Effects of Glucagon-Like Peptide-1 Receptor Agonists on \( \beta \)-Cell Function in Patients with Type 2 Diabetes

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**Abstract**

Type 2 diabetes (T2D) is characterized by insulin resistance and a progressive decline in pancreatic \( \beta \)-cell function. Over time, upregulation of insulin secretion is no longer sufficient to compensate for insulin resistance, leading to fasting hyperglycemia, while \( \beta \)-cell function continues to deteriorate. Effective treatments targeting the underlying pathophysiology of T2D involve early lifestyle interventions and a combination of therapies to counteract insulin resistance and progressive deterioration of \( \beta \) cells. Glucagon-like peptide-1 receptor agonists (GLP-1RAs) are one class of antihyperglycemic agents that target multiple pathways, mediating clinical benefits for patients with T2D. This review summarizes the effects of treatment with various GLP-1RAs, including exenatide (twice daily and once weekly), liraglutide, dulaglutide, and lixisenatide, on \( \beta \)-cell function in patients with T2D. \( \beta \)-cell function was assessed in randomized controlled trials and their extension studies using a variety of methods, including glucose and arginine clamp techniques, proinsulin-to-insulin ratio, and homeostatic model assessment of \( \beta \)-cell function (HOMA-B). Exenatide twice daily and liraglutide both improved first- and second-phase insulin secretory responses. In addition, numerous studies reported significant improvements in HOMA-B with exenatide (twice daily and once weekly; range of relative change from baseline: +28–50%; range of absolute change: +5–40%) or liraglutide (range of change vs. placebo: +13–43%). Improvements in HOMA-B were also observed with the newer GLP-1RAs dulaglutide and lixisenatide. In contrast to the effects on HOMA-B, treatment with GLP-1RAs had a lesser effect on insulin sensitivity. Taken together, the results suggest that glucagon-like peptide-1 analogs have a greater effect on \( \beta \)-cell function than insulin sensitivity.

**Keywords:** \( \beta \)-cell function; Exenatide; Glucagon-like peptide-1 receptor agonist; Type 2 diabetes

**Introduction**

Type 2 diabetes (T2D) is a chronic, complex illness characterized by insulin resistance and a progressive failure of \( \beta \)-cell function [1,2]. The natural history of T2D involves early development of postprandial hyperglycemia followed by the development of fasting hyperglycemia. Over time, upregulated insulin secretion is no longer able to compensate for increased insulin resistance, and there is continued deterioration of residual \( \beta \) cells. The pathophysiology of T2D involves deficiencies in insulin, amylin, and incretins. At diagnosis, over 80% of the \( \beta \)-cell response to glucose appears lost [1], and the loss of the first-phase insulin response is recognized as an early \( \beta \)-cell defect in T2D [3]. A multihormonal model of glucose homeostasis beyond the established anabolic and catabolic roles of insulin and glucagon, respectively, is well recognized (Table 1); additional hormones including amylin and glucagon-like peptide-1 (GLP-1) have been identified as crucial components of glucose regulation (Figure 1) [4]. GLP-1, an incretin hormone synthesized in intestinal L cells and released in response to nutrient ingestion, plays a key role in priming the \( \beta \) cell to respond to plasma glucose. Indeed, the “incretin effect” (i.e. augmented insulin secretion in response to glucose ingested in a meal vs. glucose infused intravenously) has been estimated to account for up to 70% of the \( \beta \)-cell response at mealtime [5].

Insulin responses to physiological levels of GLP-1 are severely impaired in T2D but are restored by pharmacological doses of GLP-1. Højberg and colleagues, while investigating the potentiation of glucose-stimulated insulin secretion by GLP-1, identified that physiological levels of GLP-1 resulted in an increase in insulin secretion in healthy subjects. However, in patients with T2D, the insulin response to physiological levels of GLP-1 was substantially reduced [6]. Ylvisbøll et al. investigated GLP-1 stimulation in patients with T2D and found that GLP-1 infused at pharmacological levels (1 pmol/kg/min) augmented late-phase insulin secretion to levels similar to those observed in healthy subjects [7].

GLP-1 receptor agonists (GLP-1RAs) are one class of antihyperglycemic agents that target multiple pathways, mediating clinical benefits for patients with T2D. GLP-1RAs have been shown to reduce HbA1c, fasting and postprandial plasma glucose, and body weight, and are associated with a low risk of hypoglycemia when not taken with other drugs associated with hypoglycemia such as sulfonylureas [8]. A number of GLP-1RAs are currently available, including exenatide twice daily or once weekly, liraglutide once daily, lixisenatide once daily, albiglutide once weekly, and dulaglutide once weekly, which are administered via subcutaneous (SC) injection and indicated as an adjunct to diet and exercise to improve glycemic control in adults with T2D.

GLP-1RAs lower glucose through a multifaceted mechanism of action, as these agents mimic the myriad actions of endogenous GLP-1 [9-13]. Activation of downstream signaling pathways initiated by GLP-1RAs stimulates first- and second-phase postprandial glucose-dependent \( \beta \)-cell function and suppresses inappropriately elevated

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Pancreas

α cells
- Stimulates the breakdown of stored liver glycogen
- Promotes hepatic gluconeogenesis
- Promotes hepatic ketogenesis

β cells
- Affects glucose metabolism and storage of ingested nutrients
- Promotes glucose uptake by cells
- Suppresses postprandial glucagon secretion
- Promotes protein and fat synthesis
- Promotes use of glucose as an energy source

Insulin
- Promotes hepatic glucose metabolism

Amylin
- Suppresses postprandial glucagon secretion
- Slows gastric emptying
- Reduces food intake and body weight

Intestine
L cells
- Enhances glucose-dependent insulin secretion
- Suppresses postprandial glucagon secretion
- Slows gastric emptying
- Reduces food intake and body weight
- Promotes β-cell health

GLP-1
- Slows gastric emptying
- Suppresses postprandial glucagon secretion
- Promotes use of glucose as an energy source
- Affects glucose metabolism and storage of ingested nutrients

GLP-1: glucagon-like peptide-1.
Reproduced from Aronoff SL [4]

Table 1: Effects of primary glucoregulatory hormones.

<table>
<thead>
<tr>
<th>Glucoregulatory Hormone</th>
<th>Effects</th>
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<tbody>
<tr>
<td>Glucagon</td>
<td>Stimulates the breakdown of stored liver glycogen, promotes hepatic gluconeogenesis, promotes hepatic ketogenesis.</td>
</tr>
<tr>
<td>Insulin</td>
<td>Slows gastric emptying, suppresses postprandial glucagon secretion, promotes hepatic glucose metabolism.</td>
</tr>
<tr>
<td>Amylin</td>
<td>Suppresses postprandial glucagon secretion, slows gastric emptying, reduces food intake and body weight.</td>
</tr>
<tr>
<td>GLP-1</td>
<td>Slows gastric emptying, suppresses postprandial glucagon secretion, reduces food intake and body weight.</td>
</tr>
</tbody>
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Figure 1: Glucose homeostasis: roles of insulin, glucagon, amylin, and GLP-1. The multi-hormonal model of glucose homeostasis (non-diabetic individuals): in the fed state, amylin communicates through neural pathways (1) to suppress postprandial glucagon secretion (2) while helping to slow the rate of gastric emptying (3). These actions regulate the rate of glucose appearance in the circulation (4). In animal models, amylin has been shown to dose-dependently reduce food intake and body weight (5). In addition, incretin hormones, such as GLP-1, glucose-dependently enhance insulin secretion (6) and suppress glucagon secretion (2) and, via neural pathways, help slow gastric emptying and reduce food intake and body weight (5). GLP-1: glucagon-like peptide-1. Reproduced with permission from Aronoff SL [4].

glucagon secretion [14], which in turn reduces hepatic glucose production [15]. Furthermore, GLP-1RAs slow gastric emptying, decrease food intake, and promote satiety [14]. Because native GLP-1 and GLP-1RAs were shown to reduce apoptosis and/or increase cellular proliferation and β-cell mass in rodent models of diabetes and insulin resistance [16,17], it was hypothesized that GLP-1RAs may modify β-cell function in patients with T2D. Additionally, a GLP-1RA applied to muscle and adipocyte cell lines has been shown to augment insulin sensitivity in preclinical models [18]. In humans, GLP-1RA treatment increased insulin sensitivity (assessed using hyperinsulinemic-euglycemic clamp studies) in patients with type 1 diabetes [19].

Various methods for evaluating β-cell function are available, including glucose and arginine clamp techniques, the proinsulin-to-insulin ratio, and mathematical equations describing glucose homeostasis such as the homeostatic model assessment of β-cell function (HOMA-B) [20]. HOMA-B is a surrogate measure of the ability of β cells to secrete insulin—rather than a measure of β-cell health—and should be evaluated in light of insulin sensitivity, assessed using the homeostatic model assessment of insulin sensitivity or insulin resistance (HOMA-S or HOMA-IR, respectively) [21]. The proinsulin-to-insulin ratio may be indicative of impaired β-cell secretory capacity, as deteriorating β cells are deficient in processing proinsulin and secrete abnormally high levels of proinsulin relative to insulin [20].

This review examines the impact of GLP-1RAs on β-cell function in patients with T2D by summarizing published randomized controlled trials, extension studies, and post hoc analyses that evaluated β-cell function after treatment with various GLP-1RAs. A PubMed search was conducted using the terms “exenatide,” “liraglutide,” “dulaglutide,” “albiglutide,” or “lixisenatide” and “β-cell function” and “type 2 diabetes.” Results were reviewed and selected individually.

Exenatide

Exenatide was the first GLP-1RA approved for the treatment of T2D and is available in a twice-daily (5–10 µg) SC formulation and a once-weekly (2 mg) extended-release SC formulation. Encapsulation of the active molecule, exenatide, into biodegradable poly (D, L-lactide-co-glycolide) microspheres enables the continuous release of exenatide from the once-weekly formulation [22].

Effects of Intravenous Exenatide on β-Cell Function

Fehse et al. investigated insulin secretory patterns and the effects of exenatide on insulin secretion and found that in patients with T2D (N=13), the first-phase insulin response was reduced compared with healthy control subjects (N=12) [23]. Intravenous infusion of exenatide for 300 minutes increased first-phase (0–10 minutes) and second-phase (defined in this study as 10–120 minutes) insulin secretion by approximately 180–310% (Figure 2). Thus, short-term exposure to exenatide restored the first-phase response, resulting in an insulin secretory pattern similar to that observed in healthy subjects.

Effects of Exenatide Twice Daily on β-Cell Function

Functional studies

DeFronzo et al. investigated β-cell function using hyperglycemic and euglycemic insulin clamp techniques among patients (N=50) on background metformin therapy randomized to receive exenatide twice daily, rosiglitazone, or combination therapy [24]. After 20 weeks, significant reductions from baseline in HbA1c were observed with all treatments (p<0.05). Exenatide twice daily or exenatide twice daily plus rosiglitazone significantly enhanced the first- and second-phase insulin secretory responses compared with baseline (p<0.05), whereas rosiglitazone alone had no significant effect on insulin secretion. In contrast, rosiglitazone was associated with a two-fold increase in insulin sensitivity, while exenatide twice daily had no significant insulin-sensitizing effect. β-cell function was further evaluated using the disposition index, which is calculated from hyperglycemic and hypoglycemic insulin secretory responses towards the end of the study.
was no significant effect of insulin glargine on this benchmark measure of disposition index with exenatide twice daily, whereas there was a reduction in the disposition index with insulin glargine on β-cell function posttreatment with exenatide twice daily, whereas the disposition index demonstrated a small, sustained beneficial effect on β-cell function. The disposition indices increased significantly from baseline after treatment with exenatide twice daily or combination therapy (both p<0.05), but not with rosiglitazone alone. Therefore, exenatide twice daily in combination with rosiglitazone had favorable effects on both β-cell secretory function and insulin sensitivity.

In another study, β-cell function was assessed using glucose- and arginine-stimulated hyperglycemic clamp procedures among patients (N=69) randomly assigned to receive exenatide twice daily or insulin glargine for 52 weeks [25]. After 52 weeks, exenatide twice daily significantly increased all measures of β-cell function compared with insulin glargine. First-phase glucose-stimulated C-peptide secretion was significantly greater with exenatide twice daily versus insulin glargine (geometric mean ± standard error ratio to baseline: 1.78 ± 0.11 vs. 1.17 ± 0.06; p<0.0001). Likewise, the second-phase response was enhanced with exenatide twice daily compared with insulin glargine (3.05 ± 0.22 vs. 1.08 ± 0.05; p<0.0001). Exenatide twice daily significantly increased arginine-stimulated C-peptide secretion compared with insulin glargine (3.19 ± 0.24 vs. 1.31 ± 0.07; p<0.0001). In both treatment groups, measures returned to baseline values following 4 weeks of study drug discontinuation. Thus, β-cell improvements reflected functional responses to exenatide exposure, rather than persistent β-cell modification. Thirty-six patients completed the 2-year extension study [26]. After 3 years of total exposure followed by 4 weeks of discontinuation period, glucose- and arginine-stimulated responses returned to baseline measures among patients treated with exenatide twice daily or insulin glargine. The change from baseline in the disposition index demonstrated a small, sustained beneficial effect on β-cell function posttreatment with exenatide twice daily, whereas there was a reduction in the disposition index with insulin glargine (1.43 ± 0.78 vs. −0.99 ± 0.65; p=0.028). Thus, there was a sustained improvement in the disposition index with exenatide twice daily, which was maintained 4 weeks after treatment discontinuation, whereas there was no significant effect of insulin glargine on this benchmark measure of β-cell function despite similar glycemic control. Glycemic control reverted to pretreatment levels in all patients, indicating improvements in β-cell function did not reverse glycemic dysfunction.

Recently, Xu et al. reported results from a randomized controlled trial directly comparing the effects of exenatide twice daily, premixed insulin, and pioglitazone on β-cell function in newly diagnosed, treatment-naïve patients with T2D (N=342) [27]. After 48 weeks, significant improvements in the proinsulin-to-insulin ratio, acute insulin response, and disposition index were observed across treatments, while only insulin significantly improved HOMA-B. Exenatide twice daily, insulin, and pioglitazone increased the acute insulin response from baseline by 42.4, 30.4, and 31.9 μIU mL−1 × min, respectively (exenatide twice daily and pioglitazone: p<0.001; insulin: p<0.05), and increased the disposition index from baseline by 1.3, 0.5, and 0.4 (all p<0.05). Thus, early intervention with glucose-lowering treatments improved multiple measures of β-cell function with continued treatment.

**HOMA-B**

Changes in HOMA-B after treatment with exenatide twice daily in various clinical studies are summarized in (Table 2). In order to provide a meaningful interpretation of β-cell function, it is important to evaluate function within the context of insulin resistance, which precedes β-cell deterioration and is compensated for by increased insulin secretion [28]. Thus, when assessing β-cell function using HOMA measures, it is important to also evaluate insulin resistance (measured using HOMA-IR) or the inverse, insulin sensitivity (HOMA-S). In general, exenatide twice daily had little or no effect on insulin sensitivity; however, few data exist reporting HOMA-B measures in conjunction with HOMA-S or HOMA-IR measures.

Several studies have investigated the use of exenatide twice daily early in the course of disease progression. β-cell function was assessed using HOMA-B among treatment-naïve patients randomized to receive exenatide 5 µg, exenatide 10 µg, or placebo twice daily for 24 weeks [29]. HOMA-B increased from baseline by 32% and 28% in the exenatide 5 µg and 10 µg groups, respectively, versus 6% in the placebo group. Improvements were significantly greater with exenatide twice daily versus placebo (exenatide twice daily 5 µg, p=0.002; exenatide twice daily 10 µg, p=0.010). Thus, the use of exenatide therapy early in the treatment paradigm substantially improved β-cell function. In obese Chinese patients newly diagnosed with T2D and naïve to treatment for diabetes, HOMA-B increased slightly, but not significantly, after 26 weeks of treatment with exenatide twice daily or metformin (Table 2) [30]. In contrast, HOMA-S (reflecting insulin sensitivity) increased significantly with exenatide twice daily compared with metformin (p=0.042). Relatively high levels of initial β-cell function—most likely due to the short duration of diabetes (<1 month) in these patients—may have accounted for the lack of significant improvement in HOMA-B. Nevertheless, exenatide twice daily preserved β-cell function over 26 weeks.

Changes in HOMA-B were assessed among 86 patients who received exenatide twice daily or oral sitagliptin for 4 weeks before crossing over to the other therapy for an additional 4 weeks [31]. Both treatments significantly improved HOMA-B, with a significantly greater change from baseline observed with exenatide twice daily versus sitagliptin (+32.9 ± 3.8% vs. +20.8 ± 3.9%; p=0.005).

In a randomized, placebo-controlled trial (N=194) comparing 24 weeks’ treatment with exenatide twice daily in combination with lifestyle modification versus lifestyle modification alone, HOMA-B improved significantly in both treatment groups [32].
Table 2: HOMA-B, HOMA-IR, and HOMA-S data from clinical trials of exenatide and liraglutide.
increased by 42% with exenatide twice daily plus lifestyle modification (p<0.0001) and by 23% with lifestyle modification alone (p=0.0015). There was no significant difference between treatment groups. Neither treatment had a significant effect on insulin sensitivity.

β-cell function was investigated using HOMA-B in 112 patients treated for 2 years with exenatide twice daily in a non-controlled extension of three randomized controlled trials [33]. HOMA-B data demonstrated a significant improvement in β-cell function after 2 years, with a mean relative change of almost 50% compared with baseline (p<0.01).

Among patients (N=128) randomized to receive exenatide twice daily or glibenclamide for 1 year, exenatide twice daily significantly improved HOMA-B (Table 2; p<0.01), whereas glibenclamide had no significant effect on HOMA-B [34]. HOMA-B was significantly higher with exenatide twice daily compared with glibenclamide (p<0.05), demonstrating superior improvements with exenatide twice daily. Exenatide twice daily also significantly improved HOMA-IR (p<0.01).

In a subsequent study, Derosa et al. measured β-cell function among patients (N=171) treated with exenatide twice daily or placebo, both in conjunction with metformin [35]. After 1 year, exenatide twice daily significantly increased HOMA-B compared with baseline (p<0.01) and placebo (+29%; p<0.05). Furthermore, exenatide twice daily significantly increased first- and second-phase glucose-stimulated C-peptide responses and arginine-stimulated C-peptide responses compared with baseline (all p<0.05) and placebo (all p<0.05). In addition, exenatide twice daily increased the disposition index versus baseline (+102.0 nmol/l×μmol kg⁻¹; p<0.01) and placebo (+67.5 nmol/l×μmol kg⁻¹; p<0.05). Thus, exenatide twice daily improved multiple measures of β-cell function.

Preumont et al. undertook a study (N=33) to determine whether improved glycemic control with exenatide twice daily was related to enhanced β-cell function and/or insulin sensitivity [36]. After 6 months of exenatide therapy, unadjusted HOMA-B improved significantly (p=0.0201), while no significant change in HOMA-S was observed (Table 2). Furthermore, exenatide twice daily resulted in a relative increase of 70% in β-cell function adjusted for individual insulin sensitivity (HOMA hyperbolic product; p=0.0055). The authors concluded that increased β-cell function played a larger role than insulin sensitivity in improved glycemic control.

Effects of Exenatide Once Weekly on β-Cell Function

Clinical studies investigating exenatide once weekly as part of the DURATION-1 (Diabetes Therapy Utilization: Researching Changes in treatment discontinuation. These results suggest that improvements in β-cell health were maintained while patients continued long-term treatment with exenatide once weekly.

In DURATION-3, exenatide once weekly exhibited greater increases in HOMA-B after 26 weeks (ratio to baseline: 1.69) compared with insulin glargine (1.34) [39]. DURATION-4 (N=820) compared HOMA-B measures with exenatide once weekly versus metformin, pioglitazone, and sitagliptin [40]. After 26 weeks, exenatide once weekly improved β-cell function to a greater degree versus oral comparators; the endpoint to baseline ratio of HOMA-B was 1.8 with exenatide once weekly versus 1.4, 1.3, and 1.3 with metformin, pioglitazone, and sitagliptin, respectively (all p<0.001).

Effects of Liraglutide Once Daily on β-Cell Function

Liraglutide, a once-daily GLP-1RA, is approved at SC doses of 1.2 mg and 1.8 mg in the United States and Europe and 0.9 mg in Japan. The effects of liraglutide on β-cell function have been studied in multiple clinical trials, and effects on HOMA-B specifically are summarized in Table 2.

Vilsbøll et al. measured β-cell function in patients (N=39) randomized to receive liraglutide 0.65, 1.25, or 1.9 mg/day or placebo for 14 weeks [41]. Compared with placebo, liraglutide 1.25 mg and 1.9 mg significantly increased first-phase insulin secretion by 118% and 103%, respectively (p=0.02 and p=0.05). Only liraglutide 1.25 mg significantly increased second-phase insulin secretion (by 79%) compared with placebo (p=0.005). Liraglutide 1.25 mg and 1.9 mg also significantly increased arginine-stimulated insulin secretion (by 114% and 94%, respectively; p=0.02 and p=0.01). Liraglutide had no significant effect on insulin sensitivity or fractional glucose disposal rate. Improvements in first- and second-phase insulin release, as well as arginine-stimulated insulin secretion during hyperglycemia, demonstrate a beneficial effect of liraglutide on β-cell function.

The randomized controlled LIBRA (Liraglutide and β-Cell Repair) trial investigated the effects of liraglutide on preservation of β-cell function in patients (N=51) with early T2D (duration of 2.6 years), using the Insulin Secretion-Sensitivity Index-2 (ISSI-2) on oral glucose tolerance test [42]. After 48 weeks, baseline-adjusted ISSI-2 was significantly higher with liraglutide versus placebo (339.8 vs. 229.3; p=0.008). However, 2 weeks after treatment discontinuation, the benefit with liraglutide was lost (ISSI-2: 191.9 vs. 238.1; p=0.20). The lack of response posttreatment suggests that functional improvements in response to liraglutide exposure did not translate into persistent preservation of β-cell function.

In the first LEAD (Liraglutide Effect and Action in Diabetes-1) trial, effects on β-cell function with liraglutide were compared with rosiglitazone or placebo in patients also receiving glimepiride therapy (N=1040) [43]. Improvements in the proinsulin-to-insulin ratio were significantly greater with liraglutide 1.2 mg and 1.8 mg compared with placebo or rosiglitazone (all p ≤ 0.05). Additionally, liraglutide 1.2 mg and 1.8 mg resulted in significant increases in HOMA-B of 39% and 30%, respectively, compared with rosiglitazone (both p ≤ 0.05). Liraglutide 1.2 mg significantly increased HOMA-B by a difference of 43% compared with placebo (p ≤ 0.05). Taken together, these results demonstrate that combination therapy with liraglutide and glimepiride improved β-cell function to a greater degree than rosiglitazone plus glimepiride or glimepiride alone.

In the LEAD-4 trial (N=533), liraglutide significantly improved multiple measures of β-cell function versus placebo [44]. After 26 weeks, liraglutide 1.2 mg and 1.8 mg both significantly improved the
fasting proinsulin-to-insulin ratio, fasting C-peptide, and HOMA-B compared with placebo (all p<0.05). Both doses of liraglutide resulted in an increase in HOMA-B of 27%.

Improvements in β-cell function—evaluated with the HOMA-B index, proinsulin-to-insulin ratio, and proinsulin-to-C-peptide ratio—were also observed among a Japanese population with T2D (N=664) included in two 24-week randomized controlled trials [45]. Liraglutide significantly improved HOMA-B versus placebo (liraglutide 0.6 mg: +12.5%, p=0.0470; liraglutide 0.9 mg: +20.7%, p=0.0012). In addition, liraglutide significantly improved the proinsulin-to-insulin ratio versus glibenclamide (liraglutide 0.9 mg: −0.81, p<0.0001) and placebo (liraglutide 0.6 mg: −0.57, p=0.0002; liraglutide 0.9 mg: −0.69, p<0.0001). Liraglutide significantly improved the proinsulin-to-C-peptide ratio versus glibenclamide (liraglutide 0.9 mg: −1.85, p<0.0001) and placebo (liraglutide 0.6 mg: −0.99, p=0.0016; liraglutide 0.9 mg: −1.38, p<0.0001). Thus, liraglutide improved several indicators of β-cell function compared with either glimepiride or placebo.

A meta-analysis of six large, 26-week, phase 3 liraglutide trials (LEAD-1 through LEAD-6 trials) examined the impact of liraglutide therapy on β-cell function, using measures of HOMA-B and the proinsulin-to-insulin ratio [46]. Liraglutide 1.2 mg and 1.8 mg improved HOMA-B by a mean change of 31.7% and 35.7%, respectively. With the exception of glimepiride in combination with metformin (mean change: +31.8%), comparators in the trials improved HOMA-B to a lesser extent (rosiglitazone: +9.5%, p<0.05; exenatide twice daily: +5.7%, p=0.0001; placebo: +7.5%, p<0.0001). The mean change in the proinsulin-to-insulin ratio with liraglutide (either dose: −0.08) exceeded changes observed with rosiglitazone (−0.02) and glimepiride (−0.02). Changes in the proinsulin-to-insulin ratio with exenatide and liraglutide were similar. Thus, liraglutide treatment significantly improved markers of β-cell function in this pooled population.

Effects of Newer GLP-1RAs on β-Cell Function

Dulaglutide

Dulaglutide, a once-weekly GLP-1RA, is available as 0.75-mg and 1.5-mg SC doses. Results for dulaglutide from the AWARD (Assessment of Weekly Administration of LY2189265 [dulaglutide] in Diabetes)-1 trial demonstrated significant improvements in measures of β-cell function [47]. After 52 weeks, changes from baseline for the updated homeostasis model of β-cell function (HOMA2-%B) were significantly greater with dulaglutide 1.5 mg (+35 ± 2.6%) and dulaglutide 0.75 mg (+26 ± 2.7%) versus exenatide twice daily (+14 ± 2.8%; both doses, p<0.001). Neither dulaglutide nor exenatide twice daily conferred benefits on insulin sensitivity, as measured by the updated homeostasis model of insulin sensitivity (HOMA2-%S). To date, there are no head-to-head clinical trials comparing the effects of dulaglutide and exenatide once weekly on β-cell function; however, exenatide once weekly resulted in greater improvements in β-cell function compared with exenatide twice daily [37].

In the AWARD-3 trial, dulaglutide 1.5 mg and 0.75 mg resulted in HOMA2-%B increases of 30.0 ± 3.5% and 22.6 ± 3.5%, respectively, which were significantly improved compared with metformin (+9.8 ± 3.5%; p=0.001 vs. dulaglutide 1.5 mg, p<0.05 vs. dulaglutide 0.75 mg) [48]. Changes in HOMA2-%S were smaller with dulaglutide (+5.2 ± 2.4% and +2.7 ± 2.4% with dulaglutide 1.5 mg and 0.75 mg, respectively) compared with metformin (+10.8 ± 2.4%), consistent with a lesser effect of GLP-1RAs on insulin sensitization.

Results from the AWARD-5 trial (N=1098) demonstrated significant improvements in β-cell function with dulaglutide compared with sitagliptin or placebo [49]. After 52 weeks, the change from baseline in HOMA2-%B significantly improved with dulaglutide 1.5 mg (+33.6 ± 2.5%) and dulaglutide 0.75 mg (+22.3 ± 2.5%) versus sitagliptin (+6.7 ± 2.5%; both doses p<0.001). Changes in HOMA2-%S were similar across treatment groups (dulaglutide 1.5 mg: +4.7 ± 2.4%; dulaglutide 0.75 mg: +2.3 ± 2.3%; sitagliptin: +4.3 ± 2.4%). In the AWARD-6 trial (N=599), improvements in HOMA2-%B were similar with either dulaglutide or liraglutide after 26 weeks of treatment [50].

Lixisenatide

Lixisenatide, a once-daily GLP-1RA available as 10-µg and 20-µg SC doses, is approved in Europe for the treatment of T2D in combination with oral glucose-lowering agents and/or basal insulin. Lixisenatide is also available in Australia, Japan, and several countries in Latin America. In a 24-week, randomized, placebo-controlled trial (N=680), treatment with lixisenatide 20 µg once daily (either in the morning or evening) significantly improved β-cell function compared with placebo [51]. The change from baseline for HOMA-B was +8.0 ± 2.45%, +4.8 ± 2.49%, and −4.2 ± 2.82% for lixisenatide morning, lixisenatide evening, and combined placebo treatment groups, respectively (lixisenatide morning vs. placebo: p=0.0002; lixisenatide evening vs. placebo: p=0.0071).

Measures of β-cell function with albiglutide, a once-weekly GLP-1RA, are not currently reported in the published literature.

Association of β-Cell Failure With Treatment Outcomes

A recent prospective study of 546 patients with T2D initiating GLP-1RA therapy analyzed the association between clinical markers of β-cell failure at baseline and glycemic response (change in HbA1c) after 6 months’ treatment [52]. All markers of low β-cell function, including lower fasting C-peptide, lower postmeal urine C-peptide-to-creatinine ratio, positive glutamic acid decarboxylase or islet antigen-2 autoantibodies, longer diabetes duration, and insulin cotreatment, were associated with reduced glycemic response (p ≤ 0.01 for all). Thus, consistent with the action of GLP-1RAs to potentiate β-cell insulin secretion thereby improving glycemic control, patients with low β-cell function at baseline were less responsive to GLP-1RAs.

Conclusions

There is a large body of published data from randomized controlled clinical trials, extension studies, and post hoc analyses demonstrating improvements in β-cell function with GLP-1RA therapies. Exenatide twice daily and lixisenatide both improved first- and second-phase insulin secretory responses in patients with T2D. Reductions in the proinsulin-to-insulin ratio further support the beneficial effect of GLP-1RAs on β-cell function. Numerous studies reported significant improvements in HOMA-B among patients treated with exenatide (twice daily or once weekly), liraglutide, dulaglutide, and lixisenatide, while data on the effects of albiglutide on β-cell function are not currently available. Long-term treatment with exenatide once weekly demonstrated durable benefits for β-cell function, with improvements in HOMA-B sustained for up to 3 years on treatment. However, GLP-1RA-induced improvements in β-cell function generally reversed upon treatment discontinuation, suggesting that functional augmentation of β-cell secretory capacity may not reflect long-term modulation of this system. In contrast to the effects on β-cell function, treatment with GLP-1RAs had a lesser effect on insulin sensitivity.

Overall, the evidence suggests that GLP-1RAs support β-cell
function, thereby improving glycemic control. Additional controlled studies are necessary to investigate whether continued treatment with GLP-1RAs or other glucose-lowering therapies can protect against deterioration of β-cell function if administered early in the course of T2D.

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Author Disclosures
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