Neuronal and Glial Growth in Organotypic Cultures after Vitrification

Amir Arav1*, Abraham Shahar2, Ofra Yale, Patrizio Ziv-Polat1, Yehudit Natan1 and Pasquale Patrizio1

1FertileSafe, Ltd. 11 Haharash Street Ness Ziona, Israel
2NVR 11 Haharash Street, Ness Zonia, Israel
3Yale University Fertility Center, New Haven, CT, USA

Abstract

Fetal rat dorsal root ganglia (DRG) and spinal cords (SC) slices from rat fetuses were vitrified in a new semi-automatic vitrification system, cooled in sterile slush liquid air (SLA) and stored in a special sterile sealed container in liquid nitrogen (LN). Upon warming, organotypic stationary cultures were performed using NVR-Gel (composed mainly from hyaluronic acid and laminin) and enriched with neuronal factors conjugated to iron oxide nanoparticles. Evaluation of cultures was made by daily phase-contrast microscopy observations and by immunofluorescence staining.

Results revealed that SC neurons maintained their multipolar shape and regrew dendrites and axons. The round shape DRG neurons exhibited euchromatic nuclei with prominent nucleoli and an active regeneration of nerve processes. Migration of both neurons and flat cells (fibroblasts and glia Schwann cells) started within 48 hours after seeding and intensified in the upcoming days.

In conclusion, it can be said that using a semi-automatic vitrification, sterile vitrification and sterile storage of neuronal tissues from the CNS and the PNS is a successful advanced technology for the preservation of neurons and glial cells, as shown in the regrown of a full regular growth pattern in culture. This may be an important step towards clinical use in the reconstruction of severe peripheral nerves and spinal cord injuries.

Keywords: Cryopreservation; Vitrification; Neurons; Spinal cord; Contamination; Regeneration


Introduction

Vitrification is an ice free cryopreservation method which includes a step wise exposure of the sample to high concentrations of cryoprotectants (i.e. DMSO and Ethylene glycol) followed by rapid cooling to the temperature of liquid nitrogen [1-4]. As a result, the human factor is one of the most important variables affecting the success of cryopreservation procedures by vitrification [4,5]. Furthermore, cryopreservation of stem cells and progeny tissue by direct exposure into liquid nitrogen (LN) and their storage in standard LN tanks encompass in it the risk of potential contamination and cross-contamination by viruses, bacteria, fungi and spores that survive in LN and pose a real threat [6-8].

Therefore, the use of safe cryopreservation protocols is important and highly needed for preventing contamination of biological samples by cryogenic medium and safeguard future stem cell therapies [9,10].

In order to simplify the vitrification process and to overcome these potential risks of contamination we developed three devices; (1) A semi-automatic system that allows a step wise exposure to vitrification solutions and cooling within the same device (Sarah®, FertileSafe, Nes-Ziona, Israel). (2) A device for producing clean liquid air having the same temperature and properties as LN (CLAir®, FertileSafe, Nes-Ziona, Israel). (3) A sterile storage device which enables storing samples in a sterile manner within the standard LN tanks, thus preserving the LN temperature and insulating the samples from being in contact with the surroundings (Esther®, FertileSafe, Nes-Ziona, Israel).

Materials and Methods

Preparation of organotypic DRG and SC cultures

All the experiments were carried out and authorized by the local ethics committee for animal experimentation. Stationary organotypic DRG and spinal cord SC cultures were prepared from rat fetuses (gestational day 15, Lewis inbred, Harlan, Israel). Immediately after dissection, the isolated DRG and SC slices were cut with a McIlwain tissue chopper (Ted-Pella, California, USA) into small slices (of 400 μm thickness). At a desired time, tissue were warmed and seeded in 12 well culture plates containing 1 mL of NVR-Gel (NVR laboratories, Nes-Ziona, Israel). Cultures were enriched with glial cell derived neurotrophic factor (GDNF) conjugated iron oxide nanoparticles (10 μg/mL final concentration). Once embedded and seeded in the gel, cultures were subsequently fed with the nutrient medium every 4 days. Monitoring of the DRG-SC growth pattern was done by daily phase contrast microscopic observations starting 24 hours after setting the cultures onward. Immunofluorescence staining of cultures was made at different stages of cultivation.

*Corresponding author: Amir Arav, Fertile Safe, Ltd. 11 Haharash street Ness Zonia, Israel. Tel: 3568103636; E-mail: dty2@netvision.net.il

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Culture medium

The culture medium was prepared from 90% Dulbecco's modified eagle medium-nutrient mixture F-12 (DMEM-F12), 10% heat-denatured fetal calf serum (FCS), 6 g/L D-glucose, 2 mM glutamine, 25 µg/mL gentamycin, and 50 ng/mL IGF-I (all purchased from Biological Industries, Bet-Haemek, Israel).

NVR-Gel for neuronal cultivation

NVR-Gel (NVR Laboratories, Nes-Ziona, Israel) is a proprietary gel composed of two main components: high molecular hyaluronic acid (HA, 3×10^6 Da) and laminin, both known to be inherent elements of the extracellular matrix. For cell cultivation, HA of 1% (Bio-Technology General, Kiryat Malachi, Israel) was diluted with culture medium to the concentrations of 0.3–0.5%. Laminin (Sigma, St. Louis, Missouri, USA) was then mixed with the diluted HA (final concentration of 10 µg/mL). GDNF, free or conjugated to iron oxide nanoparticles, was finally added (10 ng/mL), to complete the NVR-Gel composition.

Immunofluorescence staining

Samples were fixed in 4% paraformaldehyde. The fixed cells were permeabilized with 0.1% of Triton X-100 in PBS and then immunoblocked with a 1% bovine serum albumin (BSA) in PBS for 1 hour at room temperature. The specimens were then double stained with mouse anti S-100 antibodies (1:80, Acris Antibodies, glial cell marker) and rabbit anti neurofilament antibodies (NF, Novus Biologicals, 1: 500, neuronal cell marker). The primary antibodies were diluted in 0.1% BSA and 0.05% Tween-20 in PBS (diluents buffer) and incubated with the specimens overnight at 4°C. After rinsing with 0.05% Tween-20 in PBS (wash buffer), the DRG specimens were incubated for 1 h at room temperature with the appropriate fluorescently labeled secondary antibodies.

Vitrification, storage and warming methods

A new device for semi-automatic vitrification was used (Sarah, Fertile Safe, Ness Ziona, Israel). The device is equipped with a special capsule containing electron microscope (EM) gold grid connected to 0.25ml IMV straw (Figure 1) (IMV Technologies, L’Aigle, France). Six straws, each containing 6 tissue slices, were placed in a special mixing chamber (Figure 2). An increasing concentration of vitrification solution was continuously infused using a syringe pump (New Era Pump Systems, NY, USA) for 25 minutes with equilibration solution (ES) (Sage® Vitrification kit, Origio, Malov, Denmark). Then vitrification solution (VS) (Sage® Vitrification kit, Origio, Malov, Denmark) was flowed through the samples for 5 minutes. Finally samples were plunged into SLA (-208°C) (Clair® and VitMaster®, FertileSafe, Nes-Ziona, Israel). The samples were stored in a special sterile sealed container (Esther®, FertileSafe, Nes-Ziona, Israel) for 1 week. For the warming, straws were held for 10 second in air before being plunged into 80°C warming solution (1M sucrose) for 1 second and immediately placed in 37°C warming solution (1M sucrose) for 2.5 minutes. Then samples were moved into decreased sucrose concentrations (0.5M & 0.25M for 2.5 minutes in each solution) (Sage® vitrification warming kit, Origio, Malov, Denmark).

Results

The new semi-automatic vitrification device allows gradual increase of the vitrification solution which permits both standardization of the vitrification process and reduction of the tissue damages generally seen during the vitrification procedure. This device enables to vitrify up to 6 straws and 36 slices (6x6 tissue slices) simultaneously, thus increasing the efficiency of the process. The exposure of neuronal tissue slices to the sterile SLA (-208°C) enabled vitrification and did not damage the neurons and glia cells as seen by post warming growth. Furthermore, both neuronal cell types maintained their multipolar and round shapes (Figure 3B,C) and exhibited intensive fiber regeneration. DRG neurons exhibited one or two euchromatic nuclei with prominent nucleoli (Figure 3C). Several nerve cells showed regeneration of short nerve processes and a thin long process which might be probably the axon (Figure 3A). The flat glia and fibroblast cells migrated from the explants and divided actively (Figure 4A,B). Migration from explants of both neurons and flat cells (fibroblasts and glia Schwann cells) started within 48 hours after seeding and intensified in the upcoming days.
neuronal factors. Abraham Shahar dissected the fetal SCs and DRGs and analyzed the interpretation of the data with Arav Amir. Natan Yehudit, Arav Amir and Patrizio Pasquale wrote the manuscript.

References