

# Nutritional and Bioactive Compounds: Evaluation of *Pleurotus pulmonarius* (Fries) Quel. Fruit Bodies Grown on Different Wood Logs in Abia State, Nigeria

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## Abstract

This study was conducted to determine the nutritional and bioactive compounds composition of *Pleurotus pulmonarius* fruit bodies cultivated on tree logs of *Dacryodes edulis*, *Mangifera indica* and *Treculia africana*. Pure mycelium culture of *P. pulmonarius* was aseptically bulked in sorghum grains. Logs were cut into average length of 18 cm with inoculation holes of 3 cm × 15 mm diameter; using high speed drill (HSD) of 5 drill bit and allowed to decompose for 8 months. During mushroom cultivation, logs were soaked in tap water for 24 hrs and pasteurized at 80°C in an improvised metallic drum (IMD) for 1 hour; using cooking gas as heat source and allowed to cool overnight. 10 g of grain based spawn was inserted into 2/3 of each hole by way of inoculation and sealed with sterile polybag for mycelium incubation. Polybags were cut open after spawn run following primordial initiation. Fruit bodies were harvested at maturity, sundried ground and packed in airtight container prior to further analysis. Data was analysed using Analysis of Variance (ANOVA) and mean separation by Duncan Multiple Range Test (DMRT) while levels of significance were determined at 5%. Results indicate that *P. pulmonarius* fruit bodies harvested from various tree logs were significantly different  $p < 0.05$  in their nutritional and bioactive compounds composition. Fruit body samples were rich in protein, carbohydrates, Na, K, and Ca. It was also observed that fruit bodies contained significant amount of Alkaloids, Tannins and Saponins; and could be useful in drug synthesis. Therefore adopting this technique in oyster mushroom cultivation would lead to more jobs creation and food security; but this must be done with careful regulations to avoid indiscriminate falling of trees.

**Keywords:** *Pleurotus pulmonarius*; Logs; Nutrients; Bioactive; Fruit bodies

## Introduction

Mushrooms are unique biota which assemble their food by degrading enzymes and decompose the complex food materials present in the biomass where they grow [1]. Oyster mushrooms can be grown on various substrates due to its strong enzymatic features. Different substrates are used in each region depending on their availability [2]. Wheat straw, sawdust and other agricultural by-products resulting after processing of waste paper, Hazelnut and Tilia have been used in Oyster mushroom cultivation [3]; maize, corn, rice, elephant grass [4], sugarcane [5], coffee [6] have been examined as alternative substrates for its cultivation. These substrate materials are usually by products from industries, households, agriculture etc., and are usually considered as wastes [7]. However, these wastes are actually resources in the wrong place at a particular time and mushroom cultivation can harness them for its own benefit [8].

Kadiri and Aizai [9] showed that *Lentinus subnudus* could be cultivated on wood logs of tropical trees. According to Hyunjong and Seung [10], hard woods such as poplar, willow, beech, elm and alder are the most commonly used tree species in oyster mushroom cultivation. He noted that unlike shiitake, Oyster mushrooms do not grow well on Oak tree logs. Hyunjong and Seung [10] reported that since mushroom feed primarily on sapwood, any tree trunk selected for inoculation must have a larger sap wood area. The lighter or outermost wood of a log is the sapwood and the darker or inner wood is the heartwood.

The desirability of a food product does not necessarily bear any correlation to its nutritional values instead, its appearance, taste and aroma, sometimes can stimulate one's appetite [8]. Mushroom has been used as a food and medicine by different civilizations since immemorial

time, due to its delicious taste and dietetic qualities [11,12]. Mushrooms are also known for their medicinal properties; they are low in calories and are ideal food for diabetic and heart patients. Mushroom has qualities like lowering the blood cholesterol level, warding against cancer and invigorating hair growth. Tewari [13] reported that the fresh mushroom contains about 85-90% moisture, 3% protein, 4% carbohydrates, 0.3-0.4% fats and 1% minerals and vitamins. *Pleurotus* species are good source of protein, vitamins and minerals [8,14]. Mushroom protein is intermediate between that of animals and vegetables, but superior to most other foods, including milk and contains all the essential amino acids required by man [15-17]. Mushrooms contain appreciable quantities of crude fibres although little information exists on the total dietary fibre (TDF) content of mushrooms. Okwulehie et al. [7] reported high crude protein and carbohydrate contents in *P. ostreatus* cultivated on different substrates.

The world production of oyster mushroom is estimated to be 875,000 tons in 1997 [18]. China was responsible for 87% of world supply, oyster mushroom is the easiest to produce and least expensive to grow. Most of the world's supply of oyster mushrooms today comes

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from commercial mushroom growers [18]. For small-scale cultivation with limited budget, oyster mushroom is the clear choice for gaining entry into the mushroom industry [12]. This work aims to determine the nutritional and bioactive compounds composition of *P. pulmonarius* cultivated on tree wood logs.

## Materials and Methods

### Source of culture

Pure culture of *P. pulmonarius* (Fries) Quel. Was obtained from the laboratory of the department of plant science and biotechnology, Michael Okpara University of Agriculture, Umudike, Abia State, Nigeria.

### Spawn production

Spawn of *P. pulmonarius* was produced using sorghum grains. Grains were washed in tap water and soaked overnight. They were then boiled in water in the ratio of 1:1 (sorghum grain: water) using cooking gas for 1520 mins and drained of excess water. Completely drained sorghum grains were mixed with 4% (w/w)  $\text{CaCO}_3$  and 2% (w/w)  $\text{CaSO}_4$  to optimize pH and prevent clumping of grains respectively as described by Muhammad et al. [12]. Grains were later stuffed into 35 cl Lucozade bottles tightly plugged with cotton wool and sterilized in an autoclave at  $121^\circ\text{C}$  for 30 mins. After sterilization, the bottles were allowed to cool, before they were inoculated with actively growing mycelium of *P. pulmonarius* by grain-to-grain transfer and incubated in the dark (at  $27 \pm 2^\circ\text{C}$ ) for 10-15 days until the grains were fully colonized by mycelium [19].

### Preparation of wood logs (substrates)

Average trees size of *T. africana*, *M. indica* and *D. edulis* were fell during the Hammattern season (winter) according to the recommendations of Oei [20]. Trees were cut into logs of 18 cm using Electric wood saw (EWS); Model: Elect. 1710, Japan. Care was taken to ensure that the barks of the logs were not peeled off as instructed by Hyunjong and Seung [20].

### Inoculation holes

Holes of depth 3 cm by 15 mm diameter were made hexagonally on each log with high speed drills (HSD) of 5 drill bit in respect to log size. Average number of holes per log was determined by the formula, according to Stamets [21], Nwoko [22].

$$NH = \frac{DL(cm) \times LL(cm)}{6}$$

Where: NH=Number of holes,

DL=Diameter of log (cm),

LL=Length of Log (cm),

6=Derived constant.

### Mushroom Cultivation

Logs were laid in open field for 8-9months in alternating rains and sun to allow for decomposition. Dry weight of logs (g/kg) were determined before they were soaked in water for 24 hr. Logs were pasteurized at  $80^\circ\text{C}$  in an improvised metallic drum (IMD) for 1 hr using cooking gas as a local heat source and allowed to cool overnight, as recommended by Canford [23], Nwoko et al. [24].

Log inoculation was done by inserting about 10 g grain spawn of

*P. pulmonarius* into 2/3 of the holes and subsequently sealing the logs with transparent polybags to avoid contaminants. Mycelium recovery and colonization were clearly visible after 24 hrs; when fully colonized polythene bags were cut open to allow for fruiting [10]. Before pinhead initiation, white mycelium was visibly noticed on the cut ends of the logs. Light intensity and humidity of the air were increased to about 400 lux and 75% respectively. To achieve these, logs were watered at least morning and evening and t cropping room of the mushroom house was flooded with water. Temperature was maintained at  $27 \pm 2^\circ\text{C}$  [20,25]. Pinheads of *P. pulmonarius* were first noticed in *D. edulis* logs followed by *T. africana* and then *M. indica* logs after 9, 10 and 12 days of inoculation respectively. Mushrooms were harvested as soon as the fruit-bodies were fully matured [26].

### Proximate analysis

Proximate analysis was carried out on each of the 3 mushroom samples. Nutrients like carbohydrates, protein, fat ash, moisture and crude fiber contents were determined by using the methods outlined in the AOAC [27]. Protein determination was carried out using the Kjeldahl method [27]. Fat determination was carried out using a Soxhlet apparatus [27]. Also, determination of fiber content was done according to the enzymatic gravimetric method [27].

### Determination of minerals

Mineral compositions of dried mushroom samples were determined by wet-ashing method. The solutions of ash obtained from the samples were dissolved in a drop of trioxonitrate (V) acid made up to 50 ml with deionized water and analyzed for Calcium (Ca) and Magnesium (Mg) using vanadate ethyldiamine-tetra acetic acid (EDTA) complexometric titration method according to MFA [28]. Sodium (Na) and Potassium (K) were estimated using flame photometer while Phosphorus (P) was determined using UV-visible spectrometer after making Ammonium vanado-molybdate at 436 nm according to the established procedures of Perkin Elmer [29].

### Determination of percentage of bioactive compounds

Percentage Alkaloids were determined by the methods of AOAC [30] and Maxwell et al. [31]. Percentage Flavonoids, Saponins and Tannins were also determined by the procedures according to Cloupai-Abyazini, Peng and Kobayashi [32] while percentage Phenols were estimated by the method of Harborn [33].

### Statistical analysis

The data obtained were statistically analyzed using Analysis of Variance (ANOVA) mean separation and tests of significance were carried out by Duncan Multiple Range Test (DMRT) at  $p < 0.05$  [34].

## Results and Discussion

Results and discussion of the work on the nutritional and bioactive compounds evaluation of *P. pulmonarius* fruit bodies grown on different wood logs are presented below.

Table 1 shows the proximate composition of *P. pulmonarius* as affected by different log substrates. The results of the moisture, ash, fat, fibre, protein, carbohydrate, dry matter and free nitrogen contents of *P. pulmonarius* fruit bodies cultivated on the *D. edulis*, *M. indica* and *T. africana* are significantly different  $p < 0.05$ . This shows that the mushroom is highly nutritious when grown on these logs. This also indicates the major reason why oyster mushrooms grow naturally on already degrading logs in the wild and sometimes, around homes [8,20].

Log Substrate	MC	ASH	Fat	Fibre	Protein	CHO	DM	N <sub>2</sub>
<i>D. edulis</i>	2.63 <sup>c</sup>	9.46 <sup>a</sup>	2.69 <sup>a</sup>	6.15 <sup>c</sup>	37.17 <sup>b</sup>	41.91 <sup>c</sup>	97.38 <sup>a</sup>	5.95 <sup>b</sup>
<i>M. indica</i>	3.12 <sup>a</sup>	7.0 <sup>c</sup>	2.56 <sup>c</sup>	2.29 <sup>a</sup>	37.86 <sup>a</sup>	43.11 <sup>a</sup>	96.88 <sup>c</sup>	6.06 <sup>a</sup>
<i>T. africana</i>	2.81 <sup>b</sup>	8.48 <sup>b</sup>	2.59 <sup>b</sup>	6.24 <sup>b</sup>	37.68 <sup>a</sup>	42.21 <sup>b</sup>	97.19 <sup>b</sup>	6.03 <sup>a</sup>

Values are means of 3 replicates and values bearing the same letter are not significantly different (P>0.05). MC: Moisture Content, CHO: Carbohydrate,

DM: Dry Matter

**Table 1:** Effect of log substrates on proximate composition (%) of *P. pulmonarius* fruit bodies.

The relative high percentage of dry matter, carbohydrate and protein in the mushroom fruit bodies cultivated on the log substrates conforms to the work of Marlow and Ukoima et al. [35]. The high protein contents of the *P. pulmonarius* fruit bodies cultivated on the various logs confirms the assertion by several workers that mushroom protein is intermediate between that of animals and vegetables, but superior to most other foods, including milk and contains all the nine essential amino acids required by man [15-17]. Low fat content of the mushroom shows that the mushroom could be good for people with cardiac problems. This is in line with the reports of Okhuoya and Okigbo [36], Okwulehie and Odunze [23], who maintained that mushrooms generally contain low-oil and fat, and because of the low content of oil and fat in mushrooms, they are recommended as good supplements for patients with cardiac problems.

Table 2 represents the results of minerals compositions of *P. pulmonarius* grown on various logs. The results showed that the mushroom samples were significantly p<0.05 rich in Sodium, Potassium, Magnesium, Calcium and Phosphorus. Potassium and Phosphorus contents were higher than other minerals analyzed and also higher in mushrooms harvested from *D. edulis* logs. In this study, Sodium was found to be the lowest among other minerals analyzed in the mushroom across all log substrates. The low Sodium content in mushrooms makes them ideal for persons with certain types of heart and kidney ailments [37].

The rich minerals contents in *P. pulmonarius* fruit bodies grown on the logs as observed in this study could be because the mushroom effectively utilized the high amount of nutrients present in the sapwood as reported by Hyunjong and Seung [10]. These mineral values are higher than those reported by Ogbo and Okhuoya [38]; Okwulehie et al. [39], Okoi and Iboh [40]. *D. edulis* gave the highest constituents of all the mineral nutrients analysed while *M. indica* gave the lowest. The observed appreciable quantities of various mineral elements analysed in the three mushroom samples indicates that these logs contain the corresponding nutrients in a relative amount since the nutritional composition of mushrooms depends on the substrate where they were grown [8].

Bioactive constituents of *P. pulmonarius* as affected by different log substrates are shown in Table 3. Results show that Alkaloids, flavonoides phenols, tannins and saponins were significantly different p<0.05 at different quantities. Alkaloids were found in higher quantity than other bioactive compounds analysed. Alkaloids have powerful effect in animal physiology and are important in pharmaceutical industries for drug manufacturing [41]. Rambeli and Menini [42] reported that alkaloids are stimulants and acts by prolonging the action of several hormones. Flavonoids, phenols, tannins and saponins concentrations in *P. pulmonarius* fruit bodies cultivated on the different trees logs were higher than those reported by Okwulehie et al. [39]. Flavonoids act as anti-carcinogens, anti-bacterials [43], saponins and are implicated in the prevention of parasitic fungal diseases [44] while

Log substrate	Na	K	Mg	Ca	P
<i>D. edulis</i>	15.82 <sup>a</sup>	172.23 <sup>a</sup>	17.28 <sup>a</sup>	127.40 <sup>a</sup>	33.23 <sup>a</sup>
<i>M. indica</i>	14.94 <sup>b</sup>	171.18 <sup>b</sup>	16.52 <sup>c</sup>	126.46 <sup>c</sup>	32.16 <sup>c</sup>
<i>T. africana</i>	15.26 <sup>c</sup>	171.67 <sup>c</sup>	16.80 <sup>b</sup>	126.79 <sup>b</sup>	32.76 <sup>b</sup>

Values are means of 3 replicates and means bearing the same letter are not significantly different (P>0.05).

**Table 2:** Mineral constituents (mg/100g) of *P. pulmonarius* fruit bodies as affected by different log substrates.

Log substrate	Alkaloids	Flavonoids	Phenols	Tannins	Saponins
<i>D. edulis</i>	4.07 <sup>c</sup>	0.18 <sup>c</sup>	0.94 <sup>b</sup>	1.62 <sup>b</sup>	2.52 <sup>b</sup>
<i>M. indica</i>	4.16 <sup>b</sup>	0.21 <sup>b</sup>	0.93 <sup>b</sup>	1.52 <sup>c</sup>	2.55 <sup>b</sup>
<i>T. africana</i>	4.34 <sup>a</sup>	0.26 <sup>a</sup>	1.05 <sup>a</sup>	1.74 <sup>a</sup>	2.63 <sup>a</sup>

Values are means of 3 replicates and means bearing the same letter are not significant at (P>0.05).

**Table 3:** Effect of wood logs on bioactive compounds composition (%) of *P. pulmonarius* fruit bodies.

tannins have been used as anti-tumor agents and perform a wide range of anti-infective actions [45]. The high concentrations of these important bioactive compounds in the fruit bodies of *P. pulmonarius* with respect to their various log substrates indicate that the trees may also contain the compounds in high amount. This also shows that these mushroom samples may be considered useful in the production of certain pharmaceutical chemicals [39]. The high concentrations of these compounds may also contribute to their taste, aroma and flavour, thereby increasing their nutritional, medicinal and food value.

## Conclusion and Recommendations

*Pleurotus pulmonarius* fruit bodies were successfully cultivated on the logs of *D. edulis*, *M. indica* and *T. africana*. Nutritional and bioactive compounds analysis of fruit bodies from different log substrates showed that they were rich in nutrients and could be of high pharmaceutical importance. Therefore, efforts should be made to determine the composition of other nutrients such as vitamins and amino acids of *P. pulmonarius* with respect to the same log substrates. Commercialization of log technique of mushroom cultivation should also be encouraged since log does not easily get spent and can be repeatedly used for a long period of time. Wherever log cultivation of oyster mushroom is practiced, afforestation should be encouraged to avoid indiscriminate logging, which can lead to desertification.

## References

- Chang ST (2006) The need for scientific validation of culinary-medicinal mushroom products. *Int J Med Mush* 8: 187-195.
- Cohen R, Persky L, Hadar Y (2002) Biotechnological applications and potential of wood degrading mushrooms of the genus *Pleurotus*. *Appl Microbiol Biotechnol* 58: 582-594.
- Yildiz S, Yildiz UC, Gezer ED, Temiz A (2002) Some lignocellulosic wastes used as raw material in cultivation of the *Pleurotus ostreatus* culture mushroom. *Process Biochem* 38: 301-306.
- Obodai M, Cleland-Okine J, Vowotor KA (2003) Comparative Study on the growth and yield of *Pleurotus ostreatus* mushroom on different lignocellulosic by-products. *J Indust Microbiol Biotechnol* 30: 146-149.
- Membrillo I, Sánchez C, Meneses M, Favela E, Loera O (2011) Particle geometry affects differentially substrate composition and enzyme profiles by *Pleurotus ostreatus* growing on sugar cane bagasse. *Biores Technol* 102: 1581-1586.
- Gume B, Diriba Mulata D, Abate D (2013) Evaluation of locally available substrates for cultivation of oyster mushroom (*Pleurotus ostreatus*) in Jimma, Ethiopia. *Afr J Microbiol Res* 7: 2228-2237.



7. Okwulehie IC, Okwujiako IA, Edeoga HO (2008) Proximate, macroelement and vitamin composition of the fruit bodies of *Pleurotus ostreatus* (var. *florida*) Eger grown on different substrate and substrates supplementation. Global Sci Books 2: 184-188
8. Chang ST (2013) Training manual on mushroom cultivation Technology. Asian and Pacific Centre for Agricultural Engineering, PR China, pp: 24-122.
9. Kadiri JM, Aizai AH (2005) Cultivation of *Lentinus subnudus* (Berk) on woodlogs. Biores Technol 94: 65-69.
10. Hyunjong K, Seung WK (2004) Log cultivation in the temperate regions. Mushroom World Pub Heinart Inc, Republic of Korea, pp: 140-143.
11. Agrahar-Murrugkar D, Subbulaksmi G (2005) Nutritional value of edible wild mushrooms collected from the Khashi hills of Meghalaya. Food Chem 89: 599-603.
12. Muhammad AA, Muhammad S, Shabbier A, Asif HM (2007) Protein and fat contents of various *Pleurotus* species raised on different waste materials. Pak J Agric Sci 44: 140-143.
13. Tewari RP (1991) Effect of soaking period and spawn dose on oyster mushroom (*Pleurotus sajor-caju*) production. Adv Mushroom Sci p: 31.
14. Wahlid M, Satter A, Khan S (1988) Composition of wild and cultivated mushrooms of Pakistan. Mushroom Tropics 8: 47-51.
15. Purkayastha RP, Nayak D (2002) Analysis of protein patterns of an edible mushroom by gelelectrophoresis and its amino-acid composition. J Food Sci Tech 18: 89-91.
16. Chang ST, Miles PG (2004) Mushroom cultivation, nutritional value, medicinal effect and Environmental impact 2<sup>nd</sup> edn. CRC press, London, UK, p: 451.
17. Kurtzman RH (2009) Nutrition of *Pleurotus sapidus*: Effects of lipids. Mycologia 68: 268-275.
18. Chang ST (1999) World production of cultivated and medicinal mushrooms in 1997 with particular emphasis on *Lentinula edodes* (Berk) Sing, in China. Int J Med Mushr 1: 291-300.
19. Shyam SP, Syed AA, Sureh MT, Mirza MV (2010) The nutritional value of *Pleurotus ostreatus* (Jacq: Fr) Kumm. Cultivated on different lignocellulosic Agro waste. Innov Romanian Food Biotechnol 7: 66.
20. Oei P (2003) Mushroom cultivation, appropriate technology for mushroom growers. Back-hugs Publishers, Leiden, the Netherlands, pp: 110-150.
21. Stamets PS (2003) Growing Gourmet and medicinal mushrooms. Ten speed press, Berkeley, USA, pp: 554.
22. Nwoko MC (2015) Yield, heavy metals and nutritional evaluation of *pleurotus ostreatus* (Jacq) P. Kumm. And *Pleurotus pulmonarius* (Fries). Quel. Fruit bodies cultivate on three log substrates in Umudike, Abia state, Nigeria. An unpublished M.sc thesis of the Dept. Of plant science and Biotechnology, Michael Okpara University of Agriculture Umudike, Abia State, Nigeria, p: 19.
23. Okwulehie IC, Odunze IE (2004) Evaluation of the nutritional value of some tropical edible mushrooms. J Sust Agri Environ 6: 157-162.
24. Nwoko MC, Okwulehie IC, Achufusi JN (2016) Nutritional Composition of *Pleurotus ostreatus* (Jacq) P. kumm. Fruit Bodies Cultivated on Deciduous Tree Logs in Abia State, Nigeria. Int J Appl Res Technol 5: 102-108.
25. Chen WA (2004) Growing shiitake mushrooms. Mushrooms Publishers. Heinart Inc, Republic of Korea, pp: 248-260.
26. Okwulehie IC, Okwujiako IA (2008) The use of local Nigerian substrates for the production of *pleurotostreatus* var. *florida* (Eger) sporophores. J Dynamic Biochem, Process Biotechnol Molecular Biol 2: 38-44
27. Association of official Analytical chemistry. (AOAC) (1982) Official methods of Analysis 17<sup>th</sup> edn. Washington, USA, p: 266.
28. MFA (1982) Methods of food analysis. Nippon Shokuhin Kogyo Gakkaishi, Tokyo, p: 40.
29. Perkin-Elmer L (1964) Analytical methods for Atomic Absorption spectrometry. Perkin Elmer co-operation, USA, P: 27.
30. Association of official Analytical chemistry (AOAC) (1975) Official methods of Analysis 11<sup>th</sup> edn. Washington, USA, p: 428.
31. Maxwell A, Speeris M, Pingal R, Mootoo DR, Reynolds WF (1995) 3 beta-aminospinosolane steroidal alkaloids from *Solanum triste*. J Nat Prod 58: 625-628.
32. Peng JP, Koboyashi H (1995) Novel furostanol glycosides from *Allium macrostemon*. Planta Medica 6: 58-61.
33. Harbone M (1988) *Chemical test constituents and identification*. New York John Wiley and sons Ltd, London, UK P: 11-113.
34. Steel RGD, Torie JH (1980) Principle and procedures of statistics. McGraw Hill Co Inc, New York, USA.
35. Ukoima HN, Arikpo GE, Ikpe FN (1999) Cultural studies of mycelia of *Volvariella volvacea*, *Pleurotus tuber-regium* and *Pleurotus sajor-caju* on different culture media. Pak J Nutri 8: 1052-1054.
36. Okhuoya JA, Okogbo FO (1991) Induction of sclerotia of *Pleurotus tuber-regium* (fr). Sing. On various farm waste product. Acad Sci 71: 1-3.
37. Quimio TH, Chang ST, Royce DJ (1990) Technical guidelines for mushroom growing in the tropics. FAO Rome, pp: 155.
38. Ogbo EM, Okhuoya JA (2006) Biodegradation of aliphatic resinic and aliphatic fractions of crude oil contaminated soils by *Pleurotus tuber-regium* Fr. Singer a white-rot fungus. Afr J Biotechnol 7: 4291-4297.
39. Okwulehie IC, Nwosu CP, Okoroafor CJ (2007) Pharmaceutical and nutritional prospects of two wild macrofungi found in Nig. J Biotech 6: 567-572.
40. Okoi AI, Iboh CI (2015) The effects of different substrates on sporophore yield, mineral and nutrient composition of *Pleurotus tuberregium* fries singer in Calabar, Nigeria. Int J Agri Sci Res 4: 126-131.
41. Edeoga HO, Eriata DO (2001) Alkaloids, tanins and saponin content of some medicinal plants. J Med Aromatic Plant Sci 23: 344-349.
42. Rambelli A, Manini UG (2000) Manual on mushroom cultivation. FAO Italy, pp: 165.
43. Hilang MT, Ferraro E (1992) Phenolic compounds in food and their effects on health in Antioxidants and Cancer prevention. Am Chem Soc (ACS), Symposium series 507: 8-12.
44. Bidwell RSG (1979) Plant physiology 2<sup>nd</sup> edn. Macmillan, London, pp: 643.
45. Haslam E (1996) Plant Polyphenols (Vegetable Tannins) as drug possible of action. J Nat Prod 59: 205-215.

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