Plasma Mannoheptulose Kinetics in Adult Domestic Short-Haired Felines

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Abstract

Mannoheptulose (MH), a sugar found in avocados that inhibits glycolysis, has been investigated as a functional feed ingredient for canines. However, no studies have sought to feed MH to felines. The purpose of this study was to assess whether ingested MH appears in peripheral circulation of adult domestic short-hair felines (N = 10, 4.1 ± 0.1 kg, 1.6 y) fed a MH containing diet. The study was designed as a randomized cross-over with each cat receiving dietary treatments, control and MH. Each study period lasted 28 d and a meal challenge was performed on d 28. Felines were fasted overnight, anesthetized, and a catheter was inserted into the jugular vein for repeated blood sampling. A fasting blood sample was collected six hours after catheter placement. Subsequently, felines were offered their full daily ration of test diet and blood was collected every 2 h during the 24 h post-prandial period for analysis of plasma MH. Ingested MH appeared in the plasma within 2 h of ingestion confirming that dietary MH is absorbed and available for cellular uptake. Circulating MH was cleared from plasma within 24 h of ingestion. The differences in plasma MH kinetics between species are likely attributed to differences in feline carbohydrate metabolism relative to other mammals. No MH was detected in cats fed the control diet. These results suggest that MH is digested, absorbed and available from peripheral circulation in adult cats. Finally, MH remains in circulation longer than in dogs and may suggest that cats would only need a lower dose or fewer doses of MH per day.

Keywords: Mannoheptulose; Diet; Glucose

Introduction

In both humans and companion animals there is a growing interest in the use of plant derived dietary supplements and functional feed ingredients (“nutraceuticals”) to promote healthy weight maintenance. Mannoheptulose (MH), a sugar found in high concentrations in avocados, has been preliminarily evaluated as a functional food ingredient for dogs [1-4]. The metabolic effects of MH are due to its ability to competitively inhibit hexokinases [5,6]. The potential for MH to limit glucose availability via glycolytic inhibition may consequently lead to increases in fat utilization. With prolonged daily feeding, increased fat utilization may lead to changes in body composition. Being obligate carnivores cats have several idiosyncrasies in carbohydrate metabolism. For example, cats have notably lower hepatic glucokinase (hexokinase IV) activity [7] in comparison to omnivores. The potential for MH to further suppress hexokinase activities and proceeding metabolic consequences are unknown as no one has sought to feed MH to cats. These hypotheses remain to be tested, but make MH an appealing target for further investigation, especially in companion animals where the incidence of obesity and associated metabolic disorders is increasing. The purpose of this study was to determine the availability of ingested MH in adult cats. Specifically, we aimed to describe the appearance and disappearance circulating MH in cats fed a MH containing meal.

Methods and Materials

Animals and housing

All procedures were approved by the Institutional Animal Care and Use Committee of The Iams Company (Lewisburg, OH). Ten domestic shorthair felines (5 neutered males and 5 spayed females, 4.08 ± 0.06 kg, 1.6 y) were used in this study. All cats were deemed healthy at the initiation of the study based on a standard physical examination conducted by an accredited veterinarian. All animals resided at the The Iams Company Pet Health and Nutrition Center (The Iams Company, Lewisburg, OH). Cats were group housed with free access to water and indoor and outdoor housing areas. Indoor and outdoor areas were equipped with beds, toys, climbers and scratching posts for environmental enrichment.

Study design

This study was designed as a cross-over with each cat receiving both dietary treatments, control (CON) and mannoheptulose (MH), in random order. Seven days prior to study initiation (wash in) and for seven days between study periods (wash out) all cats received the CON diet. After the 7 day washout periods, cats were placed on one of two dietary treatments and fed treatment diets for 28 days. On the final day of the experimental period (d 28) a meal challenge was performed (Table 1).

Diets and feeding

Test diets were formulated to represent Iams’ Proactive Adult Original with Chicken Dry Cat Food, which is a commercially available diet that is nutritionally complete and balanced (17.3% fat, 35.8% protein, 35.7% carbohydrate, 4041 kcal/kg dry matter). The MH diet was made by incorporating a water-soluble extract of whole-fruit avocado (MH source; Kemin Industries, Des Moines, IA) into the CON diet to deliver a MH dose of approximately 815 mg/kg diet (as described by ref. [1]). As no studies have fed MH to cats before, the targeted dietary dose selected was similar to that given to canines by McKnight et al. [1] to allow for comparison of plasma MH kinetics between species.

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Each animal was fed to their maintenance energy requirement, based on historical records of the individual dietary energy required to maintain body weight (~63 kcal/kg BW/d). Diets were present in dry, kibble form, and felines were fed individually once daily at 07:00 and allowed one hour to eat. All remaining food was collected and weighed to account for total food refusal. Body weight was measured weekly.

**Meal challenge**

Animals were fasted overnight and anaesthetized using butorphanol (0.25 mg/kg) and dexmedetomidine (0.0125 mg/kg). An 18 gauge SurFlash® polyurethane catheter (Terumo Medical Products, Butler, Ohio) was aseptically inserted into the jugular vein and the cat’s neck was covered with stockinet to prevent the animal from coming in contact with the catheter. At the end of the procedure, each cat was administered atipamezole (0.125 mg/kg, Antisedan®, Pfizer) as a sedative reversal and placed in a heated cage to recover.

Six hours after catheter placement one fasting blood sample was collected after which each cat was offered their full daily ration of test diet (CON or MH). Cats were fed individually and given 1 h to consume their test meal. Timing for blood sampling was initiated once the animal consumed approximately half of his or her test meal (less than 15 min for all cats). Blood samples (2 mL) were collected from the catheter using 1 mL syringes, every 2 h up to 24 h post-feeding. Blood was centrifuged for 8 min at 3,000xg at 7°C and the serum was stored separately at -80°C for subsequent analysis of MH. Plasma MH was determined by HPLC tandem mass spectrometry as described by McKnight et al. [1].

**Calculations**

Plasma MH concentrations at any time point (t) were described by an apparent zero-order absorption phase with first order elimination kinetics. Graphpad Prism version 6.04 (Graphpad Software, La Jolla California, USA, www.graphpad.com) was used to estimate area under the curve (AUC) using linear trapezoid rule and Microsoft Excel (2007) to determine: first order elimination rate constant (ke), elimination half-life (t1/2 in hours), apparent zero order absorption rate constant (K0 in µg/kg/hr), and turnover time (h). Concentration at steady state (Css) was estimated from the visual inspection and where the change in slope was <10%. All data are presented as mean and SEM (Table 2).

**Results and Discussion**

The primary aim of this study was to determine the appearance of ingested MH into the peripheral circulation of adult domestic short hair cats fed a MH containing diet. MH appeared into circulation within 2 h, remained at an apparent steady state for 11 hr post meal feeding, and disappeared within 24 h of ingestion (Figure 1). MH rate of appearance was described by a zero order rate of appearance of 0.52 ± 0.13 ng/kg/hr and first order rate of disappearance (0.38 ± 0.04 h⁻¹) resulting in a half-life of 1.86 ± 0.20 (h) and turnover time of 2.68 ± 0.29 (h). These findings confirm that MH is at least partially absorbed intact and biologically available to the animal.

In order to adequately describe the appearance and disappearance of MH (or any substrate) repeated blood sampling at multiple times...

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**Table 1:** Body weight, food and mannoheptulose consumption (mg) in adult domestic short hair cats fed a single meal of a MH containing diet. Time allotted in minutes for individual felines to consume test diet.

<table>
<thead>
<tr>
<th>Animal</th>
<th>BW (kg)</th>
<th>Time (min)</th>
<th>Food Intake (g)</th>
<th>MH Intake (mg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>4.11</td>
<td>60</td>
<td>42</td>
<td>31</td>
</tr>
<tr>
<td>2</td>
<td>4.05</td>
<td>60</td>
<td>52</td>
<td>38</td>
</tr>
<tr>
<td>3</td>
<td>4.57</td>
<td>&lt;20</td>
<td>63</td>
<td>46</td>
</tr>
<tr>
<td>4</td>
<td>3.97</td>
<td>&lt;20</td>
<td>52</td>
<td>38</td>
</tr>
<tr>
<td>5</td>
<td>3.91</td>
<td>&lt;20</td>
<td>44</td>
<td>32</td>
</tr>
<tr>
<td>6</td>
<td>4.03</td>
<td>&lt;20</td>
<td>13</td>
<td>10</td>
</tr>
<tr>
<td>7</td>
<td>4.33</td>
<td>&lt;20</td>
<td>59</td>
<td>43</td>
</tr>
<tr>
<td>8</td>
<td>4.27</td>
<td>&lt;20</td>
<td>39</td>
<td>28</td>
</tr>
<tr>
<td>Mean ± SD</td>
<td>4.15 ± 0.22</td>
<td>41.2 ± 13.3</td>
<td>37.1 ± 12.6</td>
<td></td>
</tr>
</tbody>
</table>

Data are presented as means and standard error and N=8 in a complete cross-over design.

**Figure 1:** Plasma mannoheptulose (ng/mL) concentrations in adult domestic short hair felines fed a mannoheptulose containing meal at time zero. For comparison, post-prandial plasma mannoheptulose concentrations in adult Beagles are also presented (adapted from ref. [1]).
Plasma MH concentrations are sustained in the cat for considerably longer than observed in the dog. The biological significance, if any, of sustained MH concentrations is presently unclear and would warrant further study. These results may however suggest that cats need lower doses or less frequent dosing for MH as compared to dogs. Furthermore, more work to investigate the potential benefits, such as the influence of MH on circulating glucose and insulin (and C peptide) concentrations, would be logical progression of this work.

**Acknowledgements**

Zhang J and Flickinger EA helped to the experimental design and A Gerwin, G Ruschman and G Balan provided direction on the interpretation of mannoheptulose kinetic results.

**Funding**

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**Conflict of Interest**

Shoveller AK and Davenport GM were employees of The Iams Company at the time this work was completed.

**References**


<table>
<thead>
<tr>
<th>Parameter</th>
<th>Mean Estimate (SEM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>( C_{ssav} ) (ug/mL)</td>
<td>1.79 ± 0.48</td>
</tr>
<tr>
<td>AUC (µg.hr/mL)</td>
<td>26.10 ± 8.11</td>
</tr>
<tr>
<td>( K_a ) (ng/kg/hr)</td>
<td>0.52 ± 0.13</td>
</tr>
<tr>
<td>( K_e ) (h(^{-1}))</td>
<td>0.38 ± 0.04</td>
</tr>
<tr>
<td>( T_{1/2} ) (h)</td>
<td>1.86 ± 0.20</td>
</tr>
<tr>
<td>Turnover time (h)</td>
<td>2.68 ± 0.29</td>
</tr>
</tbody>
</table>

**Table 2:** Post-prandial (24 h) plasma mannoheptulose kinetics in adult domestic short hair felines fed a mannoheptulose containing diet (8 mg/kg as fed). Css: Concentration at Steady State; AUC: Area Under Curve; \( K_a \): Absorption Constant; \( K_e \): Elimination Constant; \( T_{1/2} \): Half Life.