Lessons Learned from Two Missed Prenatal Cases of Hemoglobin Bart’s Hydrops Fetalis Until Second Trimester Despite a Nationwide Screening Program

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Abstract

Hemoglobin (Hb) Bart’s hydrops fetalis is the most severe form of alpha-thalassemia and has high mortality. It is caused by deletion of all four α-globin genes leading to a severe deficiency in α-globin and to the production of γ-globin tetramers, resulting in ineffective tissue oxygen delivery. Current preconceptional or prenatal screening policies for this disorder are comprehensive, but not all pregnancies with Hb Bart’s disease can be detected early before hydrops become apparent. We report here two cases with Hb Bart’s disease that escaped being detected until second trimester after hydrops fetalis occurred. These cases remind the clinicians that the importance of this well-characterized condition (alpha-thalassemia) should be reasserted despite a nationwide screening program is available. We also review the current status of prenatal screening and diagnosis of alpha-thalassemia.

Keywords: Alpha-thalassemia; Hb Bart’s; Hydrops fetalis; Fetal ascites; Cardiomegaly

Abbreviations: Hb: Hemoglobin; SEA: Southeastern Asia; HbCS: Hb Constant Spring; HbQS: Hb Quong Sze; GA: Gestational Age; MCV: Mean Corpuscular Volume; CBC: Complete Blood Count; MCH: Mean Corpuscular Hemoglobin; MCA-PSV: Peak Systolic Velocity of Middle Cerebral Artery; CTR: Cardiothoracic Ratio; MoM: Multiples of Median; IHUV Vmax: Intrahepatic Umbilical Venous Maximum Flow Velocity; ARMS: Amplification Refractory Mutation System; PCR: Polymerase Chain Reaction

Introduction

Alpha-thalassemia is one of the most common hereditary hemoglobinopathies, which interferences with the production of α-globin chains of hemoglobin (Hb) and causes perinatal complications. Approximately 5% of the world’s population carries mutant or variant alleles over the α-globin gene, with the high frequencies throughout most of Southeastern Asia (SEA), the Mediterranean area, the Indian subcontinent, the Middle East and Africa. In SEA region, the frequency of α-thalassemia variants (αα/- -) is about 5-15% [1]. Among Taiwanese, the carrier rate of alpha-thalassemia is approximately 5-7% [2]. The disorder is associated with more than 100 known molecular defects in the gene cluster located on the short arm of chromosome 16 (16p13.3) and these defects can result in various clinical phenotypes [3]. Hb Bart’s hydrops fetalis is the most severe form of alpha-thalassemia, which is caused by deletion of all four α-globin genes (-/- -), and therefore no α-globin is produced. As a result, γ- globin tetramers (Hb Bart’s) become the dominant form of Hb and are responsible for carrying oxygen from mother to fetus. However, Hb Bart’s has a characteristic of extremely high oxygen affinity, which prevents oxygen release from the red blood cells to the tissue. This results in the occurrence of hydrops fetalis, stillbirth, or neonatal demise. Hence, it is critical to diagnose Hb Bart’s hydrops fetalis in early pregnancy as well as to identify high-risk groups for hereditary hemoglobinopathies. However, despite the current nationwide preconceptional or prenatal screening policies, not all pregnancies with Hb Bart’s disease can be detected in the first or early second trimester before hydrops become apparent. Here, we report two prenatal cases of Hb Bart’s disease, which escaped being detected by current screening program until second trimester after developing hydrops fetalis. Besides, we discuss the limitations of current “comprehensive” nationwide screening program used to screen alpha-thalassemia, as well as to review the current status and the future prospects.

Case Report

Patient 1

A 33-year-old primigravida was referred to our hospital at 20th weeks of gestational age (GA) after her obstetrician noted fetal ascites in routine antenatal care. The parents of the fetus were both Han Chinese. In the first visit to our hospital, the mother claimed that she had normal antenatal test results except iron deficiency anemia by the result of complete cell count (CBC), ferritin, serum iron and total iron binding capacity. But her obstetricians didn’t request her husband to be checked. The initial ultrasound performed at our hospital showed a live singleton fetus whose estimated fetal weight below the 10th percentile. Anatomical examination demonstrated massive fetal ascites, echogenic bowels and cardiomegaly (cardiothoracic ratio: 0.63) (Figure 1). No other structural abnormalities were noted. The amniotic fluid volume

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Received September 24, 2015; Accepted October 23, 2015; Published October 26, 2015


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was within normal limits and peak systolic velocity of middle cerebral artery (MCA-PSV) was below 1.5 multiples of median (MoM). We performed amniocentesis and rechecked parental blood samples to exclude the possibility of chromosome abnormality and intrauterine infections. All of the tests were normal, but both of the parents showed microcytic anemia (Mother: Hb: 10.6 g/dL, mean corpuscular volume (MCV): 69.5 fL; Father Hb: 11.6 g/dL, MCV: 69.0 fL). We highly suspected that the etiology of fetal ascites was Hb Bart’s disease. The DNA from the couple (from peripheral blood) and the fetus (from amniotic fluid) were thus sent for genetic testing. The results revealed that both parents are carriers of alpha-thalassemia (αα/- SEA) and the fetus is a case of Hb Bart’s hydrops fetalis (-/-SEA). After non-directive genetic counseling and consultation of neonatal and pediatric hematological specialists, the parents elected to terminate the pregnancy.

**Patient 2**

A 28-year-old primigravida admitted into our hospital at GA 24+1 weeks because of acute upper respiratory tract infection. The admission surveillance incidentally showed cardiomegaly (cardiothoracic ratio: 0.68). Reviewing her personal history, she was a victim of alpha-thalassemia trait. However, her husband was considered as a non-carrier of alpha-thalassemia only by means of electrophoresis. No genetic testing was performed for DNA studies of thalassemia. The parents of the fetus were both Han Chinese. Detailed ultrasound showed no other anatomical abnormality except echogenic bowels. The MCA-PSV was below 1.5 MoM. Clinical surveillance including intrauterine infection screening, Hb genotyping and array comparative genomic hybridization was performed for the couple and fetus. As a result, both of the parents were demonstrated to be carriers of alpha-thalassemia (αα/-SEA), and the fetus was proven to be a case of hydrops fetalis due to Hb Bart’s disease by DNA studies. Other investigation revealed normal result except an irrelevant balanced translocation was found: 46,XX,t(1;4)(p32.1;q31.3). After explanation of the possible perinatal outcome to the parents, they requested to terminate the pregnancy.

**Discussion**

In this report, we present two prenatal cases of HB Bart’s disease that escaped early detection until second trimester after developing hydrops fetalis. The results highlighted the possible pitfalls of the current program for early screening of alpha-thalassemia despite the program has been nationwide available for many years. Alpha-thalassemia is a relatively endemic hereditary hemoglobinopathy in Taiwan and about 5% of population worldwide carries an alpha-thalassemia trait. However, her husband was considered as a non-carrier of alpha-thalassemia only by means of electrophoresis. The method in the definitive identification of variant HbS. Meanwhile, the test cannot define the diagnosis of alpha-thalassemia, especially when coexistence of alpha- and beta-thalassemia. Individuals with iron deficiency anemia or co-inheritance of beta- and alpha-thalassemia mutation in a double heterozygous state would demonstrate decreased or normal the level of HbA2 [6]. Thus, when couples have microcytic anemia and have a normal level of HbA2, particularly in individuals of SEA or Mediterranean ancestry, alpha-thalassemia carrier can be rationally suspected. Genetic testing for α-globin gene is essential to establish the diagnosis.

**Genetic testing**

The only certain way to determine the alpha-thalassemia genotype of a suspicious individual is genetic testing of the globin genes. Despite the test is laborious and costly, it can identify the number of loci
deleted, whether the deletions are in the cis (both copies deleted from one chromosome) or trans (one copy deleted from each chromosome) configuration in individuals with two loci deleted, and any mutations causing non-deletion forms of alpha-thalassemia. In general, Hb Bart’s hydrops fetalis is caused by deletion of all four α-globin genes but in rare cases, the disease can involve other forms of Hb variants. For example, HbH disease (-α/-α) is usually not lethal. When α-globin involves non-deletion variants (αα+/-), particularly in the mutations called Hb Constant Spring (HbCS) or Quong Sze (HbQS), the disease can escape being detected by PCR.

Bart’s hydrops fetalis is caused by deletion of all four α-globin genes causing non-deletion forms of alpha-thalassemia. In general, Hb Bart’s hydrops fetalis syndrome is delayed in 25% of affected pregnancies. Furthermore, by the combination of cardiothoracic ratio (CTR) and MCA-PSV, it tended to increase the sensitivity at GA 12-15 weeks (100%) and decreased the false positive rate at GA 16-20 weeks (0%). Hence, screening for Hb Bart’s hydrops fetalis by ultrasound alone may be delayed.

### Ultrasonic features

Once the pregnancy affected with Hb Bart’s hydrops fetalis or HbH disease progresses, several sonographic features would become apparent, including cardiomegaly, placental edema, and hydrops fetalis (skin edema, pleural effusion and ascites). However, in most of the time, not all these classical sonographic features would appear until the late second trimester [8]. In the first trimester, cardiac axis dilatation secondary to alpha-thalassemia major may take place initially. Lam and colleagues [9] described the predictive value of the cardiothoracic ratio for alpha-thalassemia major in 1997. It should be measured under the section of 4-chamber view; a standard view should show two groups of atrioventricular valves, one complete rib, and three strong echogenic foci of the spine. The maximum cardiac is measured at end diastolic phase at the level of annulus. The cardiothoracic ratio is calculated as the ratio of the cardiac diameter to the thoracic diameter (Figure 2) [10]. A cardiothoracic ratio greater than 0.5 before GA 17 weeks or 0.52 after GA 18 weeks is considered abnormal. Although some discrepancy existed among studies reported, all of them showed that using the cardiothoracic ratio as a predictor of alpha-thalassemia major had high sensitivity (66.7-100%) and specificity (92 -100%), whereas the false-positive rate (0-9.1%) was low.

For anemic fetuses resulting from Rhesus isoimmunization, anemia can be detected by Doppler ultrasonography on the basis of an increase in MCA-PSV. Mari et al. [11] reported that a MCA-PSV of greater than 1.5 MoM could be an indicator for screening moderate to severe fetal anemia with sensitivity nearly 100% (95% confidence interval, 0.86-1.0) and the false-positive rate of 12%. In 2009, Pretlove et al. [12] performed meta-analysis on the diagnostic value of MCA Doppler flow studies for fetal anemia. The sensitivity dropped to 75.5% and the specificity was 90.8% for detecting severe anemia. Although fetal MCA-PSV is considered the best noninvasive marker in Rhesus isoimmunization, there is no enough data demonstrating that MCA-PSV is as effective in predicting alpha-thalassemia major. Leung et al. [13] reported that using MCA-PSV as a parameter in prediction of alpha-thalassemia major during GA 16-20 weeks, the diagnosis may be delayed in 25% of affected pregnancies. Furthermore, by the combination of cardiothoracic ratio (CTR) and MCA-PSV, it tended to increase the sensitivity at GA 12-15 weeks (100%) and decreased the false positive rate at GA 16-20 weeks (0%). Hence, screening for Hb Bart’s hydrops fetalis ultrasonography alone may be a choice, despite not satisfactory, to at-risk patients who decline invasive procedures.

In addition to MCA-PSV, Leung et al. [14] presented that intrahepatic umbilical venous maximum flow velocity (IHUV Vmax) was also a useful parameter in diagnosis and management of alpha-thalassemia major fetuses, when their cases demonstrated the consistent declines in value after receiving intrauterine transfusion.

### Fetal therapy for hydrops fetalis

Intrauterine transfusion seems to prolong neonatal lifespan and minimize the complications. However, the procedure usually started after completion of organogenesis. The fetuses are undoubtedly at high risk for major neurodevelopment problems and congenital anomalies. Furthermore, lifetime transfusion therapy or stem cell transplantation is required after their birth. Hence, a careful and detailed non-directive genetic counseling should be offered at prenatal stage whenever termination of the pregnancy is still possible [15].

### Conclusion

Alpha-thalassemia is an old disease with available nationwide screening programs. However, it remains a challenging task to screen all the at-risk couples and make a definitive diagnosis in early pregnancy, even in a country like Taiwan having an excellent medical standard and a comprehensive nationwide screening program for more than 20 years. The presence of microcytic anemia is a major clue and thus microcytic anemia is part of the initial evaluation in routine prenatal and preconceptional screening. Nevertheless, there are still a few cases.

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**Table 1**: Comparison and limitations of current methodologies used to screen alpha-thalassemia, including hemoglobin (Hb) Bart's hydrops fetalis.

<table>
<thead>
<tr>
<th>Cell count</th>
<th>Maternal Hb analysis</th>
<th>Genetic testing</th>
<th>Ultrasound</th>
</tr>
</thead>
<tbody>
<tr>
<td>MCV&lt;80 fL or MCH&lt;25 pg/μL</td>
<td>Abnormal level of Hb pattern</td>
<td>Deletion or mutation on the loci of Hb Bart's hydrops fetalis</td>
<td>1. Hydrops fetalis (skin edema, pleural effusion and ascites)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>2. CTR ≥ 0.5 before GA 17 weeks</td>
<td>2. CTR ≥ 0.52 after GA 18 weeks</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3. CTR ≥ 0.52 after GA 18 weeks</td>
<td>4. MCA-PSV &gt; 1.5 MoM</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>5. IHUV Vmax &gt; 95 centile</td>
</tr>
</tbody>
</table>

**MCV**: Mean Corpuscular Volume; **MCH**: Mean Corpuscular Hemoglobin; **HbCS**: Hb Constant Spring; **HbQS**: Hb Quong Sze; **CTR**: Cardiothoracic Ratio; **GA**: Gestational Age; **MCA-PSV**: Peak Systolic Velocity of Middle Cerebral Artery; **MoM**: Multiples of Median; **IHUV**: Intrahepatic Umbilical Venous Maximum Flow Velocity.

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**Keywords**: Alpha-thalassemia, ultrasound, hemoglobin Bart's hydrops fetalis, prenatal screening.

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**Tools**

<table>
<thead>
<tr>
<th>Abnormal Finding</th>
<th>Pitfall</th>
</tr>
</thead>
<tbody>
<tr>
<td>MCV &lt; 80 fL or MCH &lt; 25 pg/μL</td>
<td>1. MCV may be normal for silent carrier (αα/α-).</td>
</tr>
<tr>
<td></td>
<td>2. Falsely normal when combination with iron deficiency anemia or beta-thalassemia.</td>
</tr>
<tr>
<td>Abnormal level of Hb pattern</td>
<td>The level of HbA2 is decreased or normal when iron deficiency anemia or co-inheritance of alpha- and beta-thalassemia occurred. Hb analysis may result in a false-negative diagnosis of beta-thalassemia.</td>
</tr>
<tr>
<td>Deletion or mutation on the loci of Hb Bart's hydrops fetalis</td>
<td>Rare non-deletion mutant alleles such as HbCS, HbQS, HbG-Taichung may escape being detected by PCR.</td>
</tr>
<tr>
<td>1. Hydrops fetalis (skin edema, pleural effusion and ascites)</td>
<td>1. The sonographic features usually not appear until the late second trimester.</td>
</tr>
<tr>
<td>2. CTR ≥ 0.5 before GA 17 weeks</td>
<td>2. The sensitivity of CTR is 66.7-100%. Most of the studies are conducted before GA 18 weeks.</td>
</tr>
<tr>
<td>3. CTR ≥ 0.52 after GA 18 weeks</td>
<td>3. MCA-PSV is more effective in prediction of moderate to severe anemic cases in Rhesus isoimmunization than alpha-thalassemia major. Consequently, diagnosis of Hb Bart’s hydrops fetalis by ultrasound alone may be delayed.</td>
</tr>
<tr>
<td>4. MCA-PSV &gt; 1.5 MoM</td>
<td>5. IHUV Vmax &gt; 95 centile</td>
</tr>
</tbody>
</table>

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**References**

of fetus with Hb Bart’s hydrops fetalis remained undiagnosed until late pregnancy. Nowadays, with the development of target sequencing of maternal plasma DNA, it is feasible to detect fetal monogenic disease by noninvasive prenatal diagnosis. Lam et al. [16] successfully applied the method to determine homozygous beta-thalassemia prenatally. In addition, preimplantation genetic diagnosis is also a feasible option for at-risk couples that do not accept a 1 in 4 risk of having an affected fetus and the subsequent suffering related to abortions and termination of pregnancy [17]. Physicians should understand the benefits and disadvantages and limitations of each screening test in order to offer appropriate preconception counseling and prenatal management.

References


Table 2: A multiplex PCR panel targeted to 10 common α-globin mutations is available for rapid alpha-thalassemia genotyping of Taiwanese patients.

<table>
<thead>
<tr>
<th>Mutation type</th>
<th>Mutant allele</th>
<th>HGVS nomenclature</th>
<th>Diagnostic method</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Deletions</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>α-TSEA</td>
<td>NG_000006.1:g.26264_45564del19301</td>
<td>GAP-PCR</td>
<td></td>
</tr>
<tr>
<td>α-TLE</td>
<td>NG_000006.1:g.1664_43534del31851</td>
<td>GAP-PCR</td>
<td></td>
</tr>
<tr>
<td>α-TIAN</td>
<td>NG_000006.1:g.1066_44164del33501</td>
<td>GAP-PCR</td>
<td></td>
</tr>
<tr>
<td>α-TMED</td>
<td>NG_000006.1:g.24664_41064del16401</td>
<td>GAP-PCR</td>
<td></td>
</tr>
<tr>
<td>α-T(−2.5)</td>
<td>NG_000006.1:g.15164_37864del22701</td>
<td>GAP-PCR</td>
<td></td>
</tr>
<tr>
<td>2. Non-deletion point mutations</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HbCS</td>
<td>HBA2:c.427T&gt;C</td>
<td>ARMS-PCR</td>
<td></td>
</tr>
<tr>
<td>HbQS</td>
<td>HBA2:c.377T&gt;C</td>
<td>ARMS-PCR</td>
<td></td>
</tr>
<tr>
<td>-αG-Taichung</td>
<td>HBA1:c.223G&gt;C</td>
<td>ARMS-PCR</td>
<td></td>
</tr>
</tbody>
</table>

HGVs: Human Genome Variation Society; ARMS: Amplification Refractory Mutation System.

Figure 2: The cardiothoracic ratio (CTR) is measured as the ratio of cardiac dimension (CD) to thoracic dimension (TD) at the level of 4 chamber view.