Physicochemical Characteristics, Identification of Bacteria and Biodegradation of Industrial Effluent

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Abstract

An investigation was carried out to analyze Physicochemical parameters like colour, odour, pH, electrical Conductivity (EC), Total suspended solids (TSS), Total dissolved Solids (TDS), Biochemical Oxygen Demand (BOD), Chemical Oxygen Demand (COD), chromium and copper of untreated tannery effluent and to degrade the effluent using native and non-native bacteria. The results of the parameters analysis revealed that untreated tannery effluent was black in colour with offensive odour. pH was alkaline with high organic load such as EC, TSS, TDS, BOD and COD which were higher than the permissible since the effluent had high organic load, microbes (bacteria) present within the effluent were identified and isolated. The results of the study revealed the occurrence of 4 species of bacteria namely Escherichia coli, Klebsiella sp., Pseudomonas sp. and Staphylococcus aureus. The presence of bacteria indicates the pollutional status of the untreated tannery effluent suggesting that it should be treated before its disposal using the biological method particularly native and non-native bacteria for comparing their degrading efficiency. The results of the degradation study shows that native bacteria E. coli was found to be very much successful in reduction of toxic substances at the percentage range of 54-91% whereas non-native Bacillus sp showed reduction percentage range of 56-95% and the bio-treated water can be reused for the agricultural and aquacultural purposes.

Keywords: Untreated tannery effluent; Physico-chemical parameters; Bacterial identification; Degradation; Native E. coli; Non-native Bacillus sp

Introduction

Nature has given gifts like air, water, land, forest, minerals, fossil fuels and several resources to man. These gifts were given to improve his living standards. But unbridled exploitation gradually resulted in the release of pollutants into the environment [1]. Pollution is a major environmental issue in the world due to its adverse effect on living organism. In the past few decades, uncontrolled urbanization has caused a serious pollution problem due to the disposal of sewage and industrial effluents to water bodies [2]. Majority of industries are water based and a considerable volume of waste water emanates from them which is generally discharged into water courses either untreated or inadequately treated causing water pollution [3].

Tannery is one of the important industries causing water pollution. There are about 2161 tanneries in India excluding cottage industries, which processes 500,000 tonnes of hides and skins annually. A total annual discharge of waste water from these tanneries is 9,420,000 m³, which generates about 100,000 m³ of waste water per day (Mohan et al., 2005) and these industries spread mostly across Tamil Nadu, West Bengal, Uttar Pradesh, Andhra Pradesh, Karnataka, Rajasthan and Punjab (Lefebvre et al., 2005). In Tamil Nadu alone there are about 1120 tanneries concentrated in Vellore, RaniPET, Tiruchy, Dindugal, Erode and Pallavaram in Chennai. The effluent generated in the tanneries has high amounts of organic substances as well as high concentration of chloride, chromium, sulphide and ammonium salts used during the process.

Impact of Tannery Effluent on the Environment

The tannery industries are considered as polluting due to the inherent manufacturing processes as well as type of technology employed in the manufacture of hides and skin into leather. During the tanning process at least 30 kg of chemicals are added per ton of hides [4]. Tannery effluent when discharged into water bodies alter the physical, chemical and biological characteristics of water and depletes the dissolved oxygen, increases alkalinity, suspended solids and sulphides which are injurious to fish and other aquatic lives.

Apart from organic materials which release valuable nutrients for decomposition, tannery effluent contains chromium and pathogens mainly of faecal origin and toxic organic components, all of which pose of serious threat to the environment [4]. Heavy metals in the tannery effluent are one of the most hazardous environmental pollutants. Human beings, cattles and plants are affected when these toxic metals like Cr, Cu, Zn, Pb and Cd are incorporated into the food chain [5]. Hence tannery effluent with high pollution load should be treated before its disposal.

The treatment of industrial waste varies with its character, quantity and the nature of receiving media and the dilution available. The different methods are: Physical method, Chemical Method and Biological method. Physical and chemical methods of waste water treatment are invariably cost intensive and cannot be employed in all industries especially in developing and under developed countries. Compared with chemical/physical methods, biological processes have received more interest because of their cost effectiveness, lower sludge production and environmental friendliness. Bioremediation of tannery effluents is an attractive environment friendly, safe and cost effective alternative technology to conventional methods. Microbes in
the environment play an important role is cycling and destroying them through bio-degradation [6].

Taking into consideration of all the above said investigation carried out by many researchers pertaining to degradation of various industrial effluents using microbes especially bacteria, an attempt has been made to degrade the untreated tannery effluent using native bacteria *E. coli* sp. and non-native Bacteria, *Bacillus* sp.

### Aims and Objectives

To analyse the physico-chemical parameters of industrial effluents as a means of monitoring the pollution

- To isolate and identify the microbes present in the industrial effluent which provide clues on the ability of the microbes to survive, adapt and colonize in the polluted environment.
- To ascertain the bioremediation potential using native bacteria isolated from industrial effluent for its treatment.
- To degrade the industrial effluent using non-native bacteria which is not present in industrial effluent and obtained from Microbiological Laboratory Centre, India
- To study the comparison between the native and non-native bacteria for the degrading efficiency of the effluent.

### Materials and Methods

#### Analysis of physicochemical parameters of industrial effluent

Untreated industrial effluent was collected in polythene containers from an industry located in Chennai, Tamil Nadu of India, were brought to the laboratory, and stored for further analysis. The sample was collected for a period of 6 months (May 2011 to October 2011). The physico-chemical parameters of the effluent-pH, Electrical Conductivity (EC), Biochemical Oxygen Demand (BOD), Chemical Oxygen Demand (COD), Total Dissolved Solids (TDS), Total Suspended Solids (TSS), Total Hardness, Chloride, Sodium, Calcium and heavy metals were estimated by following the Standard methods suggested by APHA [7].

#### Isolation and identification of bacteria present in industrial effluent

Industrial effluent was collected in sterile bottles for microbial analysis, brought to the laboratory and bacterial identification was carried out by pour plate method, following the procedure of Power and Daginwala [8] and Sundararaj [9].

### Biodegradation of industrial effluent using native bacteria

Isolates of Native bacteria- *E. coli* obtained from industrial effluent was used for biodegradation of industrial effluent by following the procedure of Aftab Begum and Noorjahan [10].

### Biodegradation of industrial effluent using non-native bacteria

Isolates of Non-Native bacteria- *Bacillus* sp. which was not present in the industrial effluent was obtained from Microbiology Laboratory Centre, India and used for biodegradation of industrial effluent by following the procedure of Aftab Begum and Noorjahan [10].

### Statistical analysis

- The data obtained from the experiments were analysed and expressed as mean and standard deviation.
- The percentage change was calculated between the control and experimental for the various experiments

### Results and Discussions

#### Analysis of physicochemical parameters of industrial effluent

Results of the analysis of the physicochemical parameters of untreated industrial effluent collected for a period of 6 months (May 2011 to October 2011) are depicted in Table 1. Statistical analysis was also carried out. The results of the study revealed that colour of the untreated industrial effluent were blackish with unpleasant odour. This colour and odour could be due to decomposition of organic and inorganic matter [11]. A large number of pollutants can impart colour, taste and odour to the receiving water, thereby making them unaesthetic and unfit for domestic consumption [2,12].

The pH ranged from 7.14 ± 0.0187 (Aug. 2011) to 7.27 ± 0.0187 (May 2011) during the period of study, indicating that the pH of the tannery effluent was found to be alkaline. According to Singh et al. [11], highly alkaline water if consumed would affect the mucous membrane and may cause metabolic alkalosis. In addition, the toxicity of certain substances present in water may be enhanced due to their interaction with high or low levels of pH prevailing which may further be detrimental to aquatic organisms [13,14].

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<td>1</td>
<td>Colour</td>
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<td>6772 ± 1.8748</td>
<td>300 ± 1.7607</td>
<td>250 ± 1.7607</td>
<td>1000 ± 1.8708</td>
<td>6772 ± 1.8748</td>
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<td>6772 ± 1.8748</td>
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<td>7.27 ± 0.0187</td>
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<td>7.27 ± 0.0187</td>
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<td>Electrical Conductivity (EC) (µmhos/cm)</td>
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<td>Total Suspended Solids (TSS)(mg/l)</td>
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<td>6</td>
<td>Total Dissolved Solids (TDS)(mg/l)</td>
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<td>7</td>
<td>Biochemical Oxygen Demand (BOD)(mg/l)</td>
<td>1000 ± 1.8708</td>
<td>6772 ± 1.8748</td>
<td>300 ± 1.7607</td>
<td>250 ± 1.7607</td>
<td>1000 ± 1.8708</td>
<td>6772 ± 1.8748</td>
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<td>8</td>
<td>Chemical Oxygen Demand (COD)(mg/l)</td>
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<td>6772 ± 1.8748</td>
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<td>250 ± 1.7607</td>
<td>1000 ± 1.8708</td>
<td>6772 ± 1.8748</td>
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<td>9</td>
<td>Total Hardness (mg/l)</td>
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<td>250 ± 1.7607</td>
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<td>250 ± 1.7607</td>
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<td>Calcium (mg/l)</td>
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<td>Total Chromium (mg/l)</td>
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<td>300 ± 1.7607</td>
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<td>14</td>
<td>Copper (mg/l)</td>
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<td>6772 ± 1.8748</td>
<td>300 ± 1.7607</td>
<td>250 ± 1.7607</td>
<td>1000 ± 1.8708</td>
<td>6772 ± 1.8748</td>
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Table 1: Analysis of physicochemical parameters of Industrial effluent for a period of six months from May 2011-Oct. 2011.
Electrical conductivity of the effluent has a minimum range of 8354 μmhos/cm ± 1.8708 (Aug. 2011) and maximum range of 9148 μmhos/cm ± 1.877 (July 2011). Untreated tannery effluent showed higher level of Electrical conductivity which could reflect the presence of organic and inorganic substances and salts that would have increased the conductivity [15,16].

TSS ranged from 1138 mg/l ± 1.8756 (June 2011) to 6908 mg/l ± 1.8708 (May 2011) which was found to be beyond the permissible limits (100 mg/l) of CPCB [17]. High amounts of suspended particles have detrimental effects on aquatic flora and fauna and reduce the diversity of life in aquatic system and promote depletion of oxygen and settling in ponds during rainy season [12,18]. TDS ranged from 5768 mg/l ± 1.8794 (Aug. 2011) to 6712 mg/l ± 1.8755 (June 2011) which surpassed the CPCB [17] permissible limits (2100 mg/l).

The composition of solids present in a natural body of water depends on the nature of the area and the presence of industries nearby. High levels of TDS may be due to high salt content and also renders it unsuitable for irrigation, hence further treatment or dilution would be required [13]. Singh et al. [11] cautioned that if the TDS level of water exceeded 500 mg/l, it becomes unsuitable for bathing and drinking purposes for animals as it could cause distress in cattle and livestock.

BOD has a minimum range of 600 mg/l ± 1.8759 (August 2011) and maximum range of 1722 mg/l ± 1.8906 (May 2011) which was beyond the permissible limit (30 mg/l) of CPCB [17]. Increase in BOD which is a reflection of microbial oxygen demand leads to depletion of DO which may cause hypoxia conditions with consequent adverse effects on aquatic biota [19]. Oxygen depletion could be followed by anaerobic conditions which would result in reduced diversity and distribution of aquatic fauna. Further the presence of organic matter will promote anaerobic action leading to the accumulation of toxic compounds in water bodies [13]. Oxygen depletion could be followed by anaerobic conditions which would result in reduced diversity and distribution of aquatic fauna.

COD ranged from 2386 mg/l ± 1.8708 (Aug. 2011) to 9600 mg/l ± 2.908 (May 2011) which has exceeded the permissible limit (250 mg/l) of CPCB [17]. COD test is the best method for organic matter estimation and rapid test for the determination of total oxygen demand by organic matter present in the sample. The present investigation revealed high levels of COD. This indicates that the effluent is unsuitable for the existence of aquatic organisms due to the reduction in DO content [13].

Total Hardness has a minimum value of 1080 mg/l ± 1.8708 (May 2011) and maximum value of 5750 mg/l ± 1.8708 (September 2011) which was beyond the CPCB [17] permissible limit of 1000 mg/l. Chloride ranged from 1184 mg/l ± 1.8720 (July 2011) to 1890 mg/l ± 1.8895 (June 2011) which surpassed the permissible limit (1000 mg/l) of CPCB [17]. Sodium has a minimum range of 1200 mg/l ± 1.672 (October 2011) and maximum range of 2100 mg/l ± 1.8708 (August 2011) which was found to be beyond the permissible limit (600 mg/l) of CPCB [17] whereas Calcium ranged from 257 mg/l ± 1.8777 (May 2011) to 3750 mg/l ± 1.8708 (September 2011). The presence of heavy metals in the tannery effluent produces several adverse effects on living organisms as reported by Chukwu [20]. Thus, the physicochemical parameters analysis of untreated industrial effluent for a period of 6 months (May 2011-October 2011) confirms that the parameters surpassed the permissible limits of CPCB [17] for its disposal, indicating high pollution potential of the effluent.

**Isolation and identification of bacteria present in industrial effluent**

Microbes especially bacteria act as bio-indicator of high polluted effluents as reported by Soha Farag and Sahar Zaki [21], which prompted to analyse the native bacterial population in tannery effluent and to use it for biodegradation. The results of the analysis of isolation and identification of microbes (bacteria) present in untreated industrial effluent for a period of 6 months (May 2011 - October 2011) are represented Table 2. The results of the study revealed the occurrence of 4 species of bacteria namely *Escherichia coli*, *Klebsiella sp.*, *Pseudomonas sp.* and *Staphylococcus aureus*. *Escherichia coli* were found to be a dominant bacterial species due to the growth of maximum number of colonies (30) on the medium compared to other bacterial species.

The presence of 4 bacterial species in the tannery effluent as reported in the present study has significance in their utility as biological indicators [22]. Further as pointed out by Radha [23], the presence of native microbes in tannery effluent would be successfully exploited to remove the pollutants, a technique which is more economically and industrially effective.

**Biodegradation of industrial effluent using native and non-native microbes**

Pure culture of native bacteria, *E. coli* and non-native bacteria, *Bacillus* sp. were used for biodegradation of industrial effluent. Results of analysis of degradation of effluent using native *E. coli* and non-native *Bacillus* sp. are presented in Table 3. The colour of industrial sample is blackish before degradation but after degradation using native *E. coli* and non-native *Bacillus* sp. there is a change in colour from blackish to light brown (native *E. coli*) and black to almost colour less (non-native *Bacillus sp.*) of the industrial effluent. The odour of effluent was offensive in nature before degradation but after degrading the effluent using native *E. coli* and non-native *Bacillus sp.* samples shows odourless condition. This may be due to the action of microbes - *E. coli* and *Bacillus sp.*, which decomposed the toxic pollutants present in the effluent and made the change in colour and odour of the effluent. This is supported by the work of Sukumaran et al. [19].

The pH of effluent before degradation is alkaline in nature but after degrading the sample using native *E. coli* and non-native *Bacillus sp.* for

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<tr>
<td>1</td>
<td><em>Escherichia coli</em> (Gram negative)</td>
<td>±</td>
<td>±</td>
<td>±</td>
<td>-</td>
<td>-</td>
<td>±</td>
<td>30</td>
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<tr>
<td>2</td>
<td><em>Klebsiella sp.</em> (Gram negative)</td>
<td>±</td>
<td>±</td>
<td>±</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>16</td>
</tr>
<tr>
<td>3</td>
<td><em>Pseudomonas sp.</em> (Gram negative)</td>
<td>±</td>
<td>±</td>
<td>-</td>
<td>±</td>
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<td>20</td>
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<td>4</td>
<td><em>Staphylococcus aureus</em> (Gram Positive)</td>
<td>±</td>
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± Present - Absent

**Table 2:** Isolation and Identification of Bacteria from Industrial Effluent for a period of six months from May 2011 – October 2011.
and the percentage change of chromium is 73.5%. Copper of industrial native bacteria Bacillus E. coli degradation, native bacteria degraded chromium to 0.535 mg/l which is beyond the permissible limit (250 mg/l) of CPCB [17], but after degradation, native bacteria Bacillus sp. degraded chromium to 0.172 mg/l ± 0.00028 and the percentage change of copper is 53.9% and non-native bacteria Bacillus sp. degraded copper to 0.0094 mg/l ± 0.00158 and the percentage change of copper is 40.3%.

Thus from the above results, it can be inferred that the maximum reduction of toxic substances was recorded in bio-treated sample using native bacteria E. coli (54-91%) and non-native Bacillus sp.(56-95%) compared to untreated effluent. Thus from the foregoing discussion it is very clear that microbes play an important role in the biodegradation of organic and inorganic matter. They have enzymes that allow them to use environmental contaminants as food and hence make them ideal for biodegradation. Besides their characteristics like rapid growth, metabolism and a remarkable ability to adjust to a variety of environments make them very useful in biodegradation. How successful are the micro-organisms in degrading the environmental contaminants depends on the type of microbes, contaminant and on the nature of contaminated site. From the present study, native E. coli and non-native Bacillus sp. showed efficient degrading capabilities by degrading the contaminants as they use it for their growth and reproduction. Organic compounds are a source of carbon which forms one of the basic building blocks of new cell contaminants. In addition to the carbon source, they require nitrogen and phosphorus as primary nutrient and traces of inorganic salts through a series of complex enzymatically catalyzed reaction, the toxic organic contaminant is converted to innocuous chemical compound, obtain energy by catalyzing energy producing chemical reactions and this energy is used in the production of new cells [25] finally resulting in carbon-di-oxide and water.

Thus degradation by microbes seems to be most promising technique for 100% untreated tannery effluent as evidenced in the present investigation It is well known that water of good quality and free of pollutants are primary requirements for agricultural and piscicultural practice. After degradation the treated water could be used for crop cultivation or irrigation and aquaculture purpose.

Acknowledgement
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References

The above biodegradation results are encouraging and scale up studies for continuous treatment of wastewater at pilot scale is required. The above biodegradation results are encouraging and scale up studies for continuous treatment of wastewater at pilot scale is required.

**Table 3: Analysis of physicochemical parameters of industrial effluent before (control) and after degradation using native E. coli and non-native Bacillus sp.**

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<tbody>
<tr>
<td>1.</td>
<td>Colour</td>
<td>Colourless</td>
<td>Blackish</td>
<td>Light Brown</td>
<td>Almost colourless</td>
</tr>
<tr>
<td>2.</td>
<td>Odour</td>
<td>Odourless</td>
<td>Offensive</td>
<td>Odourless</td>
<td>Odourless</td>
</tr>
<tr>
<td>3.</td>
<td>pH</td>
<td>5.5 ± 9.0</td>
<td>8.01 ± 0.5205</td>
<td>7.0 ± 0.19235 (12.6%)</td>
<td>7.0 ± 0.5223 (12.6%)</td>
</tr>
<tr>
<td>4.</td>
<td>Electrical Conductivity (µmhos/cm)</td>
<td>400</td>
<td>8615 ± 3.1622</td>
<td>3920 ± 1.9235 (54.5%)</td>
<td>3770 ± 2.4083 (56.2%)</td>
</tr>
<tr>
<td>5.</td>
<td>Total Suspended Solids (mg/l)</td>
<td>100</td>
<td>184 ± 0.03315</td>
<td>24 ± 1.5811 (89.9%)</td>
<td>16 ± 3.1622 (91.3%)</td>
</tr>
<tr>
<td>6.</td>
<td>Total Dissolved Solids (mg/l)</td>
<td>2100</td>
<td>5950 ± 3.1622</td>
<td>2740 ± 1.5811 (53.9%)</td>
<td>2630 ± 3.1622 (55.7%)</td>
</tr>
<tr>
<td>7.</td>
<td>Biochemical Oxygen Demand (mg/l)</td>
<td>30</td>
<td>700 ± 1.5811</td>
<td>70 ± 1.5811 (90%)</td>
<td>32 ± 3.1622 (95.4%)</td>
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<tr>
<td>8.</td>
<td>Chemical Oxygen Demand (mg/l)</td>
<td>250</td>
<td>2399 ± 1.5811</td>
<td>200 ± 1.5811(91.6%)</td>
<td>297 ± 2.8635 (87.6%)</td>
</tr>
<tr>
<td>9.</td>
<td>Chromium (mg/l)</td>
<td>3</td>
<td>0.0724 ± 0.000158</td>
<td>0.535 ± 0.0015 (63.8%)</td>
<td>0.172 ± 0.028 (73.5%)</td>
</tr>
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<td>10.</td>
<td>Copper (mg/l)</td>
<td>1.5</td>
<td>0.00124 ± 3.16228</td>
<td>0.0094 ± 0.00158 (40.3%)</td>
<td>0.00023 ± 3.16228 (78.2%)</td>
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± = Standard Deviation; % = Percentage Change

96 hrs, alkaline nature of pH has changed to the neutral state indicating the efficiency of the microbes to biodegrade the effluent. This is in agreement with the reports of Noorjahan et al. [24].

EC of effluent before treatment is 8615 (µmhos/cm) ± 3.1627 which is beyond the permissible limit (400 µmhos/cm) of CPCB [17] but after degradation using native bacteria E. coli and EC degraded to 3920 µmhos/cm ± 1.9235 , percentage of degradation was found to be 54.5% and using non-native bacteria Bacillus sp.

TSS of Industrial effluent before treatment is 184 mg/l ± 3.03315 which is beyond the permissible level (100 mg/l) of CPCB [17] for disposal, but after degradation, native bacteria E. coli degraded TSS to 24 mg/l ± 1.5811 and the percentage change is 89.9% and non-native bacteria Bacillus sp. degraded TSS to 16 mg/l ± 3.1622 and the percentage change is 91.3%.

TDS of effluent before treatment is 5950 mg/l ± 3.1622 which is beyond the permissible limit (2100 mg/l) of CPCB [17] but after degradation, native bacteria E. coli degraded TDS to 2740 mg/l ± 1.5811 and the percentage change is 53.9% and non-native bacteria Bacillus sp. degraded TDS to 2630 mg/l ± 3.1622 and the percentage change is 55.7%. Since TSS and TDS are the major pollutants, the above biodegradation results are encouraging and scale up studies for continuous treatment of wastewater at pilot scale is required. The information generated would help to scale up the process and assess the economic feasibility of the technology. This is in agreement with the work of Karthikeyan et al., [18].

BOD of industrial effluent before treatment is 700 mg/l ± 1.5811 which is beyond the permissible limit (30 mg/l of CPCB) [17] for disposal but after degradation, native bacteria E. coli degraded BOD to 70 mg/l ± 1.5811 and the percentage change is 90% and non-native bacteria Bacillus sp. degraded BOD to 32 mg/l ± 3.1622 and the percentage change is 95.4%. COD of effluent before treatment is 2399 mg/l ± 1.5811 which is beyond the permissible limit (250 mg/l) of CPCB [17], but after degradation, native bacteria E. coli degraded COD to 200 mg/l ± 1.5811 and the percentage change is 91.6% and non-native bacteria Bacillus sp. degraded COD to 297 mg/l ± 2.8635 and the percentage change is 87.6%. This is supported by the work of Soha Farag and Sohar Zaki [21].

Chromium of industrial effluent before treatment is 0.0724 mg/l ± 0.00015 which is within the permissible (3 mg/l) of CPCB [17] but after degradation, native bacteria E. coli degraded chromium to 0.353 mg/l ± 0.0015 and the percentage change of chromium is 63.8% and non-native bacteria Bacillus sp. degraded chromium to 0.172 mg/l ± 0.00028 and the percentage change of chromium is 73.5%. Copper of industrial effluent before treatment is 0.00124 mg/l ± 3.1622 which is within the permissible limit (1.5 mg/l) of CPCB [17] but after degradation for 96 hours, native bacteria E. coli degraded copper to 0.0094 mg/l ± 0.00158 and the percentage change of copper is 40.37% and non-native bacteria Bacillus sp. degraded copper to 0.00023 mg/l ± 3.1622 and the percentage change of copper is 78.2%. This is in agreement with the work of Karthikeyan et al. [18].


