

Evaluation of Invasive Squamous Cell Carcinoma, Seborrheic Keratosis and Verruca Vulgaris in Superficial Shave Biopsies Using p16, p53, p63, and PHLDA1 Immunohistochemistry

Ryanne A Brown¹ and Jinah Kim^{1,2*}

¹Department of Pathology, Stanford Medicine, Stanford, CA, USA

²Department of Dermatology, Stanford Medicine, Stanford, CA, USA

Corresponding author: Jinah Kim, Department of Pathology and Dermatology, Stanford University School of Medicine, 300 Pasteur Drive, L235, Stanford, CA 94305, USA, Tel: 650-736-1068; Fax: 650-725-7409; E-mail: jinahkim@stanford.edu

Received date: February 23, 2016; **Accepted date:** April 19, 2016; **Published date:** April 25, 2016

Copyright: © 2016 Brown RA, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Occasionally, the distinction between malignant and benign is challenging in superficial shave biopsies of squamoproliferative lesions. This phenomenon is compounded by the increasing prevalence of conditions encountered that weaken the immune system, such as chemotherapy, immune deficiency diseases, and anti-rejection medications for organ transplantation that have all been shown to increase the risk of the development of squamous cell carcinoma. We collected 30 cases (10 invasive SCC, 10 SK and 10 VV) and performed immunohistochemical staining using a panel approach composed of markers important for proliferation and the cell cycle, including Ki-67, p16, p53, and PHLDA1. The results demonstrate that the invasive SCC group was enriched for high PHLDA1 (80% with PHLDA1 score=3, 100% with PHLDA1 score \geq 2) and high p53 (50% of SCC with p53 score \geq 2 vs. 60% of SK and 90% of VV with p53 score=1). The SK group was enriched for low p16 (100% with p16 score \leq 1) and high p63 scores (100% with p63 score=3). A panel approach may be utilized to help in the distinction between benign keratoses and carcinoma and may be increasingly critical to promote quality care.

Keywords: Squamous cell carcinoma; Verruca vulgaris; Seborrheic keratosis; Benign keratosis; p16; p53; PHLDA1

Introduction

Excision specimens of squamous cell carcinoma (SCC), verruca vulgaris (VV), and seborrheic keratosis (SK) demonstrate classic histopathologic features especially along the base of the lesion, precluding the necessity for ancillary diagnostic techniques such as immunohistochemistry (IHC). However, pathologists are often presented with superficial shave biopsies, some lacking basal epithelium, rendering distinction of these squamoproliferative lesions with overlapping histologic features challenging. An established immunohistochemical panel does not yet exist for this differential diagnosis. Therefore, our goal was to evaluate the diagnostic utility of immunohistochemical stains p16, p53, p63, PHLDA1, and the proliferation marker Ki-67 in differentiating SCC, VV, and SK. Although p16 has been used as a diagnostic adjunct for diagnosing human papillomavirus (HPV)-related intraepithelial neoplasia of the genital and oropharyngeal mucosa, its applications in diagnosing cutaneous SCC have not been extensively studied [1,2]. Mutations in the tumor suppressor p53 are noted to occur early and often in skin carcinogenesis [3,4]. p63 mutations have not frequently been implicated in cancers, but one of its isoforms is often upregulated in

neoplasia and is expressed in SCC [3]. PHLDA1 is a hair follicle bulge marker that has demonstrated utility in differentiating basal cell carcinoma (BCC) from trichoepithelioma and trichoblastomas [5-10]. Given the increased incidence of squamoproliferative lesions in patients treated with BRAF inhibitors [11-14], it is increasingly important to distinguish benign keratoses (SK, VV) from SCC. We evaluate the utility of IHC to assist in the distinction of these lesions and provide pathologists with a tool in approaching this potentially difficult diagnostic differential in shave biopsy specimens.

Materials and Methods

We searched the Department of Pathology database for shave biopsy cases received for evaluation between 2012 and 2014. 30 cases (10 invasive SCC, 10 SK and 10 VV) were randomly selected. Immunohistochemical staining was performed on formalin-fixed, paraffin-embedded tissue sections sliced at 5 μ m using Ki-67 (Dako monoclonal mouse anti-human antibody, 1:200 dilution, Leica instrument), p16 (Ventana E6H4, 1:2 dilution, Leica instrument), p53 (Ventana DO-7, 1:400 dilution, Ventana instrument), p63 (Biocare Medical BC4A4, 1:100 dilution, Leica instrument), and PHLDA1 (Santa Cruz Biotech RN-6E2, 1:400 dilution, Leica instrument) IHC. Positive controls were performed for each antibody with demonstration of the appropriate staining pattern.

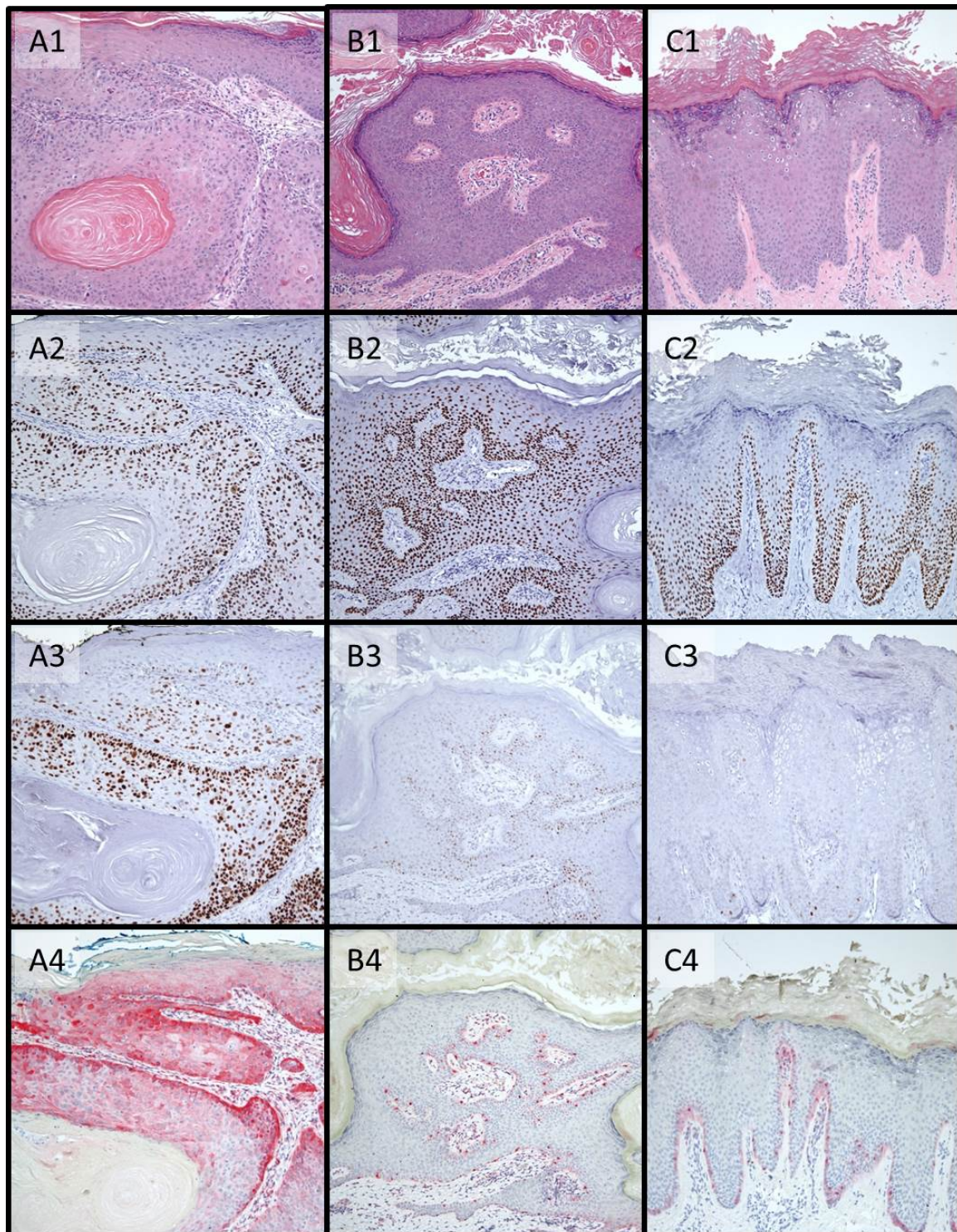


Figure 1: Histology and immunohistochemistry of squamoproliferative lesions. Staining of invasive SCC Case 7 (A), SK Case 15 (B), and VV Case 27 (C) with H&E (1), p63 (2), p53 (3), and PHLDA1 (4) immunohistochemistry. Original magnification 100X.

Each stain was assessed for percent lesional expression and, when applicable, distribution of staining. Stains were scored without knowledge of the original diagnosis as 1, 2, or 3 (p53, p63, PHLDA1,

Ki-67) or 0, 1, or 2 (p16) based on the criteria detailed in Table 1 (RB and JK). The authors then unblinded themselves to and confirmed agreement with the original histologic diagnoses. Sensitivity,

specificity, positive predictive value, and negative predictive value for each diagnosis were calculated with various combinations of staining results.

| Stain | p16 | p53 | p63* | PHLDA1 | Ki-67 |
|---------|----------------|----------------|---------------------|-----------------|--|
| Score=0 | <1% of cells | N/A | N/A | N/A | N/A |
| 1 | 1-30% of cells | <5% of cells | <50% of thickness | <10% of cells | <5% of cells or staining restricted to basal layer |
| 2 | >30% of cells | 5-30% of cells | 50-90% of thickness | 10-40% of cells | 5-30% of cells or extending above basal layer but less than 30% of cells |
| 3 | N/A | >30% of cells | >90% thickness | >40% of cells | >30% of cells |

*As measured beneath the granular layer. N/A: Not Applicable.

Table 1: Immunohistochemical scoring criteria.

Results

The results of p16, p53, p63, PHLDA1, and Ki-67 staining are shown in Tables 2-4 with histology and IHC of representative SCC, VV, and SK cases shown in Figure 1.

The invasive SCC group was enriched for high PHLDA1 (80% with PHLDA1 score=3, 100% with PHLDA1 score ≥ 2).

| Stain | p16 | p53 | p63 | PHLDA1 | Ki-67 |
|--------------|---|---------------------------|---------------------------|--|---------------------------|
| Intensity | Weak to strong | Strong | Strong | Weak to strong | Weak to strong |
| Distribution | Nuclear and cytoplasmic lesional staining | Nuclear lesional staining | Nuclear lesional staining | Cytoplasmic lesional staining with background stromal staining | Nuclear lesional staining |

Table 2: Overall immunohistochemical stain expression patterns.

| Invasive SCC | p16 | p53 | p63 | PHLDA1 | Ki-67 |
|--------------|-----|-----|-----|--------|-------|
| Case 1 | 1 | 3 | 2 | 3 | 2 |
| Case 2 | 1 | 1 | 2 | 3 | 1 |
| Case 3 | 2 | 1 | 2 | 3 | 1 |
| Case 4 | 1 | 1 | 2 | 2 | 1 |
| Case 5 | 2 | 1 | 1 | 3 | 1 |
| Case 6 | 2 | 2 | 1 | 3 | 1 |
| Case 7 | 1 | 3 | 2 | 3 | 2 |
| Case 8 | 2 | 3 | 3 | 3 | 2 |
| Case 9 | 2 | 3 | 3 | 3 | 3 |
| Case 10 | 1 | 1 | 1 | 2 | 1 |
| SK | p16 | p53 | p63 | PHLDA1 | Ki-67 |
| Case 11 | 1 | 2 | 3 | 2 | 1 |
| Case 12 | 0 | 1 | 3 | 2 | 1 |
| Case 13 | 1 | 2 | 3 | 2 | 2 |
| Case 14 | 1 | 2 | 3 | 1 | 1 |
| Case 15 | 1 | 1 | 3 | 1 | 1 |
| Case 16 | 0 | 1 | 3 | 1 | 1 |

| Case 17 | 0 | 2 | 3 | 1 | 1 |
|---------|-----|-----|-----|--------|-------|
| Case 18 | 0 | 1 | 3 | 1 | 1 |
| Case 19 | 1 | 1 | 3 | 1 | 1 |
| Case 20 | 0 | 1 | 3 | 2 | 1 |
| VV | p16 | p53 | p63 | PHLDA1 | Ki-67 |
| Case 21 | 0 | 2 | 3 | 1 | 1 |
| Case 22 | 0 | 1 | 2 | 1 | 1 |
| Case 23 | 2 | 1 | 1 | 1 | 1 |
| Case 24 | 1 | 1 | 2 | 1 | 1 |
| Case 25 | 0 | 1 | 3 | 1 | 1 |
| Case 26 | 0 | 1 | 3 | 1 | 2 |
| Case 27 | 1 | 1 | 2 | 1 | 1 |
| Case 28 | 0 | 1 | 2 | 1 | 1 |
| Case 29 | 1 | 1 | 2 | 2 | 1 |
| Case 30 | 1 | 1 | 3 | 1 | 1 |

SSC: Squamous Cell Carcinoma; SK: Seborrheic Keratosis; VV: Verruca Vulgaris

Table 3: Immunohistochemical scores by lesion type.

Compared to VV and SK, the SCC group also showed high p53 score=1). The SK group was enriched for low p16 (100% with p16 score (50% of SCC with p53 score ≥ 2 vs. 60% of SK and 90% of VV with p53 ≤ 1) and high p63 scores (100% with p63 score=3).

| Invasive SCC | p16 | p53 | p63 | PHLDA1 | Ki-67 |
|---------------------|------------|------------|------------|---------------|--------------|
| Score=0 | 0/10 | N/A | N/A | N/A | N/A |
| 1 | 5/10 | 5/10 | 3/10 | 0/10 | 6/10 |
| 2 | 5/10 | 1/10 | 5/10 | 2/10 | 3/10 |
| 3 | N/A | 4/10 | 2/10 | 8/10 | 1/10 |
| SK | p16 | p53 | p63 | PHLDA1 | Ki-67 |
| Score=0 | 5/10 | N/A | N/A | N/A | N/A |
| 1 | 5/10 | 6/10 | 0/10 | 6/10 | 9/10 |
| 2 | 0/10 | 4/10 | 0/10 | 4/10 | 1/10 |
| 3 | N/A | 0/10 | 10/10 | 0/10 | 0/10 |
| VV | p16 | p53 | p63 | PHLDA1 | Ki-67 |
| Score=0 | 5/10 | N/A | N/A | N/A | N/A |
| 1 | 4/10 | 9/10 | 1/10 | 9/10 | 9/10 |
| 2 | 1/10 | 1/10 | 5/10 | 1/10 | 1/10 |
| 3 | N/A | 0/10 | 4/10 | 0/10 | 0/10 |

SCC: Squamous Cell Carcinoma; SK: Seborrheic Keratosis; VV: Verruca Vulgaris; N/A: Not Applicable.

Table 4: Immunohistochemical score frequency by lesion type.

Sensitivity, specificity, positive predictive value, and negative predictive value for selected staining results are displayed in Table 5.

| Lesions included in analysis | SK, VV, Invasive SCC | | | SK, Invasive SCC | | | SK, VV | | |
|--------------------------------------|-----------------------------|----------|-------|-------------------------|----------|-------|--------------------|----------|-------|
| | p16<2 and p63=3 | p63=3 | p16<2 | p16<2 and p63=3 | p63=3 | p16<2 | p16<2 and p63=3 | p63=3 | p16<2 |
| Criteria for diagnosing SK | p16<2 and p63=3 | p63=3 | p16<2 | p16<2 and p63=3 | p63=3 | p16<2 | p16<2 and p63=3 | p63=3 | p16<2 |
| Sensitivity | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Specificity | 0.8 | 0.7 | 0.3 | 1 | 0.8 | 0.5 | 0.6 | 0.6 | 0.1 |
| PPV | 0.71 | 0.63 | 0.42 | 1 | 0.83 | 0.67 | 0.71 | 0.71 | 0.53 |
| NPV | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Lesions included in analysis | SK, VV, Invasive SCC | | | SK, Invasive SCC | | | Invasive SCC, VV | | |
| Criteria for diagnosing Invasive SCC | PHLDA1=3 and p53=3 | PHLDA1=3 | p53=3 | PHLDA1=3 and p53=3 | PHLDA1=3 | p53=3 | PHLDA1=3 and p53=3 | PHLDA1=3 | p53=3 |
| Sensitivity | 0.4 | 0.8 | 0.4 | 0.67 | 0.8 | 0.4 | 0.4 | 0.8 | 0.4 |
| Specificity | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| PPV | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| NPV | 0.77 | 0.91 | 0.77 | 0.83 | 0.83 | 0.63 | 0.63 | 0.83 | 0.63 |
| Lesions included in analysis | SK, VV, Invasive SCC | | | SK, VV | | | Invasive SCC, VV | | |
| Criteria for diagnosing VV | PHLDA1=1 and p53=1 | PHLDA1=1 | p53=1 | PHLDA1=1 and p53=1 | PHLDA1=1 | p53=1 | PHLDA1=1 and p53=1 | PHLDA1=1 | p53=1 |

| | | | | | | | | | |
|---|------|------|------|------|-----|-----|------|------|------|
| Sensitivity | 0.8 | 0.9 | 0.9 | 0.8 | 0.9 | 0.9 | 0.8 | 0.9 | 0.9 |
| Specificity | 0.8 | 0.7 | 0.45 | 0.6 | 0.4 | 0.4 | 1 | 1 | 0.5 |
| PPV | 0.67 | 0.6 | 0.45 | 0.67 | 0.6 | 0.6 | 1 | 1 | 0.64 |
| NPV | 0.89 | 0.93 | 0.9 | 0.75 | 0.8 | 0.8 | 0.83 | 0.91 | 0.83 |
| SK: Seborrheic Keratosis; VV: Verruca Vulgaris; SCC: Squamous Cell Carcinoma; PPV: Positive Predictive Value; NPV: Negative Predictive Value. | | | | | | | | | |

Table 5: Diagnostic statistics by stain criteria and differential diagnosis.

Discussion

p53 is a tumor suppressor that promotes cell cycle exit, senescence or apoptosis in response to DNA damage [3,15]. Soini and colleagues found an association between increased p53 expression and Ki-67 expression in benign skin lesions, including seborrheic keratosis [16]. They also noted clustering of p53 positive keratinocytes in areas of damage and inflammation. [16] Ko et al. found p53 expression in inverted follicular keratosis as well as SK [17]. Hussein et al. noted significantly increased p53 expression in invasive and in situ SCC compared to normal skin [18]. p53 expression in SK was also increased but to a lesser degree [18]. Another study found that p53 expression was highest in SK with less expression in malignant and pre-malignant squamous epithelial lesions, including SCC [19]. Bito et al. found the incidence of p53 expression to be greater in SCCs arising in sun-damaged skin [20], and p53 expression has frequently been noted in actinic keratosis (AK). A study by Gouvêa found strong p53 expression in oral SCC arising in patients with a history of proliferative verrucous leukoplakia [21]. Other studies have replicated the finding of more frequent p53 expression in SCC as compared to SK, with some evidence for increasing expression of p53 with higher grade histology. [22,23] Our results further support the finding of enriched p53 expression in invasive squamous cell carcinoma compared to SK and VV.

p63, a p53 homologue, is normally expressed in the nuclei of the basal epidermis, cells of the germinative hair matrix, and hair follicle external root sheath, and plays a role in regulating keratinocyte-specific gene expression [24-27]. Chang et al. were able to differentiate pagetoid SCC in situ from primary extramammary Paget's disease by the strong p63 positivity present in the former [28]. Takeuchi et al. demonstrated strong p63 expression in both SK and poorly differentiated SCC [29]. Other studies have noted similar findings of p63 expression in SK and SCC [30]. A trend towards increased p63 expression in less differentiated cells of invasive SCC has been noted [24]. Increased p63 expression in SK has been supported by microarray analysis [31]. Although we demonstrated p63 expression in VV and invasive SCC, only SK showed diffuse and strong positivity for p63.

PHLDA1 is a marker of matrical differentiation with demonstrated utility in identifying a subset of tumors of the hair follicle [5-10]. We found that a high level of PHLDA1 expression was moderately sensitive and highly specific for invasive SCC compared to SK and VV.

p16 expression has been described as a diagnostic tool for differentiating SCC in situ from actinic keratosis and benign squamous cutaneous lesions [32]. Hodges et al. found increasing p16 expression in the progression from AK to SCC in situ to invasive SCC [33]. Beyond its association with HPV-related oropharyngeal SCC, p16 expression in other cutaneous head and neck SCC with lymph node

metastasis can be frequent [34]. Bai and colleagues noted that a small number of vulvar and non-vulvar SKs demonstrated p16 expression with poor correlation with HPV-DNA status [35]. A study by Nakamura et al. found a subset of SK with p16 staining in all lesional cells [36]. Our findings demonstrated low p16 expression in all SK and most VV. Recently, diffuse cyclin D1 and p16 expression was demonstrated more frequently in SCCIS and SCC than in AK. Although the invasive SCC group was enriched for high p16 expression, this marker did not improve diagnostic yield above that offered by PHLDA1 and p53. Several studies have documented higher Ki-67 expression in SCC compared to non-malignant lesions including inflammatory dermatoses and SK [19,37]. Although increased Ki-67 staining was noted in a subset of invasive SCC, this marker failed to add additional diagnostic information in our cohort of cases [38,39].

This limited collection of cases provides support for the utilization of diffuse and strong p63 staining as a sensitive marker for SK with potential use of low to no p16 staining for its specificity for SK when invasive SCC is the primary differential diagnostic consideration. The combination of high PHLDA1 and p53 staining appeared highly specific for invasive SCC compared to SK and VV. Low PHLDA1 and p53 staining demonstrated moderate sensitivity for VV in this small collection of cases, but a specific marker for VV was not apparent from the markers tested [40]. Although our sample size is limited and does not include in situ SCC cases, our findings provide some support for utilization of IHC when a superficial shave biopsy specimen prompts the differential diagnosis of invasive SCC, SK, and VV. Additional studies with more cases are merited to validate these findings.

References

- Santos M, Montagut C, Mellado B, García A, Ramón y Cajal S, et al. (2004) Immunohistochemical staining for p16 and p53 in premalignant and malignant epithelial lesions of the vulva. *Int J Gynecol Pathol* 23: 206-214.
- Chaux A, Pfannl R, Rodríguez IM, Barreto JE, Velazquez EF, et al. (2011) Distinctive immunohistochemical profile of penile intraepithelial lesions: A study of 74 cases. *Am J Surg Pathol* 35: 553-562.
- Missero C, Antonini D (2014) Crosstalk among p53 family members in cutaneous carcinoma. *Exp Dermatol* 23: 143-146.
- Serdar ZA, Eren PA, Canbakan M, Turan K, Tellioglu G, et al. (2010) Dermatologic findings in renal transplant recipients: Possible effects of immunosuppression regimen and p53 mutations. *Transplant Proc* 42: 2538-2541.
- Sellheyer K, Krahl D (2011) Phlda1 (tdag51) is a follicular stem cell marker and differentiates between morphoeic basal cell carcinoma and desmoplastic trichoepitheliom. *British Journal of Dermatology* 164: 141-147.
- Battistella M, Carlson JA, Osio A, Langbein L, Cribier B (2014) Skin tumors with matrical differentiation: lessons from hair keratins, beta-catenin and PHLDA-1 expression. *J Cutan Pathol* 41: 427-436.

7. Battistella M, Peltre B, Cribier B (2014) Phlda1, a follicular stem cell marker, differentiates clear-cell/granular-cell trichoblastoma and clear-cell/granular cell basal cell carcinoma: A case-control study, with first description of granular-cell trichoblastoma. *Am J Dermatopathol* 36: 643-650.
8. Sellheyer K, Nelson P (2011) Follicular stem cell marker phlda1 (tdag51) is superior to cytokeratin-20 in differentiating between trichoepithelioma and basal cell carcinoma in small biopsy specimens. *Journal of cutaneous pathology* 38: 542-550.
9. Battistella M, Carlson JA, Osio A, Langbein L, Cribier B (2014) Skin tumors with matrical differentiation: lessons from hair keratins, beta-catenin and PHLDA-1 expression. *J Cutan Pathol* 41: 427-436.
10. Ohshima M, Terunuma A, Tock CL, Radonovich MF, Pise-Masison CA, et al. (2006) Characterization and isolation of stem cell-enriched human hair follicle bulge cells. *J Clin Invest* 116: 249-260.
11. Huang V, Hepper D, Anadkat M, Cornelius L (2012) Cutaneous toxic effects associated with vemurafenib and inhibition of the BRAF pathway. *Arch Dermatol* 148: 628-633.
12. Mandalà M, Massi D, De Giorgi V (2013) Cutaneous toxicities of BRAF inhibitors: clinical and pathological challenges and call to action. *Crit Rev Oncol Hematol* 88: 318-337.
13. Belum VR, Fischer A, Choi JN, Lacouture ME (2013) Dermatological adverse events from BRAF inhibitors: a growing problem. *Curr Oncol Rep* 15: 249-259.
14. Curry JL, Torres-Cabala CA, Kim KB, Tetzlaff MT, Duvic M, et al. (2014) Dermatologic toxicities to targeted cancer therapy: Shared clinical and histologic adverse skin reactions. *Int J Dermatol* 53: 376-384.
15. Lane D, Levine A (2010) p53 Research: the past thirty years and the next thirty years. *Cold Spring Harb Perspect Biol* 2: a000893.
16. Soini Y, Kamel D, Pääkkö P, Lehto VP, Oikarinen A, et al. (1994) Aberrant accumulation of p53 associates with Ki67 and mitotic count in benign skin lesions. *Br J Dermatol* 131: 514-520.
17. Ko CJ, Shintaku P, Binder SW (2005) Comparison of benign keratoses using p53, bcl-1, and bcl-2. *J Cutan Pathol* 32: 356-359.
18. Hussein MR, Al-Badaiwy ZH, Guirguis MN (2014) Analysis of p53 and bcl-2 protein expression in the non-tumorigenic, pretumorigenic, and tumorigenic keratinocytic hyperproliferative lesions. *Journal of cutaneous pathology* 31: 643-651.
19. Onodera H, Nakamura S, Sugai T (1996) Cell proliferation and p53 protein expressions in cutaneous epithelial neoplasms. *Am J Dermatopathol* 18: 580-588.
20. Bito T, Ueda M, Ahmed NU, Nagano T, Ichihashi M (1995) Cyclin D and retinoblastoma gene product expression in actinic keratosis and cutaneous squamous cell carcinoma in relation to p53 expression. *J Cutan Pathol* 22: 427-434.
21. Gouvêa AF, Vargas PA, Coletta RD, Jorge J, Lopes MA, et al. (2010) Clinicopathological features and immunohistochemical expression of p53, ki-67, mcm-2 and mcm-5 in proliferative verrucous leukoplakia. *Journal of Oral Pathology & Medicine* 239: 447-452.
22. Urano Y, Oura H, Sakaki A, Nagae H, Matsumoto K, et al. (1992) Immunohistological analysis of P53 expression in human skin tumors. *J Dermatol Sci* 4: 69-75.
23. Chen H, Takahara M, Xie L, Takeuchi S, Tu Y, et al. (2013) Levels of the EMT-related protein Snail/Slug are not correlated with p53/p63 in cutaneous squamous cell carcinoma. *J Cutan Pathol* 40: 651-656.
24. Reis-Filho JS, Torio B, Albergaria A, Schmitt FC (2002) p63 expression in normal skin and usual cutaneous carcinomas. *J Cutan Pathol* 29: 517-523.
25. Yang A, Schweitzer R, Sun D, Kaghad M, Walker N, et al. (1999) p63 is essential for regenerative proliferation in limb, craniofacial and epithelial development. *Nature* 398: 714-718.
26. Mills AA, Zheng B, Wang XJ, Vogel H, Roop DR, et al. (1999) p63 is a p53 homologue required for limb and epidermal morphogenesis. *Nature* 398: 708-713.
27. Della Gatta G, Bansal M, Ambesi-Impiombato A, Antonini D, Missero C, et al. (2008) Direct targets of the trp63 transcription factor revealed by a combination of gene expression profiling and reverse engineering. *Genome Res* 18: 939-948.
28. Chang J, Prieto VG, Sanguenza M, Plaza JA (2014) Diagnostic utility of p63 expression in the differential diagnosis of pagetoid squamous cell carcinoma in situ and extramammary paget disease: A histopathologic study of 70 cases. *Am J Dermatopathol* 36: 49-53.
29. Takeuchi Y, Tamura A, Kamiya M, Fukuda T, Ishikawa O (2005) Immunohistochemical analyses of p63 expression in cutaneous tumours. *Br J Dermatol* 153: 1230-1232.
30. Abbas O, Richards JE, Yaar R, Mahalingam M (2011) Stem cell markers (cytokeratin 15, cytokeratin 19 and p63) in in situ and invasive cutaneous epithelial lesions. *Mod Pathol* 24: 90-97.
31. Seo EY, Lee DH, Lee Y, Cho KH, Eun HC, et al. (2012) Microarray analysis reveals increased expression of p63 in seborrheic keratosis. *Br J Dermatol* 166: 337-342.
32. Salama ME, Mahmood MN, Qureshi HS, Ma C, Zarbo RJ, et al. (2003) p16INK4a expression in actinic keratosis and Bowen's disease. *Br J Dermatol* 149: 1006-1012.
33. Hodges A, Smoller BR (2002) Immunohistochemical comparison of p16 expression in actinic keratoses and squamous cell carcinomas of the skin. *Mod Pathol* 15: 1121-1125.
34. Beadle BM, William WN Jr, McLemore MS, Sturgis EM, Williams MD (2013) P16 expression in cutaneous squamous carcinomas with neck metastases: A potential pitfall in identifying unknown primaries of the head and neck. *Head & Neck* 35: 1527-1533.
35. Bai H, Cviko A, Granter S, Yuan L, Betensky RA, et al. (2003) Immunophenotypic and viral (human papillomavirus) correlates of vulvar seborrheic keratosis. *Hum Pathol* 34: 559-564.
36. Nakamura S, Nishioka K (2003) Enhanced expression of p16 in seborrheic keratosis; a lesion of accumulated senescent epidermal cells in G1 arrest. *Br J Dermatol* 149: 560-565.
37. Kawahira K (1999) Immunohistochemical staining of proliferating cell nuclear antigen (PCNA) in malignant and nonmalignant skin diseases. *Arch Dermatol Res* 291: 413-418.
38. Fabbrocini G, Russo N, Pagliuca MC, Delfino M, Staibano S, et al. (2000) P53, cyclin-d1, pcna, agnor expression in squamous cell cancer of the lip: A multicenter study. *Photodermatol Photoimmunol Photomed* 16: 172-177.
39. Neto PD, Alchorne M, Michalany N, Abreu M, Borra R (2013) Reduced P53 Staining in Actinic Keratosis is Associated with Squamous Cell Carcinoma: A Preliminary Study. *Indian J Dermatol* 58: 325.
40. Brasanac D, Stojkovic-Filipovic J, Botic M, Tomanovic N, Manojlovic-Gacic E (2015) Expression of G1/S-cyclins and cyclin-dependent kinase inhibitors in actinic keratosis and squamous cell carcinoma. *J Cutane Pathol* 43: 200-210.