Detection and Discrimination of Bioanalytes by Means of Colorimetric Sensor Array Based on Unmodified Gold and Silver Nanoparticles

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Abstract

A new approach for sensing bioanalytes utilizing unmodified noble metal nanoparticles to fabricate a colorimetric sensor array is described. In this array, a series of gold and silver nanoparticles with various sizes whose color change based on the interactions of bioanalytes and nanoparticles, provide the unique response patterns for bioanalytes such as protein and bacteria which can be distinguished by naked eye. This work indicates that the colorimetric sensor array based on unmodified gold and silver nanoparticles has the potential for application in medical diagnostics.

Keywords: Nanoparticles; Bacteria; Bioanalytes

Introduction

The detection and identification of bioanalytes such as protein and bacteria are very important in the field of medical diagnostics. Enzyme-linked immunosorbent assays, PCR and mass spectrum are the most commonly used methods for bioanalytes sensing which are high costs, time consuming and impossible for multianalytes or mixtures. Optical chemical sensor array called “opto-chemical nose or tongue” is a promising sensing protocol and an alternative diagnostic platforms for a variety of bioanalytes. Rotello used conjugated fluorescent polymers array for protein detection [1]. Jiang reported a sensor array based on five fluorescent probes for the identification of bacteria [2]. Fluorescence sensor array is high sensitive; however, photo bleaching and excitation light source need are problematic. Colorimetric sensor array is simple, fast, and effective way for the detection and discrimination of bioanalytes, providing optical response patterns of analytes readily discriminated by naked eye visualization. The special localized surface plasmon resonance (LSPR) effect makes noble metal nanoparticles an excellent choice of colorimetric sensor. The free electrons of gold nanoparticles (AuNPs) with different sizes and shapes can resonate with light of specific wavelength, so that they show different colors. When the condition of the solution is changed, the color will change too. Zhang developed a colorimetric sensor array for the detection of protein based on gold nanoparticles modified with aptamers [3]. But, all of the colorimetric methods required functionalized AuNPs. Quite a few studies have demonstrated that unmodified noble metal nanoparticles themselves can also interact with biological molecules such as proteins, non-specifically leading to a color change. We have developed a very simple colorimetric sensor array for the detection and discrimination of a range of bioanalytes using unmodified Nobel metal nanoparticles [4]. Here, this paper reviews the colorimetric sensor array and its application in bioanalytes sensing. Compared to previous colorimetric sensor array based on noble metal nanoparticles, this sensor array employs unmodified noble metal nanoparticles as sensing elements, which are much easier to be synthesized and cheaper. In addition, it has been demonstrated that a protein at various concentration can lead to different color changes. Therefore, this sensor array has the potential to be used for bioanalytes sensing.

Colorimetric Sensor Array based on Unmodified Gold and Silver Nanoparticles

Noble metal nanoparticles mainly include gold nanoparticles (AuNPs) and silver nanoparticles (AgNPs), having the most outstanding property is LSPR. Typically, the aggregation of AuNPs and the changes in color of AuNPs occur when the analyte interacts with AuNPs. This LSPR property has been implemented to build colorimetric biosensor and realize visualization detection for various bioanalytes. It has been reported that nanoparticles with different surface properties (e.g. size, shape, surface charge, and coating material) have different interactions with proteins [5,6]. Nanoparticles with different sizes have different surface areas and curvature which have effect on protein folding, because various surface-to-volume ratios have different non-specific interaction of amino acid residue of protein and nanoparticles surface. Therefore, we synthesized five unmodified AuNPs and two unmodified AgNPs with various sizes according to reported literatures [6] to investigate the interactions between AuNPs and bioanalytes. To synthesize AuNPs, chlorauric acid was reduced by sodium citrate. While to synthesize the AgNPs, Tollens’ reagent was reduced by glucose. The size of nanoparticles can be easily changed by adjusting the ratio of oxidant and reductant. Their diameters of AuNPs 1, AuNPs 2, AuNPs 3, AuNPs 4, AuNPs 5, AgNPs 1 and AgNPs 2 are 15 nm, 25 nm, 35 nm, 50 nm, 60 nm, 25 nm and 31 nm respectively. In fact, what is called “unmodified” AuNPs are citrate-stabilized AuNPs because they are not functionalized AuNPs or AgNPs. Five unmodified AuNPs with different sizes (NP1: 15 nm; NP2:25 nm; NP3:35 nm; NP4:50 nm; NP5: 60 nm) are chosen to study the interactions of AuNPs and proteins. When proteins including cytochrome C (Cyt C), myoglobin (Mb), trypsin (Try), pepsin (Pep), lysozyme (Lys), papain (Pap), Casein (Cas), bovine serum albumin (BSA), lipase (Lip), and hemoglobin (HB), incubated with AuNPs,
respectively, the aggregation of AuNPs led to the color changes from red to purple, which can be visualized by the naked eye. As shown in Figure 1, the color change profiles are unique fingerprints for each specific protein at given concentration. Moreover, various concentrations of proteins also result in different color patterns. Therefore, a colorimetric sensor array was designed based on five unmodified AuNPs and two unmodified AgNPs for the detection and discrimination of bioanalytes. These noble metal nanoparticles were used as the sensing elements, which served not only as the receptors but also the indicators. They could bind diversely in the presence of proteins and distinct absorbance response patterns for the proteins were created, as shown in Figure 2.

**Figure 1**: The color change patterns for five unmodified AuNPs of various sizes in the presence of different kinds and concentrations of proteins [4].

![Figure 1](image1.png)

**Figure 2**: Schematic diagrams of the interaction between protein and unmodified Nobel metal nanoparticles.

**Discrimination of Bioanalytes**

In order to evaluate the discrimination ability of the proposed sensor array, ten proteins at concentration of 0.5 μM were identified by absorbance. The proteins include Cyt C, Mb, Try, Pep, Lys, Pap, Cas, BSA, Lip and Hb, which have different molecular weights, isoelectric points (pIs) and oligomeric states. The results show that different proteins have their unique absorbance response patterns due to the interactions of the nanoparticles with proteins. The raw data obtained from the optical response patterns were subjected to linear discriminant analysis (LDA). LDA is a statistic method to recognize the linear combination of features that can differentiate different classes of objects. Herein, LDA was employed to reduce the size of the training matrix (7 sensing elements × 10 proteins × 3 replicates). Three canonical factors were obtained and visualized as a three-dimensional plot. According to this plot, all the proteins were distinguished with accuracy of 100%, as illustrated in Figure 3a.

After successful discrimination of proteins by the proposed sensor array, the next challenge was to sense the real biosamples. As we know, there are a lot of proteins on the surface of bacteria and human cancer cells. Therefore, it is assumed that this sensor array could be used for the identification of bacteria and human cancer cells. To test this assumption, seven bacteria and four human cancer cells were used as real samples to be detected by the proposed sensor array. As shown in Figure 3b and 3c, bacteria and human cells could be differentiated with accuracy of 100%. This study indicates the potential of the sensor array to be used for diagnose of cancers and detection of bacteria.

![Figure 3](image2.png)

**Figure 3**: Identification of bioanalytes [4]. (a) Canonical score plot for three factors of simplified absorbance response patterns obtained with the noble metal nanoparticle-based array against 0.5 μM proteins: Cyt C, Mb, Try, Pep, Lys, Pap, Cas, BSA, Lip and Hb. All proteins were clearly distinguished. (b) Canonical score plot for three factors of simplified absorbance response patterns against bacteria: E. coli, CRPA, Acetobacter aceti, Rhodopseudomonas, Bacillus natto, Staphylococcus, and Bacillus (0.05 OD in 200 μ L). All the bacteria were distinguished perfectly. (c) Canonical score plot for three factors of simplified absorbance response patterns against human cancer cells: Oral squamous carcinoma cells, HeLa cells, PC3 cells, and A549 human lung adenocarcinoma cells (5000 cells in 200 μL).

**Conclusion**

Based on the LSPR of noble metal nanoparticles, a colorimetric sensor array was constructed for the identification of proteins. The sensor array is composed of seven sensing elements, including five unmodified AuNPs and two unmodified AgNPs with various sizes. Ten proteins were successfully distinguished by the proposed colorimetric sensor array. The presence of proteins induced obvious color change of AuNPs, which could be easily recognized by naked eyes. Two real samples including seven bacteria and four cancer cells were also employed to test the discrimination ability of this sensor array. This sensor array is more simple and cheaper that other noble metal nanoparticles-based sensor arrays. Besides, the size of AuNPs and AgNPs can be easily adjusted, which makes it easier to select as many sensor elements as possible. One of the limitations of this sensor array
is that the sensing elements (AuNPs and AgNPs) can’t be used repeatedly. This is also a common limitation of most sensor arrays. Even though there are some limitations, this research indicates the potential of unmodified noble metal nanoparticles for the detection and discrimination of proteins and bacteria.

References


