

Human Toxicity Photodynamic Therapy Studies on DNA/RNA Complexes as a Promising New Sensitizer for the Treatment of Malignant Tumors Using Bio-Spectroscopic Techniques

A Heidari *

Faculty of Chemistry, California South University, USA

*Corresponding author: A Heidari, Faculty of Chemistry, California South University, (CSU), 14731 Comet St. Irvine, CA 92604, USA, Tel: +1-775-410-4974; E-mail: Scholar.Researcher.Scientist@gmail.com

Received date: May 09, 2016; Accepted date: May 09, 2016; Published date: May 15, 2016

Copyright: © 2016 A Heidari. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Editorial

DNA/RNA complexes play an important role in many biological and catalytic systems. The variety of their functions is due in part to the diversity of metals that bind in the “pocket” of the DNA/RNA complexes system. In this editorial, we discuss synthesis and recognition of DNA/RNA complexes and argue group theory of them. Upon metalation the DNA/RNA complexes system deprotonates, forming a dianionic ligand. The metal ions behave as Lewis acids, accepting lone pairs of electrons from the dianionic DNA/RNA ligand. Unlike most transition metal complexes, their color is due to absorption(s) within the DNA/RNA ligand involving the excitation of electrons from π to π^* ($\pi \rightarrow \pi^*$) DNA/RNA complexes orbitals. Most DNA/RNA complexes contain a metal ion in the center of the planner DNA/RNA complexes system, resulting in a kinetically inert complex. If, however, the ionic radius of the metal ions is too large to fit into the hole in the center of the macro cycle, they are located out of the ligand plane, distorting it. These kinetically labile Sitting–Atop (SAT) complexes display characteristic structural and photo induced properties that strongly deviates from those of the regular DNA/RNA complexes [1-26].

In addition, DNA/RNA complexes functional unit is located in the photosynthetic reaction center and play an important role in biological process such as light energy conversion, Oxygen transport and catalysis. Studies on water–soluble and insoluble DNA/RNA complexes have elucidated aspects of the mechanisms of metal ion incorporation into DNA/RNA to form DNA/RNA complexes. Neglecting the overall charge of the macro cycle, monomeric free–base DNA/RNA complexes in aqueous solution can add protons. The composition of the complex was recognized by bio–spectroscopic techniques such as Attenuated Total Reflectance Fourier Transform Infrared Spectroscopy (ATR–FTIR), FT–Raman, UV–Vis, HR Mass, ¹HNMR, ¹³CNMR and ³¹PNMR bio–spectroscopies. The equilibrium constant (K) was found to be 6–12. It should be noted that a possible mechanism is discussed in details.

On the other hand, DNA/RNA complexes are biochemically important, medically useful and synthetically interesting compounds. In general, there are several routes that can be followed to afford DNA/RNA complexes such as tetramerization, synthesis of DNA/RNA complexes, condensation of DNA/RNA complexes and cyclization of open chain DNA/RNA compounds. The mechanism of Photodynamic Therapy (PDT) has been mentioned as well.

Furthermore, the synthesis of water–soluble DNA/RNA complexes has been discussed. This synthesis includes condensation, followed by metal insertion, oxidation, demetallation and deboronation reactions.

This compound accumulated within human glioblastoma U87MG cells to a significant higher extent than structurally related DNA/RNA complexes and localized preferentially in the cell lysosomes. Human toxicity studies are shown that both compounds are non–toxic even at a dose of 250 mg/kg. It is concluded that the DNA/RNA complexes is a promising new sensitizer for the treatment of malignant tumors.

Moreover, reducing agents hydroxylamine hydrochloride and metallic Rhenium (Re) have catalytic effect on the rate of the Rhenium (Re) incorporation into DNA/RNA complexes. These reactions was studied at pH 2–6 and 20–30°C. Rhenium (Re) in one oxidation has large ionic radius and cannot incorporate well into the DNA/RNA complexes core. Rhenium (Re) deforms the DNA/RNA complexes plane favorably for attack of Rhenium (Re) from the back side. It should be noted that the mechanism is described in details.

References

1. Heidari A (2012) A thesis submitted to the Faculty of the Chemistry, California South University (CSU), Irvine, California, The United States of America (USA) in fulfillment of the requirements for the degree of Doctor of Philosophy (PhD) in chemistry.
2. Heidari A (2015) Simulation of interaction of light and iridium nanoparticles using 3D finite element method (FEM) as an optothermal cancer cells treatment. International Journal of Theoretical, Computational and Mathematical Chemistry 1: 11-16.
3. Heidari A, Brown C (2015) Study of composition and morphology of cadmium oxide (CdO) nanoparticles for eliminating cancer cells. Journal of Nanomedicine Research 2: 20.
4. Heidari A, Brown C (2015) Study of surface morphological, phytochemical and structural characteristics of rhodium (III) oxide (Rh₂O₃) nanoparticles, International Journal of Pharmacology, Phytochemistry and Ethnomedicine 1: 15-19.
5. Heidari A (2016) An Experimental Biospectroscopic Study on Seminal Plasma in Determination of Semen Quality for Evaluation of Male Infertility. Int J Adv Technol 7: e007.
6. Heidari A (2016) Extraction and Preconcentration of N-Tolyl-Sulfonyl-Phosphoramid-Saeure-Dichlorid as an Anti-Cancer Drug from Plants: A Pharmacognosy Study. J Pharmacogn Nat Prod 2: e103.
7. Heidari A (2016) A Thermodynamic Study on Hydration and Dehydration of DNA and RNA–Amphiphile Complexes. J Bioeng Biomed Sci 5: 006.
8. Heidari A (2016) Computational Studies on Molecular Structures and Carbonyl and Ketene Groups' Effects of Singlet and Triplet Energies of Azidoketene O=C=CH–NNN and Isocyanatoketene O=C=CH–N=C=O. J Appl Computat Math 5: e142.
9. Heidari A (2016) Study of Irradiations to Enhance the Induces the Dissociation of Hydrogen Bonds between Peptide Chains and Transition from Helix Structure to Random Coil Structure Using ATR–FTIR, Raman and ¹HNMR Spectroscopies. J Biomol Res Ther 5: e146.

10. Heidari A (2016) Future Prospects of Point Fluorescence Spectroscopy, Fluorescence Imaging and Fluorescence Endoscopy in Photodynamic Therapy (PDT) for Cancer Cells. *J Bioanal Biomed* 8: e135.
11. Heidari A (2016) A Bio-Spectroscopic Study of DNA Density and Color Role as Determining Factor for Absorbed Irradiation in Cancer Cells. *Adv Cancer Prev* 1: e102.
12. Heidari A (2016) Manufacturing Process of Solar Cells Using Cadmium Oxide (CdO) and Rhodium (III) Oxide (Rh₂O₃) Nanoparticles. *J Biotechnol Biomater* 6: e125.
13. Heidari A (2016) Anti-Cancer Effect of UV Irradiation at Presence of Cadmium Oxide (CdO) Nanoparticles on DNA of Cancer Cells: A Photodynamic Therapy Study. *Archives in Cancer Research* 4: 61.
14. Heidari A (2016) Quantitative Structure-Activity Relationship (QSAR) Approximation for Cadmium Oxide (CdO) and Rhodium (III) Oxide (Rh₂O₃) Nanoparticles as Anti-Cancer Drugs for the Catalytic Formation of Proviral DNA from Viral RNA Using Multiple Linear and Non-Linear Correlation Approach. *Annals of Clinical and Laboratory Research* 4: 76.
15. Heidari A (2016) An Analytical and Computational Infrared Spectroscopic Review of Vibrational Modes in Nucleic Acids. *Austin J Anal Pharm Chem* 3: 1058.
16. Heidari A (2016) Biochemical and Pharmacodynamical Study of Microporous Molecularly Imprinted Polymer Selective For Vancomycin, Teicoplanin, Oritavancin, Telavancin and Dalbavancin Binding. *Biochem Physiol* 5: e146.
17. Heidari A (2016) A Novel Experimental and Computational Approach to Photobiosimulation of Telomeric DNA/RNA: A Biospectroscopic and Photobiological Study. *J Res Development* 4: 144.
18. Heidari A (2016) A Combined Computational and QM/MM Molecular Dynamics Study on Boron Nitride Nanotubes (BNNTs), Amorphous Boron Nitride Nanotubes (a-BNNTs) and Hexagonal Boron Nitride Nanotubes (h-BNNTs) as Hydrogen Storage. *Structural Chemistry & Crystallography Communication* 1: 18.
19. Heidari A, Brown C (2016) Phase, Composition and Morphology Study and Analysis of Os-Pd/HfC Nanocomposites. *Nano Research & Applications* 1: 14.
20. Heidari A (2016) Biomedical Study of Cancer Cells DNA Therapy Using Laser Irradiations at Presence of Intelligent Nanoparticles. *J Biomedical Sci* 5: 2.
21. Heidari A (2016) Spectroscopy and Quantum Mechanics of the Helium Dimer (He₂⁺), Neon Dimer (Ne₂⁺), Argon Dimer (Ar₂⁺), Krypton Dimer (Kr₂⁺), Xenon Dimer (Xe₂⁺), Radon Dimer (Rn₂⁺) and Ununoctium Dimer (Uuo₂⁺) Molecular Cations. *Chem Sci J* 7: e112.
22. Heidari A (2016) Biospectroscopic Study on Multi-Component Reactions (MCRs) in Two A-Type and B-Type Conformations of Nucleic Acids to Determine Ligand Binding Modes, Binding Constant and Stability of Nucleic Acids in Cadmium Oxide (CdO) Nanoparticles-Nucleic Acids Complexes as Anti-Cancer Drugs. *Arch Cancer Res* 4: 2.
23. Heidari A (2016) A Chemotherapeutic and Biospectroscopic Investigation of the Interaction of Double-Standard DNA/RNA-Binding Molecules with Cadmium Oxide (CdO) and Rhodium (III) Oxide (Rh₂O₃) Nanoparticles as Anti-Cancer Drugs for Cancer Cells' Treatment. *Chemo Open Access* 5: e129.
24. Heidari A (2016) Simulation of Temperature Distribution of DNA/RNA of Human Cancer Cells Using Time-Dependent Bio-Heat Equation and Nd: YAG Lasers. *Arch Cancer Res* 4: 2.
25. Heidari A (2016) Measurement the Amount of Vitamin D2 (Ergocalciferol), Vitamin D3 (Cholecalciferol) and Absorbable Calcium (Ca²⁺), Iron (II) (Fe²⁺), Magnesium (Mg²⁺), Phosphate (PO₄⁻) and Zinc (Zn²⁺) in Apricot Using High-Performance Liquid Chromatography (HPLC) and Spectroscopic Techniques. *J Biom Biostat* 7: 292.
26. Heidari A, Brown C (2016) Vibrational spectroscopic study of intensities and shifts of symmetric vibration modes of ozone diluted by cumene. *International Journal of Advanced Chemistry* 4: 5-9.