was ranged between 17.21- 82.34% and 24.0- 61.0%, respectively. The *T. atomaria* crude lipids showed the highest inhibition % at all concentrations ranged from 77.21 to 82.34%. However, *T. atomaria* lipid extract had the lower IC₅₀ value against MCF-7 cell line (0.34 µg/ml) compared with that of the antitumor drug Novantron (1.40). While, *G. cylindriea* lipids caused high potential antitumor activity with IC₅₀ of 4.09 µg/ml against HepG2 cell, compared to Novantron antitumor drug (IC₅₀=4). The results of the present study demonstrated that the marine algal lipids of *T. atomaria* showed significantly higher antitumor activity against human breast carcinoma (MCF-7) and the human hepato carcinoma (HepG2) compared with that of Novantron antitumor drug. Similar results were obtained from lipids of brown algae *Sargassum marginatum*, which possessed an inhibition effect against human pro-melocytic leukemia [7]. Lipid extracts of *Ginseng marc* showed a potent inhibitory activity

Algal lipid concentration (µg/ml)	Algal Species					Reference control
	<i>U. fasciata</i>	<i>T. atomaria</i>	<i>D. fasciola</i>	<i>L. papillose</i>	<i>G. cylindriea</i>	Novantron
1.0	24.00	42.00	35.00	35.00	41.00	26.10
2.5	30.00	52.00	41.00	41.00	56.00	42.27
5.0	32.00	58.00	49.0	49.00	61.00	47.66
10.0	36.00	61.00	52.00	52.00	64.00	59.50
IC ₅₀ (µg/ml)	4.00	-	1.68	7.11	4.09	4.00

Table 5: Antitumor activity of some marine algal crude lipid extract against HepG2 after 48 h incubation period.

Algal Species	Inhibition zone (mm)			
	Bacteria		Fungi	Yeast
	<i>E. coli</i>	<i>B. subtilis</i>	<i>A. niger</i>	<i>C. albicans</i>
<i>U. fasciata</i>	-	-	8	13
<i>T. atomaria</i>	11	-	-	15
<i>L. papillose</i>	7	-	12	16
<i>G. cylindriea</i>	9	-	10	12
<i>D. fasciola</i>	13	-	7	11

Table 6: Antimicrobial activities of algal crude lipid extracts at a concentration of 100 µg/ well.

Algal lipid concentration (µg)	Algal species					Reference control	
	Mediterranean Sea		Red Sea			BHA	BHT
	<i>U. fasciata</i>	<i>T. atomaria</i>	<i>D. fasciola</i>	<i>L. papillose</i>	<i>G. cylindriea</i>		
60	25.09 ^a	12.17 ^a	66.45 ^a	28.83 ^a	22.81 ^a	52.59 ^a	51.25 ^a
80	28.90 ^b	14.50 ^a	75.85 ^b	32.81 ^b	28.72 ^b	72.25 ^b	68.13 ^b
100	39.47 ^c	19.17 ^b	90.88 ^b	35.93 ^c	32.97 ^c	88.22 ^c	88.44 ^c
LSD	2.06	2.79	13.02	1.57	2.81	8.03	3.02

The mean (n=3) difference was significant at $P \leq 0.05$

Table 7: Antioxidant activity of marine algae crude lipids assessed by DPPH radical.

on human hepatoma (HepG2, IC₅₀=41.7 µg/ml) and breast (MCF-7, IC₅₀=54.4 µg/ml) cancer cell proliferation [8]. Some macroorganisms such as mushroom lipid extract from *G. sinensis* possessed antitumor activity against leukemic (U937) and hepato carcinoma cells (HepG2). All these symptoms induced throughout the apoptotic changes in cells, including decreased cell volume and chromatin condensation [23].

Antimicrobial activity of marine algal crude lipid extracts

The antimicrobial potential of macroalgal crude lipids were assayed against one fungi (*Aspergillus niger* NRRL313), one yeast (*Candida albicans* NRRL477), one gram positive bacteria (*Bacillus subtilis* NRRL B-94) and one gram negative bacteria (*Escherichia coli* NRRL B-3703) Table 6 by measuring the inhibition zones (IZs) at the concentration of 100 µg/well. Lipid extract of *L. papillose* had antifungal effect against *A. niger* with 12.0 mm IZs followed by *G. cylindriea* (10 mm IZs). Lipid extracts of *L. papillose* and *T. atomaria* induced antiyeast effect against *C. albicans* with IZs values of 16 and 15 mm, respectively. However, lipids extract of *U. fasciata* and *D. fasciola* had a moderate activity against both microbes. *D. fasciola* lipids had the maximum antibacterial activity against *E. coli* (13.0 mm IZs) followed by *T. atomaria* (11 mm IZs). All macroalgal lipids extract did not show any effect against *Bacillus subtilis*.

The results of the present experiment showed that the algal crude lipids of *L. papillose* and *T. atomaria* induced the highest antiyeast activity, which may be attributed to the presence of unsaturated fatty acids. The results of the present study are in agreement with the findings of Ballantine [26], they found that some of the algal lipid extracts caused antifungal activity against *C. albicans*, gram-negative *P. aeruginosa* and *E. coli*. Ramadan [27] found that the lipid extract of *Spirulina platensis* induced antimicrobial activity against *A. niger* and

C. albicans at concentration of 48 µg/disc, with IZs 28.3 mm and 21.3 mm, respectively. In another study carried out by Patra [28], they found that the lipid extract of *E. compressa* exhibited antibacterial activity against *B. subtilis* and *E. coli* with 14 mm and 12 mm IZs, respectively.

Desbois and Smith [29] attributed the prime target of total lipids may be due to the constituents of fatty acids which may disrupting the electron transport chain and oxidative phosphorylation of microbe in addition to its interference with cellular energy production. Moreover, fatty acids action may be resulted from the inhibition of enzyme activity, impairment of nutrient uptake, generation of peroxidation and auto-oxidation degradation products, or direct ana lysis of bacterial cells.

Antioxidant activity of marine algal crude lipid extracts

The antioxidant effects were evaluated by scavenging of 2, 2 di-phenyl-picryl-hydrazyl (DPPH.) radical as shown in (Table 7). Most of the macroalgal lipid extracts possessed the ability of scavenging of DPPH. radical with different degrees. The scavenging % was ranged from 12.17 to 90.88% at the concentration of 100 µg algal lipids. The highest antioxidant effect was obtained with *D. fasciola* (90.88%) total lipids. Whereas, the lipid extracts of *U. fasciata* (39.47%), *L. papillose* (35.93%), *G. cylindriea* (32.97%) and *T. atomaria* (19.17%) exhibited the weakest DPPH. radical scavenging activity. Shanab [30] found that *S. dentifolium* extracted with di-chloromethane induced the greatest free radical (DPPH) scavenging activity (86%) at concentration 100 µg/ml. Lipids of *L. papillosa* caused the greatest anti-lipid peroxidation efficiency (87%) comparing with silymarin (96%) as a standard antioxidant agent. However, the highest antioxidant activity of crude lipids of several macroalgae species extracted with hexane may be due to the presence of many substances such as carotenoids, soluble lipids, phenolic compounds, steroids and terpenoids [31]. Ramadan

[25] found that the *Spirulina platensis* lipids exhibited free radical scavenging activity (from 21 to 27%) against DPPH with various degrees.

In conclusion, crude lipids of algal species under investigation was biologically evaluated. The results indicated that the algae showed different biological properties as antitumor, antioxidant, antimicrobial and antiviral with various degrees. Most of the algal crude lipids possessed an inhibition effect against human hepato carcinoma (HepG2) and human breast carcinoma (MCF-7), specially *T. atomaria* which induced the greatest antitumor activity with IC₅₀ 0.34 and 1.68 µg/ml, respectively. Moreover, it is worth mentioned that the crude lipids of *D. fasciola* showed a significant higher scavenging radical activity against the DPPH (90.88). The antioxidant activity may be due to the presence of lipophilic antioxidants (total carotenoids and α-tocopherol, phenolic and terpenoid compounds) and PUFAs. The highest antimicrobial activity of algal crude lipids was found in *T. atomaria* and *L. papillose* against *A. niger* and *C. albicans*, which might be attributed to the presence of different polyunsaturated fatty acids, phenolic and flavonoids.

References

- Hossain Z, Kurihara H, Hosokawa M, Takahashi K (2005) Growth inhibition and induction of differentiation and apoptosis mediated by sodium butyrate in Caco-2 cells with algal glycolipid. *In Vitro Cell Dev Biol Anim* 41: 154-159.
- Abd El Baky HH, El Baz FK, El-Baroty GS (2009) Natural preservative ingredient from marine alga *Ulva lactuca* L. *International J Food Sci Technol* 44: 1688-1695.
- Abd El Baky HH, El-Baroty GS (2013) The potential use of microalgal carotenoids as dietary supplements and natural preservative ingredient. *J Aquatic Food Product Technol* 4: 392-406.
- Gerasimenko IN, Busarova GN, Moiseenko PO (2010) Age dependent changes in the content of lipids, fatty acids, and pigments in brown alga *Costaria costata*. *Russ J Plant Physiol* 57: 62-68.
- Goecke F, Hernández V, Bittner M, González M, Becerra J, Silva M (2010) Fatty acid composition of three species of Codium (Bryopsidales, Chlorophyta) in Chile. *Revista de Biología Marina y Oceanografía* 45: 325-330.
- El Baz FK, El-Baroty GS, Abd El Baky HH, Abd El-Salam OI, Ibrahim EA (2013) Structural characterization and biological activity of sulfolipids from selected marine algae. *Grasasy Aceites* 64: 561-571.
- Bhaskar N, Hosokawa N, Miyashita K (2004) Growth inhibition of human promyelocytic Leukemia (HL-60) cells by lipid extract of marine algae *Sargassum marginatum* (Fucales, phaeophytae) harvested off Goa (west coast of India) with special reference to fatty acid composition. *Indian Journal Marine Science* 33: 335-360.
- Lee SD, Yoo G, Chae HJ, Hwang YK, Hwang WI, et al. (2009) Lipid-soluble extracts as the main source of anticancer activity in ginseng and Ginseng marc. *J Amer Oil Chemists Soci* 86: 1065-1071.
- Roughan PG, Bartt RD (1968) Quantitative analysis of sulfolipid (Sulfoquinovosyl Diglyceride) and galactolipids (Monogalactosyl and Digalactosyl Diglycerides) in plant tissues. *Anal Biochem* 22: 74-88.
- Luddy FE, Beerford RA, Riemen SRW (1960) Direct conversion of lipid components to their fatty acid methyl esters. *J Amer Oil Chemists Soci* 37: 447-451.
- Tebas P, Stabell EC, Olivo PD (1995) Antiviral susceptibility testing with a cell line which expresses beta-galactosidase after infection with herpes simplex virus. *Antimicrob Agents Chemother* 39: 1287-1291.
- Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, et al. (1990) New colorimetric cytotoxicity assay for anticancer-drug screening. *J Nat Cancer Inst* 82: 1107-1112.
- Greenwood D (1983) Antimicrobial chemotherapy, Part II-Laboratory Aspects of Antimicrobial Therapy Bailliere, Tindall, London.
- Ye H, Wang K, Zhou C, Liu J, Zeng X (2008) Purification, antitumor and antioxidant activities in vitro of polysaccharides from the brown seaweed *Sargassum pallidum*. *Food Chemistry* 111: 428-432.
- Gomes KA, Gomes AA (1984) Statistical procedures for agricultural research. John Wiley and Sons, New York.
- Manivannan K, Karthikai DG, Thirumaran G (2008) Biochemical composition anantharaman, seaweeds from sandapam coastal regions along southeast coast of India. *Amer Euras J Bot* 2: 42-51.
- Matanjun P, Mohamed S, Mustapha MN, Muhammad K (2009) Nutrient content of tropical edible seaweeds, *Euclima cottonii*, *Caulerpa lentillifera* and *Sargassum polycystum*. *J App Phycol* 21: 75-80.
- Araki S, Sakurai T, Oohuso T, Kayama M, Nisizawa K (1990) Content of arachidonic and eicosapentaenoic acids in polar lipids from Gracilaria (Gracilariales Rodophyta). *Hydrobiologia* 204: 513-519.
- Shehnaz, L. (2003). Comparative phytochemical investigations on a variety of marine algae from Karachi coast. Ph.D. Thesis, Kar. University Seaweed Bio.
- Orhan I, Sener B, Atici T (2003) Fatty acid distribution in the lipid extracts of various algae. *Chem Nat Com* 39: 167-170.
- Chirasuwan N, Chaiklahan R, Kittakoop P, Chanasattru W, Ruengjitchachawalya M, et al. (2009). Anti HSV-1 activity of sulphoquinovosyl diacylglycerol isolated from *Spirulina platensis*. *Science Asia* 35: 137-141.
- Novoa VA, Andrade-Wartha RSE, Linares FA, Silva MOA, Genovese IM, et al. (2011) Antioxidant activity and possible bioactive components in hydrophilic and lipophilic fractions from the seaweed *Halimeda incrassata*. *Braz J Pharmaco* 21: 53-57.
- Katz DH, Marcelletti JF, Khalil MH, Pope LE, Katz LR (1991) Antiviral activity of 1-docosanol, an inhibitor of lipid-enveloped viruses including herpes simplex. *Proc Natl Acad Sci USA* 88: 10825-10829.
- Pope LE, Marcelletti JF, Katz LR, Katz DH (1996) Anti-herpes simplex virus activity of n-docosanol correlates with intracellular metabolic conversion of the drug. *J Lipid Res* 37: 2167- 2178.
- Zhang PJ, Zheng L, Wang JH, Magnusson KE, Liu X (2008) Lipid extract from completely sporoderm-broken germinating *Ganoderma sinensis* spores elicits potent antitumor immune responses in human macrophages. *Phytother Res* 23: 844-850.
- Ballantine LD, Gerwick HW, Velez MS, Alexander E, Guevara P (1987) Antibiotic activity of lipid-soluble extracts from Caribbean marine algae. *Hydrobiologia*, 151: 463- 469.
- Ramadan FM, Asker SMM, Zeinab IK (2008) Functional bioactive compounds and biological activities of *Spirulina platensis* lipids. *Czech J Food Scie* 26: 211-222.
- Patra JK, Patra AP, Mahapatra NK, Thatoi HN, Das S, et al. (2009) Antimicrobial activity of organic solvent extracts of three marine macroalgae from Chilika Lake, Orissa, India. *Mal J Microbio* 5: 128-131.
- Desbois PA, Smith JV (2010) Antibacterial free fatty acids: activities, mechanisms of action and biotechnological potential. *Appl Microbiol Biotechnol* 85: 1629-1642.
- Shanab MMS (2007) Antioxidant and antibiotic activities of some seaweeds (Egyptian isolates). *International J Agric Bio* 9: 220-225.
- Shalaby EA (2008) Biochemical and Biotechnological studies on some marine algae. Cairo University, Egypt.