

Human bocavirus (HBoV1 and HBoV2) in Children with Acute Gastroenteritis from North India

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Abstract

Objective: *Human bocavirus* is responsible for respiratory as well as in gastroenteritis illness in several countries including India. This study was aimed to characterize the *Human bocavirus* in North Indian children suffering with acute gastroenteritis.

Methods: A total of 234 diarrheal stool specimens from the children under the age of 15 years were collected and screened for the presence of *Human bocavirus* by polymerase chain reaction targeting *NS1* and *VP1/2* gene followed by sequencing and phylogenetic analysis. Student's t test was performed for statistical analysis of quantitative clinical data.

Results: Total 7 (3%) samples were positive for *Human bocavirus* including bocavirus1 (n=4, 1.7%) and bocavirus2 (n=3, 1.3%). Median age of bocavirus positive children was 8 months (mean \pm SD; 24.14 \pm 27.58 months). Age, duration of diarrhea and episodes of diarrhea were the significant clinical factors (p values 0.0486, 0.0015 and 0.0282 respectively) for bocavirus infection. Interestingly, male were dominated than females (6:1). Phylogenetic analysis showed that Indian bocavirus sequence has similarity with China strains.

Conclusion: Detection of two bocavirus species in gastroenteritis from North India supports global presence of bocavirus. The outcome of the study facilitates more detailed studies of bocavirus infection, its prevalence and disease association in Indian population.

Keywords: Diarrhea; Viral gastroenteritis; Bocavirus; Phylogenetic analysis; North India

Introduction

Acute Gastroenteritis (AGE) is a major cause of morbidity and mortality in children worldwide. Several viruses are associated with gastroenteritis, including rotavirus, adenovirus, astrovirus and calicivirus etc. [1-3]. However, about 35-50% of non-bacterial acute Gastroenteritis is un-identifiable due to unrecognized etiologies [4,5]. Parvovirus is known to cause gastroenteritis in numerous animal species including dogs, cats and hamsters. Several different species and genotypes of Parvovirus infect humans, including the well characterized Parvovirus B19, PARV4 [6], PARV5 [7] and recently identified bocavirus.

Human bocavirus (HBoV), a well-known parvovirus was identified a decade ago in respiratory samples [8] as well as in stool samples [9]. Since then its global presence has been reported in patients with or without gastroenteritis. The *Human bocavirus* (HBoV) genome consists of approximately 5.2 kb nucleotides in length and contains three open-reading frames (ORF) encoding two non-structural proteins (NS1 and NP1) and two capsid proteins (VP1 and VP2) [6]. Based on the Phylogenetic analysis, there are four HBoV species: HBoV1, HBoV2, HBoV3 and HBoV4. HBoV1 was first discovered in respiratory samples [8]. While HBoV2, HBoV3 and HBoV4 were identified in fecal samples of children who had gastroenteritis with or

without symptoms of respiratory infection [9-11]. Very few data are available about presence of bocavirus from India but recently some studies from Delhi and Pune confirmed the association of HBoV in respiratory [12] as well as in gastroenteritis samples [13]. From North India bocavirus has not been reported in association with gastroenteritis, so this study was aimed to define the presence, prevalence and clinical association of bocavirus in North Indian pediatric patients with AGE.

Materials and Methods

Patients and clinical samples

Stool specimens of 234 children under the age of 15 years, presenting symptoms of AGE were collected during July 2012 to June 2013 from Sanjay Gandhi Post Graduate Institute of Medical Sciences, Lucknow, India. This study was approved by Institutional ethics committee. Only the patients with informed written consent from parents/guardians were enrolled in this study. Selection of patients based on the criteria of the World Health Organization (WHO) definition of gastroenteritis having three or more loose stools or any vomiting in 24 hrs. Patients having bloody diarrhea or diarrhea with more than 14 days were excluded. Since the bocavirus has a well-known presence in respiratory diseases, patients having respiratory and lung related diseases were excluded to define the association of bocavirus with gastroenteritis.

Sample processing

Sample processing was done by making 10% stool suspension in phosphate buffer saline, followed by vortexing at mechanical shaker for 20 min and centrifugation at 3000 g for 20 min to pellet particulate matter, the supernatant then passed through a 0.45 µm filter. The filtrate was transferred in cryovials and stored at -20°C until use.

PCR and sequencing

Total nucleic acids were extracted from 200 µL of stool filtrate using QiAamp DNA mini extraction kit (QIAGEN, Inc., Valencia, CA, USA) according to the manufacturer's instructions. The extracted DNA was stored at -70°C until use.

HBoV nucleic acids were detected using the previously described PCR primers and conditions for pan-Bocavirus *NS* gene (~290 bp) and *VP1/2* PCR (~570 bp) [14,11] (Table 1).

| Direction | Primer Name | Region | Sequence(5'-3') | Reference |
|-----------|-------------|----------------|------------------------------------|-----------|
| Forward | panBOVF1 | NS1 (290 bp) | 5'-TAATGCAYCARGAYTGGGTIGANCC-3' | [13] |
| Reverse | panBOVR1 | | 5'-GTACAGTCRTAYTCRTRAA RCACCA-3' | |
| Forward | panBOVF2 | | 5'-GCAYCARGAYTGGGTIGAN CCWGC-3' | |
| Forward | AK-VP-F1 | VP1/2 (570 bp) | 5'-CGCCGTGGCTCCTGCTC T-3' | [14] |
| Reverse | AK-VP-R1 | | 5'-TGTTGCCCATCACAAA GA TG TG-3' | |
| Forward | AK-VP-F2 | | 5'-GGCTCCTGCTCTAGGAA AT AAAGAG-3' | |
| Reverse | AK-VP-R2 | | 5'-CCTGCTGTTAGGTCGTT GTT GTATGT-3' | |

Table 1: Primer sets used in this study for the detection of Human Bocavirus in gastroenteritis samples.

PCR reactions include 2.5 U of Taq DNA polymerase (New England Biolabs, NEB) in 1.1 Thermopol reaction buffer with MgCl₂ (2.0 mmol/L), 50 pmol/L (each) of forward and reverse primers and 5 µL of nucleic acids (for the first round) and 0.5 µL of the first-round PCR product (for the second round) as a template in a 50 µL total volume. The products were visualized following electrophoresis on 2% agarose gel. PCR products, showing positive bands of approximately 290 bp,

corresponding to the highly conserved amplified *NS* gene fragment and 570 bp band for *VP1/2* gene (Figure 1 and Figure 2).

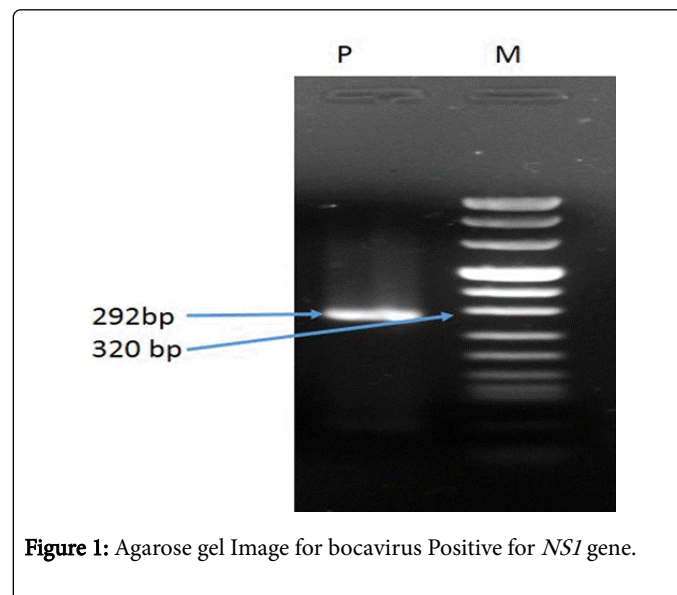


Figure 1: Agarose gel Image for bocavirus Positive for *NS1* gene.

Agarose gel electrophoresis image showing positive bands of bocavirus on 2% agarose gel. Lane M is a DNA molecular weight marker VIII (19 bp to 1114 bp). Lane P is positive band of bocavirus *NS1* gene (292 bp).

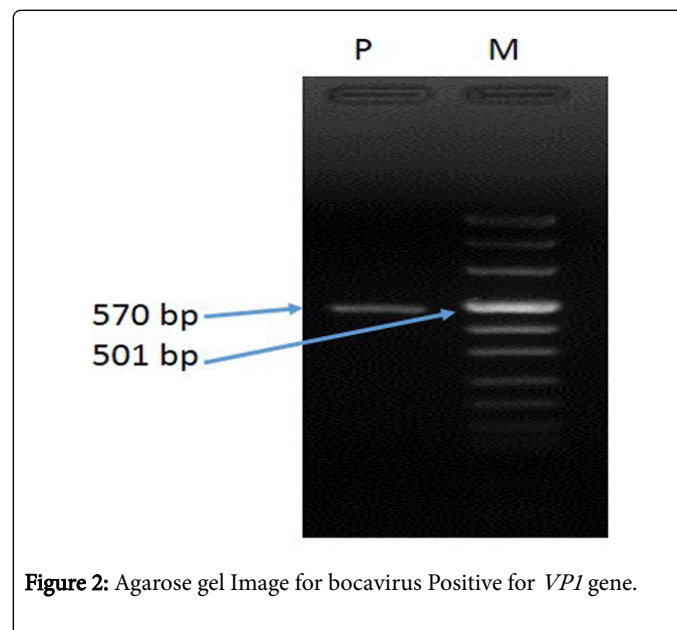


Figure 2: Agarose gel Image for bocavirus Positive for *VP1* gene.

Agarose gel electrophoresis image showing positive bands of bocavirus on 2% agarose gel. Lane M is a DNA molecular weight marker VIII (19 bp to 1114 bp). Lane P is positive band of bocavirus *VP1* gene (570 bp).

All the HBoV positive second round PCR products were treated with Exo-Sap-IT (USB) before direct sequencing. Purified PCR products were sequenced for both forward and reverse primers by ABI Sequencer (scanner v.1.0, model no 3130 I). Sequencing was done for both *NS1* and *VP1/2* genes to define the genetic makeup of North Indian bocavirus.

Sequence data obtained using forward and reverse primers was aligned and edited manually using BioEdit program. Sequences were first analyzed using NCBI Blast N ([http:// www.ncbi.nlm.nih.gov/BLAST](http://www.ncbi.nlm.nih.gov/BLAST)) analysis to find HBoV variants that were genetically most related to HBoV isolates identified in this study. Phylogenetic analysis of NS1 and VP1 sequences identified in this study with best matched NCBI blast search sequences and GenBank reference sequences was performed by Mega 5.05 using maximum likelihood method and Tamura-Nei model with 1000 replicate bootstrapping of the tree to obtain confidence in clade clustering.

Statistical analysis of all the clinical data including age, duration of diarrhea and vomiting in days, episodes of diarrhea and vomiting in 24 hours were performed using student's t test by GraphPadInStat 3.

Results

Prevalence of *Human bocavirus*

Out of 234 samples HBoV were present in 7 (3%) of which 4 were HBoV1 (1.7%) and 3 (1.3%) were HBoV2.

Demographic and clinical profiling

Of the 7 HBoV infected children 6 were male and one was female. All the HBoV infected patients were under the age of 6 years. The median age of HBoV positive cases was 8 months. No significant seasonal distribution of bocavirus has been observed in the studied period. Presence of bocavirus was almost throughout the year though its positivity was higher in the month of January n=2; 29% and March n=2; 29%) while 1 (14%) sample is positive in July, September and November individually (Figure 3).

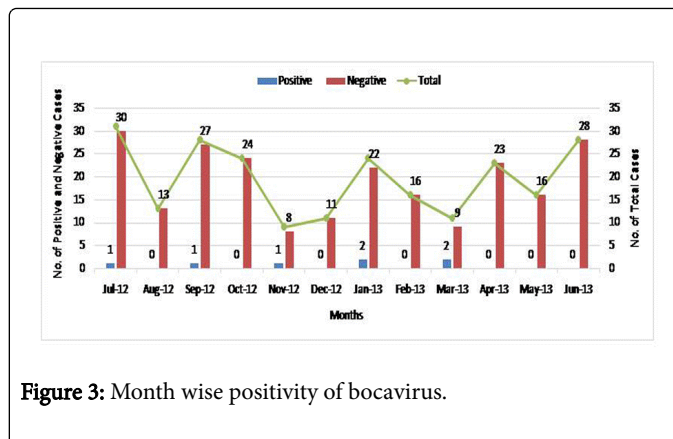


Figure 3: Month wise positivity of bocavirus.

Month wise distribution of *Human bocavirus* cases: X-axis indicates months of sample collection from July 2012 to June 2013. Y-axis indicates total number of positive and negative cases and the secondary axis shows the total number of samples. Values on the blue and red bars indicate number of positive and negative samples respectively.

All the positive samples had gastrointestinal findings having diarrhea 100%, dehydration 86%, vomiting 70%, fever 62%, and severe abdominal pain 28%. Age (p=0.0486), duration of diarrhea in days and diarrheal frequency per day (p=0.0015, p=0.0282) were the significant clinical features (p<0.05) when compared between HBoV positive and negative patients while there was no significant association with duration and frequency of vomiting per day (Table 2).

| Symptoms | Median (Mean ± SD) | | P value |
|----------------------------------|------------------------|------------------------|---------|
| | HBoV positive patients | HBoV negative patients | |
| Age | 8(24.14 ± 27.58) | 34 (45.19 ± 37.69) | 0.0486* |
| Duration of diarrhea (in days) | 5(5.85 ± 1.95) | 4(3.95 ± 1.055) | 0.0015* |
| Episodes of diarrhea (per day) | 7(7 ± 1.91) | 5(5.60 ± 2.50) | 0.0282* |
| Duration of vomiting (in days) | 1(1.71 ± 1.25) | 2(1.709 ± 1.449) | 0.4425 |
| Episodes of vomiting (per day) | 3(3.28 ± 1.79) | 3(2.70 ± 2.311) | 0.2393 |

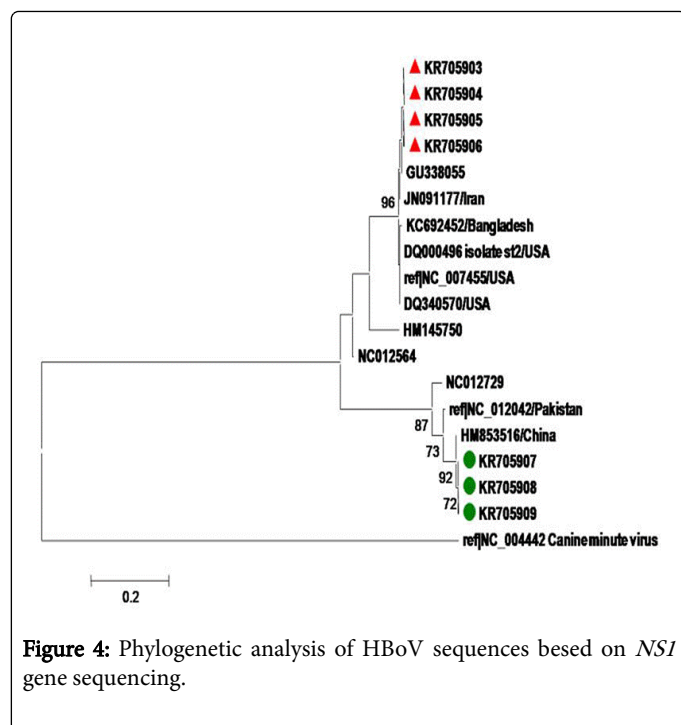
p value<0.05 considered significant and marked with asterisk*

Table 2: Comparison of clinical parameters between bocavirus positive and negative patients.

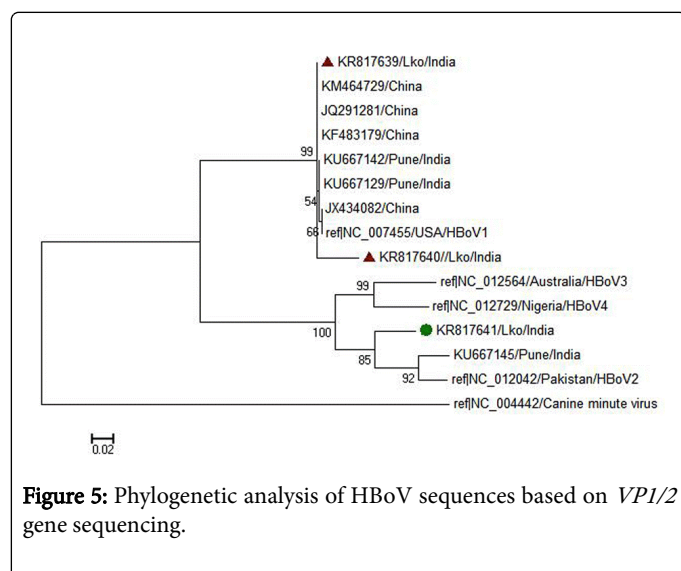
Phylogenetic analyses

Human bocavirus sequences identified in this study were submitted in GenBank (Accession No. KR705903-KR705909, KR817639-KR817641) and compared with the GenBank HBoV available bocavirus prototype strains [HBoV, NC 007455; HBoV2, NC 012042; HBoV3, NC 012564] and animal parvovirus [Canine Minute Virus,

NC 004442]. Sequence analysis showed that four HBoV isolates were related to HBoV1 and three isolates were related to HBoV2 species. Phylogenetic analysis of NS1 (Figure 4) and VP1/2 gene (Figure 5) shows the similarity of Indian *bocavirus* strain with China strains.



Phylogenetic analysis of HBoV sequences identified in this study with sequences from other countries along with GenBank reference sequences [HBoV1, NC__007455; HBoV2, NC__012042; HBoV3, NC__012564 and Canine Minute Virus, NC_004442]. Triangles shows HBoV1 and circles shows HBoV2 identified in this study with their Genbank accession id. Phylogenetic analysis was done by MEGA5 (www.megasoftware.net) using maximum likelihood method; Tamura-Nei model and 1000 replicate bootstrapping of tree was done to obtain confidence in clade clustering.



Phylogenetic analysis of HBoV sequences identified in this study with sequences from other countries along with GenBank reference sequences [HBoV1, NC__007455; HBoV2, NC__012042; HBoV3, NC__012564 and Canine Minute Virus, NC_004442]. Triangle shows HBoV1 and circle shows HBoV2 identified in this study with their

Genbank accession id. Phylogenetic analysis was done by MEGA5 (www.megasoftware.net) using maximum likelihood method; Tamura-Nei model and 1000 replicate bootstrapping of tree was done to obtain confidence in clade clustering.

Discussion

Human bocavirus species are reported as common human virus infections worldwide since their first identification in 2005 [8] and 2009 [9]. The human health relevance of their infections largely remains unknown. Differences in the presence and prevalence of viruses among diseased and healthy individuals can determine the health relevance of a virus infection. HBoV are known to infect healthy people and are frequently detected in respiratory and enteric samples. Recent reports suggest their persistent infection in the human gastrointestinal tract [14-19]. Considering these reports, development of accurate diagnostic assays is required to identify and test the association of these viruses with diseases; therefore we aimed to obtain genetic data on HBoV species infecting children from north India.

This study defines the presence, prevalence and clinical association of two HBoV species (HBoV1 and HBoV2) in pediatric gastroenteritis patients. Our findings were similar to those reported by other countries indicating HBoV detection rate of 4-5% in stools samples of AGE cases [15]. There was no significant seasonal distribution of bocavirus in the studied period. Bocavirus infection rate was relatively higher in the month of January and March like other studies [19-21]. Age and diarrhea were main significant clinical features in bocavirus positive patients.

To best of our knowledge this is the first study to define the prevalence and association of bocavirus in gastroenteritis from North India. Phylogenetic analysis reveals the similarity of Indian bocavirus with China strains. Four *bocavirus* species (HBoV1-HBoV4) were recently reported by Lasure N [13] in gastroenteritis in Pune, West India. We were unable to detect HBoV3 and HBoV4 in gastroenteritis samples from North India. The possible explanation includes primer mismatch or genetic variation or low frequency of occurrence. Further studies are required with large sample size and year round sampling for more years. Low infection rate of HBoV in stool samples of children with gastroenteritis indicates that these viruses are not a common infection in the studied population.

This study has several limitations. Firstly comparatively small sample size may be a possible reason of low prevalence rate. Since bocavirus is known to present in respiratory as well as in subjects with or without gastroenteritis the most important limitation of the study was lack of an asymptomatic control group, though the presence of bocavirus in gastroenteritis sample suggests association of these viruses with AGE.

This study supports the identification and characterization of unknown etiological agents i.e. about 30% to 50% of viral gastroenteritis. The presence and prevalence of bocavirus contribute in some part of undiagnosed etiologies.

Conclusion

This study confirms the circulation of two bocavirus species (HBoV1 and HBoV2) in North India, which are genetically similar to HBoV genotypes, found in China. To the best of our knowledge it is the first report of the presence of HBoV in stool samples from North India. Genetic analysis of data generated from HBoV variants indicates

endemic nature of these virus infections that cannot be discriminated based on their region of isolation. Our results will facilitate more detailed studies of HBoV infection, prevalence and disease association in Indian population.

Acknowledgment

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Ethical Approval

This study was approved by the Institutional Ethics Committee (A-15:PGI/BE/350/7.5.2012) of, Sanjay Gandhi Post Graduate Institute of Medical Sciences, Lucknow, UP, India. Written informed consent was received from the parent or guardian of the patients.

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