



dilutions, AKWATON killed MRSA and *E. coli* respectively at a low concentration of 0.04% and 0.005% within 1.5 minutes [19]. These authors also demonstrated that the minimum sporostatic concentration, the minimum sporicidal concentration and the time required for sporicidal activity corresponded to 0.06% (w/v), 0.08% (w/v) and 8.5 minutes, respectively [19]; and that the minimum inhibitory concentration, the minimum fungicidal concentration (MFC) and time required for the fungicidal activity of AKWATON at the MFC were 0.025% (w/v), 0.045% (w/v) and 2.5 min, respectively Oulé et al. [20].

Other previous studies have demonstrated that PHMGH, the active ingredient of the new AKWATON-based disinfectant, is an odourless, colourless and noncorrosive polymer with high solubility in water [22] and is significantly less toxic and harmless than currently used disinfectants [14,23] to humans and animals at a concentration  $\leq 1\%$ . In addition, cell cultures and the different tests carried out by Oulé et al. [20], showed that the new AKWATON-based disinfectant killed fewer animal cells than the commercial disinfectants, sparing 80% of rat pancreatic (C2C12) cells and 65% of muscle RnM5F cells, whilst some of the well-known disinfectants currently on the market killed 85-100% of those cells.

To demonstrate its potential use as a less hazardous antibacterial than the currently used disinfectants, and to validate its use as an antiseptic and an ideal disinfectant for households and hospitals, the toxic effects of AKWATON have been assessed on human tissues.

## Material and Methods

Human cells used in this study were lung cells (IMR-90 (ATCC<sup>®</sup> CCL-186<sup>™</sup>)); liver cells (Hep G2 [HEPG2] (ATCC<sup>®</sup> HB-8065<sup>™</sup>)); skin cells (A-431 (ATCC<sup>®</sup> CRL-1555<sup>™</sup>)) and eye cells (ARPE-19 (ATCC<sup>®</sup> CRL-2302<sup>™</sup>)), purchased from American Type Culture Collection (ATCC) (Manassas, Va., USA). All the reagents have been purchased from Sigma Chemical Co. (St Louis, MO, USA). Cells were maintained at 37°C under a continuous 5% CO<sub>2</sub> atmosphere.

## Assessment of the effect of disinfectants on cells

AKWATON (0.05%) and three well-known and currently marketed antimicrobial chemicals including Ethanol (70%), DEXIDIN-4, and the commercial disinfectant LYS purchased from Canadian Real Super Store (Winnipeg, MB, Canada) were used to treat the four types of human cells. Skin cell (A-431 (ATCC<sup>®</sup> CRL-1555<sup>™</sup>)) cells and eye cells (ARPE-19 (ATCC<sup>®</sup> CRL-2302<sup>™</sup>)) were plated at 7,500 cells/cm<sup>2</sup> and cultured in DMEM (Dulbecco's Modified Eagle's Medium); and Lung cells (IMR-90 (ATCC<sup>®</sup> CCL-186<sup>™</sup>)) and liver cells (Hep G2 [HEPG2] (ATCC<sup>®</sup> HB-8065<sup>™</sup>)) in EMEM (Eagle's Minimum Essential Medium).

All media were supplemented with 1% Penicillin, 1% Streptomycin, 2 mM glutamine and 10% fetal bovine serum (FBS). Cells were incubated at 37°C under a continuous 5% CO<sub>2</sub> atmosphere for 24 hour to 65-75% confluency. Before each test, cells were detached using 0.05% trypsin. Cells were exposed to AKWATON-based disinfectant and to three other well-known commercial antimicrobial chemicals for 10 minutes.

Cultures with  $\geq 95\%$  viable cells were used for experiments. The viability of cells before and after treatment was tested by trypan blue exclusion tests and MTT test. Cells were observed under a microscope and counted using a hemocytometer. Calculation: Cell

Viability (%) =  $\frac{\text{total viable cells (unstained)}}{\text{total cells (stained and unstained)}} \times 100$  [24].

## Statistical analysis

Three different batches of the AKWATON-based disinfectant and four three well-known commercial antimicrobial chemicals were used to treat the four types of cells. For AKWATON, reported data was the average of the results from the three batches tested. Each test with each chemical was performed in duplicate and repeated three times. Results were analyzed using one-way analysis of variance and Student's t-test. Differences with a value of  $P < 0.05$  were considered statistically significant.

## Discussion

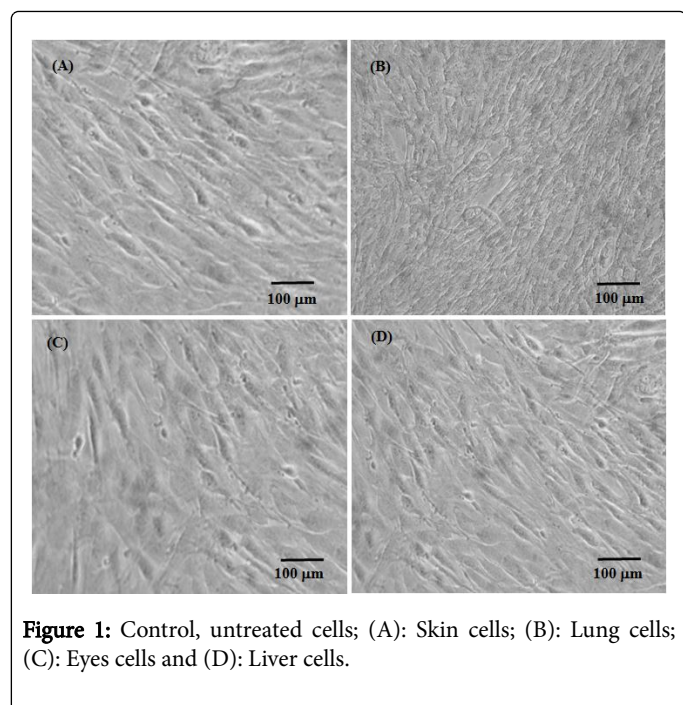
The purpose of applying disinfectants and antiseptics is to kill microorganisms or prevent their development. But an equally important thing is to ensure that the application of these antimicrobial products does not affect the human and animal health and does not affect the environment, which is a real concern with the majority of commercial chemicals currently on the market. The advent of new bacterial strains that are increasingly resistant to antibiotics and disinfectants is a serious problem that deserves more attention and to which a durable solution must be found.

This is why it is necessary to develop new molecules with powerful antimicrobial properties, but harmless for human and animal organisms and for the environment. AKWATON is a new colorless, odorless, non-corrosive antimicrobial product with excellent microbicidal power. Its antimicrobial activities have been widely documented [19,25-27].

Oulé et al. [19] have clearly demonstrated its bactericidal activity against *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Salmonella choleraesuis*, methicillin-resistant *S. aureus* (MRSA) and *E. coli*; its sporicidal activity against *Bacillus subtilis* spores [20] and its fungicidal activity against *Trichophyton mentagrophytes* [4]. In addition to its microbicidal properties, it has been demonstrated that AKWATON was much less toxic to rat cells than well-known disinfectants currently on the market. Oulé et al. [4] established that the AKWATON-based disinfectant was much less toxic to rat pancreatic and muscle cells.

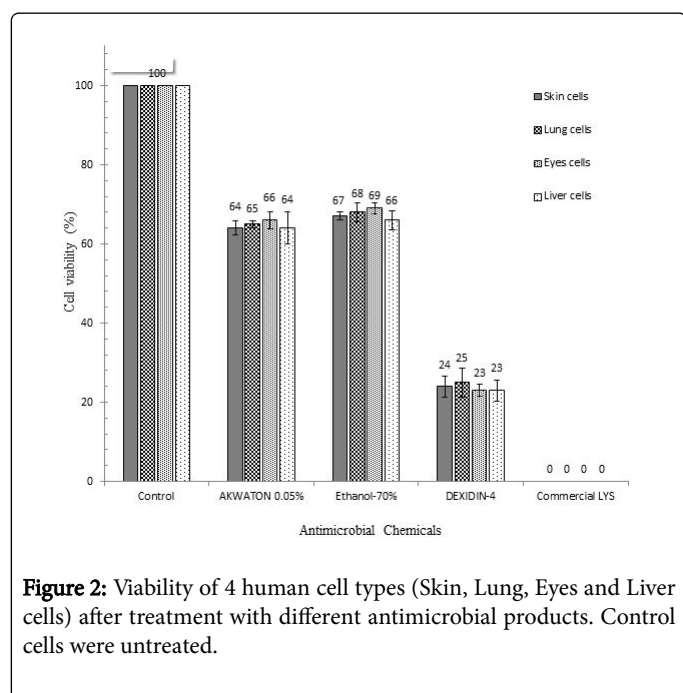
The AKWATON-based disinfectant killed fewer cells than the commercial disinfectants, sparing 80% of C2C12 (ATCC<sup>®</sup> CRL-1772<sup>™</sup>) cells and 65% of RIN-m5F (ATCC<sup>®</sup> CRL-11605<sup>™</sup>) cells, whilst LYS and MCL, currently on the market, killed 85%-100%. The purpose of this *in vitro* study is to show that AKWATON is also less toxic to human cells than commercial antimicrobial products. The *in vitro* tests with cell lines are a good alternative to the use of laboratory animals in toxicological studies. Animal protection organizations are increasingly opposed to laboratory experiments on animals, because of the cruelty inherent in these practices [28].

In this study, the toxic effects of AKWATON (0.05%) (a novel antimicrobial product), Ethanol (70%), DEXIDIN-4 (a commercial antiseptic) and LYS (a commercial disinfectant) were assessed on human cells including eyes, skin, lung and liver cells in *In-vitro* experiments. Figure 1 shows untreated cells which, have served as a control to observe and evaluate the effects of the four antimicrobial products tested. Untreated cells were numerous, healthy and evenly distributed in the visual field of the microscope.



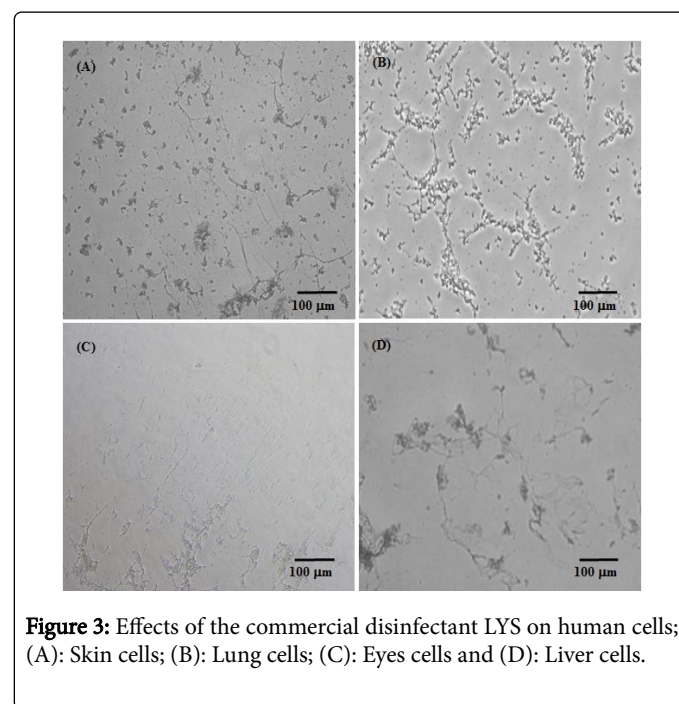
**Figure 1:** Control, untreated cells; (A): Skin cells; (B): Lung cells; (C): Eyes cells and (D): Liver cells.

The Figure 2 compares the survival of the four types of human cells exposed to various chemicals mentioned above. Each antimicrobial product killed the same percentage of cells in the four cell types. For each antimicrobial product, there was no significant difference between the cell types ( $P > 0.05$ ). However, for each cell type a significant difference was observed between antimicrobial products ( $P < 0.05$ ), except between AKWATON (0.05%) and Ethanol (70%) ( $P > 0.05$ ). Among the four antimicrobial products, LYS was highly toxic, killing 100% of each of the four cell types used, probably due to its chemical composition.



**Figure 2:** Viability of 4 human cell types (Skin, Lung, Eyes and Liver cells) after treatment with different antimicrobial products. Control cells were untreated.

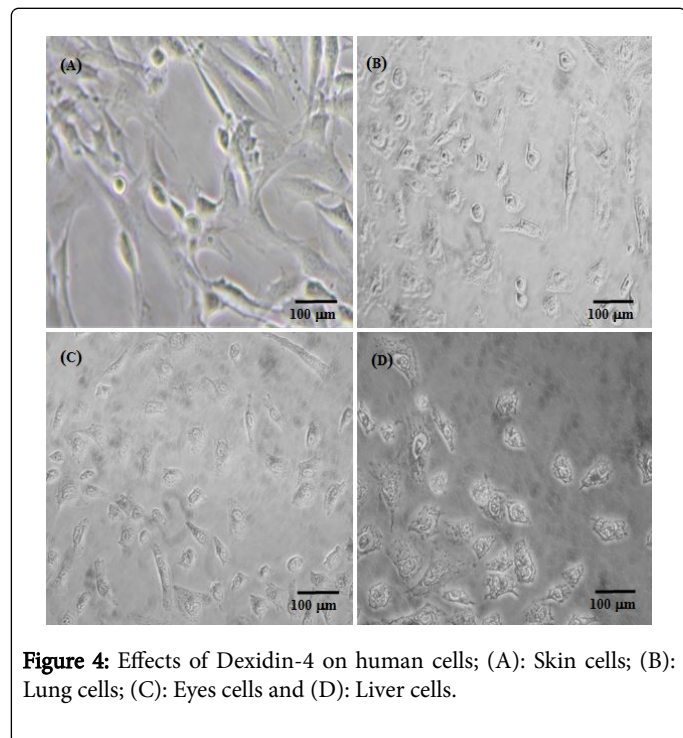
The commercial disinfectant LYS used in this study is a mixture of several active ingredients such as Ethyl alcohol, Butane, Propane and N-Alkyl-dimethyl benzyl ammonium chloride. The morphology of cells after their exposure to LYS suggests that the disinfectant attacked various targets on cells. Their appearance suggests that some active molecules in the disinfectant LYS fragmented the plasma membrane followed by penetration of other actives molecules into the cells that attack cytoplasmic components, causing loss of the cytoplasm, total collapse of the cells and resulting in an appearance of puree of the cell mass. Figure 3 shows cells after a 10 min-treatment with LYS. All cells were completely destroyed.



**Figure 3:** Effects of the commercial disinfectant LYS on human cells; (A): Skin cells; (B): Lung cells; (C): Eyes cells and (D): Liver cells.

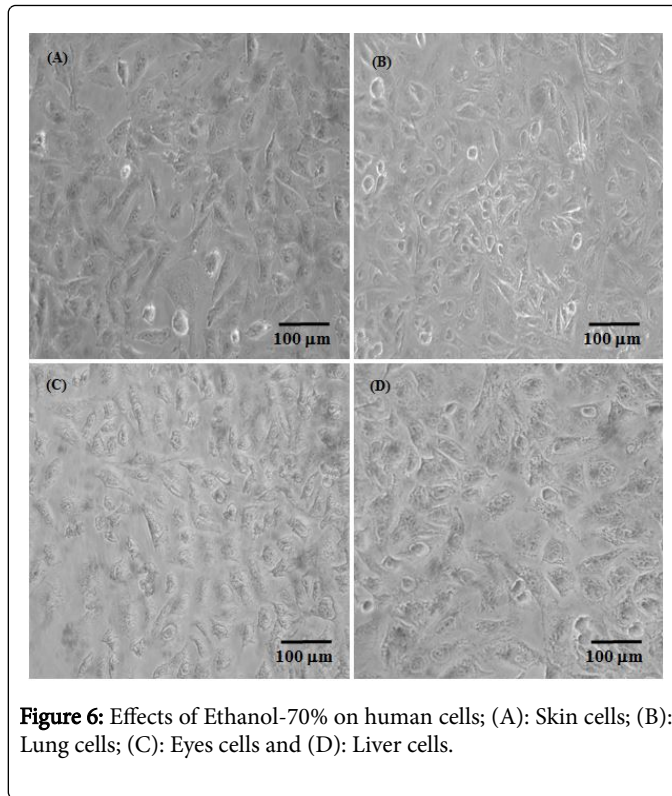
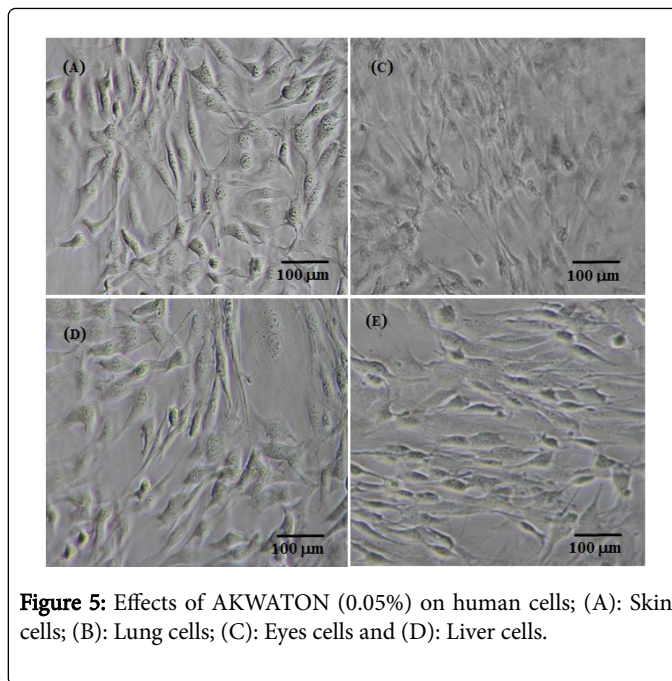
DEXIDIN-4 also caused a high percentage of cell death killing approximately 75% of cells in each cell type. Figure 4 shows cells after their treatment with DEXIDIN-4. DEXIDIN-4 was toxic to cells but less than LYS. There was a significant difference between DEXIDIN-4 and LYS ( $P < 0.05$ ). DEXIDIN-4 is used as an antimicrobial surgical cleaner; antiseptic cleaner for skin and minor injuries and for hand washing in hospitals, labs and food services. DEXIDIN-4 is composed of 4% chlorhexidine gluconate and 4% isopropyl alcohol. Depending on the concentration, each of these two chemical compounds can be toxic to mammalian cells, and their modes of action on cells are very well documented. Isopropanol is an active ingredient that can cause cell death at relatively low concentrations such as 4% in the antiseptic DEXIDIN-4. Kasajima, et al. [29] reported in an *in vitro* study that the inhibition of DNA, RNA and protein-syntheses of the mammalian cells was induced by treatment with ethyl alcohol at 1% to 10% (V/V) for 2 hours, in a dose-related manner. In DEXIDIN-4, the other main active ingredient is 4% chlorhexidine gluconate. An *in vitro* study by Flemingson et al. [30], comparing the effects of three mouth rinses on human gingival fibroblasts, showed that these three rinses were toxic to the cells, chlorhexidine being the most cytotoxic. Taner et al. [31] have also demonstrated the genotoxic and cytotoxic effects of chlorhexidine on human lymphocytes. According to Nancy and Don [32], cultured human periodontal cells treated with 2% chlorhexidine exhibited a foamy appearance in which most of the cytoplasm seemed to have been extracted from the cells. According to an *in vivo* genotoxicity

study conducted by Grassi et al. [33] in rats, chlorhexidine induced DNA damage in leukocytes, renal cells and oral mucosal cells. And in an *in vitro* study on rat oral mucosal cells and rat leukocytes, Ribeiro et al. [34] demonstrated that Chlorhexidine was highly cytotoxic, inducing oxidative stress and apoptotic and necrotic cell death. In this study, the treatment with DEXIDIN-4 resulted in the death of approximately 75% of cells of each cell type. Such high rate of cell death results directly from the elevated toxicity of DEXIDIN-4 on the cells. This high toxicity could be attributed to the combined actions of alcohol and chlorhexidine on the cells.



The other two antimicrobial products, AKWATON (0.05%) and Ethanol (70%) produced less cytotoxic effects on each cell type. When the cells were exposed to AKWATON (0.05%), respectively 64%, 65%, 66% and 64% of the skin, lung, eye and liver cells survived (Figure 1). Similar results were observed with Ethanol (70%). There was no significant difference between their effects on the cells ( $P > 0.05$ ). These two chemicals (Ethanol and AKWATON) killed about 30% to 35% of cells. Figures 5 and 6 show the appearance of the four cell types after their treatment with AKWATON-0.05% and Ethanol (70%).

In an *in vitro* study, Lingna et al. [35] examined the effects of Ethanol on mouse skin cells and they observed a dose-dependent toxic effect. When the concentration reached 50%, more than 75% of cells were killed after 2 days of exposure. In this study, we observed about 30% of cell death, probably because of the short exposure time of 10 min. The cells that survived treatment with AKWATON-0.05% (Figure 5) or with Ethanol (70%) (Figure 6) seemed to be healthy compared to untreated cells (Figure 2). The effect of Ethanol is known; it acts on the biological membrane by interdigitating the two lipid layers, thus reducing its thickness and increasing its permeability [36]. Oulé et al. [19] suggested that the main target of PHMGH, the active molecule in AKWATON, seems to be the cell envelope. PHMGH would penetrate the cell envelope, attacking the bacterial cell wall and the membrane at the same time.



The relative resistance of animal and human cells to AKWATON and Ethanol could be explained by the composition and structure of their membrane. For example, the presence of cholesterol and the quality of membrane lipids (length, saturation and level of branching) would enhance the integrity and the stability of the membrane of animal and human cells. Cholesterol is a major component of mammalian cell membranes. It contributes to the stability and the maintenance of membrane structure by intercalating between the

phospholipids, acting as a pillar of resistance to attacks on the membrane. Essentially, cholesterol intervenes in the integrity, the stability and the maintenance of membrane fluidity [37]. Cholesterol also contributes to the tightness of cell membranes, reducing their permeability to various substances [38,39]. The presence of cholesterol would reduce the penetration of PHMGH and Ethanol into cells, which would mitigate their toxicity on cells.

One wonders why the presence of cholesterol in the cell membrane could not allow the cells to resist to LYS or DEXIDIN-4. In fact, the chemical composition of LYS and DEXIDIN-4 is complex. These two antimicrobial products contain more than one active ingredient. LYS and DEXIDIN-4 are mixtures of several active components and were found to be highly toxic to the four cell types (Figure 1). The presence of several active molecules in an antimicrobial product can be effective in eliminating or reducing the spread of potentially pathogenic microorganisms, but can also generate toxic effects on mammalian cells. In AKWATON there is only one active molecule, the PHMGH, which acts on cell membranes [19]. In this study, AKWATON was shown to be much less toxic to human cells. In a previous study, Oulé et al. [4] showed that AKWATON (0.05%) was much less toxic to rat cells than commercial disinfectants. In another study, the same authors showed that AKWATON (0.04%) was very toxic for bacterial cells [19]. It is known that the membrane of the bacterial cell does not contain cholesterol. The presence of cholesterol would seem to be a determining factor for the effect of AKWATON.

## Conclusion

The growing resistance of pathogenic microorganisms to antibiotics and antimicrobial chemicals, as well as the toxicity of these products to human and animal organisms and their aggressiveness to the environment, are real concerns. AKWATON, an odourless, colourless, stainless and non-corrosive substance with high bactericidal, fungicidal and sporicidal potency, is significantly less toxic to human lung, liver, skin and eye cells than two commercial antimicrobial products currently on the market. Compared to some commercial chemicals that kill 100% human cells *in vitro*, AKWATON produces the same effects as 70% ethanol, widely known and used all over the world, killing only 30% of human cells. This study demonstrates that AKWATON with all of these properties is an ideal antimicrobial product for hospitals, laboratories, food industries and household facilities.

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