Enzymatic Responses of Ginger Plants to *Pythium* Infection after SAR Induction

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**Abstract**

A variety of enzymatic responses of ginger plants to *Pythium* infection after induction of SAR (systemic acquired resistance) have been investigated. Results of pathogenicity test of *P. aphanidermatum* on a susceptible ginger cultivar showed that disease intensity increased with time up to 28 days but Polyphenol oxidase (PPO), Lipooxygenase (LOX) and Phenyl alanine ammonia lyase (PAL) activities increased up to 14 days following inoculation and then declined whereas Peroxidase (PO) activity reached their peaks on 21st day after inoculation and then decreased sharply. To induce SAR, rhizome seeds were soaked separately in salicylic acid (SA-5 mM) and *Acalypha* leaf extract (ALE – 10%) for 1 hour prior to sowing. Significant disease reduction was observed in both SA and ALE treated plants. SA and ALE treatment enhanced activities of all four defence related enzymes in ginger leaves but the rate of increase was higher in untreated inoculated and treated non-inoculated plants in relation to their respective controls. Treated inoculated plants exhibited maximum activity for all four enzymes. SA stimulated PO and PAL more than that of ALE. Results suggest that a correlation exists between reduction of disease intensity due to SAR induction and greater stimulation of specific enzymatic activities in ginger plants although not all four enzymes are equally responsive to a defence activator.

**Keywords:** Ginger; *Pythium aphanidermatum*; Salicylic acid; *Acalypha* leaf extract; Defence related enzyme; SAR

**Introduction**

Induced resistance is defined as an enhancement of the plant defensive capacity against a broad spectrum of pathogens and pests that is acquired after appropriate stimulation. The resulting elevated resistance due to an inducing agent upon infection by a pathogen is called ISR or SAR [1]. Induction of resistance in plants to a broad spectrum of microorganisms by different plant defence activators and involvement of defence related proteins and enzymes have been discussed earlier in detail [2-5]. Various biotic and abiotic inducers can also enhance the activities of defence related enzymes. Two different biotic inducers [*Pseudomonas fluorescens* and *Pseudomonas patida*] and three different abiotic inducers [copper sulphate, indole butyric acid and potassium chloride] were tested for their efficacy in inducing resistance in *Lupinus* plants against *Fusarium* wilt disease caused by *Fusarium oxysporum* f. sp. *Lupine*. A time course of defence-related enzymes showed substantial increases in enzyme activities in induced infected seedlings compared with untreated healthy plants or infected controls [6]. Induction of defense-related marker enzyme activity, namely, peroxidase, polyphenol oxidase, β-1,3 glucanase, chitinase, and phenolics was observed in banana (*Grand Naine* variety) plants when interacting with dead or live pathogen, *Fusarium oxysporum* f. sp. *cubense*, a causative agent of Panama disease [7]. These defence related enzymes are associated with the biosynthesis of lignin, phenolic compounds and phytoalexins, which are considered as important plant, defence components. Therefore, their role in induced resistance is significant. Peroxidase (PO) oxidizes phenolics to quinones and generates H$_2$O$_2$. The latter not only is antimicrobial in itself, but it also releases highly reactive free radicals and in that way further increases the rate of polymerization of phenolic compounds into lignin like compounds. These substances are deposited in cell walls and papillae and inhibited further growth and development of pathogen. Increased activity of polyphenol oxidase (PPO) also results in accumulation of higher concentrations of toxic products of oxidation and therefore greater degree of resistance to infection occurs. In contrast to peroxidase and polyphenol oxidase, lipooxygenases (LOX) use molecular oxygen to oxygenate unsaturated fatty acids such as linoleic acid, linolenic acid producing fatty acid hydroperoxide [8,9]. Lipooxygenases are primarily induced by pathogen attack and to a lesser extent by wounding and herbivore damage [9]. This enzyme has several important roles in defense, for example, LOX are required for the synthesis of jasmonates that has a major role in defense. Phenylalanine ammonia lyase (PAL), one of the key enzymes in the phenyl propanoid pathway, has a role in phytoalexin, phenolic compound and salicylic acid synthesis. Other defense related enzymes include pathogenesis related proteins (PRs) such as β-1,3-glucanase (PR2) and chitinase (PR1) which degrade fungal cell wall and cause cell lysis.

Recent studies on host pathogen interactions revealed the existence of SAR in non-vascular plants. Andersson et al. [10] reported SA-dependent defense pathway in *Physcomitrella patens* against *Erwinia carotovora*. Ponce de Leon et al. demonstrated that treatment of whole plant of *P. patens* with elicitors or cell-free culture filtrates of the bacterium *Erwinia carotovora* or inoculation with spores of the fungus *Botrytis cinerea* altered expression of the genes PR-1, CHS, PAL, and LOX, which are all up-regulated upon pathogen attack in vascular plants. Oliver et al. [11] showed that *P. patens* activated multiple and similar responses against *Pythium irregulare* and *Pythium debaryanum*, including the reinforcement of the cell wall, induction of the defense genes CHS, LOX and PAL, and accumulation of the signaling molecules jasmonic acid (JA) and its precursor 12-oxophytodienoic acid (OPDA). This study indicated that in *P. patens*, *Pythium* infection activates common responses to those previously characterized in flowering plants. Winter et al. [12] demonstrated moss species *Amblystegium serpens* could initiate SAR like reactions upon inoculation with *Pythium irregulare*. Results of their investigation,

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together with previous studies, suggest that SAR arose prior to the divergence of vascular and non-vascular plants.

Ginger occupies an important place in plantation crops and is grown in tropical and subtropical regions of the country for its aromatic rhizomes, which are used both as a spice and as a medicine. Rhizome rot caused by *Pythium aphanidermatum* is a serious disease of ginger, which causes considerable loss in yield every year. In the present investigation an attempt has been made to study the specific enzymatic responses of ginger plants to *Pythium aphanidermatum* after induction of resistance by two plant defence activators. The purpose of this study is to determine how far defence related enzymes such as PO, PPO, LOX and PAL activities are affected by SAR induction in ginger.

**Materials and Methods**

**Source of ginger and fungal culture**

Rhizome seeds (cultivar Suprabha) were collected from Amtala seed stores, South 24 Parganas, West Bengal. The culture of *Pythium aphanidermatum* (Edson) Fitz, a causal organism of rhizome rot or soft rot disease of ginger was supplied by the Indian Institute of Spices Research, Calicut, Kerala.

**Pathogenicity test**

Healthy rhizome seeds of ginger were disinfected and sown in earthenware pots (one rhizome seed/pot of 20 cm diameter) containing non-infected sandy soil (sand: soil: 1:3). The plants were kept under ordinary conditions of daylight, temperature (28-34°C) and humidity (77-85%) of the experimental garden of the Department of Botany. Water supply was maintained once a day except during rainy days. The top soil in pots was inoculated with *P. aphanidermatum* (in sand maize meal) after 28 days of sowing rhizome seeds following the method of Karmakar et al. [13]. Disease intensity was assessed usually 4 weeks after inoculation and 8 weeks after sowing rhizome seeds. Yellowing of leaves is one of the important symptoms of rhizome rot disease of ginger and hence yellowing index (Y.I./plant) was calculated as follows:

\[
\text{Y.I.} \text{/ Plant} = \frac{\text{Total number of leaves showing yellowing symptom}}{\text{Total number of test plants / treatment}}
\]

Apart from Y.I./Plant, percentage loss in fresh weight of rhizomes due to putrefaction caused by the pathogen was also estimated in relation to non-inoculated control.

**Method of application of plant defence activators**

To induce resistance in ginger plants rhizome seeds are soaked separately in salicylic acid (SA) and 10% leaf extract of *Acalypha indica* (ALE) for 1 hour prior to sowing. Control seeds were soaked in distilled water for a similar period. The leaf extract was prepared following the method of Doubraeva et al. [14]. The leaves were homogenized with distilled water (50 g fresh weight of leaves/100 ml) in an electric blender. The homogenate was strained through two layers of muslin and the filtrate centrifuged at 5,000 x g for 15 minutes at 4°C. Usually, 100 ml of supernatant (50%) were added to 500 ml distilled water (1:5), mixed thoroughly and used for treating rhizome seeds by soaking method.

**Extraction and assay of enzymes**

**PO:** Extraction and assay of PO from ginger leaves were carried out following the method of Sadashivam and Manickam [15] with modifications. PO was extracted from ginger leaves with 0.1 M phosphate buffer, pH 7.0 (2 ml g⁻¹ fresh tissue) by grinding the leaves in a prechilled mortar and pestle with sea sand. The extract was strained through muslin and centrifuged at 15,000 rpm for 15 min at 4°C. The supernatant was used as enzyme source.

For enzyme assay, 200 µl of enzyme extract was added to 3.5 ml of phosphate buffer (pH 6.5) and to this mixture 0.1 ml freshly prepared o-dianisidine solution (1 mg ml⁻¹ methanol) was added. The assay mixture was brought to 28-30°C and the reaction started after addition of 0.2 ml (0.2 M) H₂O₂ to the mixture. The initial OD at 436 nm was noted and then readings were taken at an interval of 30 sec up to 3 min. A water blank was also included in the assay. The enzyme activity was expressed as change in OD (optical density) at 436 nm per unit time per mg protein.

**PPO:** PPO activity was measured following the method of Sadashivam and Manickam [15] with modifications. For enzyme extraction, leaf tissues of ginger were homogenized in 50 mM Tris-HCl buffer, pH 7.2 (2 ml g⁻¹ tissue). The homogenate was strained through muslin, centrifuged at 15,000 rpm for 10 min at 4°C and the supernatant was used as enzyme extract.

For enzyme assay 0.6 ml catechol solution (0.01 M) was first added to 5 ml phosphate buffer (0.1 M, pH 6.5) and then 1 ml enzyme extract was supplemented to this mixture. The change in optical density was recorded at 490 nm for every 30 seconds up to 3 min. The enzyme activity was expressed as change in OD at 490 nm per unit time per mg protein.

**LOX:** For extraction and assay of LOX activity the method of Vick and Zimmermann [16] was adopted with modifications. LOX was extracted from ginger leaves using potassium phosphate buffer (0.05 M, pH 6.0). The homogenate was filtered and centrifuged at 15,000 rpm for 15 min at 4°C and the supernatant was used for LOX assay. LOX activity was measured spectrophotometrically using linoleic acid as substrate. Hundred microlitres of crude enzyme extract was added to 2.9 ml potassium phosphate buffer (0.05 M, pH 6.0). The reaction was initiated by 20 µl of linoleic acid (8 mM) substrate at room temperature (30°C). LOX activity was measured by conjugated diene absorption of the hydroperoxide at 234 nm. The enzyme activity was expressed as µmol conjugated diene produced per unit time per mg protein.

**PAL:** For extraction and assay of PAL, the method described by Smith et al. [17] was followed. Ginger leaf tissues were homogenized in 50 mM Tris-HCl buffer (pH 8.6) containing 10 mM ascorbic acid (3 ml g⁻¹ fresh weight) with a mortar and pestle. The homogenate was filtered, centrifuged at 15,000 r.p.m for 20 min at 4°C and the supernatant dialysed against 50 mM Tris-HCl (pH 8.6) for 16 h at 4°C.

The dialysate was assayed spectrophotometrically for PAL activity after addition of 1 ml of 30 mM L-phenylalanine (dissolved in appropriate volume of 50 mM Tris-HCl, pH 8.6) and 2 ml of dialysate (protein content 1 mg/ml). Absorbance was measured at 290 nm in Shimadzu UV-160A spectrophotometer after 2 h incubation at 30°C. A control was maintained for a similar period using 10 mM D-phenylalanine instead of L-isomer. The activity was expressed as the amount of cinnamic acid (µmol) produced per unit time per mg protein.

**Result and Discussion**

**Pathogenicity test**

Disease severity was assessed after 7, 14, 21 and 28 days of inoculation. Initially, symptoms developed on leaves as slight fading of green colour followed by yellowing of leaf tips (chlorosis) that...
extended downwards resulting in withering and drying of leaves. Rhizomes became soft and gradually the internal tissues rot completely (putrefaction), plants showed wilting symptoms and finally collapsed. It appears from the results that the intensity of disease increased with time up to 28 days following inoculation. About 34% loss in fresh weight of rhizomes was noted. Yellowing index/plant also increased with time up to 28 days. Significant difference in percentage weight loss was recorded between 14 and 21 days (P<0.01) and also between 21 and 28 days (P<0.05) of inoculation. Difference was not significant between 7 and 14 days in terms of weight loss. Statistical analysis of data shows a positive correlation (P<0.05) between intensity of disease and duration of incubation time up to 28 days following inoculation (Table 1 and Figure 1).

**Effect of incubation period on enzyme activity**

Defence related enzymes (PO, PPO, LOX and PAL) were extracted from leaves of ginger plants after 7, 14, 21 and 28 days of inoculation in all cases after assessment of disease intensity. The enzyme activities were assayed as described and results are given in Figure 2A-2D.

PO activity increased in leaves of untreated inoculated plants up to 21 days of inoculation and then declined sharply (Figure 2A). Whereas PPO activity increased and reached its peak on the 14th day of inoculation and then declined gradually in leaves of untreated inoculated plants (Figure 2B). Significant differences (P<0.01) in enzyme activities were noted between 7 and 14, 14 and 21 and also between 21 and 28 days of inoculation in case of both PO and PPO. The LOX activity also increased in leaves of untreated inoculated plants up to 14 days following inoculation and then declined (Figure 2C). Differences in enzyme activities were significant (P<0.01) between 14 and 21 and also between 21 and 28 days but not between 7 and 14 days. The PAL activity also increased up to 14 days and then declined in leaves of untreated inoculated plants. There is a significant (P<0.01) decrease in enzyme activity between 14 and 21 days but not between 21 and 28 days (Figure 2D). Activities of all four enzymes increased in leaves of untreated inoculated plants as follows: PO (4-fold), PPO (4.2-fold), LOX (3.2-fold), and PAL (5-fold).

Results revealed that incubation period has significant effect on disease development and enzyme activities. But there is no direct correlation between the rate of enzyme activity and intensity of disease.

**Effects of plant defence activators (SA and ALE) on disease development**

Effects of two plant defence activators (SA and ALE) were tested on disease development and enzyme (PO, PPO, LOX and PAL) activities in ginger. Healthy rhizome seeds were soaked separately in 5 mM SA and 10% ALE for 1 h prior to sowing. Plants were inoculated as usual after 28 days of sowing and disease severity was assessed after 14 and 28 days of inoculation. Results indicated that both SA and ALE reduced disease significantly (P<0.01). The difference between SA and ALE treatments in terms of percentage loss in fresh weight of rhizomes, even after 28 days of inoculation was not significant (Table 2).

**Effects of plant defence activators (SA and ALE) on enzyme (PO, PPO, LOX and PAL) activities**

After assessment of disease intensity on 14th and 28th day of inoculation, enzyme were extracted from leaves of untreated and treated non-inoculated and inoculated ginger plants and assayed. The results are given in Table 3.

It is evident from the results that both SA and ALE enhanced PO activity in leaves of non-inoculated and inoculated ginger plants but markedly in treated inoculated ones. Enzyme activity decreased on 28th day of inoculation in all cases except untreated non-inoculated plants. The difference in PO activity between SA and ALE-treated plants was significant (P<0.01) on 14th day but not significant on 28th day of inoculation. Like PO, PPO activity also increased in leaves of SA and ALE-treated non-inoculated and inoculated plants up to 14 days of inoculation. The rate of increase was significantly higher (P<0.05)

### Table 1: Effect of incubation period on the development of rhizome rot disease of ginger.

<table>
<thead>
<tr>
<th>Incubation time (days)*</th>
<th>Yellowing index/plant</th>
<th><strong>Average fresh wt (g) of rhizomes</strong></th>
<th>% loss in fresh weight of rhizome</th>
<th>Correlation coefficient</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Non inoculated</td>
<td>Inoculated</td>
<td>Non inoculated</td>
<td>Inoculated</td>
</tr>
<tr>
<td>7</td>
<td>0</td>
<td>0</td>
<td>39.40 ± 0.57</td>
<td>38.00 ± 1.08</td>
</tr>
<tr>
<td>14</td>
<td>0</td>
<td>1.00</td>
<td>44.20 ± 1.15</td>
<td>38.95 ± 1.61</td>
</tr>
<tr>
<td>21</td>
<td>0</td>
<td>1.25</td>
<td>50.00 ± 0.77</td>
<td>37.50 ± 1.65</td>
</tr>
<tr>
<td>28</td>
<td>0</td>
<td>2.66</td>
<td>55.00 ± 1.29</td>
<td>36.15 ± 3.23</td>
</tr>
</tbody>
</table>

*Days after inoculation
**Average of 4 rhizome seeds/treatment

### Table 2: Effect of incubation period on enzyme activity of ginger plants after SAR induction.

<table>
<thead>
<tr>
<th>Incubation time (days)</th>
<th>PO (fold)</th>
<th>PPO (fold)</th>
<th>LOX (fold)</th>
<th>PAL (fold)</th>
</tr>
</thead>
<tbody>
<tr>
<td>7</td>
<td>3</td>
<td>4.2</td>
<td>3.2</td>
<td>5</td>
</tr>
<tr>
<td>14</td>
<td>4</td>
<td>4.2</td>
<td>3.2</td>
<td>5</td>
</tr>
<tr>
<td>21</td>
<td>5</td>
<td>4.2</td>
<td>3.2</td>
<td>5</td>
</tr>
<tr>
<td>28</td>
<td>6</td>
<td>4.2</td>
<td>3.2</td>
<td>5</td>
</tr>
</tbody>
</table>

*Correlation coefficient (P<0.01)
in SA-treated inoculated plants. The difference in PPO activity was not significant between SA- and ALE-treated non-inoculated plants. LOX activity increased in both SA- and ALE-treated plants up to 14 days of inoculation but markedly in SA-treated plants. PAL activity also increased in leaves of untreated inoculated as well as treated (SA or ALE) non-inoculated and inoculated plants. Activity was always higher in SA-treated than in ALE-treated plants (both non-inoculated and inoculated). Results showed that enzyme activities increased in ginger plants as a result of both treatment and inoculation. Highest activity was noted in treated inoculated plants. The rate of increase in treated inoculated plants were significantly higher in comparison to untreated inoculated plants (P<0.05) (Figure 3A-D).

In the present investigation results of pathogenicity test of *P. aphanidermatum* on a susceptible ginger cultivar showed that disease intensity increased with time up to 28 days but PPO, LOX and PAL activities increased up to 14 days following inoculation and then declined whereas PO activity reached their peaks on 21st day after inoculation and then decreased sharply. Plant defence activators SA and ALE not only reduced rhizome rot disease significantly in ginger, but they also...
### Table 3: Effect of plant defence activators (SA and ALE) on disease development and enzyme activities in ginger leaves.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>PO activity (µOD₄150 nm mg⁻¹ protein min⁻¹)</th>
<th>PPO activity (µOD₄90 nm mg⁻¹ protein min⁻¹)</th>
<th>LOX activity (µ mol conjugated diene mg⁻¹ protein min⁻¹)</th>
<th>PAL activity (µ mol cinnamic acid mg⁻¹ protein min⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>14 days*</td>
<td>28 days</td>
<td>14 days*</td>
<td>28 days</td>
</tr>
<tr>
<td>Untreated</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-inoculated</td>
<td>0.38 ± 0.02</td>
<td>0.42 ± 0.03</td>
<td>0.07 ± 0.006</td>
<td>0.05 ± 0.007</td>
</tr>
<tr>
<td>Inoculated</td>
<td>1.35 ± 0.03</td>
<td>1.12 ± 0.06</td>
<td>0.31 ± 0.008</td>
<td>0.22 ± 0.002</td>
</tr>
<tr>
<td>Treated with SA (5 mM)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-inoculated</td>
<td>0.82 ± 0.06</td>
<td>0.80 ± 0.04</td>
<td>0.15 ± 0.005</td>
<td>0.13 ± 0.005</td>
</tr>
<tr>
<td>Inoculated</td>
<td>2.05 ± 0.03</td>
<td>1.77 ± 0.01</td>
<td>0.85 ± 0.14</td>
<td>0.56 ± 0.003</td>
</tr>
<tr>
<td>Treated with ALE (10%)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Non-inoculated</td>
<td>0.72 ± 0.05</td>
<td>0.67 ± 0.06</td>
<td>0.14 ± 0.011</td>
<td>0.12 ± 0.005</td>
</tr>
<tr>
<td>Inoculated</td>
<td>1.86 ± 0.03*</td>
<td>1.65 ± 0.11</td>
<td>0.67 ± 0.015</td>
<td>0.42 ± 0.02</td>
</tr>
<tr>
<td>CD at 5%</td>
<td>0.124</td>
<td>0.174</td>
<td>0.156</td>
<td>0.035</td>
</tr>
<tr>
<td>CD at 1%</td>
<td>0.174</td>
<td>0.259</td>
<td>0.223</td>
<td>0.048</td>
</tr>
</tbody>
</table>

SA: Salicylic acid; ALE: Acalypha leaf extract
*after inoculation
**Average of 4 replicate rhizome seeds/treatment
***3 replicates/treatment

**Figure 3:** Activities of defence related enzymes in leaves of untreated inoculated and treated inoculated ginger plants.
enhanced activities of all four defence related enzymes in ginger leaves although the rate of increase was higher in untreated inoculated and treated non-inoculated plants in relation to their respective controls. The test enzymes preferentially responded to different kinds of defence activators. Systemic protection against P. aphanidermatum was induced in ginger by chemicals or specific herbal extracts. Among 12 plant defence activators tested, jasmonic acid (5 mM) and 10% leaf extract of Acalypha indica (ALE) reduced disease significantly with concomitant increase of defence related proteins [18]. In this study, treated inoculated plants exhibited maximum activity for all four enzymes. SA stimulated all the four defence related enzymes than that of ALE. It was reported earlier that nonpathogenic rhizobacteria and a fungal pathogen P. aphanidermatum stimulated PO, PPO and PAL activities in cucumber roots [19]. Schneider [20] also recorded high enzyme (PO, PPO, PAL) activities in cucumber and tobacco after 2-3 days of treatment with various biotic and abiotic inducers including salicylic acid. Aqueous leaf extract of neem provided protection to barley against leaf stripe pathogen Drechslera graminis and enhanced PAL and TAL (Tyrosine ammonia lyase) activities along with rapid accumulation of fungitoxic phenolic compounds [21]. Induction of some defence related enzymes and phenolics in roots and shoots of two different genotypes of chickpea cultivars which were susceptible (L550) and resistant (ICCV10) to wilt disease treated with salicylic acid, spermine (Spm), SA+Spm and Fusarium oxysporum f. sp. ciceri was investigated. Higher levels of polyphenol oxidase (PPO), phenylalanine ammonia-lyase (PAL), β-1,3-glucanase (PR-2) and phenolics were observed in roots and shoots of resistant cultivar than that of susceptible cultivar on treatment with elicitors and pathogen [22]. Involvement of LOX in induced resistance was also reported by a number of workers. Bohland et al. [23] observed that LOX activity increased in wheat leaves when treated with either glycopeptide elicitor from germtube of P. graminis f. sp. tritici or methyl jasmonate. Similarly, arachidonic acid (10 µM) also activated LOX activity in tuber tissues and enhanced resistance in potato against late blight [24]. Mitchell and Walters [25] reported that systemic resistance of barley to leaf stripe disease. Physiol Mol Plant Pathol 61: 3-13.

The utilization of plants owns defence mechanism is the area of major interests in the management of pests and diseases. Result of present investigation clearly indicated that defence activators SA and ALE reduced disease significantly and also induced PO, PPO, LOX and PAL activities in ginger. Although enzyme activity increased in all cases, the rate of activity varied with the nature of enzymes and defence activators. In conclusion, it could be suggested that activation of defence related enzymes are quite likely to govern some of the basic mechanisms of biochemical resistance induced by SA and ALE in ginger plants against rhizome rot disease.

Acknowledgement

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