Preparative Separation of Lappaconitine from Aconitum leucostomum by HSCCC

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Keywords: Aconitum leucostomum; Alkaloids; Preparative chromatography; High speed counter-current chromatography (HSCCC)

Abstract

This paper describes a system for the preparation of lappaconitine from Aconitum leucostomum Worosch., using HSCCC, and examines the effect of the two-immiscible solvent system (TISS) composed of chloroform- methanol-0.2 mol/L hydrochloric acid (4:1.5:2). Under the optimized condition, about 75 mg of lappaconitine, at a purity of 98% that was done by high performance liquid chromatography (HPLC), are obtained from 250 mg of the crude alkaloid extract. The compound was checked by HPLC-UV, electrospray ionization mass spectrometry (ESI-MS), 1H NMR and 13C NMR.

Keywords

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Introduction

Aconitum leucostomum Worosch., which is a kind of perennial plant (Ranunculaceae family), is popular herbs in the northwestern area of China as Xinjiang and Gansu province [1,2]. The Roots of A. leucostomum are externally used as a folk medicine to treat traumatic injury and rheumatic disease [3], and known to contain a large amount of diterpenoid alkaloids, such as N-acetylsepaconitine, anthranoyllycoctonine, N-deacetylappaconitine, delcosine, delosine, OCH3 6, 14-dimethoxyfrosetine, lappaconitine, leucotines, leucotines A, leucotines B, lycaconitine, sepaconitine, etc. [4-6]. The major alkaloid, lappaconitine, was found to exhibit anti-inflammatory, tyrosinase inhibition and analgesic actions, and has been widely used as a pain reliever with neither toxic nor addictive [7-19] (Figure 1).

At present, the major diterpenoid alkaloids in A. leucostomum are often separated with conventional isolated techniques as both column chromatography and high-performance liquid chromatography (HPLC), etc., but the methods present a challenge for irreversible adsorption, lower sample recovery and time consuming [4-7]. However, the other separation method like to HSCCC for it, which can be advantages of high efficiency and recovery, are none dealt with [20-22].

For HSCCC, extraction by the counter-current distribution principle is especially suited to the separation of components in a mixture under conditions where the differences in the partition coefficients (K) are small. Thus, it is need to applied, where the method has distinct advantages over comparable multiple chromatographic techniques, to separate an alkaloid from each other. In addition, HSCCC can provide the analysis and separation method for the diterpenoid alkaloids where have a proton affinity and strong ultraviolet-visible absorption. Although the important step for the method is based upon partitioning of desired compounds between two immiscible solvent, it is focused on the stability and solubility, the acceptable ratio, the retention of the stationary phase (R) and the suitable K for target component [21,23]. To develop a suitable for diterpenoid alkaloids, pH is adjusting based on neutralization between mobile and stationary phases [24-26].

In this paper, we reported herein the separation and structural elucidation of lappaconitine in crude alkaloid extracts from the roots of A. leucostomum. The optimization of HSCCC separation condition is also discussed.

Material and Methods

Plant material and reagents

Whole plants of Aconitum leucostomum Worosch., collected in Burqin of Xinjiang of P.R. China, were identified by Professor Sheng Guan-ming at Xinjiang Institute of Ecology and Geography Chinese Academy of Sciences.

Methanol and chloroform were HPLC-grade reagents for HPLC purchased from Fisher Scientific Company (Fair Lawn, NJ, USA) and filtered with 0.45 µm filter. Other chemicals and solvents were of analytical grade purchased from Tianjin Chemical Factory (Tianjin, China).

Instruments

HSCCC were acquired using a model TBE-300 A high-speed countercurrent chromatograph (Shanghai Tauto Biotech Co., Shanghai, China) equipped with three polytetrafluoroethylene (PTFE)

Figure 1: Chemical structure of lappaconitine.

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multilayer coils (i.d.: 2.6 mm, total volume: 290 mL) on the rotary frame 5.0 cm from the central axis of the centrifuge, and a 20-mL manual sample loop. ρ value ranged from 0.5 for the internal terminal to 0.8 for the external terminal. ρ = r/R, where r is the distance from the column holder axis to the coil and R is the distance from the column holder axis to the centrifuge axis. In addition, HSCCC was also carried out on a model TBP-50 A constant-flow pump (Shanghai Tauto Biotech, Shanghai, China) and 8823A-UV Monitor (Institute of New Technology Application, Beijing, China) at 254 nm. Model N 2000 chromatography software package (Zhejiang University, Hangzhou, China) was used for instrument control, dat acquisition and processing.

HPLC system carried out on a Dionex LC system consisting of a 4-solvent delivery system P 680 type, an ASI-100 automated sample injector, a TCC-100 thermostatted column compartment, and a UVD 170 U detector and a Chrome Leon Work Station. Analytical TLC was performed using Si gel 60 F 254 precoated glass plates (0.25 mm thickness, 10 × 10 cm).

The mass spectrometry used here was a 4000 QTRAP LC/MS/MS (Applied Biosystems, USA). The H spectra were recorded on Inova-600 superconducting NMR spectrometer (Varian, Palo Alto, CA, USA) using a 5-mm sample tube. The 13C, APT, COSY, NOESY, HSCQ, and HMBC NMR experiments were run standard pulse sequences, with TMS was used as an internal standard.

Preparation of crude alkaloid extracts

The air-dried, ground roots of A. leucostomum (1.8 kg) were powdered and extracted with 80% ethanol solution (2 × 10 L) under reflux at 80°C for 2 h. Then the extracts were combined and concentrated under reduced pressure at 60°C. The concentrated residue was suspended in water (1:3), and then partitioned with chloroform twice (each times 1 L). The organic layers were combined and concentrated under reduced pressure at 40°C to give a chloroform-soluble residue, and acidified with 5% aqueous H2SO4 solution, after removal of the acid-soluble material, re-basified with Na2CO3 to 7.0 and 10.0, and then successively partitioned with chloroform to obtain 7.89 g (in pH 7.0, C001) and 1.38 g (in pH 10.0, C002) of two chloroform-soluble crude alkaloidal residues. The crude alkaloidal residue (in pH 7.0, C001, 7.89 g) was dissolved in acetone and filtered, and then concentrated to give crude alkaloid 7.38 g.

Selection of the two-phase solvent system

HSCCC separations using of the two immiscible solvents composed of chloroform, methanol and dilute HCl at volume ratios 4:1.5 to 4:3.2, and thoroughly equilibrating in a separatory funnel at room temperature. The composition of two-phase solvent system was selected as K values, which were also determined by HPLC, is especially suitable from 0.5 to 2.0 for the desired compounds in crude alkaloid extract. About 0.5 mg of crude sample was added to 4.0 mL of an equal volume mixture of the aqueous (upper) phase and the organic (lower) phase, and dissolved. The tube agitated to allow thorough mixing, after equilibrating at room temperature, the upper and lower phase were prepared for HPLC analysis. Then, K value is evaluated by both the peak area of a desired substance in the upper phase (A_U) and the lower phase (A_L). This is shown as K = A_U/A_L.

HSCCC separation

Two immiscible solvents for HSCCC separation was used of a mixture of 4 volumes of chloroform, 1.5 volumes of methanol and 2 volumes of 0.2 mol·L⁻¹ HCl. At each operation, the solvents will be thoroughly mixed, and degassed by sonication for 30 min shortly before use. The sample solution was prepared by dissolving 60 mg of crude sample in 10 mL mixture of 5 volumes of upper phase and 5 volumes of lower phase.

HSCCC separation was carried out preparative using the following procedure. First, for the lower phase (as mobile phase) pumped into the column at a flow-rate of 4.0 mL/min, while the apparatus was run at e revolution speed of 800 r/min. For hydrodynamic equilibrium of two immiscible solvents, as the mobile phase had been eluted from the tail outlet, had been reached (about 1 h later), then sample solution was injected, the peaks of interest were online detected at 254 nm. According to the peak chromatogram, each peak fraction was manually collected and then evaporated under reduced pressure. The residue was dissolved in methanol before proceeding with chromatographic analysis of HPLC.

TLC and reverse-phase HPLC-UV analysis

In this experiment, the crude extract, and the peak fraction from HSCCC separation was analyzed by TLC, with the solvent system composed of chloroform and methanol (5:1), and HPLC. For the sample solution, were dissolved 3.0 mg of the substance to be examined in methanol, sonicate if necessary. These were detected by TLC as blue spot prior to spraying under short-wave UV light using chloroform-methanol (5:1). The analysis was performed on YWC ODS-A column (250 × 4.6 mm, 5.0 µm) at 27°C with flow-rate of 1.0 mL/min. The mobile phase was composed of methanol, water, chloroform, and TEA (70:30:20:0.1). The auto-sampler was conditioned at 4°C and injection volume was 10 µL for sample injection. The peaks of interest were monitored with UV absorbance at 238 nm of wavelengths.

Identification of target compound

In ESI-MS analysis, ESI source was operated in positive ionization mode, and mass spectrometry detector parameters were as follows: desolvant gas temperature at 300°C; nitrogen flow rate, 500 L/h; source temperature, 120°C; capillary voltage, 4.0 kV; cone voltage, 25.0 V; mass scan from m/z 100 to 1000. All date was processed using Analyst 1.4.2 software. The structural identification of the peak fraction of HSCCC was elucidated by means of 1H and 13C NMR, COSY, NOESY, HSQC, and HMBC NMR experiments.

The target compound exhibited the flowing date

Amorphous powder, M.P 218-220°C; ESI-MS m/z: 585.2 [M+H]+, 567.2 [M-H2O]+, 535.2 [M-OCH3-H]+, 406.1 [M-C6H5NO3]+, 388.1 [M-C2H11NO3]+, 161.5 [M-C5H9NO3]+; H NMR (600 MHz, CDCl3) δ: 10.84 (1 H, s, NHCOCH3), 8.67 (1 H, d, J = 8.4 Hz, H-3), 7.84 (1 H, d, J = 7.8 Hz, H-6), 7.47 (1 H, dd, J = 7.8 and 7.8 Hz, H-4), 7.03 (1 H, dd, J = 7.8 and 7.8 Hz, H-5), 3.57 (2 H, br.s, H-1, H-21), 3.56 (1 H, s, H-14), 3.48 (1 H, s, H-17), 3.46 (3 H, s, OCH3, δ: 3.29 (1 H, dd, J = 7.8 and 7.8 Hz, H-16), 3.25 (1 H, s, H-19)), 3.07 (1 H, s, H-6), 3.02 (1 H, s, H-3), 2.94 (1 H, d, J = 6.6 Hz, H-19), 2.51 (1 H, d, J = 4.8 Hz, H-13), 2.43 (1 H, d, J = 6.6Hz, H-7), 2.34 (3 H, m, H-5, H-15, H-21), 2.24 (3 H, s, NCOCH3), 2.23 (2 H, d, J = 11.4 Hz, H-12, H-15), 2.15 (1 H, s, H-12, H-22), 1.72 (1 H, m, H-6), 1.36 (1 H, d, J = 12 Hz, H-2), 1.54 (1 H, d, J = 11.4 Hz, H-3), 1.50 (3 H, s, NCH2CH3); 13C NMR (150 MHz, CDCl3) δ: 79.8 (C-1), 28.0 (C-2), 29.6 (C-3), 84.7 (C-4), 48.9 (C-5), 26.3 (C-6), 46.9 (C-7), 75.0 (C-8), 80.7 (C-9), 47.0 (C-10), 50.5 (C-11), 23.4 (C-12), 36.4 (C-13), 89.1 (C-14), 43.9 (C-15), 82.3 (C-16), 61.1 (C-17), 56.4 (C-19), 50.5 (NCH2CH3), 10.5 (NCH3), 56.4 (OCH3), 58.0 (OCH3), 56.3 (OCH3), 166.6 (OOC), 114.4 (C), T42.0 (C-2), etc.
Other factors, such as flow rate of mobile phase, column temperature, and revolution speed, were also investigated and optimized. As seen from Figure 3, it can be seen that the target compound resolution decreased as the flow rate of mobile phase from 5 to 2 increases the separation of the compound, but it took a little longer, and the more mobile phase was needed with the peak tailing of the target compound. Among those, the flow rate of mobile phase is performed more rapidly (as 4.0 mL/min) for HSCCC separation. The column temperature has significant effect on K values and Rs. In present work, it was set at 25°C. Finally, the revolution speed of the multilayer coil separation column affects Rs. In this experiment, all separations were used at 850 r/min.

In order to separate the desired compound from the alkaloid extracts from A. leucostomum by HSCCC, the two-solvent systems composed of chloroform-methanol-0.2 mol/L HCl (4:1.5:2) was chosen first. Using the optimized conditions, about 75 mg of peak 1 fraction are successfully obtained from 250 mg of alkaloid extract from A. leucostomum by preparative HSCCC with a column capacity of 290 mL in less than 3 h, and at a purity of 98% that done by HPLC, as shown in Figures 4, 5, and 6, and Rs was 76%.

**Structure identification of lappaconitine**

We isolated from the dry roots of A. leucostomum a known diterpenoid alkaloids, lappaconitine. It was identified by its physicochemical properties (M.P, ESI-MS, 1H NMR, 13C NMR) and chromatographic behaviors (TLC and HPLC).

Only a brief outline of the spectroscopic evidence for the structural assignment of lappaconitine has been published [6,7]. In a previous paper, its structure was assigned on basis of multinuclear (1H, 13C NMR) assignment of lappaconitine has been published [6,7]. In a previous paper, its structure was assigned on basis of multinuclear (1H, 13C NMR) and 1H-1H COSY, 13C-1H COSY and 1H-13C HMBC. Thus, comparison of the physical and spectral
Figure 4: Chromatogram of the crude alkaloid extract from *A. leucostomum* by HSCCC.

Figure 5: Chromatogram of lappaconitine of the crude alkaloid extract from *A. leucostomum* by HSCCC.
date obtained for the compound with literature values established the structure of this diterpenoid alkaloid as lappaconitine [3-7,17-19].

Conclusion

A rapid, efficient, and economical HSCCC method for preparative isolation of lappaconitine from A. leucostomum was established. It was identified by its physicochemical properties (MP, ESI-MS, $^1$H NMR, $^{13}$C NMR) and chromatographic behaviors (TLC and HPLC). From the results of the HSCCC and HPLC method, we can conclude that the present method can be satisfactorily used as the reference substances for chromatography or bioactivity compound.

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References