

## Marine Pollution: The Problematic of Microplastics

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Throughout the history of mankind and the conquest of the seas and oceans these water masses were always seen as unlimited sinks of wastes since they were assumed as being able to disperse, dilute and redistribute natural and synthetic substances. However, in the last few decades we have finally realized that this capacity is not unlimited [1,2]. According to literature about 6.4 millions of tons of litter are deposited in oceans and seas. Per year, there are about 8 millions of daily sewages together to 5 million tons of solid residues thrown into the marine environment by boats. Moreover, it was estimated that more than 13,000 plastic pieces are floating per each square kilometer of ocean [3]. The marine litter is a great and crescent environmental threat since it can be found in all oceans and seas even in remote places far away from obvious source of pollution. Marine litter can migrate long distances through oceans currents and winds being observed in marine and coastal environments, from poles to equator, from continental littorals to small remote islands. Islands completely made of litter already exist and the slow degradation process litter aggravates this problem [4].

The concern about the presence of plastics in marine environments comes from many years ago. Actually, in 1972 Carpenter et al. [5] warned about the increase of plastic production, which could lead to greater concentrations of plastics on sea surface. Only few months later it was reported the first case of plastics ingestion by fishes [5]. Nowadays, it is known that several million tons of plastics have been produced ever since [6,7,8] leading to the need to deal with this contamination, especially in oceans where they suffer degradation and fragmentation [6,8]. Their main sources are beach litter (contributing to about 80% of plastic debris), fishing industry (about 18%) and aquaculture [8,9]. Coastal tourism, recreational and commercial fishing and marine vessels may also be in the origin of plastic pollution [10]. Plastics debris migrates through the oceans being transformed into small fragments forming microplastics. Microplastics receive this designation due to their size smaller than 5 mm [11].

The impacts of microplastics still remains unclear nevertheless some conclusions and suspicions were already raised by recent studies. In fact, the evidences of exposure of several marine organisms enlarge, although being difficult to quantify such exposures and to establish dose/effect relationships required for setting risk limits (as PNEC values - predicted no effect concentrations), as we intended to demonstrate. Table 1 summarizes some of the studies that were performed mainly aimed in detecting exposures to microplastics and in evaluating resulting effects on different species, both under natural and laboratorial conditions. Some review papers also summarize much more information analyzing data with different points of view [12-14]. However, all the existing data suggests that for assessing the risks of microplastics, dose response curves have to be established under laboratorial conditions, and for being representative of field situations, such doses for different species have likely to be established at least based on the size, concentration of particles and on the chemical composition of the microplastics. These seems to be the most relevant factors determining their bioavailability,

chemical versus physical effects and potential for transference through trophic chains [12].

Nevertheless, the contamination of marine environments by microplastics may have other types of impacts, indirectly affecting organisms. The ingestion of microplastics by small animals may cause a decrease in food consumption due to satiation feeling and/or intestinal blockage leading to death [15]. These compounds can accumulate in the gut of filter-feeding mussels persisting for more than 48 day [16].

It is known, for example, that the presence of small plastics debris in beach sand slows the heating of the sediments [17]. The resulting reduction in temperature of sand can impact organisms with temperature-depending sex-determination, like turtles that can be affected even by a low concentration of plastic (1.5%) [18]. Further, the permeability of the sand increases with the presence of microplastics [17]. This change combined with grain size increase and desiccation stress could negatively affect the embryonic development of eggs of several organisms like crustaceans [19], mollusks [20], polychaetes [21] and fishes [22]. Permeability increase also leads to a change in trace element cycling in beach sediments. When sands have a higher permeability more water is flushed through the beaches giving more oxygen and organic matter to the small interstitial organisms. More oxygen and organic matter causes an increase in the abundance of such organisms, which, in turn, will release a higher amount of compounds resulting from their metabolism in water. More metabolites lead to changes in oxygen gradients and redox conditions impacting those environments [17,23].

Besides all the efforts applied at regional, national and international levels, marine litter continues to increase. Delays in the application and fulfillment of the already existent regulations, or either the lack of supervision or of specific regulations in several parts of the world are contributing for such increasing problem. More awareness and outreaching activities to general public are also required, to promote new behaviors related with plastics use and disposal. Such actions are of particular importance, since the effects of marine pollution

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Local	Species	Goals	Main results	References
Laboratorial exposure	<i>Lytechinus variegatus</i> (sea urchin)	Compare the effects of plastic pellets (virgin and beach stranded) on <i>Lytechinus variegatus</i> embryos development.	A 58.1 and 66.5% increase of anomalies in embryonic development were recorded for beach stranded and virgin pellets, respectively. The pellets were tested in a proportion of 1:4 (pellet:seawater)	[24]
Laboratorial exposure	<i>Mytilus edulis</i> (mussel)	Assess the uptake and translocation of microplastics (10-30-90 mm) under laboratorial conditions and the effects on energy metabolism	Organisms exposed to a high concentration of polystyrene microspheres (110 particles/mL-1 sea water). Microplastics were present in all organisms collected ( $0.2 \pm 0.3$ particles/g body weight). Ingestion and translocation of microplastics in the gut did not affect the cellular energy allocation.	[25]
Laboratorial exposure		Assess effects of polyethylene ingestion at cellular and subcellular levels.	After intake of particles with 0-80 $\mu\text{m}$ , the following effects were observed: strong inflammatory response; granulocytomas formation after lysosomal membrane destabilization in connective tissue of digestive gland. Microplastic uptake into the gills and stomach with transport to digestive gland where they accumulated in lysosomal system in 3 h.	[26]
Laboratorial exposure		Assess the effects of 30 nm polystyrene particles (0, 0.1, 0.2, and 0.3 g/L) on the feeding behavior.	Filtering activity was reduced in presence of polystyrene. Production of pseudo-feces when exposed to 0.1 g/L. The polystyrene was recognized as a low nutritional food by mussels.	[27]
Laboratorial exposure		Evaluation of the Ingestion, translocation and accumulation of microplastics debris (3.0 or 9.6 $\mu\text{m}$ ).	Microplastics accumulation in gut. Microplastics capture in hemolymph. Microplastics translocation from gut to circulatory system during 48 days.	[28]
Laboratorial exposure		Assess the presence of microplastics in soft tissues (whole body except the shell).	$0.36 \pm 0.07$ particles/g (wet weight).	[29]
Laboratorial exposure	<i>Mytilus galloprovincialis</i> (mussel)	Evaluate the effects of pyrene in presence of polyethylene and polystyrene microplastics.	Presence of microplastics in hemolymph, gills and in digestive glands. Microplastics caused DNA strand breaks in haemocytes at 20 g/L. Pyrene effects were emphasized by microplastics because they adsorb pyrene increasing its uptake and bioavailability.	[30]
Laboratorial exposure	<i>Crassostrea gigas</i> (oyster)	Assess the presence of microplastics in soft tissues (whole body except the shell).	$0.47 \pm 0.16$ particles/g (wet weight).	[29]
Laboratorial exposure	<i>Arenicola marina</i> (annelid)	Assess the uptake and translocation of microplastics (10-30-90 mm) under laboratorial conditions and the effects on energy metabolism	Organisms exposed to a high concentration of polystyrene microspheres (110 particles/g-1 sediment). Microplastics were present in all organisms collected in the field; on average $1.2 \pm 2.8$ particles/g body weight. Ingestion and translocation of microplastics in the gut did not affect the cellular energy allocation.	[25]
Laboratorial exposure	<i>Arenicola marina</i> (annelid)	Assess the bioaccumulation of polystyrene and polychlorinated biphenyl.	A low polystyrene dose increased bioaccumulation of PCBs by a factor of 1.1-3.6. Polyesterene did not accumulate in <i>A. marina</i> but it can be ingested by its predators while in the gut of <i>A. marina</i> .	[31]

Laboratorial exposure		Evaluation of the effects of microscopic unplastified polyvinylchloride (UPVC)	Energy reserves depletion after a chronic exposure to a dose of UPVC corresponding to 5% of sediment weight. Accumulation of UPVC in longer gut and inflammation with an enhanced phagocytic response after a chronic exposure.	[32]
Laboratorial exposure	Pomatoschistus microps (common goby fish)	Assess the predatory behavior of juveniles in the presence of microplastics.	Microplastics (420-500 µm size) were ingested suggesting confusion with food. Such confusion was dependent from the color of the microplastics and from the conditions of the fish juveniles.	[33]
Laboratorial exposure	Pomatoschistus microps (common goby fish)	Assess the Influence of microplastics on chromium toxicity in juveniles.	In presence of microplastics (0.216 mg/L), chromium (1.8 – 28.4 mg/L inhibited acetylcholinesterase activity.	[34]
North Western Mediterranean basin	Zooplankton	Evaluation of the ratio of microplastic to zooplankton in neustonic waters collected in 40 sampling stations	Presence of microplastics of different types (filaments, polystyrene, thin plastic films) in 90% of the sampling stations with sizes ranging 0.3-0.5 mm and an average weight of 1.81 mg/particle. A ratio of 1:5 (microplastic/zooplankton) was recorded in neustonic water samples thus representing a high risk to filter feeding organisms.	[35]
Cepola macrophthalma (bandfish)	Southwest of Plymouth, United Kingdom	Assessment of plastic ingestion. (The study documents microplastics in 10 species of fish from the English Channel.)	Microplastics ingestion (<40 pieces/particles). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts.'	[36]
Southwest of Plymouth, United Kingdom	Callionymus lyra (common dragonet fish)	Assessment of plastic ingestion. (The study documents microplastics in 10 species of fish from the English Channel.)	Microplastics ingestion (< 50 pieces/particles). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts.'	[36]
Southwest of Plymouth, United Kingdom	Buglossisium luteum (yellow sole)	Assessment of plastic ingestion. (The study documents microplastics in 10 species of fish from the English Channel.)	Microplastics ingestion (< 20 pieces/particles). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts).'	[36]
Southwest of Plymouth, United Kingdom	Microchirus variegatus (sole)	Assessment of plastic ingestion. (The study documents microplastics in 10 species of fish from the English Channel.)	Microplastics ingestion (< 20 pieces/particles). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts).'	[36]
Southwest of Plymouth, United Kingdom	Aspitrigla cuculus (red gurnard fish)	Assessment of plastics ingestion. (The study documents microplastics in 10 species of fish from the English Channel).	Microplastics ingestion (< 70 pieces). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts).'	[36]
Mediterranean Sea (Pelagos Sanctuary)	Balaenoptera physalus (fin whale)	Detection of MP and phthalates in surface neustonic/planktonic samples. Detection of phthalates in stranded fin whales.	56% of the surface neustonic/planktonic samples contained microplastic particles. Portofino MPA (Ligurian Sea) with the highest abundance of microplastics (9.67 items/m <sup>3</sup> ). High concentrations of phthalates (1.00 – 4.32 ng/g fw) were detected in the neustonic/planktonic samples. Phthalates were in bubbler of stranded fin whales suggesting that they can be used as a tracer of the intake of microplastics.	[37]

with microplastics still few evident for the society, thus resulting, for example, in misinterpretations of the taxes applied to plastic bags. Nevertheless, there is still hope that, as it happened with other

dangerous contaminants such as tributyltin [23], the legislation will contribute to prevent the catastrophe presently envisaged to the marine environment due to pollution with microplastics.

Mediterranean Sea		Evaluation of phthalate levels in this species.	Presence of phthalates in bubbler (1.48 – 377.82 ng/g lipid basis). This species can be a potential bioindicator of the presence of microplastics in pelagic environments.	[38]
Ireland	Mesoplodon mirus (beaked whale)	Evaluation of exposures through the analysis of stomach and gut contents.	Presence of microplastics in stomachs. Top oceanic predatory species are exposed to plastics; exposure pathways still unclear.	[39]
Mediterranean Sea	Cetorhinus maximus (basking shark)	Evaluation of the exposure to phthalates.	High concentrations of phthalates in muscle (11.17 – 156.67 ng/g lipid basis). This species can be a potential bioindicator of microplastics in pelagic environments.	[38]
Southwest of Plymouth, United Kingdom	Merlangius merlangus (whiting fish)	Assessment of plastic ingestion. (Study documents microplastics in 10 species of fish from the English Channel).	Microplastics ingestion (< 30 pieces/particles). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts.	[36]
Southwest of Plymouth, United Kingdom	Micromesistius poutassou (blue whiting fish)	Assessment of plastic ingestion. (The study documents microplastics in 10 species of fish from the English Channel.)	Microplastics ingestion (~ 30 pieces). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts.	[36]
Southwest of Plymouth, United Kingdom	Trisopterus minutus (poor cod fish)	Assessment of plastic ingestion. (The study documents microplastics in 10 species of fish from the English Channel.)	Microplastics ingestion (~ 40 pieces/particles). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts.	[36]
Central Mediterranean Sea	Xiphias gladius (swordfish), Thunnus alalunga (tuna albacore) and Thunnus thynnus (tuna fish)	Evaluation of the presence of plastic debris in stomach.	Microplastics ingestion: 29 particles were found in the stomach of 22 fish. Plastic fragments with different colors and shapes. Swordfish: dominance of mesoplastics (44.4%); Albacore: dominance of microplastics (75%); Tuna fish: meso and macroplastics ingested in the same proportion. A relation between fish size and plastic size was found.	[40]
Southwest of Plymouth, United Kingdom	Zeus faber (fish)	Assessment of plastics ingestion. (The study documents microplastics in 10 species of fish from the English Channel.)	Microplastics ingestion (< 60 pieces/particles). Presence of polyamide, semi-synthetic cellulosic material and rayon in gastrointestinal tracts.	[36]

**Table 1:** Collection of some microplastics exposure and effects in animal species under both natural and laboratorial conditions.

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