Efficient Direct Electron Transfer for a Highly Oriented PQQ-GDH Immobilized Electrode for Bioanode

Ayako Koto, Saki Taniya, Hiroaki Sakamoto*, Takenori Satomura, Haruhiko Sakuraba, Toshihisa Oshima and Shin-ichiro Suye

Abstract

A bioanode with improved enzyme orientation was developed to achieve an efficient enzyme reaction and electron transfer on an electrode surface. A highly stable PQQ-dependent glucose dehydrogenase (PQQ-GDH) isolated from a hyper-thermophilic archaeon was employed as an electron conversion element. PQQ-GDH is expected to maintain battery properties and to have a long battery life. To immobilize the enzyme onto the electrode with appropriate orientation, we introduced a His-tag to the N-terminal of PQQ-GDH by a genetic technique and utilized the affinity bond between His-tag and Cu atoms. The catalytic current density in the presence of substrate was 18.6 μA/cm² without a mediator. The current density of the oriented electrode was approximately 90 times higher than that of the non-oriented electrode. By immobilizing the enzyme with orientation, the accessibility between the enzyme and substrate for enzyme reaction increased because the active site of PQQ-GDH is located opposite the electrode. Because enzymes have different orientations at the surface of the non-oriented electrode, the efficiency of the electrode was lower than that of the high-orientation electrode. The results of the present study present a potentially promising finding for application to practical bioelectric devices, such as bio-fuel cells and biosensors.

Keywords: Bioanode; PQQ-GDH; Oriented immobilization; Direct electron transfer

Introduction

Bio-fuel cells catalyzed by enzymes are expected to provide next-generation energy supply sources for microelectronic devices because they are safe to use and are light in weight and can be easily miniaturized [1-4]. The principle of bio-fuel cells is based on the redox reaction between the enzyme and substrate. The substrates are prepared from biomass and include glucose, methanol, ethanol, and lactate. Previously, many researchers have developed enzymatic bio-fuel cells [5-13] that were shown to have current and power densities to some extent, but were limited by major issues including low output of the battery because of inefficient redox reaction on the electrode surface and lower active lifetime because of low stability of enzyme [14,15]. Therefore, it has been considered difficult to achieve a practical bio-fuel cell thus far.

In the present study, in order to overcome the above issues, the following two strategies were used.

The first strategy was to increase the orientation of the enzyme immobilized onto the electrode surface. Because electron-transfer between the enzyme and electrode, and enzyme reactivity on the electrode surface, depend on the orientation of enzyme [16-19], controlled and oriented immobilization of enzyme onto the electrode surface is required [20-25]. However, it is not easy to modulate macromolecules, e.g., enzymes and peptides, on the electrode surface. Because macromolecules have an intricate chemical and physical structure, the enzyme orientation on the electrode surface is the major rate-limiting factor for enzyme function. In the past, various approaches, such as covalent cross-linking [26,27], Layer-By-Layer (LBL) [28-30], and Self-Assembled Monolayers (SAM) [31-34], have been reported for the immobilization of macromolecules on solid substrates. Almost all of these techniques have been based on covalent cross-linking through chemical reactions that lack site selectivity. Using the above immobilization methods, specificity of the reaction site in the macromolecule cannot be obtained because the macromolecules have more than one reactive group on their surface.

The second approach is to employ PQQ-GDH obtained from the hyperthermophilic archaeon, Pyrobaculum aerophilum [35]. PQQ-GDH from hyperthermophiles, which can grow at or near the boiling point of water, is much more stable than that obtained from mesophilic microorganisms or eukaryotic organisms. In addition, PQQ-GDH can utilize artificial dyes such as 2,6-Dichloroindophenol (DCIP) and ferricyanide as electron acceptors. PQQ-GDH has high potential for utilization as a specific element for a bioanode electrode because electrons from D-glucose can be introduced to the electrode using an artificial dye as the mediator. Use of PQQ-GDH from the hyperthermophilic archaeon provided a long cell life and high power output.

In the present study, we developed a bio-anode electrode by highly oriented immobilization of PQQ-GDH. To immobilize the enzyme with orientation, we employed the interaction between His-tag (consisting of six successive histidine residues) and the Cu atom. As the His-tag acts as an immobilization site on the electrode surface, the enzyme can be immobilized with a high degree of orientation (Figure 1). We have introduced the His-tag into the PQQ-GDH from Pyrobaculum aerophilum by using a genetic technique. The PQQ-GDH-(His-tag) was immobilized onto a Cu-deposited electrode with a high degree of orientation. The highly oriented immobilized PQQ-GDH electrodes were investigated using electrochemical techniques to obtain information on the properties of the constructed electrode.

*Corresponding author: Hiroaki Sakamoto, Department of Frontier Fiber Technology and Science, Graduate School of Engineering, University of Fukui, Fukui, Japan, Tel: 0776-27-9753; E-mail: hi-saka@u-fukui.ac.jp

Received January 29, 2014; Accepted March 27, 2014; Published April 02, 2014


Copyright: © 2014 Koto A, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Material and Methods

**Chemicals**

D(+)-Glucose, sodium chloride, copper(II) sulfate pentahydrate (CuSO₄·5H₂O), nickel(II) sulfate hexahydrate (NiSO₄·6H₂O), imidazole, 1-Ethyl-3-(3-Dimethylaminopropyl) Carbodiimide Hydrochloride (EDC) and Pyrroloquinoline quinone disodium salt (PQQ) were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan). Potassium chloride (KCl) and 2-[4-(2-Hydroxyethyl) pipеразин-1-yl] ethanesulfonic acid (HEPES) were obtained from Nacalai Tesque, Inc. (Kyoto, Japan). 2-Morpholinoethanesulfonic acid, monohydrate (MES) was purchased from Daidojo Laboratories (Kumamoto, Japan). The water used in the present experiment was first deionized and passed through a Milli-Q water purification system from Millipore Co. (Bedford, MA, USA).

**Preparation of PQQ-GDH-(His-tag)**

To prepare purified PQQ-GDH-(His-tag) by nickel affinity chromatography, a six-His sequence was fused at the N-terminal of the PQQ-GDH protein. The pET15b/PaeAsd plasmid encoding PQQ-GDH-(His-tag) was expressed in *Escherichia coli* BL21- CodonPlus(DE3)-RIL grown in LB medium containing ampicillin. Recombinant PQQ-GDH-(His-tag) proteins were induced by adding 1 mM isopropyl-β-D-thiogalacto-pyranoside at OD₆₀₀ = 0.6, and cultivation was continued for an additional 4 h. Cells were harvested by centrifugation at 10,000×g, suspended in lysis buffer (20 mM imidazole, 1 mM CaCl₂), and disrupted by ultrasonication. The cell debris was removed by centrifugation (12,000×g for 10 min) and the supernatant solution was used as the crude extract. The enzyme solution was incubated at 50°C on an ALS electrochemical analyzer (BAS Inc., Tokyo, Japan). The working electrode was gold (geometrical area: 0.02 cm², BAS Inc.), the counter electrode was a platinum wire, and the reference electrode was Ag/AgCl. All potential values below are reported with respect to the Ag/AgCl reference electrode. The electrolyte solution was 50 mM HEPES buffer at pH 7.5. The CV measurements were recorded at a potential scan rate of 0.01 V/s. The working electrode was sonicated in 50 mM H₂SO₄, and then the surface was polished with a micro-cloth and alumina slurry suspension (q = 0.1 μm). The electrode was scanned 50 times at a potential scan rate of 0.1 V/s, at potentials ranging from -1.0 V to 1.0 V. Finally, the electrode was sonicated in Milli-Q water.

**Electrode modification**

Au/Cu/ PQQ-GDH-(His-tag): CuSO₄ was dissolved in 100 mM sodium chloride to a final concentration of 100 mM. Copper deposition was controlled by applying a potential of -0.3 V vs. Ag/AgCl to the Au electrode for 5 min. Then, the electrode was rinsed with Milli-Q water and immersed in 0.98 mg/ml PQQ-GDH-(His-tag) solution (at room temperature, 5 min) to immobilize the PQQ-GDH-(His-tag). The electrode was rinsed again with 50mM HEPES buffer at pH 7.5.

C / PQQ-GDH-(His-tag): The freshly polished carbon electrode (0.07 cm²) was immersed in 100 mM H₂SO₄ for 1 h, washed, and electrocytically oxidized at +1.2 V in 5% K₂Cr₂O₇ in 10% HNO₃ to introduce carboxyl groups on the electrode surface. The electrode was immersed in freshly prepared 400 mM EDC solution in 10 mM MES buffer at pH 4.5 for 1 h, and then immersed in 0.98 mg/ml enzyme solution overnight at room temperature.

**Quantification of the immobilized enzyme**

To remove the enzyme from electrode surface, the Au/Cu/ PQQ-GDH-(His-tag) was expressed in *Escherichia coli* BL21- CodonPlus(DE3)-RIL grown in LB medium containing ampicillin. Recombinant PQQ-GDH-(His-tag) proteins were induced by adding 1 mM isopropyl-β-D-thiogalacto-pyranoside at OD₆₀₀ = 0.6, and cultivation was continued for an additional 4 h. Cells were harvested by centrifugation at 10,000×g, suspended in lysis buffer (20 mM imidazole, 1 mM CaCl₂), and disrupted by ultrasonication. The cell debris was removed by centrifugation (12,000×g for 10 min) and the supernatant solution was used as the crude extract. The enzyme solution was incubated at 80°C for 10 min. Denatured proteins were separated by centrifugation (12,000×g for 10 min). The resultant supernatant was used for further purification. To reconstitute the apoenzyme with PQQ, a 10-fold molar excess of PQQ was added into the enzyme solution and incubated it for 16 h at 4°C.

To obtain purified PQQ-GDH-(His-tag), the holoenzyme, after reconstitution of the apoenzyme with PQQ, was purified by using Ni-NTA chromatography (HisTrap HP column; GE Healthcare UK Ltd, Little Chalfont, Bucks, UK). The enzyme was loaded onto the column equilibrated with 20 mM potassium phosphate buffer at pH 7.5 and eluted from the column using an elution buffer (300 mM NaCl, 500 mM imidazole, 20 mM potassium phosphate buffer, at pH 7.0). The purified PQQ-GDH-(His-tag) proteins were dialyzed to remove imidazole and replace the buffer (50 mM HEPES pH 7.5), and stored at room temperature until further use.

The activity of PQQ-GDH-(His-tag) was measured using 2,6-Dichlorophenolindophenol (DCIP) as described previously [37].

Cyclic Voltammetry (CV) was conducted using a three-electrode cell at 50°C on an ALS electrochemical analyzer (BAS Inc., Tokyo, Japan). The working electrode was gold (geometrical area: 0.02 cm², BAS Inc.), the counter electrode was a platinum wire, and the reference electrode was Ag/AgCl. All potential values below are reported with respect to the Ag/AgCl reference electrode. The electrolyte solution was 50 mM HEPES buffer at pH 7.5. The CV measurements were recorded at a potential scan rate of 0.01 V/s. The working electrode was sonicated in 50 mM H₂SO₄, and then the surface was polished with a micro-cloth and alumina slurry suspension (q = 0.1 μm). The electrode was scanned 50 times at a potential scan rate of 0.1 V/s, at potentials ranging from -1.0 V to 1.0 V. Finally, the electrode was sonicated in Milli-Q water.

**Estimation of electrode area**

CV was conducted with the Au/Cu electrode and the carbon electrode in the 25 mM potassium ferricyanide/25 mM potassium ferrocyanide solution. Potassium ferrocyanide and potassium ferricyanide react as shown in Eq. (1). The peak reflecting the number of molecules that have reacted at the electrode surface obtained from CV and the electrode area (A) was calculated using the peak and Eq. (2). Iₚ is the peak current, n is the number of electrons, DR is the diffusion coefficient, v is the scan rate, and C is the concentration.

\[
\frac{[\text{Fe(CN)}_6]^{3-}}{[\text{Fe(CN)}_6]^{4-} + e^-} \quad \text{Fe CN Fe CN e} \\
\]

\[
A = \frac{0.5 \cdot \text{I}_p \cdot \text{D}^{1/2} \cdot \text{C}}{2699 \text{v}^{1/2}} \\
\]

**Figure 1: Schematic illustration of PQQ-GDH-(His-tag) immobilization onto the Cu deposited electrode surface. Depoated Cu (orange spheres) and His-tag (green arcs) are forming affinity binding.**
Using this reaction, we can immobilize enzymes with a high degree of orientation via a His-tag, without requiring modification involving NTA. Therefore, we prepared the PQQ-GDH-(His-tag) modified electrode on the deposited Cu electrode (Au/Cu/PQQ-GDH-(His-tag)). Figure 3 shows the CV of the catalytic reaction of the Au/Cu/PQQ-GDH-(His-tag) electrode. Because deposited Cu atoms are not oxidized at less than 0 V, CV was performed from -0.3 V to 0 V. When using the Au/Cu/PQQ-GDH-(His-tag) in the absence of substrate, a small peak was observed at -0.1 V, which represents oxidation of PQQ in the enzyme by DET, which confirmed DET between the enzyme and electrode, as previously reported [36-38]. When 5 mM D-glucose existed in the electrolyte solution, the catalytic current arising from the oxidation reaction catalyzed by PQQ-GDH-(His-tag) was observed in the vicinity of -0.1 V vs. Ag/AgCl. When PQQ was not incorporated in the GDH and enzymes were not immobilized on the electrode, a similar peak was not observed, even after addition of D-glucose. These findings prove that a catalytic oxidation current was obtained from the reaction of immobilized enzymes on the Au/Cu electrode.

**Confirmation of highly oriented immobilization**

We have confirmed that the PQQ-GDH-(His-tag) is immobilized to the Au/Cu electrode via the His-tag. His-tag is a short peptide chain consisting of six histidine residues that coordinates to the transition metal by sharing the electron density of the nitrogen of the imidazole ring of histidine in the His-tag. Upon imidazole treatment, imidazole molecules act antagonistically to the imidazole ring of histidine residues of His-tag; then, the His-tag leaves the Cu, and the protein is removed from the electrode surface. In the CV measurements using electrodes treated with imidazole, the peak value of the catalytic current of oxidation of D-glucose was dramatically decreased (Figure 4). With an increase in the concentration of imidazole, reduction of the peak value was also observed. Because PQQ-GDH-(His-tag) is definitely eliminated by imidazole treatment, the enzyme definitely binds to the electrode via Cu atoms. Therefore, the immobilized enzymes may achieve a high degree of orientation on the electrode.
The oxidation peak current of PQQ (IpPQQ) was determined using the electrode reaction of potassium ferricyanide. In this study, the His-Cu affinity bond and (His-tag) shows DET properties derived from the redox of PQQ in CV measurements (Figure 3). In this study, the His-Cu affinity bond and (His-tag) shows DET properties derived from the redox of PQQ in CV measurements (Figure 3). To determine the efficiency of enzyme reaction on the electrode surface, the properties of the electrode (electrode area and total amount of electroactive coverage on electrodes) were analyzed.

The electrode area of the Cu-deposited Au electrode (Au/Cu) was determined using the electrode reaction of potassium ferricyanide. The value of -0.925 mA was obtained for the reduction peak; thus, the electrode area was estimated to be 0.40 cm² by using equation (2). Before deposition, the geometric area of the Au electrode was 0.02 cm², so the electrode area was increased by 20-fold compared with the bare Au electrode.

To determine the total amount of immobilized enzyme on the electrode, we modified gold electrodes as described above and then washed off the His-tagged enzymes by immersing the electrode in the imidazole solution. The total amount of protein in the imidazole solution was measured. The protein concentration was determined according to the BSA calibration curve. PQQ-GDH-(His-tag) is immobilized to Cu by a His-tag attached to the N-terminal of the enzyme. It is expected that the catalytic reaction is facilitated because accessibility of the substrate to the active site of the enzyme is located opposite to the electrode surface (Figure 5b). Because the enzyme molecules had different orientations on the non-oriented electrode surface, the efficiency was lower than that of the highly oriented electrode. These results demonstrated in this study will allow high-performance bio-fuel cells to be designed and constructed, and lead to practical usage.

<table>
<thead>
<tr>
<th>Immobilization Method</th>
<th>Random Immobilization</th>
<th>Covalent Bond</th>
<th>High-oriented Immobilization</th>
<th>(His-tag affinity)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Current Density (µA/cm²)</td>
<td>0.20</td>
<td>0.15</td>
<td>4.19x10⁻¹¹</td>
<td>18.6</td>
</tr>
<tr>
<td>IpPQQ (µA)</td>
<td>0.73</td>
<td>4.19x10⁻¹¹</td>
<td></td>
<td>0.73</td>
</tr>
<tr>
<td>Electroactive molecules (mol/cm²)</td>
<td>5.10x10⁻¹¹</td>
<td>4.19x10⁻¹¹</td>
<td></td>
<td>5.10x10⁻¹¹</td>
</tr>
</tbody>
</table>

**Table 1:** Comparison of electrochemical properties in each electrode system.

**Effect of the high degree of orientation of the enzyme**

We compared the efficiency of a highly oriented electrode containing a His-tag (Au/Cu/PQQ-GDH-(His-tag)) and a non-oriented electrode functionalized by EDC (C/PQQ-GDH-(His-tag)). To evaluate the efficiency of enzyme reaction on the electrode surface, the properties of the enzyme (electrode area and total amount of electroactive coverage on electrodes) were analyzed.

The electrode area of the Cu-deposited Au electrode (Au/Cu) was determined using the electrode reaction of potassium ferricyanide. The value of -0.925 mA was obtained for the reduction peak; thus, the electrode area was estimated to be 0.40 cm² by using equation (2). Before deposition, the geometric area of the Au electrode was 0.02 cm², so the electrode area was increased by 20-fold compared with the bare Au electrode.

To determine the total amount of immobilized enzyme on the electrode, we modified gold electrodes as described above and then washed off the His-tagged enzymes by immersing the electrode in the imidazole solution. The total amount of protein in the imidazole solution was measured. The protein concentration was determined according to the BSA calibration curve. PQQ-GDH-(His-tag) is immobilized to Cu by a His-tag attached to the N-terminal of the enzyme. It is expected that the catalytic reaction is facilitated because accessibility of the substrate to the active site of the enzyme is located opposite to the electrode surface (Figure 5b). Because the enzyme molecules had different orientations on the non-oriented electrode surface, the efficiency was lower than that of the highly oriented electrode. These results demonstrated in this study will allow high-performance bio-fuel cells to be designed and constructed, and lead to practical usage.

**Conclusion**

In the present study, we developed an anode to realize a highly efficient reaction by focusing on affinity binding between Cu and His-tag. In the study of electrodes for bio-fuel cell, the stability of the enzyme itself is a major challenge, but enzymes from hyperthermophiles can solve this problem because they have high thermostability and long-term stability. In the PQQ-GDH used in this study, PQQ is present...
inside the enzyme, and electron transfer is performed through the PQQ to provide a current density of 18.6 μA/cm² without mediators. We determined the amount of enzyme that existing on the surface of the electrode, and showed that a highly efficient electrode reaction was achieved by orientation of the enzymes. The His-Cu affinity bond allowed proper orientation of the active site, thereby permitting an effective electrode reaction. By employing His-PQQGDH from the hyperthermophilic archaean P. aerophilum with a high degree of orientation, it may become possible to develop a practical bioanode with long-term stability.

**Acknowledgment**

This work was supported by the Japan Society for the Promotion of Science (JSPS) KAKENHI (23510127) Grant-in-Aid for Scientific Research (C).

**References**


