

## Photobioreactor for CO<sub>2</sub> Sequestration: Possibilities and Challenges

Pushap Chawla<sup>1</sup>, Anushree Malik<sup>1\*</sup> and Sreekrishnan SR<sup>2</sup>

<sup>1</sup>Applied Microbiology Laboratory, Centre for Rural Development and Technology, Indian Institute of Technology Delhi, Hauz Khas, New Delhi-110016, India

<sup>2</sup>Waste Treatment Laboratory, Department of Biochemical Engineering and Biotechnology, Indian Institute of Technology Delhi, Hauz Khas, New Delhi-110016, India

\*Corresponding author: Anushree Malik, Applied Microbiology Laboratory, Centre for Rural Development and Technology, Indian Institute of Technology Delhi, Hauz Khas, New Delhi-110016, India, Tel: (+) 91-11-2659-1158; E-mail: [anushree\\_malik@yahoo.com](mailto:anushree_malik@yahoo.com)

Rec date: May 19, 2014, Acc date: Jul 14, 2014, Pub date: Jul 18, 2014

Copyright: © 2014 Chawla P, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### Abstract

Global warming due to increased CO<sub>2</sub> emission has become a major issue today. In this regard, algae offer a plethora of opportunities due to its photosynthetic route of conversion of CO<sub>2</sub> to biomass which is manifold higher than that of plants. In this regard, this paper highlights the issues, challenges and opportunities generated through algal sequestration of CO<sub>2</sub>. The paper talks about various factors that enhance the efficiency of algal consumption as well as accumulation of CO<sub>2</sub>. The parameters like mode of cultivation, photobioreactors design considerations and harvesting of algal biomass have been given due importance. The perspectives offered will provide a deep insight into development of an integrated technology for making CO<sub>2</sub> sequestration commercially viable.

**Keywords:** Photobioreactor; Carbon dioxide; Sequestration; Algae; Global warming; Climate change

### Introduction

Increased energy and water demand has resulted in degradation of the environment. Current energy demands are mostly met by conventional sources often leading to emission of greenhouse gases [1]. Across the globe, from developed economies to developing ones, there is a lot of interest generated in finding sustainable solutions to increased CO<sub>2</sub> emissions. Algal mediated wastewater treatment and CO<sub>2</sub> sequestration could provide impetus to counter concern for clean water, air and energy, effectively treating even complex contaminants like heavy metals and polycyclic aromatic compounds [2-4]. Moreover, algae that have high rate of CO<sub>2</sub> sequestration are now preferred for biodiesel production due to their high rate of lipid/oil accumulation going as high as 58,700 L oil per hectare [5,6].

Several models have indicated the techno-economic feasibility of integrated algal use by utilizing nutrients from wastewater, carbon from waste gases and subsequent production of biofuel [7]. The key problem associated with the current research is that the investigations are mostly confined to laboratory experiments where often the conditions are controlled which deviate a lot from the real ones. On a larger scale, the applicability of lab findings itself become a challenge. In some cases, the size has been increased to a mere pilot scale of 1000 L but owing to difficulties in scale up of the given design, less breakthroughs have been achieved in design of bioreactor where scale up is taken to industrial level in case of simultaneous CO<sub>2</sub> sequestration and waste water treatment [8].

### Strain selection and mode of cultivation

There are multiple parameters to be taken care of for algal mediated CO<sub>2</sub> sequestration. The very basic and primary step is strain selection. Certain desirable qualities include tolerance to high CO<sub>2</sub> concentration, high biomass productivity, tolerance to variations in temperature and most importantly high CO<sub>2</sub> fixation rates. Most commonly used algal species are depicted in Table 1.

Strain selection is directly linked with mode of cultivation. The conditions of cultivation of algae have a peculiar effect on end result. The photoautotrophic cultivation mode utilizes light for energy requirement and inorganic carbon as carbon source. In this mode, there are large variations in lipid content which can be tilted towards user's advantage by increasing the lipid accumulation through nitrogen limiting condition [5,9]. Totally opposite mode of cultivation is heterotrophic condition where algae utilize organics for both energy as well as nutrient requirement. Since it does not require light, hence it is under the dark conditions [10,11]. Another mode of cultivation is the mixotrophic growth where light is used as an energy source and depending upon the availability, both organics as well as inorganics are used as nutrient source. This is particularly required in cases where algal cultures are cultivated in wastewater with CO<sub>2</sub> injection. In such cases, algae utilizes light for energy, CO<sub>2</sub> as carbon source while simultaneously utilizing organics for treatment of wastewater hereby making the process more economically viable [7,12,13].

Reactor type	Algal cultures	Inlet CO <sub>2</sub> (%)	CO <sub>2</sub> Fixation / utilization efficiency	References
Three-stage serial tubular Photobioreactor	<i>Spirulina</i> sp.	6% (v/v) and 12% (v/v)	53.29% and 5.61% respectively	[14]

Membrane Photobioreactor	<i>Chlorella vulgaris</i>	1% (v/v) (3 L/min)	80 to 260 mg l <sup>-1</sup> h <sup>-1</sup>	[15]
Vertical tank Photobioreactor	<i>Chlorella</i> sp.	2%, 5%, 10% and 15%, at 0.25 vvm	0.261, 0.316, 0.466 and 0.573 g h <sup>-1</sup>	[16]
Outdoor open Photobioreactor	<i>Chlorella</i> sp.	Flue gas (6–8% v/v CO <sub>2</sub> )	10-50 %	[17]
Membrane Carbonation Photobioreactor (bubble-less)	<i>Synechocystis</i> sp. PCC6803	29 g C/m <sup>3</sup>	88%	[18]
Tubular Photobioreactor	<i>Chlamydomonas</i> sp.	10%-40% (Variable flow)	3-12% respectively	[19]
Air lift Photobioreactor	<i>Scenedesmus obliquus</i>	Flue gas (18% CO <sub>2</sub> )	67%	[20]
Internally illuminated Photobioreactor	<i>Nannochloropsis salina</i>	0.035-2%	0.57-2.35 g CO <sub>2</sub> 1-1 d-1	[21]

**Table 1:** Performance of various algal cultures and photobioreactors with respect to CO<sub>2</sub> fixation

### Photobioreactor design considerations

Once the strain selection is made, well designed photobioreactor becomes need of the hour. Cultivation could be done in open as well as closed system. Open systems, like unstirred ponds, circular ponds, raceways and modified raceways offer ease of operation and are more economical than closed system [22]. Major constraints offered by open systems include poor light utilization efficiency, huge evaporative losses and biological contamination. For high rate of CO<sub>2</sub> sequestration, open system are not viable as gas mixing becomes a challenge due to low depth of ponds [23]. When the efficacy of algal mediated CO<sub>2</sub> sequestration was tested in outdoor open photobioreactor, the results indicated that flue gas decarbonisation decreased with increasing flue gas injection rate into culture [17].

For high rate of CO<sub>2</sub> sequestration, mainly closed type photobioreactors are employed. The various design parameters for closed type photobioreactors include type of reactors, height to diameter ratio, mode of delivery, minimizations in temperature variations, optimum intensity of light, maximum utilization of light, proper mixing, optimum light - dark cycles, minimization of CO<sub>2</sub> losses. As depicted in Table 1, multiple designs of photobioreactors present multiple opportunities and challenges. These include mainly tubular type photobioreactors (horizontal and vertical), stirred tank photobioreactors and flat panel photobioreactors. Tubular photobioreactors are fairly easy to operate with decent biomass productivities due to high illumination surface area [22]. This system incorporates gas sparging as a means to provide overall mixing and high gas transfer efficiency [24]. Other variants include bubble column photobioreactor and airlift photobioreactor [25]. In a three stage tubular reactor, though the CO<sub>2</sub> fixation rate was high yet the problems associated with flocculating *Spirulina* sp. highlighted the importance of strain selection [14]. In the study concerning vertical tank reactors, it was noted that the algal sequestration of CO<sub>2</sub> could be increased by multiplicity of reactors hereby presenting an insight with regard to scale up [16]. In case of membrane photobioreactor, gas (CO<sub>2</sub>) exchange efficiency improved from the introduction of membrane module hereby resulting in minimization of losses [15]. While in membrane carbonation photobioreactor the bubble less gas transfer through membranes allows precise control of CO<sub>2</sub> delivery hereby minimizing the loss of CO<sub>2</sub> to atmosphere [18]. There is also need to highlight the maximized fixation of CO<sub>2</sub> by optimizing the light intensity for minimization of CO<sub>2</sub> losses [20].

Light intensity can also be increased by better designs (internally illuminated) where light utilization rate is significantly higher as compared to bubble column and airlift reactors [21]. Studies have exhibited equal importance of dark cycles for algal growth. The dark cycle allows for re-oxidation of electron transporters of photosynthetic apparatus for sustained productivities [26]. Even the importance of optimum concentration where one has to remove the excess biomass continuously in order to provide maximum spread area to volume ratios, can't be ignored [27].

### Harvesting of algal biomass post CO<sub>2</sub> sequestration

Keeping in mind the process economics, significant economical addition to the entire process can be done through harvesting of algal biomass and then use of algal biomass for bioenergy generation [7]. Depending upon the scale, size and density of algal culture, various harvesting techniques like centrifugation, flocculation, gravity sedimentation, filtration, electrophoresis etc. can be employed. Centrifugation can result in high harvesting efficiency but the process is costly and energy intensive [28]. Flocculation is a process where particles form heavy aggregates which make their settling easily [29]. This could be done via increase in pH (autoflocculation) through base (NaOH) addition or through chemical flocculants like ferric chloride (FeCl<sub>3</sub>) or aluminium sulphate Al<sub>2</sub>(SO<sub>4</sub>)<sub>3</sub> [24]. Success of gravity sedimentation highly depends upon size and density of algal cultures [30]. Efficiency of gravity sedimentation can be increased using flocculation [29]. Filtration involves passing of culture through screen of particular pore size. Major associated problem with filtration is high concentration of algal biomass often results in blocking of filtering material [31]. Other techniques include electrophoresis where electric current induced electrostatic field forces the movement of charged algal cells out of solution [32].

### Recent large scale developments in the field of CO<sub>2</sub> sequestration

The ENN group from China has developed a technology where a microalga is used to fix CO<sub>2</sub> released from coal production and subsequently biofuel is produced. The pilot plant consists of equipment required for microalgae cultivation with CO<sub>2</sub> absorption, oil extraction and bio-diesel production. This pilot system can absorb 110 tonnes of CO<sub>2</sub> and produce 20 and 5 tonnes of bio-diesel and proteins per year, respectively. Based on this pilot plant, a demonstration project has been initiated by ENN group in Dalate

(Mongolia) in 2010 where microalgae will be utilized to absorb CO<sub>2</sub> emitted from flue gas of coal-derived methanol and coal derived dimethylether production equipment and will produce biodiesel and feed.

Another group from Sweden, has set up a pilot plant in eastern Germany where algae is used to absorb greenhouse gas emissions from a coal-fired power plant. The gas emitted from the brown-coal-fired plant is pumped through a broth into the plastic tanks where algae is cultivated. The resultant algal biomass is used to produce biodiesel, to feed biogas power plants and as a nutrient supplement for fishes.

## Conclusion

There is a need for informed and effective design of photobioreactor which could replicate the real conditions and could provide an alternative solution. In this cost driven world, this idea of algal use is bound by importance of light as a parameter in algal growth. Across the world, there are places with abundance of sunlight available almost throughout the year. This advantage is still underutilized mainly because of lack of bioreactors which maximizes algal productivity and CO<sub>2</sub> sequestration. While the CO<sub>2</sub> sequestration offers huge environmental benefit, one has to make effort in the direction to make it economically viable. This can be made possible by integration of CO<sub>2</sub> sequestration with wastewater treatment and biofuel generation.

## Acknowledgement

Authors are grateful to Ministry of New and renewable Energy, Government of India for financial support.

## References

1. Intergovernmental Panel on Climate Change (IPCC) Summary for Policymakers, 2007: Climate Change: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, United Kingdom and New York, NY, USA.
2. Rahman A, T Ellis J (2012) Bioremediation of Domestic Wastewater and Production of Bioproducts from Microalgae Using Waste Stabilization Ponds. Journal of Bioremediation and Biodegradation 03: 06.
3. Siva Kiran RR MG, Satyanarayana SV, Bindya P (2012) Bioaccumulation of Cadmium in Blue Green Algae Spirulina (Arthrospira) Indica. J Bioremed Biodegrad 3.
4. El-Sheekh MM GM, EL-Souod GW (2012) A Biodegradation of Phenolic and Polycyclic Aromatic Compounds by Some Algae and Cyanobacteria. J Bioremed Biodegrad 3.
5. Mata TM, Martins AA, Caetano NS (2010) Microalgae for biodiesel production and other applications: A review. Renewable & Sustainable Energy Reviews 14: 217-232.
6. Chisti Y (2007) Biodiesel from microalgae. Biotechnol Adv 25: 294-306.
7. Prajapati SK, Kaushik P, Malik A, Vijay VK (2013) Phycoremediation coupled production of algal biomass, harvesting and anaerobic digestion: possibilities and challenges. Biotechnol Adv 31: 1408-1425.
8. Chinnsamy S, Bhatnagar A, Hunt RW, Das KC (2010) Microalgae cultivation in a wastewater dominated by carpet mill effluents for biofuel applications. Bioresour Technol 101: 3097-3105.
9. Huang G, Chen F, Wei D, Zhang X, Chen G (2010) Biodiesel production by microalgal biotechnology. Ap En 87: 38-46.
10. Cheng Y, Zhou W, Gao C, Lan K, Gao Y, et al. (2009) Biodiesel production from Jerusalem artichoke (*Helianthus Tuberosus* L.) tuber by heterotrophic microalgae *Chlorella protothecoides*. J Chem Technol Biotechnol 84: 777-781.
11. Xiong W, Li X, Xiang J, Wu Q (2008) High-density fermentation of microalga *Chlorella protothecoides* in bioreactor for microbio-diesel production. Appl Microbiol Biotechnol 78: 29-36.
12. Liang Y, Sarkany N, Cui Y (2009) Biomass and lipid productivities of *Chlorella vulgaris* under autotrophic, heterotrophic and mixotrophic growth conditions. Biotechnol Lett 31: 1043-1049.
13. Mandal S, Mallick N (2009) Microalga *Scenedesmus obliquus* as a potential source for biodiesel production. Appl Microbiol Biotechnol 84: 281-291.
14. de Morais MG, Costa JA (2007) Biofixation of carbon dioxide by *Spirulina* sp. and *Scenedesmus obliquus* cultivated in a three-stage serial tubular photobioreactor. J Biotechnol 129: 439-445.
15. Cheng L, Zhang L, Chen H, Gao C (2006) Carbon dioxide removal from air by microalgae cultured in a membrane-photobioreactor. Sep Purif Technol 50: 324-329.
16. Chiu SY, Kao CY, Chen CH, Kuan TC, Ong SC, et al. (2008) Reduction of CO<sub>2</sub> by a high-density culture of *Chlorella* sp. in a semicontinuous photobioreactor. Bioresour Technol 99: 3389-3396.
17. Doucha J, Straka F, Livanský K (2005) Utilization of flue gas for cultivation of microalgae *Chlorella* sp.) in an outdoor open thin-layer photobioreactor. J Appl Phycol 17: 403-412.
18. Kim HW, Marcus AK, Shin JH, Rittmann BE (2011) Advanced control for photoautotrophic growth and CO<sub>2</sub>-utilization efficiency using a membrane carbonation photobioreactor (MCPBR). Environ Sci Technol 45: 5032-5038.
19. Hadiyanto H, Sumarno S, Rostika R, Handayani N (2012) Biofixation of Carbon dioxide by *Chlamydomonas* sp. in a Tubular Photobioreactor. Int Journal of Renewable Energy Development 1: 5.
20. Li FF, Yang ZH, Zeng R, Yang G, Chang X, et al. (2011) Microalgae Capture of CO<sub>2</sub> from Actual Flue Gas Discharged from a Combustion Chamber. Ind Eng Chem Res 50: 6496-6502.
21. Pegallapati AK, Nirmalakhandan N (2013) Internally illuminated photobioreactor for algal cultivation under carbon dioxide-supplementation: Performance evaluation. Renewable Energy 56: 129-135.
22. Ugwu CU, Aoyagi H, Uchiyama H (2008) Photobioreactors for mass cultivation of algae. Bioresour Technol 99: 4021-4028.
23. Ho SH, Chen CY, Lee DJ, Chang JS (2011) Perspectives on microalgal CO<sub>2</sub>-emission mitigation systems—a review. Biotechnol Adv 29: 189-198.
24. Razzak SA, Hossain MM, Lucky RA, Bassi AS, de Lasa H (2013) Integrated CO<sub>2</sub> capture, wastewater treatment and biofuel production by microalgae culturing—A review. Renewable and Sustainable Energy Reviews 27: 622-653.
25. Kumar A, Ergas S, Yuan X, Sahu A, Zhang Q, et al. (2010) Enhanced CO<sub>2</sub> fixation and biofuel production via microalgae: recent developments and future directions. Trends Biotechnol 28: 371-380.
26. Sforza E, Simionato D, Giacometti GM, Bertucco A, Morosinotto T (2012) Adjusted light and dark cycles can optimize photosynthetic efficiency in algae growing in photobioreactors. PLoS One 7: e38975.
27. Vasumathi KK, Premalatha M, Subramanian P (2012) Parameters influencing the design of photobioreactor for the growth of microalgae. Renewable and Sustainable Energy Reviews 16: 5443-5450.
28. Molina Grima E, Belarbi EH, Acien Fernández FG, Robles Medina A, Chisti Y (2003) Recovery of microalgal biomass and metabolites: process options and economics. Biotechnol Adv 20: 491-515.
29. Eisenberg DM, Koopman B, Benemann JR, Oswald WJ (1981) Algal biofloculation and energy conservation in microalgal sewage ponds. Biotechnology bioeng. 11: 429-448.
30. Brennan L, Owende P (2010) Biofuels from microalgae-A review of technologies for production, processing, and extractions of biofuels and co-products. Renewable and Sustainable Energy Reviews 14: 557-577.
31. Molina Grima E, Belarbi EH, Acien Fernandez FG, Robles Medina A, Chisti Y (2003) Recovery of microalgal biomass and metabolites: process options and economics. Biotechnol Adv 20: 491-515.

32. Azarian GH, Mesdaghinia AR, Vaezi F, Nabizadeh R, Nematollahi D (2007) Algae removal by electro-coagulation process, application for treatment of the effluent from an industrial wastewater treatment plant. Iranian Journal of Public Health 36: 57-64.