

Review of *Mansonella ozzardi* and its Vectors in the Neotropical Region with Emphasis on the Current Situation in Haiti

Christian P. Raccurt*

Department of Infectious and Communicable Diseases, University Notre-Dame of Haiti, Port-au-Prince, Haiti

*Corresponding author: Christian P. Raccurt, Department of Infectious and Communicable Diseases, University Notre-Dame of Haiti, Port-au-Prince, Haiti, E-mail: raccurt@yahoo.fr

Received date: February 17, 2017; Accepted date: March 03, 2017; Published date: March 08, 2017

Copyright: © 2017 Raccurt CP. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Mansonella ozzardi (Nematoda: Onchocercidae) is an understudied filarial nematode. This human parasite transmitted by two families of dipteran vectors, biting midges (most of them members of the genus *Culicoides*) and black flies (genus *Simulium*), is endemic to the Neotropic regions of the New World. With a patchy geographic distribution from southern Mexico to northwestern Argentina, human infection with *M. ozzardi* is highly prevalent in some of the Caribbean islands, along riverine communities in the Amazon Basin, and on both sides of the border between Bolivia and Argentina. Studies conducted in Haiti between 1974 and 1984 allowed the first complete description of adult worm and permitted to clarify the taxonomic position of this filarial species. In this paper, the author reports the known geographic distribution of *M. ozzardi* in neotropical regions of America and Caribbean, and focuses on the current situation in Haiti where this filariasis remains a completely neglected public health problem.

Keywords: *Mansonella ozzardi*; Neotropic; Haiti

Introduction

Mansonella ozzardi is a filarial nematode from the neotropical region, endemic as small foci in South America (Amazon Basin), Central America and the West Indies. Upon its discovery, *M. ozzardi* was the subject of confusion to some extent. Indeed, Sir Patrick Manson has described it under two different names: *Filaria demarquayi*, for the microfilariae found in the inhabitants of Saint Vincent Island, in the Lesser Antilles and *F. ozzardi*, for those found among the Indians of Guiana. With little research interest, due to the lack of serious pathogenic effects easily recognizable in humans, this filarial nematode has remained not very well-known for a long time, especially as the adult form was difficult to find. In the decade 1974-1984, studies were carried out on this parasite and its vectors in Haiti. They have resulted in the definitive description of the adult form of the filarial nematode [1], which allows to clarify the taxonomic position of the species [2] and to accumulate morphological and biological information on this parasite of the Onchocercidae family [3]. Singularly, *M. ozzardi* has adapted himself to two families of Diptera, possible intermediate hosts or effective vectors, depending on the geographical regions. Furthermore, the blood microfilariae are able to leave the vascular bed to migrate into the dermal interstitial fluid so that they can be detected by skin biopsy as well as by taking venous or capillary blood sample. Thus *M. ozzardi* establishes a kind of link between the human filarial species with blood microfilariae (*M. perstans*) and those with dermal microfilariae (*M. streptocerca*).

This article aims to highlight the knowledge gained over *M. ozzardi* and its vectors in Haiti for the last forty years, to review the literature, and to generate a renewed interest among researchers, policy makers, and donors in this filarial nematode which still very much neglected.

Historical Review

At the end of the nineteenth century, Manson described *M. ozzardi* under the name *Filaria ozzardi* from blood samples taken from the Indians of Guyana by Ozzard [4,5]. Daniels remarked that the microfilariae with tapered posterior extremities were very common among the Indians of this country. By performing autopsies, he thought he had discovered the adult forms [6,7]. But these adult worms, supposed to belong to the species *M. ozzardi*, were in fact *Wuchereria bancrofti* [8]. The first adult specimens of this new species (five female) were found in 1899 during the autopsy of a Caribbean from Saint Lucia in the Lesser Antilles. The genus *Mansonella* was created in 1929, because the morphological characteristics of the microfilariae and those given by the incomplete description of the adult made it impossible to link this species to any other known genus of the time [9]. Due to the success of experimental infection of the monkey *Erythrocebus patas* with third stage larvae from Haiti [10], the complete description of male and female adult worms was achieved almost a century after discovery of microfilariae [1] and the definitive taxonomic position of this nematode finally established [2].

Taxonomic position

The last review of the genus *Mansonella* [9], which includes low-pathogenic filaria living in subcutaneous tissues and in the muscular fascia of mammals, including humans, transmitted by Ceratopogonidae and/or Simuliidae, and which brings up to date the classification of this group of parasites, makes it possible to definitively classify this species (Table 1) [11]. Its scientific name is: *Mansonella* (*Mansonella*) *ozzardi* (Manson, 1897) Orihel & Eberhard, 1982 [1,4].

Geographical Distribution

Mansonella ozzardi is strictly a neotropical filaria. Since its discovery in Guyana and St. Vincent Island in the Caribbean, its presence has been reported in almost all Latin American countries,

from southern Mexico to northern Argentina, and in some fifteen islands of the Caribbean archipelago [12,13].

| | |
|--------------|-----------------------|
| Class | Nematoda |
| Subclass | Secernentea |
| Order | Spirurida |
| Suborder | Camallania |
| Super-family | Filarioidea |
| Family | Onchocercidae [8] |
| Subfamily | Onchocercinae [8] |
| Genus | <i>Mansonella</i> [9] |
| Subgenus | <i>Mansonella</i> [2] |
| Species | <i>ozzardi</i> [1,4] |

Table 1: Classification of *Mansonella ozzardi*.

In fact, the actual distribution of the parasite in the Western Hemisphere is only partially known because of the immensity of the territories concerned, access difficulties, the relatively limited number of epidemiological surveys carried out, limited interest raised by this low or non-pathogenic parasite, precarious and sometimes non-existent health infrastructures in some regions still underserved. Moreover, geographical distribution deserves to be periodically updated due to the spontaneous or induced disappearance of certain foci and the emergence of other foci due to migrations of infected populations. Three main endemic areas persist, or have existed in the Neotropical area where the importance of source of infection is not always well-known.

On the South American continent

M. ozzardi is located in the forest areas of the Amazon Basin [14] and the Orinoco Basin [15] where there are small foci scattered along the banks of the rivers. The countries concerned are Northern Brazil [16-33], the three Guianas : French Guiana [34], Surinam [35] and Guyana [36,37], Venezuela [15,38-41], Eastern Colombia [42-45], and Northeastern Peru [46,48].

Three other forest regions relate to this important group:

In the North, the focus straddling Northwestern Colombia and Eastern Panama with extensions to the Atlantic and Pacific coastal regions of Colombia. Information about this endemic area is old [49] and deserves to be verified and updated.

In the center of Brazil exist some foci that go up along the Xingu River in the North of Mato Grosso [50].

In the South, the focus straddling Southern Bolivia and Northern Argentina is still active, as illustrated by recent publications [51-54].

In these forest regions, the parasite infects predominantly Amerindian populations established along streams in small, scattered communities that are difficult to access. The prevalence is usually very high.

In Central America

The foci of the Yucatan peninsula in Mexico has been known since 1930 [55] and carriers of microfilariæ *M. ozzardi* have been reported at the same time in Panama among the Indians of the province of Darien, near the Colombian border [56]. *Mansonella ozzardi* has been reported in the three Mexican regions of Campeche, Yucatan and Quintana Roo [57-59]. No recent information relates to these Mexican foci which may have disappeared. The presence of *M. ozzardi* would also have been reported in Guatemala by O'Connor in 1937 [60]. An update of the situation in Central America and Mexico is therefore necessary.

In the Caribbean

Small foci have been reported over the last century in almost all the Lesser Antilles from the island of Viéques, East of Puerto [61], to Trinidad on the Northern coast [62-65]. The latter source of contamination was successfully treated with ivermectin [66,67]. In Guadeloupe, it seems that Le Dantec first reported the presence of *M. ozzardi* in 1924. In 1929, in the British Overseas Territories of the Caribbean, the endemic nature of the filaria was confirmed for the first time in Saint-Vincent [68]. Subsequently, *M. ozzardi* was regularly found in the French West Indies (St. Bartholomew, Guadeloupe, Desirade, Marie-Galante, Martinique) and in the Lesser Antilles (Antigua, Nevis, Dominica, Saint Lucia, Saint Vincent, Grenadine and St. Kitts) [3]. The contamination of a US blood donor returning from a cruise in these popular tourist destinations was also the subject of a report in 1978 [69]. The foci of the Lesser Antilles, particularly in the French departments of Guadeloupe and Martinique, seem to have spontaneously disappeared with the economic development of these islands. The last cases reported go back to about fifty years [70]. In the Greater Antilles, the presence of the parasite has curiously never been reported in Puerto Rico, Cuba, Jamaica, Dominican Republic. It has been reported only in two lepers from the Turks Islands, an archipelago located in the North of Haiti [71]. In Haiti, *M. ozzardi* was reported for the first time by the Rockefeller mission in 1920 [72] and then in an article on lymphatic filariasis in Puerto Rico [61]. Haiti has important foci in coastal areas colonized by the mangroves [3,73], a situation that persists until today [74,75]. A review of cases of carriers of microfilariæ - detected between November 1964 and June 1965 (144 blood smears) and February 1969 to February 1976 (280 blood smears) by the Service National d'Éradication de la Malaria (SNEM) and the medical laboratory of the hospital "Le Bon Samaritain", in Limbé, in the North of Haiti, from July 1969 to February 1976 (83 blood smears) - has been carried out by Ripert et al. [76]. Five hundred and seven slides showed microfilariæ with the specific morphological characteristics of *M. ozzardi*: absence of sheath, irregular and imbricated nuclei, posterior tapered extremity free of nuclei in its terminal portion. All subjects with microfilariæ came from localities on the coast, in a plain or valley close to the coastline at an altitude of less than 100 m. The distribution of the cases is shown on the map of Figure 1. They are grouped into small coastal foci of varying size.

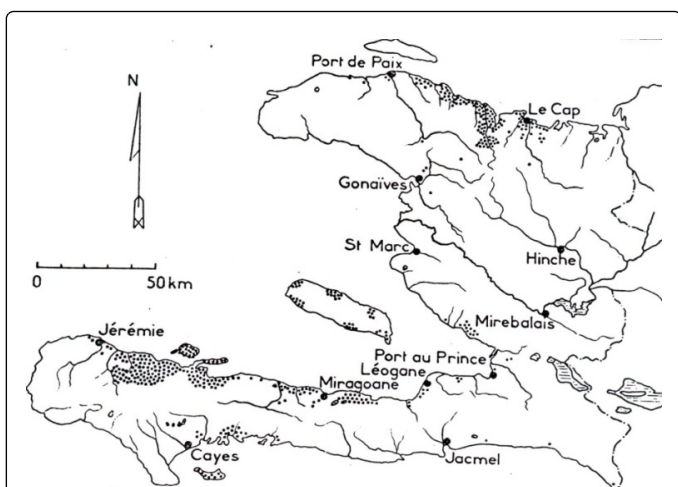


Figure 1: Map of the distribution of *Mansonella ozzardi* cases in Haiti (1964 – 1976) according to C. Ripert, C Raccurt and PL Douyon.

The two main foci are located in the Northern part of Haiti (coastal foci between Port-de-Paix and Cap Haitian, and along the Limbé river valley) and in the South-West of Haiti where numerous cases concentrate along the coast between Jérémie and Petit-Trou-de-Nippes, including the Baradères Peninsula and the Cayemittes Islands.

Small foci are observed on the peripheries of the island of La Gonâve, around Cabaret, Gressier, Leogane, in the Gulf of Gonâve, around Miragoâne, from Petit-Goâve to Anse-à-Veau, on the coasts of Nippes, around Saint-Louis-du-Sud, and on the island of Ile à Vache.

The two most important foci are found in marshy coastal areas where grown the mangroves, particularly favorable to the proliferation of *Culicoides furens* and *C. barbosai*, vectors of the parasite.

Vectors of *Mansonella ozzardi*

Depending on the region, the vectors of *M. ozzardi* belong to two groups of Diptera (suborder Nematocera). The morphological

| | Ceratopogonidae | Simuliidae | |
|-------------------------------|----------------------------------|--------------------------------|---|
| <i>Culicoides furens</i> | Caribbean, Yucatan | <i>Simulium gr. amazonicum</i> | Guyana |
| <i>Culicoides barbosai</i> | Haiti | <i>Simulium amazonicum</i> | Amazon Basin (Brazil, Colombia and Venezuela) |
| <i>Culicoides phlebotomus</i> | Trinidad | <i>Simulium argentiscutum</i> | Brazilian and Colombian Amazon |
| <i>Culicoides insinuatus</i> | Colombia | <i>Simulium sanguineum</i> | Colombia |
| <i>Culicoides guttatus</i> | Surinam | | |
| <i>Culicoides paraensis</i> | French Guiana, Brazil, Argentina | | |
| <i>Leptoconops bequaerti</i> | Haiti | | |

Table 2: Ceratopogonidae and Simuliidae species recognized as vectors of *Mansonella ozzardi* by geographic region.

Current Situation of Mansonellosis in Haiti

A study was conducted in June-July 2013 in the Corail region, in Grande Anse, by Raccurt et al. [75]. Finger-prick blood samples were

similarly of adult filariae found at autopsy of monkeys experimentally infected with third stage larvae from the Haitian *M. ozzardi* strain transmitted by culicoides and the Colombian strain transmitted by simulium confirmed that it was indeed a single species of filaria [10]. Similarly, the optical microscopy and electron microscopy studies of *M. ozzardi* microfilariae from Haiti and Colombia [77,78], showed no significant difference in their respective morphology and ultrastructure. We are therefore dealing with only one and the same species of filaria transmitted by two groups of competent vectors belonging to two kinds of Diptera: the Ceratopogonidae and the Simuliidae.

In 1933, a Ceratopogonidae, *C. furens*, was first recognized as vector of the parasite in St. Vincent, in the Caribbean [79,80], while in the Amazon Basin [81], Simuliidae have been recognized as the competent vector of *M. ozzardi*.

Culicoides furens is the main vector of *M. ozzardi* in the Yucatan peninsula in Mexico [59], and in the Caribbean, especially in Haiti [82]. Other culicoides ensure the transmission of the parasite such as *C. phlebotomus* in Northern Trinidad, [63,83,84] or *C. barbosai* in Southern Haiti [85].

At least one species has been identified as valid for larval maturation up to the third stage of *M. ozzardi* in South America: *C. insinuatus* in Colombia [86].

Taxonomic studies of *Simulium* fauna in Amazonia have shown that the species of Simuliidae, intermediate hosts and vectors of *M. ozzardi* in this region, belong to the amazonicum group [87-91].

From the data from the literature, we can list the vectors currently known according to the foci and in which *M. ozzardi* accomplishes larval development up to the infesting stage Table 2. Among the Ceratopogonidae, seven species are currently recognized as vectors, and four among the Simuliidae.

carried out in 462 people who provided their consent: 76 showed the presence of *M. ozzardi* microfilariae, an overall prevalence of 16.5%. In 70% of infected subjects, microfilaraemia was less than 10 microfilariae

per 20 microliters of capillary blood. The highest microfilaraemia (227 mf/20 μ L) was found in a 70-year-old woman. Among adults, males were more frequently infected (23%) than females (21%). These results are quite comparable to those found 35 years ago, in the Bayeux focus located in the Northern part of Haiti [73]. On the other hand, the site of Corail where this infection is developed is similar to that of the fishing village of Bayeux: the houses are constructed of materials that are permeable to the vectors living around and within the mangroves where two efficient vectors are proliferated: *C. furens* and *C. barbosa*.

Conclusion

In 45 years, the situation of Mansonellosis has remained stable in Haiti where *Mansonella ozzardi* persists in localized coastal foci. No action has been taken either to control filariasis by appropriate treatment or to combat culicoides proliferation during this time. Authorities have never paid attention to this filariasis known to be non-pathogenic, while affected individuals complain of fever, headache and chronic pruritus [92]. This situation is also explained by the fact that the economic situation in Haiti is very precarious and has not improved significantly over the last 50 years, especially in peripheral regions where poverty and illiteracy remain at very high levels. As already mentioned Raccurt et al. [74], it would be useful to place this neglected disease on the agenda to the Ministry of Public Health, and to provide ivermectin to the health facilities to treat the inhabitants, fishermen or cultivators, carriers of microfilariae and who complain of chronic symptoms such as fever, headache, pruritus. Surveillance campaigns for *M. ozzardi* among exposed populations should also be carried out to better control this endemic disease. Moreover, following the current efforts to develop tourism in Haiti, it would be urgent to pay serious attention to this completely neglected Filariasis. If tourists come back carrying the parasite after a stay in Haiti, this would undoubtedly have unfortunate consequences for the future of tourism in this country and who knows for potential new additional foci of this Filariasis in other parts of the world.

References

- Orihel TC, Eberhard ML (1982) *Mansonella ozzardi*: a redescription with comments on its taxonomic relationships. Am J Trop Med Hyg 31: 1142-1147.
- Eberhard ML, Orihel TC (1984) The genus *Mansonella* (syn. *Tetrapetalonema*): a new classification. Ann Parasitol Hum Comp 59: 483-496.
- Raccurt CP (1984) Contribution à l'étude de *Mansonella ozzardi* (Nematoda, Onchocercidae) et de ses vecteurs (Diptera, Ceratopogonidae) en Haïti. Thèse n° 84.43, Université Claude Bernard Lyon I, France.
- Manson P (1897) On certain new species of Nematode haematozoa occurring in America. Br Med J 2: 1837-1838.
- Ozzard AT (1897) A supposed new species of *Filaria sanguinis hominis* found in the interior of British Guiana. Br Guiana Med A 9: 24-27.
- Daniels CW (1898) The *Filaria ozzardi* and their adult forms. Br Guiana Med A 10: 1-15.
- Daniels CW (1899) The Probable Parental form of the Sharp-tailed filaria found in the blood of the aboriginals of British Guiana. Br Med J 1: 1459.
- Leiper RT (1913) Observations on certain helminths of man. Trans R Soc Trop Med Hyg 6: 265-297.
- Faust EC (1929) Human Helminthology. 1st Edn, Lea & Febiger, Philadelphia.
- Orihel TC, Lowrie RC Jr, Eberhard ML, Raccurt C, Kozek WJ, et al. (1981) Susceptibility of laboratory primates to infection with *Mansonella ozzardi* from man. Am J Trop Med Hyg 30: 790-794.
- Bain O, Mutafchiev Y, Junker K, Guerrero R, Martin C, et al. (2015) Review of the genus *Mansonella* Faust, 1929 sensu lato (Nematoda: Onchocercidae), with descriptions of a new subgenus and a new subspecies. Zootaxa 3918: 151-193.
- Sasa M (1976) Human filariasis. A global survey of epidemiology and control. Univ Tokyo Press: 819.
- Hawking F (1979) The distribution of human filariasis throughout the world part IV. America. Trop Dis Bull 76: 693-710.
- Shelley AJ (1975) A preliminary survey of the prevalence of *Mansonella ozzardi* in some rural communities on the river Purus, state of Amazonas, Brazil. Ann Trop Med Parasitol 69: 407-412.
- Medrano CE, Volcán GS, Godoy GA (1992) Mansonellosis in the southeast Venezuelan Orinoquia region. Rev Inst Med Trop Sao Paulo 34: 61-70.
- Richou RG (1957) Distribuição geográfica das filarioses humanas no Brasil. Rev Bras Malariol Coenças Trop 9: 79-100.
- Oliveira WR (1963) Filial infection in inhabitants of Vila Pereira, Territory of Roraima (Brazil). Rev Inst Med Trop Sao Paulo 5: 287-288.
- Lage HA (1964) Mansonellosis in Indians of the Aruak group of the Icana River region. Hospital (Rio J) 66: 557-564.
- Moraes MA, Almeida MM, Lovelace JK, Chaves GM (1978) *Mansonella ozzardi* among Ticuna Indians of the State of Amazonas, Brazil. Bol Oficina Sanit Panam 85: 16-25.
- Moraes MA, Shelley AJ, Luna Dias AP (1985) *Mansonella ozzardi* in the Federal Territory of Roraima, Brazil. Distribution and finding of a new vector in the area of Surumu River. Mem Inst Oswaldo Cruz 80: 395-400.
- Martins M, Pessoa FA, de Medeiros MB, de Andrade EV, Medeiros JF (2010) *Mansonella ozzardi* in Amazonas, Brazil: prevalence and distribution in the municipality of Coari, in the middle Solimões River. Mem Inst Oswaldo Cruz 105: 246-253.
- Basano S de A, Camargo J de S, Vera LJ, Velasques SN, Ogawa GM, et al. (2011) Investigation of the occurrence of *Mansonella ozzardi* in the State of Rondônia, Western Amazonia, Brazil. Rev Soc Bras Med Trop 44: 600-603.
- Basano S de A, Fontes G, Medeiros JF, Aranha Camargo JS, Souza Vera LJ, et al. (2014) Sustained clearance of *Mansonella ozzardi* infection after treatment with ivermectin in the Brazilian Amazon. Am J Trop Med Hyg 90: 1170-1175.
- Basano S de A, Medeiros JF, Fontes G, Vieira G de D, Camargo JS, et al. (2016) Occurrence of *Mansonella ozzardi* diagnosed using a polycarbonate membrane in a riverside population of Lábrea in the Western Brazilian Amazon. Rev Soc Bras Med Trop 49: 115-118.
- Medeiros JF, Py-Daniel V, Barbosa UC, Ogawa GM (2008) Current profile of *Mansonella ozzardi* (Nematoda: Onchocercidae) in communities along the Ituxi river, Lábrea municipality, Amazonas, Brazil. Mem Inst Oswaldo Cruz 103: 409-411.
- Medeiros JF, Py-Daniel V, Barbosa UC, Izzo TJ (2009) *Mansonella ozzardi* in Brazil: prevalence of infection in riverine communities in the Purus region, in the state of Amazonas. Mem Inst Oswaldo Cruz 104: 74-80.
- Medeiros JF, Py-Daniel V, Barbosa UC (2011) Prevalence of *Mansonella ozzardi* among riverine communities in the municipality of Lábrea, State of Amazonas, Brazil. Rev Soc Bras Med Trop 44: 186-190.
- Medeiros JF, Costa CA, Lima AM, Pessoa FA (2014) *Mansonella ozzardi* (Nematoda: Onchocercidae) in the riverine population of the Tefé River, State of Amazonia, Brazil. Rev Soc Bras Med Trop 47: 113-115.
- Medeiros JF, Pessoa FAC, Camargo LMA (2014) Mansonellosis: a Brazilian neglected disease. Rev Patol Trop 43: 1-6.
- Medeiros JF, Rodrigues MS, Katsuragawa TH, Costa CA, Pessoa FA (2014) *Mansonella ozzardi* in the municipality of Tefé, Amazonas, Brazil, 60 years after the first report: an epidemiologic study. Mem Inst Oswaldo Cruz 109: 480-483.
- Adami YL, Moraes MAP, Lanfredi RM, Main-Herzog M (2008) An atypical microfilaria in blood samples from inhabitants of Brazilian Amazon. Parasitol Res 104: 95-99.

32. Adami YL, Rodrigues G, Alves MC, Moraes MA, Banic DM, et al. (2014) New records of *Mansonella ozzardi*: a parasite that is spreading from the state of Amazonas to previously uninfected areas of the state of Acre in the Purus River region. Mem Inst Oswaldo Cruz 109: 87-92.
33. Ta-Tang TH, Luz SL, Merino FJ, de Fuentes I, López-Vélez R, et al. (2016) Atypical *Mansonella ozzardi* microfilariae from an endemic area of Brazilian Amazonia. Am J Trop Med Hyg 95: 629-632.
34. Floch H, Abonnenc E (1950) Présence de *Wuchereria bancrofti*, *Mansonella ozzardi* et *Acanthocheilonema perstans* en Guyane française. Cah Med Union Fr 5: 623-626.
35. Bruijning CF (1957) Notes on the common species of *Culicoides* (Diptera: Ceratopogonidae) from Surinam in relation to *ozzardi filariasis*. Doc Med Geogr Trop 9: 169-172.
36. Orihel TC (1967) Infections with *Dipetalonema perstans* and *Mansonella ozzardi* in the aboriginal Indians of Guyana. Am J Trop Med Hyg 16: 628-635.
37. Nathan MB, Tikasingh ES, Munroe P (1982) Filariasis in Amerindians of western Guyana with observations on transmission of *Mansonella ozzardi* by a *Simulium* species of the *amazonicum* group. Tropenmed Parasitol 33: 2119-2122.
38. Beaver PC, Neel JV, Orihel TC (1976) *Dipetalonema perstans* and *Mansonella ozzardi* in Indians of southern Venezuela. Am J Trop Med Hyg 25: 263-265.
39. Godoy GA, Volcan G, Medrano C, Teixeira A, Matheus L (1980) *Mansonella ozzardi* infections in Indians of the Southwestern part of the state of Bolivar, Venezuela. Am J Trop Med Hyg 29: 373-376.
40. Formica S, Botto C (1990) Filariasis focus due to *Mansonella ozzardi* and *Mansonella perstans* in the Amazon Federal Territory of Venezuela. J Trop Med Hyg 93: 160-165.
41. Gómez J, Guerrero R (2000) Environmental factors and the distribution of mansonelliasis in southern Venezuela. Parasite 7: 71-76.
42. Marinkelle CJ, German E (1970) Mansonelliasis in the Comisaría del Vaupes of Colombia. Trop Geogr Med 22: 101-111.
43. Kozek WJ, D'Alessandro A, Silva J, Navarette SN (1982) Filariasis in Colombia: prevalence of mansonellosis in the teenage and adult population of the Colombian bank of the Amazon, Comisaria del Amazonas. Am J Trop Med Hyg 31: 1131-1136.
44. Kozek WJ, Palma G, Henao A, García H, Hoyos M (1983) Filariasis in Colombia: prevalence and distribution of *Mansonella ozzardi* and *Mansonella (= Dipetalonema) perstans* infections in the Comisaría del Guanía. Am J Trop Med Hyg 32: 379-384.
45. Kozek WJ, Palma G, Valencia W, Montalvo C, Spain J (1984) Filariasis in Colombia: prevalence of *Mansonella ozzardi* in the Departamento de Meta, Intendencia del Casanare, and Comisaría del Vichada. Am J Trop Med Hyg 33: 70-72.
46. Marcos LA, Arrospide N, Recuenco S, Cabezas C, Weil GJ, et al. (2012) Genetic characterization of atypical *Mansonella (Mansonella) ozzardi* microfilariae in human blood samples from northeastern Peru. Am J Trop Med Hyg 87: 491-494.
47. Vargas-Herrera J, Arróspide-Velasco N, Gutierrez-González S, Celis-Salinas JC, Huamani-Solano D, et al. (2013) Reporte de cuatro casos clínicos de filariasis en Alto Nanay, Loreto. Rev Peru Med Exp Salud Publica 30: 506-511.
48. Vargas J, Arróspide N, Gutiérrez S, Obregón C, Valencia P, et al. (2015) Mansonellosis by *Mansonella ozzardi* in volunteers undergoing screening for malaria in the Peruvian Amazon. Rev Peru Med Exp Salud Publica 32: 265-271.
49. Mc Coy OR (1933) Occurrence of microfilaria *ozzardi* in Panama. Am J Trop Med Hyg 13: 297-306.
50. D'Andretta Jr, Pio da Silva CM, Kameyna F (1969) Ocorrência da mansonelose entre índios do alto Xingu. Rev Soc Bras Med Trop 3: 11.
51. Shelley AJ, Coscarón S (2001) Simuliid blackflies (Diptera: Simuliidae) and ceratopogonid midges (Diptera: Ceratopogonidae) as vectors of *Mansonella ozzardi* (Nematoda: Onchocercidae) in northern Argentina. Mem Inst Oswaldo Cruz 96: 451-458.
52. Dantur Juri MJ, Veggiani Aybar CA, Ortega ES, Galante GB, Zaidenberg MO (2013) *Plasmodium vivax* and *Mansonella ozzardi* co-infection in north-western Argentina. Malar J 12: 248.
53. Veggiani Aybar CA, Dantur Juri MJ, Zaidenberg MO (2016) *Mansonella ozzardi* in Neotropical region of Argentina: Prevalence through time (1986-2010). Acta Tropica 153: 1-6.
54. Lima NE, Veggiani Aybar CA, Dantur Juri MJ, Ferreira MU (2016) *Mansonella ozzardi*: a neglected New World filarial nematode. Pathog Glob Health 110: 97-107.
55. Hoffman CC (1930) Nota sobre la existencia de Microfilaria *ozzardi* en la Peninsula de Yucatan. An Inst Biol Univ Mex 1: 55-57.
56. Petersen JL, Bawden MP, Wignall FS, Latorre CR, Johnson CM, et al. (1984) *Mansonella ozzardi* in Darién (Panama). Rev Med Panama 9: 236-246.
57. Mazzotti L (1942) Comprobacion de la existencia de la *Microfilaria ozzardi* en Mexico. Rev Inst Salubr Enferm Trop Mex 3: 223-228.
58. Biagi F (1956) Observations on mansonelliasis in the Peninsula of Yucatan. I. Frequency. below Medicina (Mex) 36: 521-526.
59. Biagi F, Tay J, De B De Biagi AM (1958) Observations on mansonellosis in the Yucatan peninsula. V. *Culicoides* furen as vectors. Medicina (Mex) 38: 377-379.
60. Brumpt LC (1949) Présence de *Mansonella ozzardi* chez une femme de la Guadeloupe. C R Séance Soc Biol 143: 468-469.
61. Hoffman WA, Marin RA, Burke AMB (1932) Filariasis in Puerto-Rico. Puerto Rico J Pub Health Trop Med 7: 321-362.
62. Ashcroft MT (1965) A history and general survey of the helminth and protozoal infections of the West Indies. Ann Trop Med Parasitol 59: 478-493.
63. Nelson GS, Davies JB (1976) Observations on *Mansonella ozzardi* in Trinidad. Trans R Soc Trop Med Hyg 70: 16-17.
64. Nathan MB, Tikasingh ES, Nelson GS, Santiago A, Davies JB (1979) The prevalence and distribution of *Mansonella ozzardi* in coastal north Trinidad, W.I. Trans R Soc Trop Med Hyg 73: 299-302.
65. Chadee DD, Rawlins SC, Doon R, Baboolal S (1994) Presence of annular rings on *Mansonella ozzardi* microfilariae. Trans R Soc Trop Med Hyg 88: 356.
66. Chadee DD, Tilluckdharry CC, Rawlins SC, Doon R, Nathan MB (1995) Mass chemotherapy with diethylcarbamazine for the control of Bancroftian filariasis: a twelve-year follow-up in northern Trinidad, including observations on *Mansonella ozzardi*. Am J Trop Med Hyg 52: 174-176.
67. Gonzalez AA, Chadee DD, Rawlins SC (1999) Ivermectin treatment of mansonellosis in Trinidad. West Indian Med J 48: 231-234.
68. Cameron TW (1929) Observations on a Parasitological tour of the Lesser Antilles. Proc R Soc Med 22: 933-941.
69. Weller PF, Simon HB, Parkhurst BH, Medrek TF (1978) Tourism-acquired *Mansonella ozzardi* microfilaremia in a regular blood donor. JAMA 240: 858-859.
70. Courmes E, Fauran P, Lespinasse JP (1968) Contribution to the study of human filariasis in Guadeloupe. Bull Soc Pathol Exot Filiales 61: 234-245.
71. Stafford JL, Hill KR, De Montaigne EL (1955) Microfilariasis in the Turks Islands; report of two cases. West Indian Med J 4: 183-187.
72. Raccurt CP (1999) Filariasis in Haiti: a century of history. Bull Soc Pathol Exot 92: 355-359.
73. Raccurt C, Lowrie RC Jr, McNeeley DF (1980) *Mansonella ozzardi* in Haiti. I. Epidemiological survey. Am J Trop Med Hyg 29: 803-808.
74. Raccurt CP, Brasseur P, Boncy J (2014) Mansonelliasis, a neglected parasitic disease in Haiti. Mem Inst Oswaldo Cruz 109: 709-711.
75. Raccurt CP, Brasseur P, Cicerón M, Boncy J (2014) Epidemiologic survey of *Mansonella ozzardi* in Corail, Haiti. Am J Trop Med Hyg 90: 1167-1169.
76. Ripert C, Raccurt C, Douyon PL (1977) La filariose *Mansonella ozzardi* en Haïti (Grandes Antilles). Premières données épidémiologiques. Bordeaux Médical 10: 689-696.

77. Kozek WJ, Eberhard ML, Raccurt C (1983) Comparative morphology of *Mansonella ozzardi* microfilariae from Colombia and Haiti. A light microscope study. Tropenmed Parasit 34: 33-37.
78. Kozek WJ, Raccurt C (1983) Ultrastructure of *Mansonella ozzardi* microfilariae, with a comparison of the South American (Simulid-transmitted) and the Caribbean (Culicoid-transmitted) forms. Tropenmed Parasit 34: 38-53.
79. Buckley JJC (1933) A Note on the development of *Filaria ozzardi* in *Culicoides furens* Poey. J Helminth 11: 257-258.
80. Buckley JJC (1934) On the development, in *Culicoides furens* Poey, of *Filaria* (= *Mansonella*) *ozzardi* Manson, 1897. J Helminth 12: 99-118.
81. Cerqueira NL (1959) Sobre a transmissao de *Mansonella ozzardi*. Note 1 et 2. Jorn Bras Med 1: 885-914.
82. Lowrie RC Jr, Raccurt C (1981) *Mansonella ozzardi* in Haiti II. Arthropod vector studies. Am J Trop Med Hyg 30: 598-603.
83. Nathan MB (1978) *Culicoides phlebotomus*, a vector of *Mansonella ozzardi* in coastal North Trinidad, West Indies. Trans R Soc Trop Med Hyg 72: 436-437.
84. Nathan MB (1980) Transmission of the human filarial parasite *Mansonella ozzardi* by *Culicoides phlebotomus* (Williston) (Diptera: Ceratopogonidae) in coastal North Trinidad. Bull Ent Res 71: 97-105.
85. Lowrie RC Jr, Raccurt C (1984) Assessment of *Culicoides barbosai* as a vector of *Mansonella ozzardi* in Haiti. Am J Trop Med Hyg 33: 1275-1277.
86. Tidwell MA, Tidwell MA (1982) Development of *Mansonella ozzardi* in *Simulium amazonicum*, *S. argentiscutum* and *Culicoides insinuatus* from Amazonas, Colombia. Am J Trop Med Hyg 31: 1137-1141.
87. Shelley AJ, Dias AP (1980) *Simulium argentiscutum* sp. nov. (Diptera: Simuliidae), a member of the *S. amazonicum*-group of species: description of adults, pupa and larva. Mem Inst Oswaldo Cruz 75: 105-111.
88. Tidwell MA, Peterson BV, Ramirez Perez J, Tidwell M, Lacey LA (1981a) Notas y claves preliminares de los jejenes neotropicales de los grupos *Simulium amazonicum* y *S. sanguineum* (Diptera : Simuliidae) incluyendo los vectores de *Onchocerca volvulus* y *Mansonella ozzardi*. Boln Dir Malar San Amb 21: 77-87.
89. Tidwell MA, Tidwell M, Peterson BV (1981b) A redescription of the female of *Simulium sanguineum* Knab and description of the male, pupa, and larva (Diptera: Simuliidae). Proc Ent Soc Wash 83: 13-27.
90. Ramirez Perez J, Peterson BV (1981) Estudio del complejo *Simulium amazonicum-sanguineum* en Venezuela. Descripcion de tres nuevas especies. Boln Dir Malar San Amb 21: 150-160.
91. Shelley AJ, Pinger RR, Moraes MAP (1982) The taxonomy, distribution, biology, and medical importance of *Simulium amazonicum* Goeldi (Diptera: Simuliidae) with a review of related species. Bull Br Mus Nat Hist, Ent ser 44: 1-29.
92. McNeeley DE, Raccurt CP, Bony J, Lowrie RC Jr (1989) Clinical evaluation of *Mansonella ozzardi* in Haiti. Trop Med Parasitol 40: 107-110.