Sub-acute Toxicity of Aqueous Extracts of Tephrosia vogelii, Vernonia amygdalina and Senna occidentalis in Rats

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Abstract

There is an intense search for new plant-derived medicines to match the ever increasing prevalence of drug resistance. Toxicity profiles of three commonly used plants against helminthiosis in Uganda will support their optimum use.

Objective: To evaluate the sub-acute toxicity of aqueous leaf extracts of Tephrosia vogelii (TV), Vernonia amygdalina (VA) and Senna occidentalis (SO).

Methods: One hundred Wistar white albino rats in 10 groups were dosed orally daily with 200, 400 and 600 mg/kg of TV, VA and SO for 28 days or Goodwin’s physiological solution. Daily changes in behaviour, feed/water intake and weight (weekly) were monitored. On day 29, blood for haematology, serum for biochemical analysis and organs (lungs, liver, kidney, heart and small intestines) for histopathology were collected.

Results: Continued exposure of rats to 600 mg/kg doses of SO and VA extracts led to elevation of ALT, AST and ALP and cholesterol in the treatment groups. SO and VA at 600 mg/kg elevated urea and uric acid; depressed bilirubin while VA and TV 600 mg/kg caused the reverse. Histopathology findings (villi sloughing, pneumonitis, liver and kidney degeneration) further supported these results.

Conclusion: The very low toxicity of the aqueous leaf extracts of TV, VA and SO at high doses makes them safe at currently non-standardised doses used for animal treatment.

Keywords: Tephrosia vogelii; Vernonia amygdalina; Senna occidentalis; Toxicity; Rats

Background

Uganda has 80% of its population depending on agriculture and about 50% are engaged in livestock farming [1]. In small-scale livestock production, majority of the farmers use a combination of conventional and ethnopharmacological formulations to treat diseases and parasitic infections [2,3]. Use of plants in animal disease treatment is practiced traditionally [2-4,6], albeit with non-standardised doses as a major shortfall [4,7,8]. This is further compounded by the basic belief that plant extracts are non-toxic [2,7,9]. In an earlier study mapping ethnopharmacological practices in Mpiigi and Guru districts in Uganda, farmers prioritized Vernonia amygdalina (VA), Tephrosia vogelii (TV) and Senna occidentalis (SO) for detailed evaluation [3].

Tephrosia vogelii Hook (Family Leguminosae; synonym Cracca vogelii, Hook F; ILDIS record 5085), is a soft woody branching herb, 1-4 m tall [10]. The plant is widely promoted in eastern and southern Africa for its insecticidal properties [11], although some people use it as a fish poison [12]. Vernonia amygdalina Delile (Family Asteraceae; synonym Calacila amygdalina (http://www.theplantlist.org/tpl/record/gcc-84295) Kuntze, TICA Id 2949) is a shrub that grows predominantly in tropical Africa [2,10]. The leaf extracts of VA are used to treat malaria, gastrointestinal ailments [13-16] and reducing blood sugar levels in dogs [17]. Senna occidentalis Linn (Family Caesalpinicaceae; synonym Cassia occidentalis L.; ILDIS id 1087), is a 0.8-1.5 m tall, smooth leaved perennial plant important in traditional medicine in the tropics [10,18]. It has good antimicrobial, anti-mutagenic, anti-inflammatory, anti-diabetic and anti-malarial activity [18-20]. Acute toxicity studies of leaves of TV [21], VA [15,22,23] and seeds of SO [24-28] have been studied in different animals. The objective of this study was to determine the sub-acute toxicity due to prolonged administration of these three commonly used plants in rats.

Materials and Methods

Plant collection and extract preparation

Fresh leaves of all plants were collected in January 2013 from wild shrubs at Mbarara Zonal Agricultural Research Development Institute farm, located 274 km south-west of Kampala, the capital of Uganda. The voucher specimens were confirmed by a botanist and deposited in Makerere University Herbarium (numbers SO - NI 02, VA - NI 03 and TV - NI 04). The leaves were dried under shade in the laboratory for two weeks. They were ground to fine powder with an automated electric grinder (Grindomix, Retsch, GM 200). For aqueous extraction, 500 g of powder from each plant was infused in 2 l of hot distilled water for two weeks. They were ground to fine powder with an automated electric grinder (Grindomix, Retsch, GM 200). For aqueous extraction, 500 g of powder from each plant was infused in 2 l of hot distilled water for 24 hours with regular stirring (metallic magnet). After filtration with a 4 mm Whatmann filter paper, the filtrate was freeze dried at -82°C. The dried extracts were weighed and stored at 4°C until needed.

Animal dosing and monitoring

After determining the lethal doses and safety margins of the aqueous extracts, the safest dose was used to calculate doses for the sub-
acute study. In the protocol, OECD test guidelines were followed [29]. National Academies guide for the care and use of laboratory animals [30] were adapted for the animal protocol in this study and was approved by the institutional review board (study number IRBCOVAB/2013/009). Briefly, 100 Wistar white albino rats (50 females and 50 males), of about 6 to 8 weeks old weighing 70-100 g were procured from Uganda Virus Research Institute and kept for two weeks to acclimatize to the new laboratory conditions. Each extract was reconstituted in Goodwin’s physiological solution to a stock concentration of 200 mg/ml. The rats were weighed and randomly assigned to 10 treatment groups i.e. of each extract, 200 mg/kg, 400 mg/kg and 600 mg/kg body weight plus the negative control (given Goodwin’s physiological solution w/v). In each group, the number of females and males was equal. Each group was kept in 80x50 cm wooden cages, fed on commercial rodent feed (Unga Limited, Kenya) and given water ad libitum. The rats were treated once daily, while body weights were taken once a week and on the 29th day, before termination of the experiment.

Sample collection

On the 29th day, the rats were weighed, anaesthetised with 70% chloroform (trichloromethane, E08A/0808/2104/13, Fine-Chem, India) for ten minutes before blood and tissue collection. Whole blood collected by cardiac puncture was transferred into labelled plain and EDTA vacutainer tubes (BD Plymouth, UK). EDTA blood was kept at 4°C while non-EDTA blood was centrifuged at 1500 g for three minutes to separate serum, which was kept at -20°C. The rats were then dissected to collect the liver, kidneys, small intestine sections, heart and lungs; which were weighed and kept in 10% buffered formalin for two weeks. The rats were processed in the Central Diagnostic Laboratory of the College of Veterinary Medicine at Makerere University. Sections of the liver, kidney, lung, small intestines and heart were prepared as described previously [31,32]. Sections were stained with Haemotoxylin and Eosin (H&E), and then microscopically examined for pathological changes at x10, 20 and 40 magnifications.

Histopathology

Whole rat organs kept in 10% buffered formalin for two weeks were processed in the Central Diagnostic Laboratory of the College of Veterinary Medicine at Makerere University. Sections of the liver, kidney, lung, small intestines and heart were prepared as described previously [31,32]. Sections were stained with Haemotoxylin and Eosin (H&E), and then microscopically examined for pathological changes at x10, 20 and 40 magnifications.

Data analysis

Laboratory data was entered in Microsoft Excel and analysed in Minitab 16.0 (Minitab Inc., USA). Means, standard deviation, standard error of the mean, median and 95% confidence intervals of continuous variables were calculated and plotted by a pre-designed Excel sheet [33]. One-way analysis of variance (ANOVA) and Tukey’s high square difference test were used to compare mean differences for different parameters in the treatment and control groups. Proportions and percentages were computed and compared across groups using the Mann-Whitney, Newman-Keuls and Chi-square tests. The level of significance was set at P<0.05.

Results

Physical signs and gross necropsy findings

In all treatment groups, we observed no changes in feed and water intake. There were three deaths (one each at 600 mg/kg of TV, SO and VA) after 15, 24 and 26 days after treatment respectively. The immediate cause was wounds due to cannibalism, but after housing the rats individually, no more deaths were registered. No pathological gross necropsy findings were detected in treated and control rats during organ collection and processing.

Effect on body, heart, liver and kidney weights

There was a gradual increase in body weight in rats across all treatment groups but mean weight of SO 600 mg/kg was lower than other groups at day 0 and 14. However, the difference was non-existent by day 21 and at the end of experiment (day 29). There was less variation of mean post-mortem weight of the kidney and liver in the treatment groups. Although the liver weights ranged from 4.6-9.7 g, there was no statistical difference in mean weights due to treatment.

<table>
<thead>
<tr>
<th>Treat group &amp; dose (mg/kg)</th>
<th>WBC (10^3/µl)</th>
<th>RBC (10^6/µl)</th>
<th>HCT (%)</th>
<th>MCV (fL)</th>
<th>MCH (pg)</th>
<th>MCHC (g/dL)</th>
<th>RDWCV (%)</th>
<th>PLT (10^3/µl)</th>
<th>MPV (fL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>19.6 ± 10.5</td>
<td>7.4 ± 0.3</td>
<td>13.5 ± 0.4</td>
<td>44.9 ± 2.3</td>
<td>60.7 ± 2.3</td>
<td>18.4 ± 0.4</td>
<td>30.4 ± 0.8</td>
<td>19.5 ± 0.6</td>
<td>733.8 ± 34.8</td>
</tr>
<tr>
<td>TV200</td>
<td>9.9 ± 2.4</td>
<td>7.0 ± 0.5</td>
<td>13.2 ± 0.7</td>
<td>42.2 ± 2.3</td>
<td>60.7 ± 1.1</td>
<td>18.3 ± 0.4</td>
<td>30.2 ± 0.7</td>
<td>23.1 ± 1.1</td>
<td>362.5 ± 95.3</td>
</tr>
<tr>
<td>TV400</td>
<td>14.6 ± 4.3</td>
<td>5.9 ± 1.1</td>
<td>11.4 ± 1.4</td>
<td>39.3 ± 6.1</td>
<td>67.1 ± 2.4</td>
<td>19.7 ± 1.2</td>
<td>29.3 ± 0.9</td>
<td>22.7 ± 2.4</td>
<td>397.7 ± 78.9</td>
</tr>
<tr>
<td>VA200</td>
<td>10.5 ± 0.7</td>
<td>7.3 ± 0.5</td>
<td>13.0 ± 0.9</td>
<td>46.0 ± 3.6</td>
<td>63.1 ± 1.1</td>
<td>17.8 ± 0.2</td>
<td>28.3 ± 0.7</td>
<td>22.9 ± 0.7</td>
<td>489.0 ± 87.6</td>
</tr>
<tr>
<td>VA400</td>
<td>11.4 ± 0.5</td>
<td>7.7 ± 0.9</td>
<td>14.3 ± 0.8</td>
<td>49.6 ± 5.6</td>
<td>64.4 ± 1.4</td>
<td>18.5 ± 0.3</td>
<td>28.8 ± 0.3</td>
<td>21.1 ± 1.0</td>
<td>943.0 ± 56.3</td>
</tr>
<tr>
<td>VA600</td>
<td>13.1 ± 2.9</td>
<td>8.1 ± 0.3</td>
<td>14.8 ± 0.1</td>
<td>50.9 ± 1.2</td>
<td>63.2 ± 1.1</td>
<td>18.0 ± 0.6</td>
<td>28.5 ± 0.4</td>
<td>21.3 ± 0.8</td>
<td>714.0 ± 109.0</td>
</tr>
</tbody>
</table>

* Mean Platelet values significantly different from control and other treatment groups; P=0.005

Table 1: Haematological parameters (Mean ± SEM) of 100 Wistar white albino rats in nine treatment groups and a placebo control group.
Effect on biochemical and haematological parameters

The extracts caused varied effects on haematology and biochemistry (Table 1). For example, there was depression of haematocrit in a dose-response pattern, whereas the WBC counts were much higher in the control (19.6 ± 10.5x10^3/µl) compared to other groups (<15 x10^3/µl). A reverse relationship was observed with RDCHV; showing elevated levels with increasing doses for all plant extracts but there was minimal change in MCHC. At increasing doses, TV had a significant reducing effect on the mean platelet counts compared to other treatment groups and the controls (F_{9,89}=3.4; P=0.005).

Levels of liver function enzymes; ALP, ALT, AST and cholesterol significantly differed across the treatment groups (F_{9,89}=2.88, P = 0.035; 2.55, 0.022; 2.32, 0.035 and 3.35, 0.004 respectively). At 600 mg/kg, TV elevated the AST enzyme to 355.6 U/L while SO 600 mg/kg depressed it to 182.0 U/L, compared to 227.1 U/L in the control group (Figure 1). Cholesterol levels also differed in SO 600 mg/kg (3.55), VA 600 mg/kg (3.38) and TV 200 mg/kg (2.13), TV 600 mg/kg (2.36) compared to 2.93 in the control group (Figure 1A and 1B). Furthermore, albumin protein was elevated in SO and VA in dose-response manner compared to TV, which led to a small change (Table 2). The elevation was significant at 600 mg/kg of SO (44.9 g/L) and VA (47.8 g/L) compared to TV (38.2 g/L) and control group (34.8 g/L).

Histopathology and microscopic effects of the treatments

As shown in Table 3 and Figure 2A-2F, there were varied effects of treatments on the histopathology of the liver, kidney, lung, heart and small intestines. TV caused more toxic changes in the organs

![Image]

Figure 1: A: Comparison of mean (U/L) and 95% CI of liver function enzymes - Alanine amino-transferase (ALT) and Aspartate amino-transferase (AST) in 9 treatment groups with non-treated (control) rats. B: Mean and 95% CI of albumin (g/L) protein levels in blood of 100 rats in 9 treatment groups with non-treated (control) rats. *Control rats given 0.5 ml of Goodwin’s physiological solution daily

\*Mean significantly different at P<0.05
\*Mean different between VA 600 and all groups with letter b

Table 2: Mean and 95% CI of albumin (g/L) protein levels in blood of 100 rats in nine treatment groups compared to placebo control group.

<table>
<thead>
<tr>
<th>Treatment Group (mg/kg)</th>
<th>Mean (g/L)</th>
<th>Std. Error</th>
<th>95% Confidence Interval for Mean</th>
</tr>
</thead>
<tbody>
<tr>
<td>SO 200</td>
<td>35.5a,b,c</td>
<td>2.6</td>
<td>28.8 - 42.2</td>
</tr>
<tr>
<td>SO 400</td>
<td>39.3</td>
<td>3.7</td>
<td>23.6 - 55.0</td>
</tr>
<tr>
<td>SO 600</td>
<td>44.9a,b,c</td>
<td>0.9</td>
<td>41.9 - 47.8</td>
</tr>
<tr>
<td>Control</td>
<td>34.3a,b,c</td>
<td>6.0</td>
<td>18.8 - 49.7</td>
</tr>
<tr>
<td>TV 200</td>
<td>35.5</td>
<td>1.1</td>
<td>32.8 - 38.3</td>
</tr>
<tr>
<td>TV 400</td>
<td>35.4</td>
<td>2.4</td>
<td>29.3 - 41.6</td>
</tr>
<tr>
<td>TV 600</td>
<td>38.2</td>
<td>2.5</td>
<td>31.1 - 45.2</td>
</tr>
<tr>
<td>VA 200</td>
<td>38.3b,c</td>
<td>5.7</td>
<td>13.7 - 62.9</td>
</tr>
<tr>
<td>VA 400</td>
<td>41.7b,c</td>
<td>2.5</td>
<td>34.8 - 48.6</td>
</tr>
<tr>
<td>VA 600</td>
<td>47.8b,c</td>
<td>2.4</td>
<td>37.6 - 58.1</td>
</tr>
</tbody>
</table>

Discussion

The geographical location, season, soil type, climate differences, agronomic practices, plant part and age affect the concentration of phytochemicals, usually leading to differences in bioactivity and toxicity [7,16,18]. Using proteomics, metabolomics and transcriptomics in recent advances to determine nephrotoxic, genotoxic and teratogenic effects of natural plant products has been recommended [34]. These in silico, in vitro and in vivo methods which may explain the mechanisms of action, are however still unstandardised [34]. Following the established protocol [29,33], the recommended number of animals used supported statistical analysis.

The difference in concentration of phytochemicals extracted by various solvent systems may account for differences in observed toxicity of a plant extract at the same doses. For example, saponins are incriminated to cause haemolysis [35]. Although VA has significant levels of saponins [36,37], there was a dose-dependent increase in RBC counts. On the contrary, a reduction in RCB counts with increasing doses of SO and TV and PCV was noted. This is a possible mechanism of toxicity for the latter two extracts. Although no specific study on effect of extract on platelet count was done, we observed a marked reduction in platelets in higher dose groups of TV (363 and 398 compared to 734x10^3/µl in control). When platelets are significantly reduced, there may be multiple organ failure, eventually leading to toxicity signs or death.

Specifically, rotenone present in TV [11,38], causes similar signs of poisoning in humans and is one of the possible chemicals responsible for the observed acute toxicity [21]. In insects and fish, rotenone interferes with mitochondrial electron transport system, whereas mammals are more tolerant [38]. Rotenone however, is less soluble in water and easily disintegrates in the environment [12] which explains the presently observed reduced toxicity by aqueous extracts. Anthracenosides and anthocyanosides have been incriminated as toxic phytochemicals previously. However, qualitative phytochemistry (results not presented here) showed both anthracenosides and anthocyanosides in the aqueous extract of TV and VA.
The seeds of SO are reportedly toxic to many vertebrates and the unnamed toxic albumin is incriminated [18]. This study however, considered only the leaf extracts and thus some of these toxic principles may have been in limited amounts (no specific tests done presently). Some of the phytochemicals in SO are reported to be hepatoprotective [18,39]. In chicken, studies demonstrated lower phosphorylation ratios and respiratory control ratios and oxygen use in liver mitochondria [18]. Pneumonitis, mononuclear cell infiltration and lymphoid necrosis as observed in some cases in this study lead to compromised gaseous exchange and eventual death. Underlying toxicity mechanisms at cellular level should be undertaken with standardised proteomics, metabolomics and transcriptomics.

## Conclusion

SO and VA were hepatoprotective while TV was slightly hepatotoxic in a dose response manner. The very low toxicity of the aqueous leaf extracts of TV, VA and SO at high doses provide for its safety within the irregular and unstandardized doses used for human and animal treatment.

## Acknowledgement

We are grateful for the tireless input into the laboratory work by Mr. Nduki James especially during daily dosing of the rats. The invaluable laboratory support by Mr. Kisekka Magid and Dr. Afayo Mathias is appreciated. Special thanks go to Swedish International Development Agency (Sida) for the financial support.

## Author contributions

Denis Mugizi, John Katerega, Deogracious Oilla and Immaculate Nabukenya contributed to the study design, data collection and manuscript writing. Chris Rubaire-Akiiki, Deogracious Oilla and Johan Höglund supervised the laboratory work and contributed to manuscript writing.

## References


10. The plant list.


16. The plant list.


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