

Histone Deacetylase Inhibitors (HDACIs): Untapped Therapeutic Potential in Cancer Treatment

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Introduction

Editoria

Epigenetics is the study of changes in phenotype without the corresponding changes in genotype, and historically it includes all the "inexplicable" changes in heritability of phenotype from a single cell to an organism [1]. More recently, a branch of epigenetics examines non-genetic tumor genesis and progression. This branch stems from the notion that not all cancers are generated by genetic alterations, rather epigenetic changes that lead to silencing of certain genes, while allowing transcription machinery access to other genes, resulting in divergence from the parent cell line, loss of cell regulation, and cell immortalization [2-8].

A group of key players in transcriptional regulation are histone acetyltransferases (HATs) and histone deacetylases (HDACs) (Figure 1). These two classes of enzymes work in opposing direction by either catalyzing the transfer of acetyl groups from acetyl coenzyme A (HATs) or by removing acetyl groups (HDACs) from lysine residues of histone tails [2,3,5-11]. Besides acetylation, histones are capable of being methylated, phosphorylated, ubiquinated, sumoylated, poly-ADP ribosylated, carbonylated and glycosylated [2,12].

Mechanisms of action of HDACIs

In eukaryotic cells, DNA wraps around histone proteins, creating complexes called nucleosomes that are packaged inside the nucleus. How tightly the negatively charged DNA is wrapped around the histones partly depends on the acetylation state on histone lysine residues, particularly on histones H3 and H4 [1,8,13]. HATs add an acetyl group from acetyl coenzyme A to the lysine ɛ-amino group, which reduces the positive charge on the histones. When histones are acetylated, the DNA is more loosely wrapped, thus leading to gene activation. HDACs, on the other hand, cause deacetylation, resulting in positively charged histones that are more tightly wrapped by DNA [1,6,8]. Histone deacetylation is correlated with gene repression. The mechanism of action of HDACI in cancer therapy seems to be multifaceted. HDACIs are thought to reduce the deacetylation levels of histone proteins that are overexpressed in cancerous cells. HAT/ HDAC enzymes are capable of regulating both histone and non-histone proteins; therefore, HDACIs are able to exert their therapeutic activity through both histone and non-histone pathways [2,3,7,8,10-12,14-20].

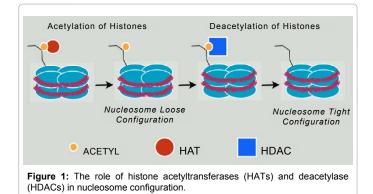
Epigenetic therapy

On the market: Although HDACIs have been investigated against many different types of cancers in both *in vitro* and *in vivo* systems, the clinical understanding and applications of these therapeutic agents have been limited. More success was noted with liquid tumors but there is a shift towards examining the potential use of HDACIs in solid tumors [3,6,8,10,11,13,14,16,21-23]. Fewer studies in solid tumors have reached the clinical trial phase of development (www. ClinicalTrials.gov). On the market, only two HDACIs have been approved by the FDA, namely vorinostat (Zolinza, approved 2006) and romidepsin (Istodax, approved 2009) for the treatment of cutaneous T-cell lymphoma (CTCL) [3,10,11]. Both of these HDACIs bind to the zinc-finger motif of histone deacetylases, resulting in the acetylation

of histones. Vorinostat is considered a pan-inhibitor while romidepsin inhibits HDAC Classes I and II, with significantly increased affinity [2,9].

In clinical trials: Several HDACIs, including valproic acid (an antiepileptic drug), vorinostat, entinostat, and panobinostat, are presently examined in clinical trials for various types of cancers [24]. In a Phase I clinical trial study, the bioavailability of vorinostat ranged from approximately 35% to 52% in patients with advanced cancers [25]. Several clinical trials showed the oral bioavailability of vorinostat and belinostat (PXD101) to be about 33% [26,27]. Valproic acid shows good oral bioavailability [28]. It is currently tested for the treatment of breast, thyroid, lung, ovarian, bladder, head and neck, pancreatic, brain and leukemia cancers. Panobinostat (LBH589) is an HDACI that is similar to vorinostat in its mechanism of action but was found to be more potent [3,5,8,9,11]. It is being tested against Hodgkin's lymphoma and cutaneous T-cell lymphoma as well as other types of cancers [24]. Additionally, there are a number of studies examining the enhanced therapeutic efficacy of using HDACIs with conventional cancer treatment strategies. Table 1 shows examples of some current clinical trials involving HDACIs in breast cancers, as found from www. ClinicalTrials.gov [24].

Challenges and future outlook: The clinical use of HDACIs for cancer therapy is limited. HDACIs have a broad range of applications and can reverse the aberrant epigenetic changes in cancers. The mechanisms by which HDACIs exert their effects are not completely



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HDACI	Study	Phase	Identifier Number
Entinostat (SNDX-275)	Azacitidine and Entinostat in Treating Patients With Advanced Breast Cancer	П	NCT01349959
Entinostat	Exemestane With or Without Entinostat in Treating Postmenopausal Patients With Recurrent Hormone Receptor-Positive Breast Cancer That is Locally Advanced or Metastatic	111	NCT02115282
Entinostat	Entinostat, Lapatinib Ditosylate and Trastuzumab in Patients With Locally Recurrent or Distant Relapsed Metastatic Breast Cancer Previously Treated With Trastuzumab Only	I	NCT01434303
Panobinostat (LBH589)	Re-expression of Estrogen Receptor (ER) in Triple Negative Breast Cancers	1/11	NCT01194908
Vorinostat	Vorinostat in Treating Patients With Stage IV Breast Cancer Receiving Hormone Therapy	Pilot	NCT01720602
Vorinostat	Vorinostat, Paclitaxel, and Bevacizumab in Treating Patients With Metastatic Breast Cancer and/or Breast Cancer That Has Recurred in the Chest Wall and Cannot be Removed by Surgery	1/11	NCT00368875
Vorinostat	Ixabepilone and Vorinostat in Treating Patients With Metastatic Breast Cancer	I	NCT01084057

Table 1: Examples of some clinical trials of combination therapies using HDACIs in breast cancers (obtained from www.ClinicalTrials.gov in May 2014) [24].

understood. Interestingly, HDACIs have been shown to sensitize tumors to their respective therapies [14-16,22,23,29]. Potentially, this approach may lead to improving the treatment of therapy resistant cancers. Studies have shown that HDACIs interact with both histone and non-histone proteins [2,3,7,8,10-12,14-20]. These therapeutic agents cause no or little damage to normal cells [5,19], which is highly desired. Additionally, a major advantage to using HDACIs is that at therapeutic doses, they do not show any major side effects [30]. More recent efforts are focused on examining the enhanced efficacy of HDACIs in combination therapy with conventional cancer treatment approaches [3,8,10,11,15,16,20-23,29]. Additionally, promising results have been shown for using HDACIs as chemopreventive agents [1,3,5,13,15,31-35]. Although additional studies are warranted, epigenetic therapy using HDACIs appears as a novel and effective treatment approach for cancers.

References

- 1. Goldberg AD, Allis CD, Bernstein E (2007) Epigenetics: a landscape takes shape. Cell 128: 635-638.
- Bolden JE, Peart MJ, Johnstone RW (2006) Anticancer activities of histone deacetylase inhibitors. Nat Rev Drug Discov 5: 769-784.
- 3. Bose P, Dai Y, Grant S (2014) Histone deacetylase inhibitor (HDACI) mechanisms of action: Emerging insights. Pharmacol Ther .
- Cruz FD, Matushansky I (2012) Solid tumor differentiation therapy is it possible? Oncotarget 3: 559-567.
- Dokmanovic M, Clarke C, Marks PA (2007) Histone deacetylase inhibitors: overview and perspectives. Mol Cancer Res 5: 981-989.
- Koutsounas I, Giaginis C, Patsouris E, Theocharis S (2013) Current evidence for histone deacetylase inhibitors in pancreatic cancer. World J Gastroenterol 19: 813-828.
- Marks PA, Xu WS (2009) Histone deacetylase inhibitors: Potential in cancer therapy. J Cell Biochem 107: 600-608.
- Xu WS1, Parmigiani RB, Marks PA (2007) Histone deacetylase inhibitors: molecular mechanisms of action. Oncogene 26: 5541-5552.
- Bieliauskas AV, Pflum MK (2008) Isoform-selective histone deacetylase inhibitors. Chem Soc Rev 37: 1402-1413.
- West AC, Johnstone RW (2014) New and emerging HDAC inhibitors for cancer treatment. J Clin Invest 124: 30-39.
- Zhang L, Han Y, Jiang Q, Wang C, Chen X, et al. (2014) Trend of Histone Deacetylase Inhibitors in Cancer Therapy: Isoform Selectivity or Multitargeted Strategy. Med Res Rev.
- Singh BN, Zhang G, Hwa YL, Li J, Dowdy SC, et al. (2010) Nonhistone protein acetylation as cancer therapy targets. Expert Rev Anticancer Ther 10: 935-954.
- Meeran SM, Patel SN, Li Y, Shukla S, Tollefsbol TO (2012) Bioactive dietary supplements reactivate ER expression in ER-negative breast cancer cells by active chromatin modifications. PLoS One 7: e37748.
- Bangert A, Cristofanon S, Eckhardt I, Abhari BA, Kolodziej S, et al. (2012) Histone deacetylase inhibitors sensitize glioblastoma cells to TRAIL-induced

apoptosis by c-myc-mediated downregulation of cFLIP. Oncogene 31: 4677-4688.

- Carrier F (2013) Chromatin Modulation by Histone Deacetylase Inhibitors: Impact on Cellular Sensitivity to Ionizing Radiation. Mol Cell Pharmacol 5: 51-59.
- He G, Wang Y, Pang X, Zhang B (2014) Inhibition of autophagy induced by TSA sensitizes colon cancer cell to radiation. Tumour Biol 35: 1003-1011.
- Kerr E, Holohan C, McLaughlin KM, Majkut J, Dolan S, et al. (2012) Identification of an acetylation-dependant Ku70/FLIP complex that regulates FLIP expression and HDAC inhibitor-induced apoptosis. Cell Death Differ 19: 1317-1327.
- Kramer OH, Baus D, Knauer SK, Stein S, Jager E, et al. (2006) Acetylation of Stat1 modulates NF-kappaB activity. Genes Dev 20: 473-485.
- Lee JH, Choy ML, Ngo L, Foster SS, Marks PA (2010) Histone deacetylase inhibitor induces DNA damage, which normal but not transformed cells can repair. Proc Natl Acad Sci U S A 107: 14639-14644.
- Miller CP, Singh MM, Rivera-Del Valle N, Manton CA, Chandra J (2011) Therapeutic strategies to enhance the anticancer efficacy of histone deacetylase inhibitors. J Biomed Biotechnol 2011: 514261.
- Frew AJ, Lindemann RK, Martin BP, Clarke CJ, Sharkey J, et al. (2008) Combination therapy of established cancer using a histone deacetylase inhibitor and a TRAIL receptor agonist. Proc Natl Acad Sci U S A 105: 11317-11322.
- Seo SK, Jin HO, Woo SH, Kim YS, An S, et al. (2011) Histone deacetylase inhibitors sensitize human non-small cell lung cancer cells to ionizing radiation through acetyl p53-mediated c-myc down-regulation. J Thorac Oncol 6: 1313-1319.
- Wang D, Jing Y, Ouyang S, Liu B, Zhu T, et al. (2013) Inhibitory effect of valproic acid on bladder cancer in combination with chemotherapeutic agents in vitro and in vivo. Oncol Lett 6: 1492-1498.
- 24. http://www.clinicaltrials.gov/ct2/results?term=histone+deacetylase+inhibitor+c ancer&Search=Search
- Kelly WK, Richon VM, O'Connor O, Curley T, MacGregor-Curtelli B, et al. (2003) Phase I clinical trial of histone deacetylase inhibitor: suberoylanilide hydroxamic acid administered intravenously. Clin Cancer Res 9: 3578-3588.
- Northfelt D, Marschke Jr R, Bonnem E, Ooi C, Gerwien R, et al. (2007) A phase Ib/II study of PXD101 alone and in combination with 5-fluorouracil in patients with advanced solid tumors. J Clin Oncol (Meeting Abstracts) 25: 3501.
- 27. Steele N, Vidal L, Plumb J, Attard G, Rasmussen A, et al. (2005) A phase 1 pharmacokinetic (PK) and pharmacodynamic (PD) study of the histone deacetylase (HDAC) inhibitor PXD101 in patients (pts) with advanced solid tumours. J Clin Oncol (Meeting Abstracts) 23: 3035.
- Peterson GM, Naunton M (2005) Valproate: a simple chemical with so much to offer. J Clin Pharm Ther 30: 417-421.
- Konstantinopoulos PA, Wilson AJ, Saskowski J, Wass E, Khabele D (2014) Suberoylanilide hydroxamic acid (SAHA) enhances olaparib activity by targeting homologous recombination DNA repair in ovarian cancer. Gynecol Oncol 133: 599-606.
- Elaut G, Rogiers V, Vanhaecke T (2007) The pharmaceutical potential of histone deacetylase inhibitors. Curr Pharm Des 13: 2584-2620.
- 31. Jakubikova J, Cervi D, Ooi M, Kim K, Nahar S, et al. (2011) Anti-tumor activity

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and signaling events triggered by the isothiocyanates, sulforaphane and phenethyl isothiocyanate, in multiple myeloma. Haematologica 96: 1170-1179.

32. Kelloff GJ, Lippman SM, Dannenberg AJ, Sigman CC, Pearce HL, et al. (2006) Progress in chemoprevention drug development: the promise of molecular biomarkers for prevention of intraepithelial neoplasia and cancer--a plan to move forward. Clin Cancer Res 12: 3661-3697.

33. Li Y, Wicha MS, Schwartz SJ, Sun D (2011) Implications of cancer stem cell

theory for cancer chemoprevention by natural dietary compounds. J Nutr Biochem 22: 799-806.

- 34. Li Y, Zhang T, Korkaya H, Liu S, Lee HF, et al. (2010) Sulforaphane, a dietary component of broccoli/broccoli sprouts, inhibits breast cancer stem cells. Clin Cancer Res 16: 2580-2590.
- Pham TX, Lee J (2012) Dietary regulation of histone acetylases and deacetylases for the prevention of metabolic diseases. Nutrients 4: 1868-1886.