

RAF-MEK-MAPK Pathway Targeted by Tumor Suppression and Anticancer Therapeutic Agents

Zhang X^{1*}, Zhou J¹, Li T³, Zheng B², He Z^{1*} and Huang Z^{1*}

¹Department of Pathophysiology, School of Basic Medical Science, and Chinese American Collaborative Cancer Institute, Provincial Key Laboratory of Medical Molecular Diagnostics, People's Republic of China

²Department of Microbiology, School of Laboratory Medicine, Guangdong Medical University, Dongguan, Guangdong, People's Republic of China

³Department of Thoracic Surgery, Hospital of Traditional Chinese Medicine of Zhonshan, Zhong Shan, Guangdong, People's Republic of China

Abstract

RAS-RAF-MEK-MAPK pathway comprises a group of kinases which regulates the activities of effector proteins in growth, proliferation and apoptosis. The extracellular signals from growth factors, cytokines and other stimuli transmitted by surface receptors and upstream signaling molecules are integrated by this cascade of kinases whose activity is regulated by the interaction of oncoproteins and tumor suppressors. The anomaly of the signaling pathway would lead to occurrence of malignancies. The RAS-RAF-MEK-MAPK pathway is therefore targeted by anticancer therapeutic agents. The present paper discussed the interaction of individual component with tumor suppressors and the impact of their inhibitor on the efficacy of anticancer therapy, and improvement on small molecule inhibitor of RAS-RAF-MEK-MAPK pathway with modified targeting has been proposed.

Keywords: Oncogene; RAS; RAF-MEK-MAPK pathway; Tumor suppression; Tyrosine Kinase Inhibitor (TKI); Anti-cancer therapy

Introduction

Kinases, notably those regulated the action of RAF-MEK-MAPK and AKT-mTOR-PI3K pathways have attracted increasing attention to identify anticancer therapeutic targets, in view of their activity in integration of signals of growth, proliferation, angiogenesis and apoptosis. Oncoprotein RAS activates the RAF/MEK/ERK (Extracellular Signal-Regulated Kinases) pathways, involving proteins of MAPK family as end effector; and pathways of PI3K (Phosphatidylinositol 3-Kinase)/Akt/NF-KappaB (Nuclear Factor-Kappa B) pathway, p120-GAP/p190-B/Rac/NF-Kappa-B, and Raf/MEK1/IKK (I-Kappa-B Kinase)/I-Kappa-B/NFKappa-B pathway activate transcription factor NF kappa-B through signaling molecules PI3K and Akt [1]. RAS proteins are encoded by members of oncogene RAS family with H-, Ki- and N-RAS, whose mutational activation has been seen more than 50% of human cancer cases [2], leading to cancerous cell growth. Inhibition of the kinases has demonstrated efficacy in therapy against cancers, and is also targeted by tumor suppressor genes (TSGs) when exerting their anti-oncogenic activities, through downregulation of cell cycle entry and angiogenesis, and potentiation of apoptosis. Mitogen-activated protein kinase (MAPK) is a serine/threonine kinase, which activates transcription factors and other cytoplasmic factors leading to mitogenesis [3]. The modulation of RAF-MEK-MAPK pathway in the context of oncogenes-TSGs interaction, and of intervention of anticancer drugs is to be discussed in the present paper.

The Implications of RAF-MEK-MAPK Pathway in Promotion of Cell Growth and Proliferation

The family of RAS gene comprises of a group of oncogenes that are frequently mutated in human tumors like pancreas, lung, and colorectal cancers and neuroblastoma. The prominent members of the family include N-Ras (neuroblastoma cell line), H-Ras (Harvey murine sarcoma virus), and the alternatively spliced K-Ras (Kirsten murine sarcoma virus). Among these, K-Ras is most frequently constitutively activated in human cancers [2]. The genes of this family code for RAS proteins, which reversely binds guanidine nucleotides of GDP and GTP. The metabolic forms of guanidine phosphate correspond with functional statuses of RAS, a small molecule G protein. RAS is

activated when recruiting adapter proteins such as Grb2 that in turn engages guanine nucleotide exchange factors (GEFs) like SOS to the cell membrane, GDP bound to RAS is replaced by GTP transferred by factors SOS [4,5].

RAS is normally activated in response to the binding of extracellular signals, such as growth factors, RTKs (Receptor Tyrosine Kinases), TCR (T-Cell Receptors) and PMA (Phorbol-12 Myristate-13 Acetate). The GTP associated Ras triggers the activation of a sequential three-kinase phosphorylation cascade through RAF, MEK, and ERK. RAF-MEK-ERK is essential for the regulation of cellular proliferation and survival [6]; the pathway integrates a wide range of signals into major cellular programs such as proliferation, differentiation, or apoptosis. And half of all human malignancies display aberrations in the RAS-RAFMEK-ERK pathway.

RAF is a downstream effector kinase of RAS, and is found in three isoforms: A, B and C-RAF (also called RAF-1 or C-RAF-1). Many studies showed the role of RAF kinase as a potential cellular oncogene for cancer therapy [7]. RAF-1 is a 74 kDa mitochondrial protein, ubiquitously expressed in adult tissues, with highest expression in muscle, cerebellum, and fetal brain. It was the first RAF isoform identified. The agents targeting the RAF family as a whole or C-RAF extensively examined in many pre-clinical studies and more recently some of them are in clinical trials [8-11].

B-RAF is a 94 kDa mitochondrial protein identified as second RAF isoform which acts as mutational target in various human cancers

***Corresponding author:** Dr. Xiangning Zhang, M.D., Ph.D.; Zhiwei He, M.D., Ph.D.; Zunnan Huang, Ph.D., Department of Pathophysiology, Guangdong Medical University, 1 Xincheng Avenue, Songshan Lake Scientific and Industrial Park, Dongguan, Guangdong 523808, People's Republic of China, Tel: 00867692896405, Fax: 008676922896100; E-mail: zhangxn_2006@126.com

Received April 18, 2017; Accepted May 09, 2017; Published May 12, 2017

Citation: Zhang X, Zhou J, Li T, Zheng B, He Z, et al. (2017) RAF-MEK-MAPK Pathway Targeted by Tumor Suppression and Anticancer Therapeutic Agents. J Mol Genet Med 11: 264 doi:10.4172/1747-0862.1000264

Copyright: © 2017 Zhang X, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

[1,12]. It is the strongest RAF kinase in terms of induction of MEK activity. A-RAF isoform is the weakest activator of MEK, and can only activate MEK1 but not MEK2. At present, no mutations in A-RAF have been found in human cancers [13,14].

MEK activates MAPK, the members of the family include extracellular signal regulated kinase (ERK) or MAPK, p38 MAPK, and JNK [15]. While different MAPK family members involve the same pathway with similar components with similar activities, the downstream effects notably regulators of cell cycle are not completely overlapped between different molecules.

In the field of anticancer drug, success in screening drugs targeting to the upstream factor RAS have been limited [16]. RAF and MEK, however, are important intermediates in the MAPK pathway [17], researchers invested efforts to screen inhibitors of RAF and MEK as agents in anticancer therapy.

The Regulation of RAF-MEK-MAPK Involving Interactions between Oncogenes and Tumor Suppressor Genes

The pathway is activated by oncoproteins, and targeted by tumor suppressors while exerting their anticancer potential, and in some context, the activity involves interactions between TSGs and oncogenes. TSGs are implicated in the genesis of malignancies in case of inactivation, through loss of heterozygosity (LOH) mutation, or epigenetic inactivation, in a manner of loss-of-function. As a result, the transformed cells are no longer harnessed in growth and proliferation by the regulation of cell cycle progression [18,19]. In fact, cell cycle progression, and hence cell proliferation is regulated by cyclin dependent kinases (CDKs) in complexed with cyclins transcriptionally activated mostly by RAS-RAF-MEK-MAPK pathway [20]. Tumor suppressors have been shown to downregulate the signaling axis of MAPK-cyclin, for example, JNK-cyclin D1, and some through interactions with RAS proteins [21-23].

FHIT is tumor suppressor mapped on 3p14, a frequently lost chromosomal region in human cancers; its inactivation is an early event in development of the cancer [24-26]. Data obtained from lung cancer lines have not just indicate the importance of FHIT in carcinogenesis but also its potential to serve as an early biomarker for lung cancer [27]. A new role of FHIT in down-regulating the Ras/Rho GTPase-associated oncogenic signaling pathway has been suggested [28].

We have reported that BLU, a TSG mapped on the same chromosomal region as RASSF1, i.e. 3p21 which is frequently lost in nasopharyngeal carcinoma (NPC) and a variety of human tumors, mainly of epithelial origin, suppressed the signaling of JNK pathway, and reduced the level of cyclin D1 to arrest cell cycle at G1 phase when it is re-expressed in negative NPC cells [29]. Structurally, BLU protein contains a zinc finger MYND domain on its amino-terminus. The molecular mechanisms underlying its downregulation of JNK-cyclin D1 axis remain to be elucidated.

Transcription factor AP1 is formed by, heteromerization of c-FOS and c-JUN, whose phosphorylation is catalyzed by JNK and another protein c-FOS. AP1 binds to the genomic DNA sequences upstream to the coding portion of a number of genes coding for cell cycle regulator, notably CCND1 coding for cyclin D1. Known as product of proto-oncogene, cyclin D1 promotes proliferation in malignancy through interaction with oncogenic molecules [30]. It has been reported that over-expressed cyclin D1 facilitates the infection of nasopharyngeal epithelial cells by a lymphotropic human herpesvirus, Epstein-Barr virus (EBV), a ubiquitous human virus that is tightly associated with

the occurrence of NPC and Burkitt lymphoma (BL) [31]. It has been proposed that MAPK pathway and cyclin D1 forms a signaling axis to regulate cell proliferation, and amplification of the chromosomal region that harbors CCND1 is a frequent anomaly at cytogenetic level during the occurrence of human tumors [32,33].

We reported that re-expression of BLU downregulated JNK signaling through reducing phosphorylation on JNK and inhibiting formation of AP1. It is reasoned that the effect was due to the inhibition of upstream kinase(s) by reducing their levels via epigenetic mechanism. In fact, we have shown that BLU inhibited the expression of IKK alpha, reduced the level of NFkappaB and hence NFkappaB dependent anti-apoptotic factors, so as to promote death receptor induced apoptosis [34]. It is speculated that BLU binds HDACs or SIRT, to repress transcription of genes coding for kinases in the pathway of RAF-MEK1/2-MAPK to downregulate the pathway and exert tumor suppression.

Previous study has shown that lymphoid-specific helicase (LSH), a SNF2-SWI chromatin remodeler, plays an essential role in cancer progression via regulation of fumarate hydratase (FH) [35]. Mechanistically, together with histone methyltransferase G9a, LSH is critical for the normal development of mammals and is involved in the establishment and maintenance of DNA methylation [36]. Since apart from depositing of H3K9me2 [37], G9a and its partner modifier GLP also interact with DNA methyltransferases (DNMTs) and protect proper DNA methylation at certain loci [38], LSH might also directly interact with DNMTs and affect the patterns of DNA methylation in the cells. Therefore, in human NPC cells, investigation of chromatin loading of LSH and the patterns of DNA methylation at the BLU locus might shed light on the mechanisms that required for the regulation of BLU and the progression of these malignant carcinomas.

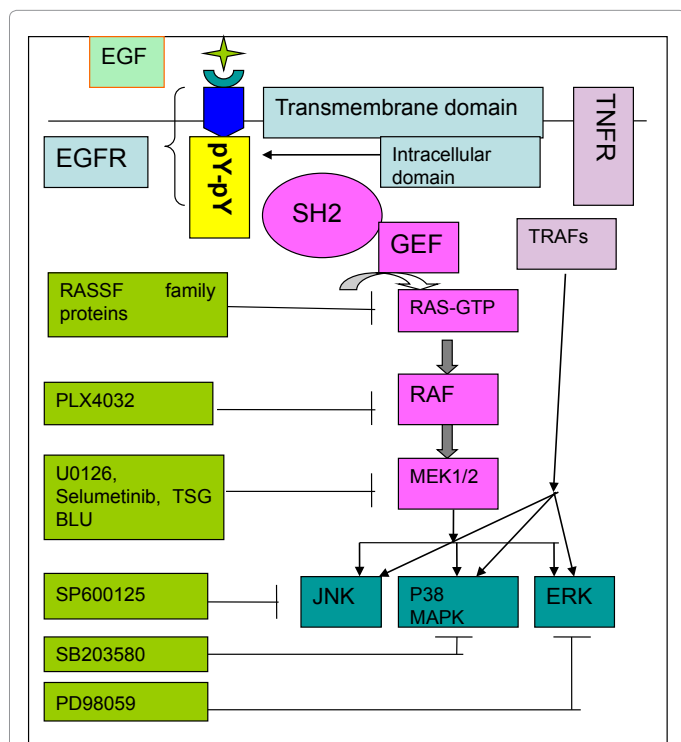
The tumor suppression of a family of proteins RASSF is exerted by downregulation of RAS-RAF-MEK1/2-ERK pathway. RASSF1A, the founding member of the RASSF family and RASSF5/NORE inhibits tumor growth and proliferation by targeting to signaling molecules of the MAPK family [23,35]. Phosphorylated ERK (pERK) is a key downstream component of the Ras/Raf/MEK/ERK signaling pathway. After phosphorylation, it translocates to the nucleus, and regulates various transcription factors such as Ets family transcription factors (Elk-1) [39].

The Implications of RAF-MEK-MAPK on Malignant Transformation of Cells

The aberrant activation of the RAF/MEK/MAPK signaling pathway is correlated with the occurrence of hepatocellular carcinoma (HCC) and a variety of human cancers. The activity of the RAF/MAPK signaling pathway was significantly higher, and the activity of ERK1/2 and MEK1 were upregulated threefold to fourfold in neoplastic liver specimens when compared to normal liver tissue adjacent to the HCC lesions [40,41]. Furthermore, it has been reported that the over-expression of RAF-1 could be regarded as an indicator of HCC prognosis [42]. These data suggest that the RAF/MEK/MAPK pathway may serve as an attractive target in the therapy of HCC.

The Applications of Components of RAF-MEK-MAPK Pathway in Anticancer Therapy

Epidermal growth factor receptor (EGFR) signaling is triggered by the binding of its ligand, resulting in the dimerization of EGFR molecules or heterodimerization with other closely related receptors, such as HER2/neu. EGFR is overexpressed in 40-80% of non-small



EGFR transmits growth signals when dimerizing triggered by EGF. Its intracellular domain of protein tyrosine kinase (PTK) catalyzes phosphorylation of tyrosine residues (p-Y), and the altered spatial configuration of the intracellular portion recruits adapter protein with SH2 motif, and then guanidine exchange factor to transfer GTP to RAS proteins. Through a cascade activation of RAS-RAF-MEK kinases, the effectors JNK, p38 MAPK and ERK are activated to induce transcription factor activity and then gene transcription is initiated. It is indicated that activity of components of this pathway are regulated by TSGs and small molecule inhibitors. MAPK family proteins are activated by trimerized TNFR triggered by its corresponding ligands, in a TRAF dependent manner with kinase activation [55].

Figure 1: Scheme of regulation of the signaling of RAS-RAF-MEK-MAPK pathway.

Targeting component	Small molecule inhibitor	References
EGFR Tyrosine Kinase	Genistein; AG 1478; gefitinib, erlotinib; afatinib, Icotinib	[62,63]
Mutant BRAF	PLX4032; dabrafenib, sorafenib	[64,65]
MEK1/2	PD 98059; Pimasertib U0126, Selumetinib (AZD6244, ARRY-142886) cobalt(II) complex 2	[55,56,66-69]
P38 MAPK	SB202190, SB203580	[70,71]
JNK	SP600125	[72]
ERK	PD98059	[73]

Table 1: Small molecules generated for targeting the components of the RAS-RAF-MEK-MAPK signaling pathway.

cell lung cancers (NSCLC) and many other epithelial cancers [43]. Besides the mutations in EGFR and HER2, KRAS genes are generally identifiable in over 40% of NSCLCs [44,45]. Tyrosine kinase inhibitors (TKIs), gefitinib and erlotinib acting as reversible inhibitors for binding to the ATP pocket of the protein tyrosine kinase were clinically approved agents targeting EGFR. In NSCLC, gefitinib was shown to induce partial responses (PR) in approximately 10% of cases [46]. The downstream kinases were also tested as target for therapeutic intervention, resulting in the development of several potent MEK inhibitors [47,48]. Patients with lung cancer whose tumors harbor EGFR and HER2 mutations

respond to EGFR TKIs but develop drug resistance can still benefit from the use of second generation tyrosine kinase inhibitors [49-51].

The downstream signaling that EGFR triggers is the RAS/RAS/MEK/ERK pathway. While RAF-MEK-MAPK pathway is targeted to be downregulated by TSGs, it has always been thought as an attractive pathway for anticancer therapy because it plays a central role in regulating various cellular processes from a broad spectrum of human tumors [52]. The activities of the pathway are often aberrant in tumors it therefore serves as a potential target for inhibitors of small molecule inhibition [53]. The blocking the post-translational modifications that promote Ras membrane association have been adopted as a strategy of anticancer therapy [54]. In recent years, various inhibitors have been developed for RAS downstream effector signaling, with efforts focused on the ERK/MAPK pathway (Figure 1).

In the recent years, small molecule intervention has been used targeting for the members of the RAS-RAF-MEK-ERK signaling pathway. The potential of AZD6244, a selective MEK1/2 inhibitor [55,56] which exerts anti-proliferative effects on NSCLC cell lines was characterized [57,58]. The dual inhibition of MEK and EGFR or MEK and signal transducer and activator of transcription 3 (STAT3) signaling pathways may constitute a potent therapeutic strategy for the treatment of KRAS mutant NSCLCs. But AZD6244 displays insufficient efficacy in non-small cell lung cancers (NSCLCs) due to deregulated expression and/or mutations of PIK3CA and PTEN [59-63]. A rational basis has been validated for choosing the inhibitor to best combine with the MEK inhibitor in cells on the basis of the expression and mutations of several oncogenes [64-74].

Emerging problems of using kinase inhibitors to treat cancer include acquired resistance in cancer cells [75]. A strategy to overcome to obstacle so as to achieve efficacy was to administer the drugs according to the gene profile of the individual patient, notably the status of EGFR, RAS and PTEN. Alternatively, combined administration of the inhibitor has been proved to be effective; for example, the use of BRAF inhibitor dabrafenib and MEK inhibitor trametinib in treating melanoma patients with BRAF V600E mutation has significantly improved progression-free survival of the patients [76]. We have identified glabridin, an isoflavone isolated from licorice as BRAF and MEK double targeting, based on computational modeling and shown that the drug suppressed the proliferation of HCC cells by inhibiting the phosphorylation of MEK1/2 and the downstream molecules including ERK1/2 and transcription factors ATF1 and CREB [77] (Table 1).

Conclusion

In conclusion, small molecule inhibitors to RAS-RAF-MEK-MAPK pathway have been validated as effective in therapy against a variety of cancers, and have gained wide application (Table 1). To circumvent the problems like acquired resistance, agents with multiple targeting are to be developed, and modality with combination of agents should be considered based on cancer related gene profile of the individual patients.

Acknowledgments

Our work is supported by Medical Science Research Fund, Guangdong Provincial Commission of Health and Family Plan (2014A267).

References

- Wellbrock C, Karasarides M, Marais R (2004) The Raf proteins take centre stage. Nat Rev Mol Cell Biol 5: 875-885.
- Matallanas D, Crespo P (2010) New druggable targets in the Ras pathway? Curr Opin Mol Ther 12: 674-683.

3. Gay B, Suarez S, Caravatti P, Furet P, Meyer T, et al. (1999) Selective GRB2 SH2 inhibitors as anti-Ras therapy. *Int J Cancer* 83: 235-241.
4. Agathangelou A, Cooper WN, Latif F (2005) Role of the Ras-association domain family 1 tumor suppressor gene in human cancers. *Cancer Res* 65: 3497-3508.
5. Rowinsky ER, Windle JJ, Von Hoff DD (1999) Ras protein farnesyltransferase: A strategic target for anticancer therapeutic development. *J Clin Oncol* 17: 3631-3652.
6. Arozarena I, Calvo F, Crespo P (2011) Ras, an actor on many stages: Posttranslational modifications, localization, and site-specified events. *Genes Cancer* 2: 182-194.
7. Hagemann C, Rapp UR (1999) Isotype-specific functions of Raf kinases. *Exp Cell Res* 253: 34-46.
8. Schreck R, Rapp UR (2011) Raf kinases: oncogenesis and drug discovery. *Int J Cancer* 119: 2261-2271.
9. Sridhar SS, Hedley D, Siu LL (2005) Raf kinase as a target for anticancer therapeutics. *Mol Cancer Ther* 4: 677-685.
10. Strumberg D, Seeber S (2005) Raf kinase inhibitors in oncology. *Onkologie* 28: 101-107.
11. Thompson N, Lyons J (2005) Recent progress in targeting the Raf/MEK/ERK pathway with inhibitors in cancer drug discovery. *Curr Opin Pharmacol* 5: 350-356.
12. Garnett MJ, Marais R (2004) Guilty as charged: B-RAF is a human oncogene. *Cancer Cell* 6: 313-319.
13. Nantel A, Huber M, Thomas DY (1999) Localization of endogenous Grb10 to the mitochondria and its interaction with the mitochondrial-associated Raf-1 pool. *J Biol Chem* 274: 35719-35724.
14. Yuryev A, Ono M, Goff SA, Macaluso F, Wennogle LP (2000) Isoform specific localization of A-RAF in mitochondria. *Mol Cell Biol* 20: 4870-4878.
15. Schaeffer HJ, Weber MJ (1999) Mitogen-activated protein kinases: Specific messages from ubiquitous messengers. *Mol Cell Biol* 19: 2435-2444.
16. Wong KK (2009) Recent developments in anti-cancer agents targeting the Ras/Raf/MEK/ERK pathway. *Recent Pat Anticancer Drug Discov* 4: 28-35.
17. Sebolt-Leopold JS, Herrera R (2004) Targeting the mitogen-activated protein kinase cascade to treat cancer. *Nat Rev Cancer* 4: 937-947.
18. Herman JG, Baylin SB (2003) Gene silencing in cancer in association with promoter hypermethylation. *N Engl J Med* 349: 2042-2054.
19. Baylin SB, Chen WY (2005) Aberrant gene silencing in tumor progression: implications for control of cancer. *Cold Spring Harb Symp Quant Biol* 70: 427-433.
20. Albanese C, Johnson J, Watanabe G, Eklund N, Vu D (1995) Transforming p21 ras mutants and c-Ets-2 activate the cyclin D1 promoter through distinguishable regions. *J Biol Chem* 270: 23589-23597.
21. Whang YM, Kim YH, Kim JS, Yoo YD (2005) RASSF1A suppresses the c-Jun-NH2-kinase pathway and inhibits cell cycle progression. *Cancer Res* 65: 3682-3690.
22. Yoo YA, Na AR, Lee MS, Yoon S, Kim JS (2006) RASSF1A suppresses oncogenic H-Ras-induced c-Jun N-terminal kinase activation. *Int J Oncol* 29: 1541-1547.
23. Yi M, Yang J, Chen X, Li J, Li X (2011) RASSF1A suppresses melanoma development by modulating apoptosis and cell-cycle progression. *J Cell Physiol* 226: 2360-2369.
24. Croce CM, Sozzi G, Huebner K (1999) Role of FHIT in human cancer. *J Clin Oncol* 17: 1618-624.
25. Huebner K, Croce CM (2003) Cancer and the FRA3B/FHIT fragile locus: It's a HIT. *Br J Cancer* 88: 1501-1516.
26. Huebner K (2001) Tumor suppressors on 3p: A neoclassic quartet. *Proc Natl Acad Sci USA* 98: 14763-14765.
27. Pekarsky Y, Palamarchuk A, Huebner K, et al. (2002) FHIT as tumor suppressor: mechanisms and therapeutic opportunities. *Cancer Biol Ther* 1: 232-236.
28. Jayachandran G, Sazaki J, Nishizaki M, Xu K, Girard L, et al. (2007) Fragile histidine triad-mediated tumor suppression of lung cancer by targeting multiple components of the Ras/Rho GTPase molecular switch. *Cancer Res* 67: 10379-10388.
29. Zhang X, Liu H, Li B, Huang P, Shao J, et al. (2012) Tumor suppressor BLU inhibits proliferation of nasopharyngeal carcinoma cells by regulation of cell cycle, c-Jun N-terminal kinase and the cyclin D1 promoter. *BMC Cancer* 12: 267.
30. Yu Z, Wang C, Wang M, Li Z, Casimiro MC, et al. (2008) A cyclin D1/microRNA 17/20 regulatory feedback loop in control of breast cancer cell proliferation. *J Cell Biol* 182: 509-517.
31. Tsang CM, Yip YL, Lo KW, Deng W, To KF, et al. (2012) Cyclin D1 overexpression supports stable EBV infection in nasopharyngeal epithelial cells. *Proc Natl Acad Sci USA* 109: E3473-E3482.
32. Liu Y, Hock JM, Sullivan C, Fang G, Cox AJ, et al. (2010) Activation of the p38 MAPK/Akt/ERK1/2 signal pathways is required for the protein stabilization of CDC6 and cyclin D1 in low-dose arsenite-induced cell proliferation. *J Cell Biochem* 111: 1546-1555.
33. Qin L, Yang YB, Yang YX, Gong YZ, Li XL (2014) Inhibition of smooth muscle cell proliferation by ezetimibe via the cyclin D1-MAPK pathway. *J Pharmacol Sci* 125: 283-291.
34. Zhou J, Huang Z, Wang Z, Liu S, Grandien A (2016) Tumor suppressor BLU promotes TRAIL-induced apoptosis by downregulating NF- κ B signaling in nasopharyngeal carcinoma. *Oncotarget* 14126.
35. He X, Yan B, Liu S, Jia J, Lai W, et al. (2016) Chromatin Remodeling Factor LSH Drives Cancer Progression by Suppressing the Activity of Fumarate Hydratase. *Cancer Res* 76: 5743-5755.
36. Myant K, Termanis A, Sundaram AY, Boe T, Li C (2011) LSH and G9a/GLP complex are required for developmentally programmed DNA methylation. *Genome Res* 21: 83-94.
37. Shinkai Y, Tachibana M (2011) H3K9 methyltransferase G9a and the related molecule GLP. *Genes Dev* 25: 781-788.
38. Zhang T, Termanis A, Özkan B, Bao XX, Culley J (2016) G9a/GLP Complex Maintains Imprinted DNA Methylation in Embryonic Stem Cells. *Cell Rep* 15: 77-85.
39. Moshnikova A, Frye J, Shay JW, Minna JD, Khokhlatchev AV (2006) The growth and tumor suppressor NORE1A is a cytoskeletal protein that suppresses growth by inhibition of the ERK pathway. *J Biol Chem* 281: 8143-8152.
40. Roberts PJ, Der CJ (2007) Targeting the Raf-MEK-ERK mitogen-activated protein kinase cascade for the treatment of cancer. *Oncogene* 26: 3291.
41. McKillop IH, Schmidt CM, Cahill PA, Sitzmann JV (1997) Altered expression of mitogen-activated protein kinases in a rat model of experimental hepatocellular carcinoma. *Hepatology* 26: 1484-1491.
42. Ito Y, Sasaki Y, Horimoto M, Wada S, Tanaka Y, et al. (1998) Activation of mitogen-activated protein kinases/extracellular signal-regulated kinases in human hepatocellular carcinoma. *Hepatology* 27: 951-958.
43. Chen L, Shi Y, Jiang CY, Wei LX, Wang YL (2011) Expression and prognostic role of pan-Ras, Raf-1, pMEK1 and pERK1/2 in patients with hepatocellular carcinoma. *Eur J Surg Oncol J Eur Soc Surg Oncol British Asso Surg Oncol* 37: 513-520.
44. Thomas RK, Baker AC, Debiassi RM, Winckler W, Laframboise T, et al. (2003) High-throughput oncogene mutation profiling in human cancer. *Nat Genet* 2007; 39: 347-351. Arteaga CL. ErbB-targeted therapeutic approaches in human cancer. *Exp Cell Res* 284: 122-130.
45. Adjei AA (2008) K-ras as a target for lung cancer therapy. *J Thorac Oncol* 3: S160-S163.
46. Fang B (2016) RAS signaling and anti-RAS therapy: Lessons learned from genetically engineered mouse models, human cancer cells, and patient-related studies. *Acta Biochim Biophys Sin* 48(1): 27-38.
47. Kris MG, Natale RB, Herbst RS, Lynch TJ, Prager D, et al. (2003) Efficacy of gefitinib, an inhibitor of the epidermal growth factor receptor tyrosine kinase, in symptomatic patients with non-small cell lung cancer: A randomized trial. *J Am Med Assoc* 290: 2149.
48. Ding L, Getz G, Wheeler DA, Mardis ER, McLellan MD, et al. (2008) Somatic mutations affect key pathways in lung adenocarcinoma. *Nature* 455: 1069-1075.
49. de La Rouge MT, Galluzzi L, Olausson KA, Zermati Y, Tasdemir E, et al. (2007)

- A novel epidermal growth factor receptor inhibitor promotes apoptosis in non-small cell lung cancer cells resistant to erlotinib. *Cancer Res* 67: 6253–6262.
50. Engelman JA, Zejnullahu K, Gale CM, Lifshits E, Gonzales AJ, et al. (2007) PF00299804, an irreversible pan-ERBB inhibitor, is effective in lung cancer models with EGFR and ERBB2 mutations that are resistant to gefitinib. *Cancer Res* 67: 11924–11932.
51. Li D, Shimamura T, Ji H, Chen L, Haringsma HJ, et al. (2007) Bronchial and peripheral murine lung carcinomas induced by T790M-L858R mutant EGFR respond to HKI-272 and rapamycin combination therapy. *Cancer Cell* 12: 81–93.
52. Mc Cubrey JA, Steelman LS, Chappel WL, Abrams SL, Wong EWT, et al. (2007) Role of the Raf/MEK/ERK pathway in cell growth, malignant transformation and drug resistance. *Biochim Biophys Acta* 1773: 1263-1284.
53. Ripple MO, Kim N, Springett RJ (2013) Acute mitochondrial inhibition by mitogen-activated protein kinase/extracellular signal-regulated kinase kinase (MEK) 1/2 inhibitors regulates proliferation. *J Biol Chem* 288: 2933-2940.
54. Gandhi J, Zhang J, Xie Y, Soh J, Shigematsu H, et al. (2009) Alterations in genes of the EGFR signaling pathway and their relationship to EGFR tyrosine kinase inhibitor sensitivity in lung cancer cell lines. *PLoS One* 4(2): e4576.
55. Takada Y, Ichikawa H, Pataer A, Swisher S, Aggarwal BB (2007) Genetic deletion of PKR abrogates TNF-induced activation of I κ B kinase, JNK, Akt and cell proliferation but potentiates p44/p42 MAPK and p38 MAPK activation. *Oncogene* 26: 1201–1212.
56. Singer G, Oldt R III, Cohen Y, Wang BG, Sidransky D, et al. (2003) Mutations in BRAF and KRAS characterize the development of low-grade ovarian serous carcinoma. *J Natl Cancer Inst* 95: 484–486.
57. Adjei AA, Cohen RB, Franklin W, Morris C, Wilson D, et al. (2008) Phase I pharmacokinetic and pharmacodynamic study of the oral, small-molecule mitogen-activated protein kinase kinase 1/2 inhibitor AZD6244 (ARRY-142886) in patients with advanced cancers. *J Clin Oncol* 26: 2139–2146.
58. Meng J, Peng H, Dai B, Guo W, Wang L, et al. (2009) High level of AKT activity is associated with resistance to MEK inhibitor AZD6244 (ARRY-142886). *Cancer Biol Ther* 8: 2071–2078.
59. Engelman JA, Chen L, Tan X, Crosby K, Guimaraes AR, et al. (2008) Effective use of PI3K and MEK inhibitors to treat mutant Kras G12D and PIK3CA H1047R murine lung cancers. *Nat Med* 14: 1351–1356.
60. Pratilas CA, Hanrahan AJ, Halilovic E, Persaud Y, Soh J, et al. (2008) Genetic predictors of MEK dependence in non-small cell lung cancer. *Cancer Res* 68: 9375–9383.
61. Wee S, Jagani Z, Xiang KX, Loo A, Dorsch M, et al. (2009) PI3K pathway activation mediates resistance to MEK inhibitors in KRAS mutant cancers. *Cancer Res* 69: 4286–4293.
62. Yoon YK, Kim HP, Han SW, Oh DY, Im SA, et al. (2010) KRAS Mutant Lung Cancer Cells Are Differentially Responsive to MEK Inhibitor Due to AKT or STAT3 Activation: Implication for Combinatorial Approach. *Molec Carcinog* 49: 353–362.
63. Li D, Ambrogio L, Shimamura T, Kubo S, Takahashi M, et al. (2008) BIBW2992, an irreversible EGFR/HER2 inhibitor highly effective in preclinical lung cancer models. *Oncogene* 27: 4702–4711.
64. Shi Y, Zhang L, Liu X, Zhou C, Zhang L, et al. (2013) Icotinib versus gefitinib in previously treated advanced non-small-cell lung cancer (ICOGEN): a randomised, double-blind phase 3 non-inferiority trial. *Lancet Oncol* 14(10): 953–961.
65. Hauschild A, Grob JJ, Demidov LV, Jouary T, Gutzmer R, et al. (2012) Dabrafenib in BRAF-mutated metastatic melanoma: a multicentre, open-label, phase 3 randomised controlled trial. *Lancet* 380: 358–365.
66. Long GV, Trefzer U, Davies MA, Kefford RF, Ascierto PA, et al. (2012) Dabrafenib in patients with Val600Glu or Val600Lys BRAF-mutant melanoma metastatic to the brain (BREAK-MB): A multicentre, open-label, phase 2 trial. *Lancet Oncol* 13: 1087–1095.
67. Mohr S, McCormick TS, Lapetina EG (1998) Macrophages resistant to endogenously generated nitric oxide-mediated apoptosis are hypersensitive to exogenously added nitric oxide donors: Dichotomous apoptotic response independent of caspase 3 and reversal by the mitogen-activated protein kinase kinase (MEK) inhibitor PD98059. *Proc Natl Acad Sci USA* 95: 5045–5050.
68. Morgillo F, Cascone T, D'Aiuto E, Martinelli E, Troiani T, et al. (2011) Antitumor efficacy of MEK inhibitors in human lung cancer cells and their derivatives with acquired resistance to different tyrosine kinase inhibitors. *Br J Cancer* 105: 382–392.
69. Favata MF, Horiuchi KY, Manos EJ, Daulerio AJ, Stradley DA, et al. (1998) Identification of a novel inhibitor of mitogen-activated protein kinase kinase. *J Biol Chem* 273: 18623–18632.
70. Li H, Zhou T, Liu H, Xu F, Niu Y, et al. (2017) Discovery of a cobalt complex with high MEK1 binding affinity. *Bioorg Med Chem Lett* S0960-894X (17) 30255-X.
71. Sicard P, Clark JE, Jacquet S, Mohammadi S, Arthur JS, et al. (2010) The activation of p38 alpha, and not p38 beta, mitogen-activated protein kinase is required for ischemic preconditioning. *J Mol Cell Cardiol* 48: 1324–1328.
72. Nemoto S, Xiang J, Huang S, Lin A (1998) Induction of apoptosis by SB202190 through inhibition of p38beta mitogen-activated protein kinase. *J Biol Chem* 273: 16415–16420.
73. Shin M, Yan C, Boyd D (2002) An inhibitor of c-jun aminoterminal kinase (SP600125) represses c-Jun activation, DNA-binding and PMA-inducible 92-kDa type IV collagenase expression. *Biochim Biophys Acta*. 1589: 311–316.
74. Kumar B, Sinclair J, Khandrika L, Koul S, Wilson S, et al. (2009) Differential effects of MAPKs signaling on the growth of invasive bladder cancer cells. *Int J Oncol* 34: 1557–1564.
75. Dobbstein M, Moll U (2014) Targeting tumour-supportive cellular machineries in anticancer drug development. *Nat Rev Drug Discov* 13: 179–196.
76. Flaherty KT, Infante JR, Daud A, Gonzalez R, Kefford RF, et al. (2012) Combined BRAF and MEK inhibition in melanoma with BRAF v600 mutations. *N Engl J Med* 367: 1694–1703.
77. Wang Z, Luo S, Wan Z, Chen C, Zhang X, et al. (2016) Glabridin arrests cell cycle and inhibits proliferation of hepatocellular carcinoma by suppressing braf/MEK signaling pathway. *Tumor Biol* 37: 5837–5846.