



## Significance of Bitter Leaf (*Vernonia Amagdalina*) In Tropical Diseases and Beyond: A Review

Clement Egharevba<sup>1</sup>, Erharuyi Osayemwenre<sup>2</sup>, Vincent Imieje<sup>2</sup>, Joy Ahomafor<sup>3</sup>, Christopher Akunyuli<sup>2</sup>, Anthony Adeyanju Udu-Cosi<sup>2</sup>, Onyekaba Theophilus<sup>3</sup>, Osakue James<sup>4</sup>, Iftikhar Ali<sup>5</sup> and Abiodun Falodun<sup>6</sup>\*

<sup>1</sup>Department of Pharmacognosy, Faculty of Pharmacy, University of Benin, Nigeria

<sup>2</sup>Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Benin, Nigeria

<sup>3</sup>Department of Pharmaceutical Chemistry, Faculty of Pharmacy, Delta State University, Abraka, Delta State, Nigeria

<sup>4</sup>Department of Physiology, School of Basic Medical Sciences, University of Benin, Benin City, Nigeria

<sup>5</sup>Institute of Chemistry, University of Rostock, Albert-Einstein-Str. 3A, 18059 Rostock, Germany

<sup>6</sup>Department of Pharmacognosy, School of Pharmacy, University of Mississippi, 38655 Oxford, Mississippi, USA

\*Corresponding author: Abiodun Falodun, Department of Pharmacognosy, School of Pharmacy, University of Mississippi, 38655 Oxford, Mississippi, USA, Tel: 662-638-5786, +2348073184488; E-mail: afalodun@olemiss.edu

Received date: April 04, 2014; Accepted date: June 3, 2014; Published date: June 10, 2014

Copyright: © 2014 Abiodun F et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

### Abstract

*Vernonia amygdalina* Delile (VA), family Asteraceae or Compositae is plants that is consumed locally as food and serve important ethnomedicinal uses. It grows throughout tropical Africa to a height of about 1 – 5 metres and it is indigenous to many West African Countries. Many parts of the plants are useful, they are used locally for the treatment of fever, Stomach disorder, jaundice, worm infestation, constipation, malaria, hiccups, kidney problems, amoebic dysentery, schistosomiasis, cough, wounds, diabetes, laxative, venereal diseases and other bacterial and protozoal infection. This review examines, discusses and summarizes the current evidence of ethnomedicinal uses, phytochemistry and biological activities as well as toxicity of this species with a view to identifying its therapeutic relations and possible contradictions, inconsistencies and gaps that may have arisen in the research literature. This review is based on literature study on journals and books of scientific origin from library, both manual and electronic sources such as PubMed, Science Direct, Elsevier, ACS, google Scholar etc using various combinations of search words. *V. amygdalina* is a tropical plant with a lot of interesting biological and medicinal uses. The plants are relatively not toxic, safe for consumption and possess a great potential as pharmaceutical leads for the treatment of diseases and beyond. This review will stimulate further research in the pharmacology and phytochemistry of *V. amygdalina*.

**Keywords:** *Vernonia amygdalina*; Plant extract; Pharmacology; Medicinal uses; Phytochemistry; Pharmacotherapy

### Introduction

The Asteraceae (Compositae) are herbs, shrubs, or less commonly trees and is arguably the plants largest family of flowering and has approximately 1,620 genera and more than 23,600 species [1]. *Vernonia* is a genus of about 1,000 species of forbs and shrubs of which *V. amygdalina* is the most prominent specie and one of the pan tropical tribes of the family Asteraceae [2]. It grows predominantly in tropical Africa especially in Nigeria, Zimbabwe and South Africa and it is domesticated in parts of West Africa [3-5]. It is popularly called bitter leaves because of its bitter taste and is used as vegetables or as flavour decoction soups. The bitter taste of VA is as a result of its anti-nutritional components such as alkaloids, saponins, glycosides and tannins [6-8]. In Nigeria, it is known by several local names such as "Ewuro" in Yoruba language, "Onugbu" in Igbo language, "Oriwo" in Bini language, "Ityuna" in Tiv language, "Chusar doki or fatefate" in Hausa language and "Etidot" in Ibibio [9]. The roots and leaves decoction of VA are commonly used in ethno medicine to treat fevers, hiccups, kidney problems and stomach discomfort among other several uses [10,11]. It is also used in the treatment of diarrhea, dysentery hepatitis and cough and as a laxative and fertility inducer

[11]. The leaves of VA are also commonly used as a treatment against nematodes in humans and chimpanzees as well as other intestinal worms [12,13]. In addition, extracts of the plants have been reported to be used in Nigerian herbal homes as tonic, in the control of tick and treatment of hypertension [14-16]. The reported activity of VA is attributable to the complex active secondary plant compounds that are pharmacologically active [17]. Besides, this review also discusses the toxicity of this species and it gave a reasonable information that no abnormality or toxicity was caused by the administration of some of the extracts as well as single compounds on the organs of the animal samples. Nevertheless, there is need to carry out further study on chemical constituents and their mechanisms of action in other to fully comprehend its full phytochemical profile and complex pharmacological effects. Furthermore, clinical studies on the toxicity of all the plant parts extracts and the compounds isolated from this plant are required to ensure that they are eligible as sources of drugs.

### Medicinal Properties

The aqueous and alcoholic crude extracts of the leaves, bark, stem and roots are reported to be widely used as antimalarial, for the treatment of eczema and as a purgative [18-20]. The roots and the leaves of VA are used in traditional medicine to treat fever, stomach discomfort, hiccups and kidney problems [21]. It is known as quinine

substitute because it is widely used for the treatment of fevers [21]. The wood, particularly those from the root is a tooth cleaner, an appetizer, fertility inducer and also for gastrointestinal upset [22]. The root infusion is taken in Nigeria for the treatment of intestinal worms as well as for enteritis and rheumatism [22]. Wild Chimpanzees have been observed to eat both the leaves and stems of the plant as a medication for self-deparasitization [23]. Other documented medicinal uses include the treatment of schistosomiasis, amoebic

dysentery, treatment of malaria, wound healing, venereal diseases, hepatitis and diabetes [24,25]. Fresh leaves of VA have been reported to have abortifacient and purgative activities [26]. It is used in some part of Africa to prepare cough remedy [27]. The chopped roots of VA are used for the treatment of sexually transmitted diseases in parts of Zimbabwe [28]. The root of VA is used for its antifertility effect and for the treatment of amenorrhoea [28] (Table 1).

Part	Medicinal uses	References
Whole Plant	Snakebite and insect sting treatment, dyspepsia, influenza, dysentery, malaria and respiratory infections.	Kirtikhar and Basu, Chopra et al.
Leaf	Fever, colic pain, loss of appetite irregular stools and diarrhea, common cold, cough, fever, hepatitis, tuberculosis, mouth ulcers, bronchitis gastro-intestinal disorder and sores .	Selena et al. Panossian et al. Bensky and Gamble, Dymock et al.
Aerial part	Common cold, hypertension, diabetes, cancer, malaria and snakebite, urinary tract infection.	Panossian et al. Bensky and Gamble, Dymock et al. Perry,
Root	Febrifuge, tonic, stomachic and anthelmintic	Chopra et al.

Table 1: Medicinal uses of VA.

### Phytochemistry

The phytochemical studies of *Vernonia amygdalina* reveals the presence of saponins, flavonoids, alkaloids, terpenes, steroids,

coumarins, phenolic acids, lignans, xanthenes, anthraquinones, edotides and sesquiterpenes [29] (Table 2).

Compound	Type	Plant part	Reference
Andrographolide	Diterpenoid lactone	Leaves/aerial	Reddy et al. Klaijipool, Puri et al. Matsuda et al. Cheung et al. Du et al. Kumar et al.
Neoandrographolide	Diterpenoid lactone	Leaves/aerial	Chan et al. Rao et al. Jain et al. Yang et al.
14-deoxyandrographolide	Diterpenoid lactone	Aerial parts	Matsuda et al. Kumar et al. Rao et al. Wu et al.
Andrographoside	Diterpene	Leaves/aerial parts	Matsuda et al. Rao et al.
14-deoxy-11, 12-didehydroandrographolide	Diterpenoid lactone	Aerial parts	Balman and Connolly, Wu et al. Jain et al.

19-O-β-D-glucopyranosyl-ent-labda-8(17), 13-dien-15, 16, 19-triol	Ent-labdane diterpenoid lactone	Aerial parts	Zou et al.
8α-methoxy-14-deoxy-17β-hydroxyandrographolide	Ent-labdane diterpenoid lactone	Aerial parts	Ma et al.
Andrographolactone	Diterpenoid lactone	Aerial parts	Xou et al.
3, 13, 14, 19-tetrahydroxy- ent-labda-8(17), 11-dien-16, 15 olide and 3, 19 isopropylidene- 14-deoxy- ent-labda-8(17), 13-diene-16, 15-olide	Diterpenoid lactone	Aerial parts	Xou et al.
14-deoxy-15-isopropylidene-11,12-didehydroandrographolide	Unusual Terpenoid	Aerial parts/roots	Reddy et al.
3,7,19-trihydroxy-8,11, 13- ent-labdatriene-15, 16-olide and 8α,17β-epoxy-3, 19-dihydroxy-11,13-ent-labdatrien-15, 16-olide	Diterpene lactone	Aerial parts	Ma et al.
Andrograpanin	Diterpene	Leaves	Liu et al.

Table 2: Compounds of VA.

### Sesquiterpenes

Different types of sesquiterpenes lactones have been isolated from VA, they include vernolide, vernodalol, vernolepin, vernodalin, vernomygdin, hydroxyvernolide, vernodalinol, vernomenin, vernolic, 11, 13-dihydrovernodalol, 11, 13-dihydrovernoderoline, 4, 15-dihydrovernodalol, 1, 2, 3, 15, 11, 13, 2', 3'-octahydrovernodalol and epivernodalol [17,29,30-33].

### Stigmastane-type steroid glucosides:

These includes vernoniosides A1 (C<sub>35</sub>H<sub>52</sub>O<sub>10</sub>, MW 632), vernoniosides A<sub>2</sub> (C<sub>35</sub>H<sub>52</sub>O<sub>10</sub>, MW 632), vernoniosides A<sub>3</sub> (C<sub>35</sub>H<sub>50</sub>O<sub>10</sub>, MW 630), vernoniosides A<sub>4</sub> (C<sub>35</sub>H<sub>52</sub>O<sub>11</sub>, MW 648),

vernoniosides B<sub>1</sub> (C<sub>35</sub>H<sub>52</sub>O<sub>10</sub>, MW 632), vernoniosides B<sub>2</sub> (C<sub>36</sub>H<sub>52</sub>O<sub>12</sub>, MW 680), vernoniosides B<sub>3</sub> (C<sub>37</sub>H<sub>54</sub>O<sub>11</sub>, MW 674), vernoniosides D (C<sub>35</sub>H<sub>52</sub>O<sub>12</sub>, MW 664), venonioside D<sub>2</sub> (C<sub>35</sub>H<sub>50</sub>O<sub>10</sub>) and vernoniosides E (C<sub>37</sub>H<sub>58</sub>O<sub>11</sub>) [34-37].

### Steroid alcohol:

7, 24(28)-stigmastadien-3-β-ol [38].

### Flavonoids:

These are; luteolin, luteolin 7-O-β glucoside and luteolin 7-O-glucuronoside, Myricetin [39,40] (Table 3).

Compound	Type	Plant part	Reference
5, 7, 2/, 3/-tetramethoxyflavonone	Flavonone	Whole plant	Rao et al.
5-hydroxy-7, 2/, 3/-trimethoxy flavones	Flavone	Whole plant	Rao et al.
5-hydroxy-7,2/,6/trimethoxyflavone	Flavone	Root	Rao et al.
7-O-methyldihydrowogonin	Flavone	Root/aerial part	Redi et al. Rao et al.
7-O-methylwogonin	Flavone	Root/aerial part/whole plant	Redi et al. Rao et al. Gupta et al. Kuroyanagi et al.
Flavone-1, 2/methylether	Flavone	Root/aerial part/whole plant	Redi et al. Rao et al. Jalal et al.
7-O-methylwogonin-5-glucoside	Flavones	Root/aerial parts	Redi et al. Rao et al. Kuroyanagi et al.
Dihydroskullcapflavone	Flavone	Whole plant	Hari et al.

5-hydroxy-7, 8, 2, 3/ tetramethoxyflavone	Flavone	Whole plant	Kuroyanagi et al. 1987 Rao et al.
---	---------	-------------	--------------------------------------

Table 3: Flavonoids of VA.

### Peptides:

The peptides are known as edotides and were first isolated [41] (Table 4).

Arabinogalactan	Protein	Herbs	Singh et al.
1, 8-dihydroxy-3, 7-dimethoxy-xanthone	Xanthone	Root	Dua et al.
4,8-dihydroxy-2,7-dimethoxy-xanthone	Xanthone	Root	Dua et al.
1,2-dihydroxy-6, 8-dimethoxy-xanthone	Xanthone	Root	Dua et al.
3,7,8-trimethoxy-1-hydroxy-xanthone	Xanthone	Root	Dua et al.
Andrographidoid A	Noriridoid	Root	Xu et al.
Andrographidoid B	Noriridoid	Root	Xu et al.
Andrographidoid C	Noriridoid	Root	Xu et al.
Andrographidoid D	Noriridoid	Root	Xu et al.
Andrographidoid E	Noriridoid	Root	Xu et al.

Table 4: Peptides of VA.

### Essential oil:

Essential oil obtained from hydrodistillation of the aerial parts yields 1, 8 - cineol (eucalyptol) (25.11%),  $\beta$  - pinene (14.54%), myrtanal (6.52%), trans-pinocarveol (6.24%), linalool (4.28%) and  $\alpha$ -pinene (4.93%) as the major components as well as other minor components [42]. Palmitic acid (22%),  $\alpha$ -linoleic acid (Omega-3, 21.5%) and linoleic acid (Omega-6, 15.8%) were the major fatty acids obtained from hexane/ Isopropanol extract of VA leaves which had a yield of 0.31% w/w [43].

### Pharmacology

The Pharmacological properties of VA have been investigated with a view to validate the wide traditional uses of the plant as a therapeutic agent. Several research has shown that VA possesses the following activities; antidiabetic, antiplasmodial, cathartic, hepatoprotective, antimicrobial, antioxidant, chemoprotective and cytotoxic, antihelmintic, hypolipidaemic, anti-platelet and abortifacient activities.

### Anti-diabetic:

A number of studies have demonstrated the anti-diabetic properties of VA. Erasto et al. [44] demonstrated the *in vitro* anti-diabetic properties. The *in vivo* anti-diabetic have also been proven. [45-49]. While the clinical studies have also showed that VA improved glucose tolerance, fasting blood sugar and postprandial blood glucose levels in

normoglycemic subjects [50]. Ethanolic extract of the leaves of VA improves glucose tolerance in streptozotocin-induced diabetic and normal Wistar rats at a dose of 400 mg/kg, 500 mg/kg metformin and ethanol vehicle served as positive and negative controls respectively. In the experiment, pre-treatment of the Wistar rats test group with 400 mg/kg of VA before inducing diabetics brought about 18.4% increase in the blood glucose of the rats (relative to blood glucose at time zero) compared to 36.6% increase in Diabetic control rat group after 1 hour of glucose loading, though metformin performed better at improving the glucose tolerance by reducing the blood glucose level to 5%, [51]. The possible antidiabetic mechanism of action of VA could be due to the fact that the aqueous extract of VA has been known to enhanced glucose utilization and uptake of muscles and liver cell cultures [44].

### Antihelmintic activities:

Hot water extract of VA has shown no significant *in vitro* anthelmintic activity against *Haemonchus Contortus* eggs at concentrations up to 11.2 mg/ml which gave up to 97.5% hatch rate as against 0.0% at concentration of 25 $\mu$ g/ml for albendazole standard [52]. This also corroborates an earlier *in vivo* trial [53]. Though it has been suggested that wild chimpanzees and gorillas eat VA for self-deparasitization [54] the above *in vitro* and *in vivo* studies does not seem to support the claim. However, since the primates eat the whole plant. It may therefore be argued that it is the purgation after ingesting the plant material that potentially causes the mechanical agitation and expulsion of helminthes.

### Antioxidant Properties:

The antioxidant properties of VA are associated with its chemical constituents. Several researchers [55,56] have previously established that the antioxidant activities of fruits and vegetables like VA are related to their phenolic compounds. Natural polyphenols scavenges free radicals, chelates metal catalysts, activates antioxidant enzymes, reduces  $\alpha$ -tocopherol radicals and inhibits oxidases, all of which have chain-breaking antioxidant activities and contributing to the prevention of degenerative diseases, cancer and atherosclerosis [57] has established that the DPPH radical scavenging activities of the leaves of VA fractions were significantly higher (P<0.05) than that of Glutathions. They also established that the polar polyphenolic compounds from VA were significantly better (P>0.05) at DPPH radical scavenging than the non-polar compounds (which mostly contain chlorophyll) present in non-polar acetone eluate. Fasakin et al. [58] demonstrated that leaf extract fractions of VA displayed weaker superoxide scavenging activities against superoxide radicals produced from pyragallol autoxidation when compared to Glutathione. It was also established that the 80% acetone extract fraction (Chlorophyll-enriched) of VA exhibited stronger superoxide radical scavenging ability than 70% ethanol and 70% methanol. This study is important because, though superoxide ions cannot directly initiate lipid oxidation, they are potential precursors of highly reactive species such as hydroxyl radical. Also 80% acetone fraction of VA leaves displayed significantly higher hydroxyl scavenging abilities than Glutathione, unlike ethanol fractions which displayed lower hydroxyl scavenging

activities than Glutathione. Polyphenolic fractions VA also showed a higher metal chelating ability than Glutathione [59-61]. The polyphenolic compounds in the acetone extract leaves of VA extract are good hydrogen and electron donors, this was demonstrated by its reduction of  $Fe^{3+}$ -ferricyanide complex to the ferrous state and measured at 700 nm and was found to be directly proportional to the reducing at power of VA [61]. The result showed that the ferric reducing ability of the VA extract fraction is similar to that of Glutathione extract fraction.

### **Hypolipidemic:**

Hypercholesterolemia is a risk factor for cardiovascular diseases such as myocardial infarction and atherosclerosis, which is a common cause of mortality and morbidity [62,63]. Even though several factors such as age, lifestyle, diet rich in cholesterol and hypertension, have been reported to cause heart failure [64], increased levels of cholesterol, especially low density lipoprotein cholesterol, are mainly responsible for hypercholesterolemia. Several researches have shown that hypercholesterolemia is associated with enhanced oxidative stress related to increased lipid peroxidation. High generation of oxidized LDL is a major factor in the vascular damage associated with high cholesterol levels. Thus, the inhibition of oxidative stress under hypercholesterolemia is considered a vital therapeutic approach [65-68]. The screening of VA as hypolipidemic drug was borne out of the desire to discover new drugs from nature that can serve as a credible alternative to the present synthetic drugs such as fibrates, bile acids sequestrants and statins which are currently used for such purpose, but however, has severe adverse effect such as rhabdomyolysis which can be caused by taking a statin with another lipid-lowering drug, particularly fibrates [69]. The lipid-lowering effect of methanol extract of *Vernonia amygdalina* (MEVA) leaves in rats fed on high cholesterol diet, and compared with a standard hypolipidemic drug, Questran have been investigated. The effects of MEVA on the lipid profile were assessed by measuring the levels of total cholesterol, LDL cholesterol, HDL cholesterol, triglyceride, lipid peroxidation (LPO), phospholipid, and glutathione (GSH) in the plasma and liver of the rats. Cholesterol administered at a dose of 30 mg/ 0.3 ml, five times in a week for nine consecutive weeks resulted in a significant increase ( $p < 0.05$ ) in plasma and post mitochondrial fraction (PMF) cholesterol levels by 33% and 55%, respectively. But, treatment with MEVA at doses of 100 mg/kg and 200 mg/kg caused a dose dependent reduction in plasma and PMF cholesterol by 20%, 23% and 23%, 29%, respectively. These reductions in cholesterol were similar to the ones obtained in Questran-treated rats. VA enhanced the cholesterol-induced decrease in PMF glutathione levels of the rats [70]. In general, it can be deduced that these results suggest the lipid-lowering effects of VA and as a result could serve as potential natural product for the treatment of hyperlipidaemia.

### **Anticancer activity:**

The different extracts of VA have been investigated against different types of cancer cell lines. Various fractions from different extracts of VA were investigated against cells derived from human carcinoma of the nasopharynx in tissue culture. The compounds from VA were cytotoxic sesquiterpene lactones; vernodalin, vernomygdin and vernolide with activity of ED<sub>50</sub> of 1.8, 1.5 and 2.0  $\mu$ g/ml respectively against KB cell culture [15]. Ethanol extract of VA have also been reported to possess considerable *in vitro* cytotoxic activity (IC<sub>50</sub>=60.33) against non-cancerous vero cell line (C-1008 kidney fibroblast from African green monkey) by neural red uptake method

[71]. Also, the *in vitro* growth inhibitory and cytotoxic evaluation of the epivernodalol, a known sesquiterpene lactone compound from VA leaves against skin melanoma cell line (HT-144) by the sulforhodamine B (SRB) assay have been investigated [72]. It was demonstrated that various concentrations (3 – 100 mg/ml) of water soluble extract of VA potently inhibited extracellular cellular Signal-Regulated Kinase (ERK) activities, DNA synthesis and cell growth in a dose-dependent manner. The results showed that VA in least concentrations of up to 10 mg/ml exhibited cytostatic action to retard the growth of human breast cell cancer cells, both in the presence or absence of serum [41].

Further studies conducted by Adesanoye et al. investigated the *in vitro* mechanisms of VA leaf extracts against breast cancer. The study demonstrated that, in a concentration and time dependent manner, VA-induced cytotoxicity and apoptosis in MCF-cells (breast cancer cell lines) involved phosphatidyl serine externalization followed by secondary necrotic cell death [73]. Kupchan et al. reported that vernomygdin, vernodalin showed inhibitory effects against human nasopharynx carcinoma cell line. Vernomenin, vernodalin and vernolepin also showed inhibitory activity against P-388 and L-1210 mouse leukemic cancer cell lines, while only vernodalin possessed antitumor activity against nasopharynx carcinoma KB cancer cell lines [30]. VA extract inhibited cell proliferation and DNA synthesis of BT-549 Breast cancer cells in *in vitro* antiproliferative study [74,75].

### **Cathartic effect:**

The methanol extract of VA produced significant ( $p < 0.05$ ) promotion of gastrointestinal motility of charcoal meal in mice, gastric emptying of gastrointestinal contents was also promoted. Though these effects were not as pronounced as that of carbachol (1mg/kg), VA extract treatment produced a dose-dependent increase of the total number of faeces in rats. The reference drug used, senna, produced defecation to a greater degree than the extract. The increased gastrointestinal motility was attributed to the presence of saponin which may act by direct gastric mucosa irritation [76] or reasonable content of calcium ions [77] which is essential for various physiological responses including glandular secretion and muscle contraction; these findings support the traditional use of VA for constipation and stomach upset [78].

### **Abortifacient:**

The methanol extract of VA when administered to pregnant mice caused abortion within 24 hours [26].

### **Antifertility:**

Recent report indicated that 95% ethanol crude extract of the leaves of VA possesses an *in vitro* anti-implantation effect at doses of 0.385, 0.5 and 1.0 g/kg body weight of mice in isolated mouse uterus compared to control agonist Acetylcholine (1 g/kg). It caused a significant reduction in mean number of implantation sites compared to the control, there was also a significant ( $p < 0.05$ ) reduction in the number of live fetuses and survival percentages between the controls and test groups at a dose of 3 mg/kg [78].

### **Antimicrobial:**

A methanol leaf extract (60%) of VA has been found to be active at 25 mg/ml against a panel of microorganisms (8 bacterial isolates and some fungi, but with absence of marked activity against *Candida*

*albicans* [27]. The secondary plant metabolites such as saponins, flavonoids and alkaloids found in the plant are thought to be responsible for this observed antimicrobial inhibitory activity effective in wound healing. Jisaka et al. showed that vernolepin and vernomenin possessed antibacterial effect against *B. subtilis* and *M. lutea*. 4, 15-dihydrovernolalin demonstrated the highest antibacterial activity against *B. subtilis* and *M. lutea* when compared to vernolepin, vernolide, vernodalin and vernomenin [17,32]. Vernodalol is also active against *B. cereus*, *S. epidemidus*, *S. aureus*, *M. kristinae* and *S. pyrogens* (gram positive bacteria), except for *S. pooni* (inhibited at 0.5 mg/ml), which was found to be inactive against gram negative bacteria [79].

#### **Antiplatelet and anticoagulant:**

An antiplatelet drug is a member of class of pharmaceuticals that decrease platelet aggregation [80]. Vernolepin is a sesquiterpene lactone compound isolated from VA alcoholic leaves extract and has demonstrated *in vitro* platelet anti-aggregating and disaggregating activity at an optimum concentration of  $1 \times 10^{-5}$  g/ml. it showed a 'stabilizing effect' towards rabbit platelets during freeze-thawing, it inhibited arachidonic acid, ADP and collagen-induced platelet aggregation as well as interferes with ATP-release. Electromicroscopy reveals platelet protection by vernolepin against adhesion and a disaggregating effect. These activities gave a steep dose response relationship and were time dependent [81]. VA extract caused a reduction in blood pressure, vernolepin isolated from this plant was identified to be responsible for its antiplatelet activity. Methanol extract of VA at doses of 100 mg/kg and 200 mg/kg induced a 40% and 50% inhibition against thrombosis in mice [39].

#### **Anti-malarial:**

The anti-plasmodial activities of VA have been investigated by several researchers. Isoamyl alcohol fractions of VA at concentrations less than 3 µg/ml showed *in vitro* antiplasmodial activity. The *in vitro* antiplasmodial activity was evaluated by dissolving 5 mg of VA in 5 ml ethanol to obtain a polar extract which was diluted in distilled water to give a series of test concentrations ranging from 0.5 to 500 µg/ml which were tested in triplicate against chloroquine sensitive *Plasmodium falciparum* infected human blood, Quinine 2HCl was used as an antimalarial reference product, with IC<sub>50</sub> value of 0.25 µg/ml. Krafi et al. demonstrated that ethanol extract of VA showed a high antiplasmodial activity (9.82 µg/ml) against 3D7 chloroquine sensitive clone of NF-54 isolate of *Plasmodium falciparum* using a 48 hours microassay technique [82]. The aqueous and hydroethanol extract however demonstrated significant antiplasmodial activity at IC<sub>50</sub> of 41.690 µg/ml and 44.03 µg/ml. It was reported that the significant *in vitro* antiplasmodial activities of vernodalin and vernodalol from *Vernonia cololata*; a related species to VA, at IC<sub>50</sub> values of 1.1-4.8 µg/ml were also determined. Sesquiterpene lactones such as vernolepin, vernolin, vernolide, vernodalin and hydroxyvernodalol isolated from VA leaves have been reported to exhibit antiplasmodial activities (IC<sub>50</sub><4 µg/ml) against *P. falciparum* strains [82,83].

#### **Hepatoprotective:**

Oral administration of methanol extract of leaves of VA brought about a modulatory effect on the hepatotoxicity of carbon tetrachloride (CCl<sub>4</sub>) [84]. It brought about a liver protective effect against CCl<sub>4</sub> - induced hepato-toxicity at the tested concentration.

Hepatotoxicity was induced in albino Wistar rats by the oral administration of CCl<sub>4</sub> at a dose of 1.2 g/kg body weight 3 times a week for 3 weeks, this treatment induced a significant hepatic injury as shown by increased activity of the serum enzymes AST, ALT, SALP and Y-GT. Methanol extract of VA administered 5 times a week for 2 weeks prior to CCl<sub>4</sub> treatment at 250 and 500 mg/kg doses of the extract reduced the increase in the activities of these enzymes. In the same way, the methanol extract of VA reduced the CCl<sub>4</sub>-induced increase in the concentrations of cholesterol, phospholipid and triglyceride by 37.8%, 8.5% and 30.6% respectively, as well as reduction in the cholesterol/ phospholipid ratio. Pretreatment with methanol VA extract at a dose of 500 mg/kg reduced CCl<sub>4</sub>-induced lipid peroxidation by 57.2%. In a similar way, it increased the activities of superoxide dismutase, glutathione S-transferase, but reduced glutathione concentration significantly at 500 mg/kg (p<0.05) and catalase activity at 500-1000 mg/kg doses, suggesting that VA leaves possesses protective effect against CCl<sub>4</sub> - induced hepatotoxicity by antioxidant mechanism of action, it was hypothesized that the increase in the activity of aspartate aminoferase alone may be of extra hepatic origin [85].

#### **Analgesic activity:**

Ethanol extract of VA has been found to induce a significant (p<0.05) dose-dependent analgesic activity in acetic acid induced writhing test in mice, the highest percentage inhibition (71.9%) was found at 100 mg extract/kg against the acetic acid-induced writhing in mice. The activity was higher than that of the standard analgesic; ketonal (67.5%) used at 10 mg/kg [86,87]. The study results tend to justified the ethnomedicinal uses of VA in the management of toothache, gingivitis, rheumatism since it was found to have analgesic activity. Agbaje et al. examines the antinociceptive potential of aqueous leaf extract (50-200 mg/kg), using the acetic acid - induce writhing, formalin test and tail flick test models [88]. It was observed that the extract significantly inhibited acetic acid-induced writhing and formalin test in mice but did not give any significant effect in the tail - flick test, suggestive of central and peripheral analgesic properties of the extract [89].

#### **Anti-inflammatory:**

The roots and leaves extracts, and the saponin fractions of VA possessed significant anti-inflammatory activity against xylene-induced acute inflammation in the ear of Wistar rats. The anti-inflammatory response was dose-dependent and the percentage inhibition was higher with the leaf than the root extract. The VA-saponin extract gave a relatively lower, but also significant inhibitory activity compared to the extracts. The anti-inflammatory activity of the extracts were comparable in magnitude to activity of dexamethasone and may partly be explained to be due to the presence of flavonoids, tannins, glycosides and trace elements; copper, manganese and zinc which were shown to be present in previous study [84]. Other studies have reported the anti-inflammatory activity of these substances [90-93].

#### **Anti-pyretic activity:**

Investigations proved that the anti-pyretic activities of the leaf, root and saponin fraction from VA. Pyrexia was induced using the procedure demonstrated Oboh et al. 20 hours after the administration of *Saccharomyces cerevitae* (Brewer's yeast) induces pyrexia, the anal temperatures of the animals (Wistar's rat), the anal temperature

reading of each animal was taken before dose administration [94]. The anal temperature reading of each animal was repeated 4 hours after dose administration; this procedure was used for the evaluation of the leaf and root aqueous extract, and the VA saponin fractions. Normal saline (5 ml/kg) was used as placebo and Acetyl Salicylic Acid 250 mg/kg as standard analgesic. The result showed that, just as observed with ASA, all doses of aqueous extracts of leaves and roots and VA-saponin fraction gave a significant anal temperature decrease (but to a lesser extent than ASA), except at 50 mg/kg of VA-saponin B fraction. Anti-pyretic activity was observed to be higher in the leaf than the root extract, but the saponin B fraction showed much lower activity than the root extract at similar dose of 200 mg/kg (108).

### Haemolytic properties:

Recent reports investigated the *in vitro* haemolytic properties of VA, results showed that infusion of VA induced a significant ( $p < 0.05$ ) haemolysis of human erythrocyte. Human genotype SS (1024) were highly susceptible to haemolysis induced by VA infusion, genotype-AS (512) were moderately susceptible, but genotype-AA (256) were highly resistant to haemolysis induced by the same infusion [95]. However, Alawa et al. reported a non-significant effect of methanol extract of VA in a 30-day treatment of rats on red blood cells (RBC) counts and other indices such as Hemoglobin concentration (Hb), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC) which are related to it, this is in relation to the control [96]. Also, there was no significant changes in the Total White Blood Cell count (TWBC), platelet, neutrophil, lymphocyte, eosinophil, and monocytes values relative to their respective controls.

### Antimutagenicity:

Obaseiki –Ebor et al. demonstrated that the Petroleum ether, methanol and ethylacetate extracts of VA were able to significantly (more than 60%) inhibit His-to His+ mutation induced by ethylmethanesulfonate on *Salmonella typhimurium* TA100 [97]. Petroleum ether extract was the most active, followed by methanol extract and then the ethylacetate fraction.

### Anti-leishmanial activity:

Methanol, hexane and aqueous extract of VA leaves have demonstrated both *in vitro* and *in vivo* experiments to suppress the infection rate of leishmaniasis [32], it delayed onset of the disease with significant reduction in lesion size and attenuation of the histopathological outcome characterized by intact epidermis and less tissue destruction in skin, spleen and liver after inoculation with metacyclic promastigotes from *Leishmania major* parasite in Balb/c mice. The methanol extract containing the highest concentration of flavonoids produced the highest activity, thus suggesting that, the flavonoids are responsible for the anti-leishmanial activity of VA (120) [32]. Hydroxylvernonolide, a compound isolated from the leaves of VA showed a significant antileishmanial effect on *Leishmania infantum* [32].

### Spermatogenic effect:

Recent report showed that the aqueous extract of the leaves of VA significantly ( $p < 0.05$ ) improved sperm concentration, motility, percentage normal morphology and percentage number of live sperm at doses of 50 mg/kg and 100 mg/kg in dose dependent manner in groups of male Wistar rats treated over a duration of 30 days when

compared to the control group of rats. This observation suggests that the administration of aqueous extracts of the leaves of VA successfully improved the sperm qualities [98] since there were significant improvements observed in all the sperm parameters of rats treated with the extract. The possible mechanism of VA induced improvement in sperm parameters could be as a result of the ability of VA to increase glucose metabolism, leading to the production of pyruvate which is known to be the preferred substrate essential for the activity and survival of the sperm cells [99,100]. It could also be as a result of or in addition to its antioxidant potential, the flavonoids and vitamins in VA leaf extract could maintain sperm morphology, sperm survival and sperm function, thus, could be regarded as a source of supply of additional nutrients to the treated group. However, there was a decrease in the level of Follicle Stimulating Hormone (FSH), but no significant increase in the level of Leutinizing Hormone (LH) and testosterone. In contrast, administration of VA at higher dose of 200 mg/kg for longer period of time provoked varying degrees of testicular degeneration, ranging from significant reduction in sperm motility, concentration, percentage normal morphology, percentage number of live sperm, to a significant increase in number of percentage of abnormal sperm. Though it was suggested that the possible mechanism by which higher doses of VA exert untoward effect against spermatogram and its relevant hormone may be as a result of its alkaloidal content which releases its metabolites that binds to cell molecules and cross-linked DNA thereby causing cytotoxicity, but the exact mechanism of action remains unknown [98,101].

### Effect on CD4+ cell count (HIV/AIDS):

Aqueous extract of leaves of VA showed a dose-dependent increase in CD4+ cells in a statistically significant manner ( $p < 0.005$ ) when compared to the control group. The exact mechanism by which this is done is currently not known, though, it could be related to its antioxidant contents of tannins, saponins and flavonoids, each of which has been individually shown to possess antioxidant property. Another possible mechanism of action could be as a result of enhanced early maturation and releases of leucocytes, since transient interaction between leucocytes have long been known to be critical for the normal function of the immune system [102]. It was therefore concluded that since doses of 200-800 mg/kg body weight of aqueous extract of leaves of VA for as long as 21 days will have a positive effect on the CD4+ cells of the Wistar rats used in the study when compared to standard group of Wistar rats, it could be advised that aqueous extract of the leaves of VA can be used as immune booster in immune compromised health conditions [103].

### Toxicity:

Lower doses of ethanol extract of leaves of VA (100 mg/kg per oral) showed no significant toxicity in the testis of male albino rats, higher doses (300 mg/kg and 600 mg/kg per oral), however, showed significant ( $p < 0.05$ ) testicular toxicity as demonstrated by reduction in tubular diameter, cross-sectional area, number of tubular profiles per unit area and the mean numerical density of seminiferous tubules [89]. The experimental model used showed that the toxicity limit was far higher than the limit for substances considered highly toxic (toxicity at less than 1mg/kg. Adiukwu et al. reported that VA caused no clinical signs of toxicity or adverse toxicological effects of VA at doses of 500-2000 mg/kg/day for 14 consecutive days [89].

## Conclusion

The review summarizes the pharmacotherapeutic effects of VA. The phytochemistry and pharmacology was also discussed in detail. VA is a potent ethnomedicinal plant that may be used in the management of cancer and other tropical diseases. The full potential of this plant has not been fully exploited. Hence, this review will stimulate further scientific research into the biological activities, with the view to discovering novel or lead pharmaceutical agents.

## Acknowledgement

This work was in part supported by a US-Senior Fulbright Award granted to Dr. A. Falodun for study at University of Mississippi, USA, CIESCs for the Fulbright award, TETFUND 2013 research grant and URPC VC 23.

## References

1. Vicki AF, Alfonso S, Tod FS, Harold R (2009) Classification of Compositae. The International Compositae Alliance.
2. Johri RK, Singh C (1997) Medicinal uses of *Vernonia* species. *Journal of Medicinal and Aromatic Plant Science*. 19: 744-752.
3. Igile GO, Oleszek W, Jurzysta M (1995) Vernoniosides D and E, two novel saponins from *Vernonia amygdalina*. *J. Nat. Prod.* 58: 1438-1443.
4. Farombi EO (2003) African indigenous plants with chemotherapeutic potentials and biotechnological approach to the production of bioactive prophylactic agents. *Afr. J. Biotech.* 2: 662-671.
5. Erasto P, Grierson DS, Afolayan AJ (2006) Bioactive sesquiterpene lactones from the leaves of *Vernonia amygdalina*. *J Ethnopharmacol* 106: 117-120.
6. Butter GW, Builey RW (1973) *Chemistry and Biochemistry of herbage*, Vol. 1 Accident Press London and New York.
7. Ologunde MO, Akinyemi AO, Adewusi SRA, Afolabi OA, Shepard RL et al. (1992) Chemical evaluation of exotic seed planted in the humid lowlands of West Africa. *Trop. Agric* 69: 106-110.
8. Afolabi OA, Oke OL (1981) Preliminary studies on the nutritive value of some cereal-like grains. *Nutr. Rep. Int.* 24: 389-394.
9. Igile GO, Oleszek W, Jurzysta M, Burda S, Fafunso M et al. (1994). Flavonoids from *Vernonia amygdalina* and their antioxidant activities. *J. Agric. Food Chem.* 42: 2445-2448.
10. Burkill HM (1985) *The Useful Plants of West Tropical Africa* (2nd edn) Royal Botanical Gardens, Kew, Vol. 1. Ainslie J.R., "List of Plants Used in Native Medicine in Nigeria," Imperial Forestry Institute, Oxford.
11. Hamowia AM, Safran AM (1994) Pharmacological Studies on *Vernonia amygdalina* (Del) and *Tithonia Diversifolia* (Gray). *J.Vet. Medicine.* 42: 91-97.
12. Huffman MA, Seifu M (1989) Observations on illness and consumption of a possibly medicinal plant *Vernonia amygdalina* (Del.), by a wild Chimpanzee in the Mahale Mountains National park, Tanzania. In: *Primates* 30 : 51-63.
13. Krief S, Hladik CM, Haxaire C (2005) Ethnomedicinal and bioactive properties of plants ingested by wild chimpanzees in Uganda. *J Ethnopharmacol* 101: 1-15.
14. Regassa A (2000) The use of herbal preparations for tick control in western Ethiopia. *J S Afr Vet Assoc* 71: 240-243.
15. Kambizi L, Afolayan AJ (2001) An ethnobotanical study of plants used for the treatment of sexually transmitted diseases (njovhera) in Guruve District, Zimbabwe. *J Ethnopharmacol* 77: 5-9.
16. Amira CA, Okubadejo NU (2007) Frequency of complementary and alternative medicine utilization in hypertensive patients attending an urban tertiary care centres in Nigeria. *BMC Compl. Alternative Med.* 7: 30-48.
17. Jisaka M, Ohigashi H, Takegawa K, Hirota M, Irie R et al. (1993) Steroid glucosides from *Vernonia amygdalina*, a possible chimpanzee medicinal plant. *Phytochemistry* 34: 409-413.
18. Oliver B (1960) Medicinal Plants in Nigeria. Nigerian college of Arts, Science and Technology; Zaria, Nigeria.
19. Kupcham SM (1971) Drugs from Natural products. Plant source in drugs discovery, science and development. *Am. Chem. Soc.* 6: 311-318.
20. Masaba SC (2000) The antimalarial activity of *Vernonia amygdalina* Del (Compositae). *Trans R Soc Trop Med Hyg* 94: 694-695.
21. Challand S, Willcox M (2009) A clinical trial of the traditional medicine *Vernonia amygdalina* in the treatment of uncomplicated malaria. *J Altern Complement Med* 15: 1231-1237.
22. Ainslie JR (2001) *List of Plants Used in Native Medicine in Nigeria*. Imperial Forestry Institute, Oxford.
23. Huffman MA, Koshimizu K, Ohigashi H (1996) Ethnobotany and zoopharmacognosy of *Vernonia amygdalina*, a medicinal plant used by humans and chimpanzees. *Biol. Utilization.* 2: 351-360.
24. Akah PA, Ekekwe RK (1995) Ethnopharmacology of some of the asteraceae family used in the Nigerian traditional medicine. *Fitoterapia.* 66:352-355.
25. Vlietinck AJ, Van Hoof L, Totté J, Lasure A, Vanden Berghe D et al. (1995) Screening of hundred Rwandese medicinal plants for antimicrobial and antiviral properties. *J Ethnopharmacol* 46: 31-47.
26. Ojukwu EM, Onuora GI, Iwu MM (1983) Effects of extracts of fresh leaves of *Vernonia amygdalina* (DEL) in pregnant local albino mice. *Bull. anim. health Product. Africa* 30.
27. Akinpelu DA (1999) Antimicrobial activity of *Vernonia amygdalina* leaves. *Fitoterapia* 70: 232-234.
28. Van Wyk, BE, Gericke N (2000) *People's Plants: a guide to useful plants of southern Africa*. Briza Publications, Pretoria.
29. Owoeye O, Yousuf S, Akhtar MN, Qamar K, Dar A et al. (2010) Another Anticancer Elemanolide from *Vernonia amygdalina* Del. *Int. J. Biol. Chem. Sci.* 4: 226-234.
30. Kupchan SM, Hemingway RJ, Karim A, Werner D (1969) Tumor inhibitors. XLVII. Vernodalin and vernomygdin, two new cytotoxic sesquiterpene lactones from *Vernonia amygdalina* Del. *J Org Chem* 34: 3908-3911.
31. Jisaka M, Ohigashi H, Takagaki T, Nozaki H, Tada T et al. (1992) Bitter steroids glucosides, vernoniosides A1, A2, and A3 and related B1 from a possible medicinal plant, *Vernonia amygdalina* used by wild chimpanzees. *Tetrahedron* 48: 625-632.
32. Koshimizu K, Ohigashi H, Huffman MA (1994) Use of *Vernonia amygdalina* by wild chimpanzee: possible roles of its bitter and related constituents. *Physiol Behav* 56: 1209-1216.
33. Lu F, Foo LY (2001) Antioxidant activities of polyphenols from sage (*Salvia officinalis*). *Food Chem.* 75: 197-202.
34. Ohigashi H, Jisaka M, Takagaki T, Nozaki H, Tada T et al. (1991) Bitter principle and a related steroid glucoside from *Vernonia amygdalina*, a possible medicinal plant for wild chimpanzees. *Agr. Biol. Chem.* 55: 1201-1203.
35. Kamperdick C, Breitmaier E, von Radloff MA (1992) A new steroid saponin from *Vernonia amygdalina* (Compositae). *J Prakt Chem.* 334: 425-428.
36. Aregheore EMK, Makkar HPS, Becker K (1998) Feed Value of Some Browse Plants from the Central Zone of Delta State. *Nig Trop Sci* 38: 97-104.
37. Schmittmann T, Rotscheidt K, Breitmaier E (1994) Drei neue steroidsaponine aus *Vernonia amygdalina* (Compositae). *J fur praktische Chemie* 336: 225.
38. Arene EO (1972) 7,24(28)-Stimastadien-3 $\beta$ -ol from *Vernonia amygdalina* *Phytochemistry* 11: 2886-2887.
39. Tona L, Cimanga RK, Mesia K, Musumba CT, Bruyne TDe, Apers S et al. (2004) In vitro antiplasmodial activity of extracts and fractions from seven medicinal plants used in the Democratic Republic of Congo. *J Ethnopharmacol.* 93: 27-32.

40. Obute GC, Godswill O, Adubor GO (2007) Chemicals Detected in Plants Used For Folk Medicine in South Eastern Nigeria. *Ethnobotanical Leaflets* 11: 173-194.
41. Izevbigie EB, Bryant JL, Walker A (2004) A novel natural inhibitor of extracellular signal-regulated kinases and human breast cancer cell growth. *Exp Biol Med* (Maywood) 229: 163-169.
42. Asawalam EF, Hassanali A (2006) Constituents of the essential oil of *Vernonia amygdalina* as maize weevil protectants. *Trop. Subtrop. Agroecosyst*, 6: 95-102.
43. Erasto P, Grierson DS, Afolayan AJ (2007) Antioxidant constituents in *Vernonia amygdalina* leaves. *Pharm Biol*. 45: 195-199.
44. Erasto P, Venter M, Roux S, Grierson DS, Afolayan AJ (2009) Effect of extracts of *Vernonia amygdalina* on glucose utilization of chang-liver, C2C12 muscle and 3T3-L1 cells. *Pharm. Biol.* 47: 175-181.
45. Akah PA, Okafor CI (1992) Blood Sugar lowering effect of *vernonia amygdalina* (del) in an experimental rabbit model. *Phytother. Res.* 6: 171-173.
46. Nwanjo HU (2005) Efficacy of aqueous leaf extract of *vernonia amygdalina* on plasma lipoprotein and oxidative status in diabetic rat models. *Niger J Physiol Sci* 20: 39-42.
47. Abraham AO (2007) Effects of *Vernonia amygdalina* and chlopropamide on blood glucose. *Med. J.Islamic World Acad. Sci.* 16: 115-119.
48. Ebong PE, Atangwho IJ, Eyong EU, Egbung GE (2008) The antidiabetic efficacy of combined extracts from two continental plants: *Vernonia amygdalina* (A. Juss) and *Vernonia amygdalina* (Del.) (African Bitter Leaf). *Am. J. Biochem. Biotech.* 4: 239-244.
49. Eteng MU, Bassej BJ, Atangwho IJ, Egbung GE, Eyong EU et al. (2008) Biochemical indices of macrovascular complication in diabetic rat model: compared effects of *Vernonia amygdalina*, *Charantus roseus* and chlopropamide. *Asian J Biochem.* 3: 228-234.
50. Uchenna VO, Chinwe EO, John MO, Ijeoma OE (2008) Hypoglycemic indices of *Vernonia amygdalina* on postprandial blood glucose concentration of healthy humans. *Afr J Biotechnol.* 7: 4581-4585.
51. Ong KW, Hsu A, Song L, Huang D, Tan BK (2011) Polyphenols-rich *Vernonia amygdalina* shows anti-diabetic effects in streptozotocin-induced diabetic rats. *J Ethnopharmacol* 133: 598-607.
52. Alawa CB, Adamu AM, Gefu JO, Ajanusi OJ, Abdu PA et al. (2003) In vitro screening of two Nigerian medicinal plants (*Vernonia amygdalina* and *Annona senegalensis*) for anthelmintic activity. *Vet Parasitol* 113: 73-81.
53. Alawa CBI, JO, Chiezey NP, Abdu PA, Magaji SO, Eduvie LO et al. (2000) Screening of *Vernonia amygdalina* for anthelmintic properties. In: Gefu, Abdu, P.A. and Alawa, C.B. (eds). *Ethno-veterinary practices research and development. Proceedings of the international workshop on ethno-veterinary practices held on 14 - 18 August, 2000 at Kaduna, Nigeria* 49-55.
54. Huffman MA (2001) Self-medicative behavior in the African great apes: An evolutionary perspective into the origins of human traditional medicine. *Boisei.* 51: 651-661.
55. Fraga CG (2007) Plant polyphenols: how to translate their in vitro antioxidant actions to in vivo conditions. *IUBMB Life* 59: 308-315.
56. Halliwell B, Rafter J, Jenner A (2005) Health promotion by flavonoids, tocopherols, tocotrienols, and other phenols: Direct or indirect effect? Antioxidant or not? *The Am. J. Clin. Nutria.* 81: 268S-276S.
57. Roginsky V (2003) Chain-breaking antioxidant activity of natural polyphenols as determined during the chain oxidation of methyl linoleate in Triton X-100 micelles. *Arch Biochem Biophys* 414: 261-270.
58. Fasakin F, Aluko CC (2011) E.R. Antioxidant properties of chlorophyll-enriched and chlorophyll-depleted polyphenolic fractions from leaves of *Vernonia amygdalina* and *Gongronema latifolium*. *Food Res. Intern.* 44: 2435-2441.
59. Oboh G, Raddatz H, Helen T (2008) Antioxidant properties of polar and non-polar extracts of some tropical green leafy vegetables. *J. Sci. Food Agric.* 88: 2486-2492.
60. Xie Z, Huang J, Xu X, Jin Z (2008) Antioxidant activity of peptides isolated from alfalfa leaf protein hydrolysate. *Food Chem.* 111: 370-376.
61. Pownall TL, Udenigwe CC, Aluko RE (2010) Amino acid composition and antioxidant properties of pea seed (*Pisum sativum* L.) enzymatic protein hydrolysate fractions. *J Agric Food Chem* 58: 4712-4718.
62. Wald NJ, Law MR (1995) Serum cholesterol and ischaemic heart disease. *Atherosclerosis* 118 Suppl: S1-S5.
63. Krieger M (1998) The "best" of cholesterol, the "worst" of cholesterol: a tale of two receptors. *Proc Natl Acad Sci U S A* 95: 4077-4080.
64. Schaefer EJ, Lichtenstein AH, Lamon-Fava S, McNamara JR, Ordovas JM (1995) Lipoproteins, nutrition, aging, and atherosclerosis. *Am J Clin Nutr* 61: 726S-740S.
65. Pritchard KA Jr, Groszek L, Smalley DM, Sessa WC, Wu M et al. (1995) Native low-density lipoprotein increases endothelial cell nitric oxide synthase generation of superoxide anion. *Circ Res* 77: 510-518.
66. Hu SH, Liang ZC, Chia YC, Lien JL, Chen KS et al. (2006) Antihyperlipidemic and antioxidant effects of extracts from *Pleurotus citrinopileatus*. *J Agric Food Chem* 54: 2103-2110.
67. Tomotake H, Yamamoto N, Yanaka N, Ohinata H, Yamazaki R et al. (2006) High protein buckwheat flour suppresses hypercholesterolemia in rats and gallstone formation in mice by hypercholesterolemic diet and body fat in rats because of its low protein digestibility. *Nutrition* 22: 166-173.
68. Visavadiya NP, Narasimhacharya AV (2007) Ameliorative effect of *Chlorophytum borivilianum* root on lipid metabolism in hyperlipaemic rats. *Clin Exp Pharmacol Physiol* 34: 244-249.
69. Miller CA (2001) Update on statins and other lipid-lowering drugs. *Geriatr Nurs* 22: 276-277.
70. Adaramoye O, Ogungbenro B, Anyaegbu O, Fafunso M (2008) Protective effects of extracts of *Vernonia amygdalina*, *Hibiscus sabdariffa* and vitamin C against radiation-induced liver damage in rats. *J Radiat Res* 49: 123-131.
71. Omoregie S, Pal A, Sisodia B (2011) In vitro Antimalarial and Cytotoxic Activities of Leaf Extracts of *V. amygdalina* (Del.) *Nig. J. Basic Applied Sci.* 19: 121-122.
72. Farombi EO, Owoeye O (2011) Antioxidative and chemopreventive properties of *Vernonia amygdalina* and *Garcinia biflavonoid*. *Int J Environ Res Public Health* 8: 2533-2555.
73. Adesanoye OA, Farombi EO (2010) Hepatoprotective effects of *Vernonia amygdalina* (astereaceae) in rats treated with carbon tetrachloride. *Exp Toxicol Pathol* 62: 197-206.
74. Oyugi DA, Luo X, Lee KS, Hill B, Izevbigie EB (2009) Activity markers of the anti-breast carcinoma cell growth fractions of *Vernonia amygdalina* extracts. *Exp Biol Med* (Maywood) 234: 410-417.
75. Gresham LJ, Ross J, Izevbigie EB (2008) *Vernonia amygdalina*: anticancer activity, authentication, and adulteration detection. *Int J Environ Res Public Health* 5: 342-348.
76. Tyler VC, Brady LR, Robbers EJ (1981) *Pharmacognosy*. (11th ed). London: Baillere Trindall. 295.
77. Udosen EO (1995) Proximate and mineral composition of some Nigerian vegetables. *Discov. Nino.* 7: 383-386.
78. Chung KT, Wong TY, Wei CI, Huang YW, Lin Y (1998) Tannins and human health: a review. *Crit Rev Food Sci Nutr* 38: 421-464.
79. Ganjian I, Kubo I, Fludzinski P (1983) Insect antifeedant elemanolide lactones from *Vernonia amygdalina*. *Phytochemistry* 22: 2525-2526.
80. Yedjou CG, Izevbigie EB2, Tchounwou PB1 (2013) *Vernonia amygdalina*-Induced Growth Arrest and Apoptosis of Breast Cancer (MCF-7) Cells. *Pharmacol Pharm* 4.
81. Igboechi AC, Anuforo DC (1984) Anticoagulant activities of extracts of *Eupatorium odoratum* and *Vernonia amygdalina*. *Nig J Pharm* 20: 313-320.
82. Kraft C, Jenett-Siems K, Siems K, Jakupovic J, Mavi S et al. (2003) In vitro antiplasmodial evaluation of medicinal plants from Zimbabwe. *Phytother Res* 17: 123-128.

83. Phillipson JD, Wright CW, Kirby GC, Warhurst DC (1993) Phytochemistry of some plants used in traditional medicine for the treatment of protozoal diseases. Symposium of the Phytochemical Society of Europe; University of Lausanne: Lausanne, Switzerland.
84. Ibrahim ND, Abdurahman EM, Ibrahim G (2001) Elemental analysis of the leaves of *Vernonia amygdalina* and its biological evaluation in rats. *Niger. J. Natl. Prod. Med.* 5: 13-16.
85. Adesanoye OA, Farombi EO (2009) Hepatoprotective effects of *Vernonia amygdalina* (Asteraceae) in rats treated with carbon tetrachloride. *Experimental Toxicol. Pathol.* 16: 891-895.
86. Ahmadiani A, Hosseiny J, Semnani S, Javan M, Saeedi F et al. (2000) Antinociceptive and anti-inflammatory effects of *Elaeagnus angustifolia* fruit extract. *J Ethnopharmacol* 72: 287-292.
87. Njan AA, Adzu B, Agaba AG, Byarugaba D, Díaz-Llera S et al. (2008) The analgesic and antiplasmodial activities and toxicology of *Vernonia amygdalina*. *J Med Food* 11: 574-581.
88. Agbaje EO, Adeneye AA, Delete TI (2008) Anti-nociceptive and anti-inflammatory effects of a Nigerian Polyherbal Tonic (PHT) extract in rodents. *Afr J Compl Alt Med* 5: 399-408.
89. Adiukwu PC, Amon A, Nambatya G, Adzu B, Imanirampa L et al. (2012) Acute toxicity, antipyretic and antinociceptive study of the crude saponin from an edible vegetable: *Vernonia amygdalina* leaf. *Int J Biol Chem Sci* 6: 1019-1028.
90. Bittar M, de Souza MM, Yunes RA, Lento R, Delle Monache F et al. (2000) Antinociceptive activity of I3,II8-binarigenin, a biflavonoid present in plants of the guttiferaceae. *Planta Med* 66: 84-86.
91. DiSilvestro RA, Marten JT (1990) Effects of inflammation and copper intake on rat liver and erythrocyte Cu-Zn superoxide dismutase activity levels. *J Nutr* 120: 1223-1227.
92. Kim HP, Son KH, Chang HW, Kang SS (2004) Anti-inflammatory plant flavonoids and cellular action mechanisms. *J Pharmacol Sci* 96: 229-245.
93. Okokon JE, Onah MI (2004) Pharmacological studies on root extract of *Vernonia amygdalina* Nig *J Prod Med* 8: 59-61.
94. Obboh G (2006) Nutritive value and haemolytic properties (in vitro) of the leaves of *Vernonia amygdalina* on human erythrocyte. *Nutr Health* 18: 151-160.
95. Oyedeji KO, Bolarinwa AF, Akintola AM (2013) Effect of Methanol Extract of *Vernonia Amygdalina* on Haematological and Plasma Biochemical Parameters in Male Albino Rats. *Journal of Dental and Medical Sciences* 3, 64-67.
96. Alawa JN, Carter KC, Nok AJ, Kwanashie HO, Adebisi SS et al. (2012) Infectivity of macrophages and the histopathology of cutaneous lesions, liver and spleen is attenuated by leaf extract of *Vernonia amygdalina* in *Leishmania major* infected BALB/c mice. *J Complement Integr Med* 9: Article 10.
97. Obaseiki-Ebor EE, Odukoya K, Telikepalli H, Mitscher LA, Shankel DM (1993) Antimutagenic activity of extracts of leaves of four common edible vegetable plants in Nigeria (west Africa). *Mutat Res* 302: 109-117.
98. Saalu LC, Akunna GG, Oyewopo AO (2013) The histomorphometric evidences of *Vernonia amygdalina* leaf extract-induced testicular toxicity. *Int. J. Morphol.* 31: 662-667.
99. Egbunike GN, Branscheid W, Pfisterer J, Holtz W (1986) Changes in porcine sperm lactate dehydrogenase isoenzymes during sperm maturation. *Andrologia* 18: 108-113.
100. Di Carlo GI, Mascolo N, Izzo AA, Capasso F (1999) Flavonoids: old and new aspects of a class of natural therapeutic drugs. *Life Sci* 65: 337-353.
101. Longe OG, Farinu GO, Fetuga BL (1983) Nutritional value of the fluted pumpkin (*Telfaria occidentalis*). *J Agric Food Chem* 31: 989-992.
102. Momoh MA, Adiukwu MU, Oyi AR (2010) *Vernonia amygdalina* extract and CD4+ Cell Counts: An Immune Study. *Global J Biotech Biochem* 5: 92-96.
103. Ibrahim G, Abdurahman EM, Ibrahim H, Ibrahim NG, Magaji MG (2011) Toxicity and Analgesic Effects of *Vernonia Amygdalina* Del. (Asteraceae) Leaf Extract on Mice. *Int J Adv Pharm Biol Sci.* 1:1-4