Synthesis and Biological Activities of Novel 1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidines

Amine Karoui[1], Monia Deghrigue[2], Gilbert Kirsch[3], Abderrahman Bouraoui[2], Fakher Chabchoub[1] and Fatma Allouche[1*]

[1]Laboratoire de Chimie Appliquée, Hétérocycles, Corps Gras et Polymères, Faculté des Sciences de Sfax, Université de Sfax, 3018 Sfax, Tunisie
[2]Laboratoire de développement chimique, galénique et pharmacologique des médicaments (LR12ES09). Equipe de Pharmacologie marine, Faculté de pharmacie de Monastir, Université de Monastir, 5000 Monastir, Tunisie
[3]Laboratoire d'Ingénierie Moléculaire et Biochimie Pharmacologique (EA3940), Institut Jean Barriol, Université de Lorraine, Metz, France

Abstract

A straightforward method has been developed for the synthesis of new anti-inflammatory 1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5 from aminocyanopyrazole. These compounds were screened for their anti-inflammatory, gastroprotective, analgesic, antioxidant and anticandidal activities. Studies of structure-activity relationships have led to selection of compound 6-(4-methoxyphenyl)-3-methyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine, 5a which exhibited the most potent activities. The structures of all new compounds were elucidated using IR, ^{1}H NMR, ^{13}C NMR and HRMS.

Keywords: Anti-inflammatory; Gastroprotective; Antioxidant; Anticandidal; Analgesic; Pyrazolo[3,4-d]pyrimidine, 1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidines

Introduction

The growing incidence of drug-resistant infection diseases has stimulated the need for the development of new drugs. Since the few last decades, synthetic chemistry has been recognized to be rich source of bioactive metabolites with varied biological and pharmacological activities. Currently, Non-steroidal anti-inflammatory drugs (NSAIDs) are used throughout the world for the treatment of inflammation, pain and fever; however most of these produce several adverse reactions such as ulcers and hemorrhage [1]. In addition, reactive oxygen species (ROS) and free radicals play important roles in degenerative or pathological processes leading to many health disorders including inflammatory and cancer diseases [2]. The harmful effect of the free radicals can however, be blocked by synthetic antioxidants such as butylated hydroxytoluene (BHT), butylated hydroxyanisole (BHA), tert-butylhydroquinone (TBHQ) and propyl gallate (PG) [3]. However, due to their adverse side effects, search for more effective antioxidants has become crucial. Pyrazolopyrimidines, class of sedative and anxiolytic drugs [4], and its derivatives constitute a rich source of a wide variety of structurally unusual metabolites and seem to be an endless source of new chemical constituents.

In order to determine the role of methyl group and methoxy group of pyrazolopyrimidines in the antioxidant, anti-inflammatory, gastroprotective, analgesic and anticandidal activities, we have synthesized new 1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f] pyrimidines 5 and evaluate their pharmacological activities. Interestingly, we found that the activity of new compounds is dependent on the location of the methyl and methoxy group. Our results provided new evidence for the relationship between chemical structure and pharmacological activities as the case of 5a and 5b.

Materials and Methods

Chemistry

Phenyl hydrazine, malononitrile, triethylorthoester, ammoniac, α-Bromoacetophenone, PTSA, CH3COOH and solvents used in this work were obtained from Aldrich and Fluka and were used without further purification.

Melting points were measured on an Electrothermal apparatus.

Progress of their actions was monitored with TLC using aluminium sheets with silica gel 60 F254 from Merck. Spectra IR were recorded on a Perkin-Elmer PARAGON FT-IR spectrometer covering field 400-4000 cm^{-1}. The spectra of ^{1}H NMR and ^{13}C NMR was recorded in solution in CDCl$_3$ or in dimethylsulfoxide (DMSO-d$_6$) on a Bruker spectrometer (^{1}H at 400 MHz, ^{13}C at 100 MHz) and High resolution mass spectrometry (HRMS). The chemical shifts are expressed in parts per million (ppm) by using tetramethylsilane (TMS) as internal reference. The multiplicities of the signals were indicated by the following abbreviations: s, singlet; d, doublet; t, triplet; q, quadruplet; m, multiplet, and coupling constants are expressed in Hertz.

Synthesis and spectral data of compounds 2-5: The general synthetic procedure employed for 1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidines 5 are outlined in Scheme 1.

5-amino-4-cyano-N^1-phenyl pyrazoles (2): 5-Amino-4-cyano-1-N^1-phenyl pyrazoles prepared via a standard addition of hydrazine derivatives to ketene ethoxymethylene compounds following the reported procedure. Recrystallization from ethanol afforded pure 2 in good yields [5,6].

4-cyano-N^1-phenyl pyrazolo-5-imidates (3): The required 5-amino-4-cyano-N^1-phenyl pyrazole (1.0 mmol) was treated with triethylorthoester (6.0 mmol) and a catalytic amount of acetic acid and the mixture was refluxed for 24 h. After cooling, the reaction mixture was evaporated. The product was filtered, washed with diethyl ether then purified by recrystallization (ethanol).
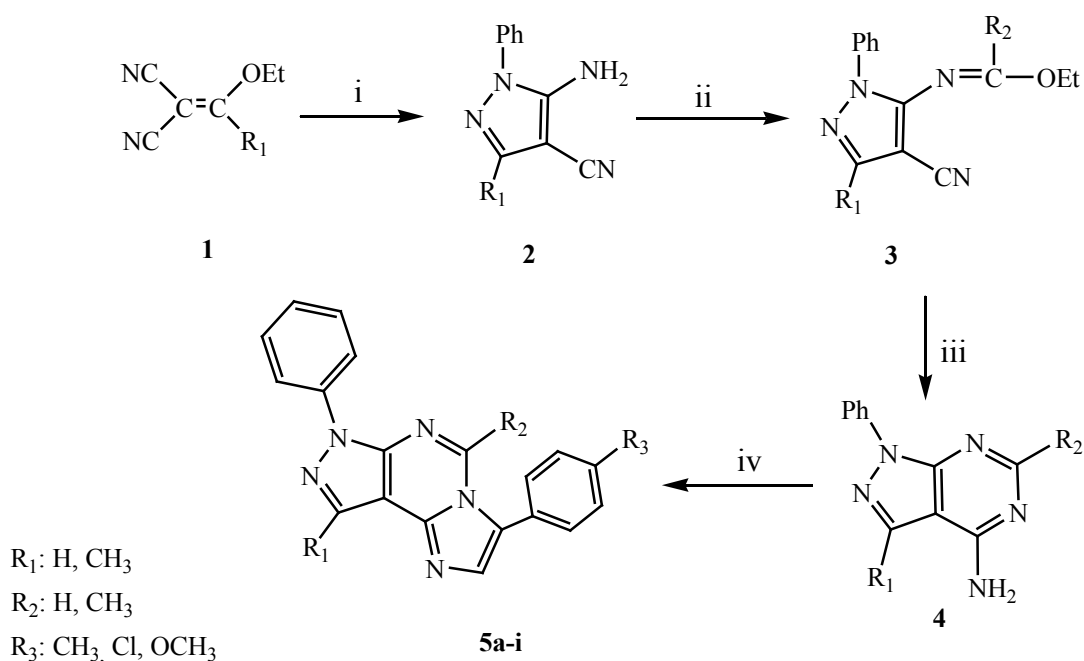
4-amino-N^1-phenyl pyrazolo[3,4-d] pyrimidine (4): A solution of imidate 3 (1.0 mmol) in dry ethanol (5 mL) was treated with ammoniac (2.0 mmol) and a catalytic amount of acetic acid. The reaction mixture

Scheme 1: Synthetic procedure of 1,7-dihydropyrazolo[3,4-*d*]imidazo[1,2-*f*]pyrimidines **5a-i**.

was refluxed for 6 h, and the formed solid was collected by filtration, dried and recrystallized from ethanol to give compound 4.

α) **4-amino-N^1-phenyl-1H-pyrazolo[3,4-d]pyrimidine 4a:** Yield 83 %; mp 228 °C; IR (cm^{-1}): ν_{NH2} 3100, 3283; $\nu_{C=N}$ 1480, 1500, 1590 cm^{-1}; RMN ^{1}H (δ ppm, DMSO-d$_6$): 4.69 (2H, s, NH$_2$), 7.36 (1H, t, J=7.3Hz, ArH$_4$), 7.48 (2H, t, J=7.3Hz, ArH$_3$ and ArH$_5$), 7.52 (2H, d, J=7.3Hz, ArH$_2$ and ArH$_6$), 7.60 (1H, s, H$_3$), 7.72 (1H, s, H$_6$), ^{13}C RMN (δ ppm, DMSO-d$_6$): 114.14 (C-3a), 124.27 (C-2' and C-6'), 129.00 (C-4'), 129.58 (C-3' and C-5'), 130.04 (C-3), 136.94 (C-1'), 141.36 (C-7a), 149.83 (C-6), 156.84 (C-4); HRMS Calculated for C$_{11}$H$_9$N$_5$: 211.0858, found: 211.0859.

β) **4-amino-3-methyl-N^1-phenyl-1H-pyrazolo[3,4-d] pyrimidine 4b:** Yield 68 %; mp 192 °C; IR (cm^{-1}) ν_{NH2}: 3083, 3317, $\nu_{C=N}$ 1626, 1647, 1665; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.76 (3H, s, CH$_3$), 5.97 (2H, s, NH$_2$), 7.33 (1H, t, J=7.1Hz, ArH$_4$), 7.57 (2H, t, J=7.1Hz, ArH$_3$ and ArH$_5$), 8.16 (2H, d, J=7.1Hz, ArH$_2$ and ArH$_6$), 8.46 (1H, s, H$_3$); RMN^{13}C (δ ppm, DMSO-d$_6$): 14.89 (CH$_3$), 101.23 (C-3a), 121.49 (C-2' and C-6'), 126.37 (C-4'), 129.19 (C-3' and C-5'), 138.81 (C-3), 141.83 (C-1'), 154.41 (C-7a), 156.48 (C-4), 158.40 (C-6); HRMS Calculated for C$_{12}$H$_{11}$N$_5$: 225.1014, found: 225.1018.

χ) **4-amino-6-methyl-N^1-phenyl-1H-pyrazolo[3,4-d] pyrimidine 4c:** Yield 70 %; mp 160 °C; IR (cm^{-1}) ν_{NH2}: 3090, 3320; $\nu_{C=N}$ 1597, 1638, 1663; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.65 (3H, s, CH$_3$), 4.28 (2H, s, NH$_2$), 7.28 (1H, t, J=7.3Hz, ArH$_4$), 7.56 (2H, t, J=7.3Hz, ArH$_3$ and ArH$_5$), 8.19 (2H, d, J=7.3Hz, ArH$_2$ and ArH$_6$), 8.29 (1H, s, H$_6$); RMN^{13}C (δ ppm, DMSO-d$_6$): 14.44 (CH$_3$), 100.24 (C-3a), C$_{arom}$ 120.24

(C-2' and C-6'), 124.67 (C-4'), 129.16 (C-3' and C-5'), 138.80 (C-3), 142.79 (C-1'); C$_3$ 154.14 (C-7a), 156.51 (C-4), 158.58 (C-6); HRMS Calculated for C$_{12}$H$_{11}$N$_5$: 225.1014, found: 225.1016.

1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidines 5a-i: To a mixture of 4-amino-N^1-phenyl-1H-pyrazolo[3,4-d]pyrimidine 4 (1mmol) and α-bromoacetophenone (1mmol) in 3 mL of ethanol, was added p-TsOH (5%) then refluxed for 12 h. After completion of the reaction, as indicated by TLC (EtOAc–hexane, 90:10), the precipitate product was separated by filtration and washed with ethanol and was crystallized from a suitable solvent to obtain pure product.

a) **6-(4-methoxyphenyl)-3-methyl-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5a:** Yield 74 %; mp 210 °C; IR (cm^{-1}): $\nu_{C=N}$ 1505, 1542, 1593; RMN (δ ppm, DMSO-d$_6$): 2.80 (3H, s, CH$_3$); 3.84 (3H, s, CH$_3$); 7.07 (2H, d, J=8.7Hz, ArH$_3$' and ArH$_5$'); 7.39 (1H, t, J=7.2Hz, ArH$_4$); 7.60 (2H, t, J=5.4Hz, ArH$_3$ and ArH$_5$); 7.95 (2H, d, J=8.7Hz, ArH$_2$ and ArH$_6$); 8.15 (2H, d, J=7.8Hz, ArH$_2$' and ArH$_6$'); 8.42 (1H, s, H$_5$); 9.29 (1H, s, H$_8$). RMN^{13}C (δ ppm, DMSO-d$_6$): 14.37 (CH$_3$); 55.17 (CH$_3$); 106.62 (C-3a); 114.23 (C-3" and C-5"); 121.22 (C-5); 121.28 (C-3' and C-5'); 125.22 (C-6); 126.52 (C-1"); 126.79 (C-4'); 127.14 (C-2" and C-6"); 129.18 (C-2' and C-6'); 139.80 (C-1'); 140.20 (C-8); 142.20 (C-4a); 151.35 (C-3); 154.57 (C-9a); 159.35 (C-4"); HRMS Calculated for C$_{21}$H$_{17}$N$_5$O: 355.1433, found: 355.1489.

b) **6-(4-methoxyphenyl)-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5b:** Yield 78 %; mp 171 °C; IR (cm^{-1}): $\nu_{C=N}$ 1511, 1575, 1595; RMN ^{1}H (δ ppm, DMSO-d$_6$): 3.82 (3H, s, CH$_3$); 7.09 (2H, d, J=7.2Hz, ArH$_3$' and ArH$_5$'); 7.46 (1H,

t, J=7.5Hz, ArH$_4$); 7.63 (2H, t, J=8.4Hz, ArH$_3$ and ArH$_5$); 7.94 (2H, d, J=8.7Hz, ArH$_2$ and ArH$_6$); 8.13 (2H, d, J=8.0Hz, ArH$_2$' and ArH$_6$'); 8.54 (1H, s, H$_5$); 8.69 (1H, s, H$_3$); 8.41 (1H, s, H$_8$). RMN^{13}C (δ ppm, DMSO-d$_6$): 55.26 (CH$_3$); 107.50 (C-3a); 114.14 (C-3''), 114.49 (C-5''), 121.79 (C-5); 121.86 (C-3' and C-5'), 122.67 (C-6), 127.27 (C-1''), 127.38 (C-4'), 128.73 (C-2'' and C-6''), 129.36 (C-2' and C-6'), 133.02 (C-3), 138.13 (C-1'), 139.06 (C-8), 139.95 (C-4a), 144.87 (C-9a), 159.79 (C-4''); HRMS Calculated for C$_{20}$H$_{15}$N$_5$O: 341.1277, Found: 341.1341.

c) 6-p-tolyl-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5c: Yield 81 %; mp 166 °C; IR (cm^{-1}): $\nu_{C=N}$ 1519, 1550, 1594; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.37 (3H, s, CH$_3$); 7.04 (2H, d, J=8.4Hz, ArH$_3$' and ArH$_5$'); 7.34 (1H, t, J=7.5Hz, ArH$_4$); 7.61 (2H, t, J=5.9Hz, ArH$_3$ and ArH$_5$); 7.92 (2H, d, J=6.0Hz, ArH$_2$ and ArH$_6$); 8.13 (2H, d, J=7.5Hz, ArH$_2$' and ArH$_6$'); 8.56 (1H, s, H$_5$); 8.71 (1H, s, H$_3$); 9.42 (1H, s, H$_8$). RMN^{13}C (δ ppm, DMSO-d$_6$): 21.38 (CH$_3$); 108.63 (C-3a); 120.35 (C-5); 121.67 (C-3' and C-5'), 122.45 (C-6), 126.23 (C-4'), 127.83 (C-2'' and C-6''), 129.85 (C-3'' and C-5''), 129.89 (C-2' and C-6'), 130.06 (C-1''), 136.45 (C-3), 138.70 (C-4''), 139.54 (C-1'), 141.50 (C-8), 145.25 (C-4a), 149.10 (C-9a); HRMS Calculated for C$_{20}$H$_{15}$N$_5$: 325.1327, found: 325.1364.

d) 6-p-tolyl-3-methyl-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5d: Yield 69 %; mp 206 °C; IR (cm^{-1}): $\nu_{C=N}$ 1519, 1550, 1594; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.28 (3H, s, CH$_3$); 2.36 (3H, s, CH$_3$); 7.05 (2H, d, J=7.5Hz, ArH$_3$' and ArH$_5$'); 7.40 (1H, t, J=8.5Hz, ArH$_4$); 7.59 (2H, t, J=5.4Hz, ArH$_3$ and ArH$_5$); 7.92 (2H, d, J=6.3Hz, ArH$_2$ and ArH$_6$); 8.01 (2H, d, J=5.1Hz, ArH$_2$' and ArH$_6$'); 8.51 (1H, s, H$_5$); 9.34 (1H, s, H$_8$). RMN^{13}C (δ ppm, DMSO-d$_6$): 14.90 (CH$_3$); 21.39 (CH$_3$); 108.02 (C-3a); 120.95 (C-5); 121.76 (C-3' and C-5'), 122.24 (C-6), 126.23 (C-4'), 127.09 (C-2'' and C-6''), 129.50 (C-3'' and C-5''), 129.88 (C-2' and C-6'), 130.10 (C-1''), 138.19 (C-4''), 139.35 (C-1'), 140.41 (C-8), 142.77 (C-3), 145.48 (C-4a), 149.53 (C-9a); HRMS Calculated for C$_{21}$H$_{17}$N$_5$: 339.1484, found: 339.1421.

e) 6-(4-methoxyphenyl)-8-methyl-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5e: Yield 62 %; mp 212 °C; IR (cm^{-1}): $\nu_{C=N}$ 1500, 1526, 1594; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.78 (3H, s, CH$_3$); 3.86 (3H, s, CH$_3$); 7.02 (2H, d, J=7.3Hz, ArH$_3$' and ArH$_5$'); 7.41 (1H, t, J=7.5Hz, ArH$_4$); 7.68 (2H, t, J=5.4Hz, ArH$_3$ and ArH$_5$); 8.03 (2H, d, J=7.8Hz, ArH$_2$ and ArH$_6$); 8.21 (2H, d, J=7.5Hz, ArH$_2$' and ArH$_6$'); 8.39 (1H, s, H$_5$); 8.87 (1H, s, H$_3$). RMN^{13}C (δ ppm, DMSO-d$_6$): 14.21 (CH$_3$); 55.43 (CH$_3$); 106.58 (C-3a); 114.64 (C-3'' and C-5''); 121.36 (C-5); 121.45 (C-3' and C-5'), 125.51 (C-6), 126.64 (C-1''), 126.81 (C-4'), 127.22 (C-2'' and C-6''), 129.18 (C-2' and C-6'), 133.19 (C-3), 139.74 (C-1'), 142.68 (C-4a), 153.76 (C-9a), 159.82 (C-4''), 162.17 (C-8); HRMS Calculated for C$_{21}$H$_{17}$N$_5$O: 355.1433, found: 355.1457.

f) 6-p-tolyl-8-methyl-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5f: Yield 58 %; mp 205 °C; IR (cm^{-1}): $\nu_{C=N}$ 1512, 1557, 1596; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.29 (3H, s, CH$_3$); 2.34 (3H, s, CH$_3$); 7.05 (2H, d, J=8.4Hz, ArH$_3$' and ArH$_5$'); 7.42 (1H, t, J=8.4Hz, ArH$_4$); 7.60 (2H, t, J=5.4Hz, ArH$_3$ and ArH$_5$); 8.03 (2H, d, J=7.5Hz, ArH$_2$ and ArH$_6$); 8.11 (2H, d, J=8.7Hz, ArH$_2$' and ArH$_6$'); 8.49 (1H, s, H$_5$); 8.74 (1H, s, H$_3$). RMN^{13}C (δ ppm, DMSO-d$_6$): 14.36 (CH$_3$); 22.70 (CH$_3$); 108.83 (C-3a); 120.12 (C-5); 121.32 (C-3' and C-5'), 122.68 (C-6), 126.46 (C-4'), 127.25 (C-2'' and C-6''), 129.26 (C-3'' and C-5''), 129.83 (C-2' and C-6'), 130.91 (C-1''), 134.15 (C-3), 138.94 (C-4''), 139.50 (C-1'), 145.12 (C-4a), 149.67 (C-9a), 163.22 (C-8); HRMS Calculated for C$_{21}$H$_{17}$N$_5$: 339.1484, found: 339.1452.

g) 6-(4-Chlorophenyl)-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5g: Yield 79 %; mp 168 °C; IR (cm^{-1}): $\nu_{C=N}$ 1501, 1556, 1595; RMN ^{1}H (δ ppm, DMSO-d$_6$):

7.45 (1H, t, J=7.3Hz, ArH$_4$); 7.54 (2H, d, J=8.4Hz, ArH$_3$' and ArH$_5$'); 7.62 (2H, t, J=8.1Hz, ArH$_3$ and ArH$_5$); 8.05 (2H, d, J=8.4Hz, ArH$_2$ and ArH$_6$); 8.14 (2H, d, J=7.8Hz, ArH$_2$' and ArH$_6$'); 8.58 (1H, s, H$_5$); 8.66 (1H, s, H$_3$); 9.35 (1H, s, H$_8$). RMN^{13}C (δ ppm, DMSO-d$_6$): 108.66 (C-3a); 120.25 (C-5); 121.77 (C-3' and C-5'), 122.68 (C-6), 127.12 (C-4'), 127.40 (C-2'' and C-6''), 128.85 (C-3'' and C-5''), 129.27 (C-2' and C-6'), 131.83 (C-1''), 132.55 (C-4''), 138.37 (C-3), 139.83 (C-1'), 140.08 (C-8), 142.98 (C-4a), 144.46 (C-9a); HRMS Calculated for C$_{19}$H$_{12}$ClN$_5$: 345.0781, found: 345.0789.

h) 6-(4-Chlorophenyl)-3-methyl-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5h: Yield 69 %; mp 192 °C; IR (cm^{-1}): $\nu_{C=N}$ 1504, 1561, 1591; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.18 (3H, s, CH$_3$); 7.30 (1H, t, J=7.5Hz, ArH$_4$); 7.44 (2H, d, J=7.5Hz, ArH$_3$' and ArH$_5$'); 7.59 (2H, t, J=7.8Hz, ArH$_3$ and ArH$_5$); 7.67 (2H, d, J=7.3Hz, ArH$_2$ and ArH$_6$); 7.79 (2H, d, J=7.5Hz, ArH$_2$' and ArH$_6$'); 8.48 (1H, s, H$_5$); 9.21 (1H, s, H$_8$). RMN^{13}C (δ ppm, DMSO-d$_6$): 14.42 (CH$_3$); 108.29 (C-3a); 120.74 (C-5); 121.32 (C-3' and C-5'), 122.64 (C-6), 127.19 (C-4'), 127.51 (C-2'' and C-6''), 128.87 (C-3'' and C-5''), 129.12 (C-2' and C-6'), 132.11 (C-1''), 132.83 (C-4''), 135.61 (C-3), 139.83 (C-1'), 143.59 (C-4a), 146.28 (C-9a), 162.31 (C-8); HRMS Calculated for C$_{20}$H$_{14}$ClN$_5$: 359.0938, found: 359.0923.

i) 6-(4-Chlorophenyl)-8-methyl-N^1-phenyl-1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidine 5i: Yield 67 %; mp 189 °C; IR (cm^{-1}): $\nu_{C=N}$ 1511, 1553, 1594; RMN ^{1}H (δ ppm, DMSO-d$_6$): 2.24 (3H, s, CH$_3$); 7.18 (1H, t, J=7.0Hz, ArH$_4$); 7.35 (2H, d, J=7.5Hz, ArH$_3$' and ArH$_5$'); 7.54 (2H, t, J=8.0Hz, ArH$_3$ and ArH$_5$); 7.68 (2H, d, J=7.5Hz, ArH$_2$ and ArH$_6$); 7.74 (2H, d, J=7.5Hz, ArH$_2$' and ArH$_6$'); 8.51 (1H, s, H$_5$); 8.82 (1H, s, H$_3$). RMN^{13}C (δ ppm, DMSO-d$_6$): 14.64 (CH$_3$); 108.43 (C-3a); 120.68 (C-5); 121.36 (C-3' and C-5'), 122.85 (C-6), 127.07 (C-4'), 127.23 (C-2'' and C-6''), 128.69 (C-3'' and C-5''), 129.81 (C-2' and C-6'), 131.54 (C-1''), 132.76 (C-4''), 139.84 (C-1'), 141.22 (C-8), 143.36 (C-4a), 143.76 (C-3), 145.84 (C-9a); HRMS Calculated for C$_{20}$H$_{14}$ClN$_5$: 359.0938, found: 359.0941.

Pharmacology

Chemicals and drugs: Carrageenan (BDH Chemicals Ltd., Poole, England), ranitidine (Medis, Tunis, Tunisia) and diclofenac (Medis, Tunis, Tunisia) were purchased from Central pharmacy of Tunisia.

Animals: Westar rats of either sex, weighing 150-180 g of both sexes were obtained from Pasteur Institute (Tunis, Tunisia). Housing conditions and *in vivo* experiments were approved according to the guidelines established by the European Union on Animal Care (CCE Council 86/609).

Antioxidant activities:

DPPH radical-scavenging activity: The free radical-scavenging activity of compounds (5a, b, c, d) were evaluated using the stable radical DPPH, according to the method of Kim et al. [7]. One milliliter of diluted sample (1 mg/mL) was added to 1 mL of the ethanolic DPPH solution. The mixture was then shaken and allowed to stand at room temperature in the dark. After 30 min, the decrease in absorbance at 517 nm was measured against a blank (ethanol solution) by using a UV–Vis spectrophotometer. A mixture consisting of 1 mL of ethanol and 1 mL of DPPH solution was used as the control. The radical-scavenging activity of test samples, expressed as percentage inhibition of DPPH, was calculated according to the formula:

% inhibition=$[(A_B-A_A)/A_B] \times 100$, where A_B and A_A are the absorbance values of the control and of the test sample, respectively. The compound concentration providing 50% inhibition (IC$_{50}$) was calculated from the graph of inhibition percentage plotted against

test samples concentration. DPPH radical-scavenging activity of compounds (5a, b, c, d) were compared with ascorbic acid used as standard.

Ferric reducing antioxidant power (FRAP): The ferric reducing power (FRAP) of compounds (5a, b, c, d) was evaluated using the method described by Zaouali et al. [8]. Briefly, one milliliter of diluted sample (1 mg/mL) was mixed with 2.5 mL of Potassium phosphate buffer (0.1 M, pH 6.6) and 2.5 mL of 1% (w/v) potassium ferricyanide. The mixture was incubated at 50 °C for 20 min, thereafter 2.5 mL of 10% (w/v) trichloroacetic acid was added, and subsequently centrifuged at $1000 \times g$ for 10 min. 2.5 mL of the supernatant was mixed with equal volume of water and 0.5 mL of 0.1% (w/v) ferric chloride. The solution was incubated at ambient temperature for 30 min for color development. The absorbance of all sample solutions was measured at 700 nm and compared with ascorbic acid used as standard.

Carrageenan-induced rat paw oedema: Wistar rats were divided into groups of six animals. Oedema was induced by injecting 0.05 mL of 1% carrageenan subcutaneously into the sub-plantar region of the left hind paw. Compounds 5a, b, c and d (50 and 100 mg/ kg) were administered intraperitoneally (i.p.). The control group received the vehicle (Tween 80/absolute ethanol/saline solution (0.9 %) in the ratio 1:1:18) (2.5 mL/ kg, i.p.). The reference group received dichlofenac (25 mg/ kg, i.p). All drugs were administrated 30 min before the injection of carrageenan. Measurement of paw size was done by means of volume displacement technique using plethysmometer (Ugo Basile no.7140) immediately before carrageenan injection and 1, 2, 3, 4 and 5 h after carrageenan injection. Percentages of inhibition in our anti-inflammatory tests were obtained for each group using the following ratio: $[(V_t-V_o) \text{ control}-(V_t-V_o)\text{treated}] \times 100/(V_t-V_o)\text{Control}$

Where, V_t is the average volume for each group and V_o is the average volume obtained for each group before any treatment.

Gastroprotective activity: Westar rats were initially treated with compounds 5a, b, c, d (50 and 100 mg/kg, i.p.). After 30 min, gastric damage was induced in the experimental groups by 150 mM HCl/EtOH (40:60, v/v) orally administration (1 mL/100 g), while the control group received vehicle and the reference group received ranitidine (60 mg/ kg, i.p). One hour later, the animals were sacrificed and their stomachs rapidly removed and opened via an incision along the greater curvature [9]. Gastric damage was determined by measuring the extent of the ulcerative lesions. The summative length of these lesions was recorded (mm) as lesion index.

Acetic acid writhing test in mice: The analgesic activity was performed according to the method of Ayed et al. [10]. Swiss mice (20–30 g) were selected one day prior to each test and were divided into six groups of six mice each. One group served as control (vehicle 10 ml/kg) by subcutaneous injection (s/c). The second group was given the acetylsalicylate of lysine (ASL) (200 mg/kg) by the same route, as a reference drug. The remaining group was treated with compounds 5a, b, c, d (50 and 100 mg/kg, (s/c)). All animals received 10 ml/kg (i.p.) of 1% acetic acid 30 min after treatment. The number of writhing was recorded during 30 min commencing 5 min after the acetic acid injection. A writhe is indicated by abdominal constriction and stretching of at least one hind limb.

Anticandidal activity: The antifungal effect of compounds 5a, b, c, d was tested against three *Candida* strains (*Candida albicans* ATCC 90028, *Candida glabrata* ATCC 90030 and *Candida parapsilosis* ATCC 22019). The minimal inhibitory concentration (MIC) preventing visible fungal growth was measured by the broth dilution method (microdilution using 96-well microplates), following the procedure of Hammer et al. [11]. Compounds 5a, b, c, d solutions were prepared by dissolution in 10% dimethyl sulfoxide (DMSO). The compounds concentrations tested ranged from 0.1 to 10 mg/ml. The MIC of each compound was defined as the lowest concentration which inhibited candidal growth after incubation at 37°C between 18 and 24 h. The minimal fungicidal concentration (MFC) was determined by subculture on blood agar at 37°C between 18 and 24 h. Amphotericin B was used as anticandical positive control.

Statistical analysis

Data are presented as the mean ± standard error of the mean (s.e.m). Statistical analysis was performed using Student's *t*-test. The significance of difference was considered to include values of $P<0.05$.

Results and Discussion

Chemistry

The 5-amino-4-cyano-N^1-phenylpyrazole 2, used as a starting material, was prepared in two steps following a similar method reported by Petrie and al [12-14]. The first step involves acid catalyzed condensation of orthoester with malonate to form ethoxymethylene malononitrile 1. This later reacts then with substituted hydrazine to give the aminocyanopyrazole 2. Treatment of 2 with orthoester in the presence of catalytic amount of acid furnished the corresponding cyano-pyrazoloimidates 3 which subsequently were transformed to the corresponding amino pyrazolopyrimidines 4 upon treatment with ammoniac [5,15-17]. Reaction of compound 4 with α-bromoacetophenone in the presence of a catalytic amount of PTSA (5 mol%) in refluxing ethanol furnished the 1,7-dihydropyrazolo[3,4-*d*] imidazo[1,2-*f*]pyrimidines derivatives 5. 1,7-dihydropyrazolo[3,4-*d*] imidazo[1,2-*f*]pyrimidines derivatives 5 were isolated as stable compounds in good yields.

The reaction occurs by a primary amino group intercepts a bromine atom liberating HBr, followed by an intracyclisation and elimination of a water molecule to give 1,7-dihydropyrazolo[3,4-*d*]imidazo[1,2-*f*] pyrimidines 5a-i. It is interesting to note that time reaction and yield of products are directly related to the nature of substituent (R_1, R_2 and R_3). The yields of compounds 5a and 5c are 74 and 81 %, respectively. When R is a hydrogen substituent, the product is obtained with superior yield in short time (e.g., Compounds 5b and 5c and 5g.). From these results, we conclude that the electronic nature of the substituent on the bromoacetophenone has a significant role on the reaction outcome. The correct identity of compound as 1,7-dihydropyrazolo[3,4-*d*] imidazo[1,2-*f*]pyrimidines was confirmed by ^{1}H NMR, ^{13}C NMR and HRMS (Scheme 1; Table 1).

Pharmacology

Antioxidant activities: Free radicals have an important role in pathogenesis of a wide range of diseases including inflammation. Antioxidants can prevent biological and chemical substances from free radical induced oxidative damage and stress. Consequently, multipotent antioxidants have gained a great attention from scientists for their potential in treatment of many diseases [18]. In this study, two methods were used to evaluate the antioxidant activity of compounds 5a, b, c, d; the DPPH radical-scavenging activity and ferric reducing antioxidant power (FRAP). Figure 1 show that the radical-scavenging activity of compounds (5a, b, c, d) and ascorbic acid on DPPH radicals increased in dose-dependent manner. The IC_{50} values calculated from the graph (Figure 1) show that compound 5a exhibited significant antioxidant activity with IC_{50} values of 0.037 mg/mL, whereas, compound 5b exhibited moderate activity (IC_{50} value of 0.049 mg/mL). Compounds

Compounds	R_1	R_2	R_3	Yields (%)	Mp (°C)	Reaction time (h)
5a	CH_3	H	4-OCH_3	74	210	12
5b	H	H	4-OCH_3	78	171	7
5c	H	H	4-CH_3	81	166	5
5d	CH_3	H	4-CH_3	69	206	12
5e	H	CH_3	4-OCH_3	62	212	24
5f	H	CH_3	4-CH_3	58	205	18
5g	H	H	4-Cl	79	168	5
5h	CH_3	H	4-Cl	69	192	10
5i	H	CH_3	4-Cl	67	189	17

Table 1: Synthesis of 1,7-dihydropyrazolo[3,4-d]imidazo[1,2-f]pyrimidines **5a–i.**

Sample	IC_{50}^a of DPPH radical-scavenging activity (mg/ml)	IC_{50}^b of reducing power (mg/ml)
5a	0.037 ± 0.01	0.042 ± 0.02
5b	0.049 ± 0.03	0.065 ± 0.03
5c	-	-
5d	-	-
Ascorbic acid	0.013 ± 0.02	0.025 ± 0.01

Values are expressed as mean ± SEM of triplicate measurement.
[a] IC_{50} means the concentration of sample that can decrease DPPH concentration by 50%.
[b] IC_{50} is the concentration for which the absorbance at 700 nm is 0.5.

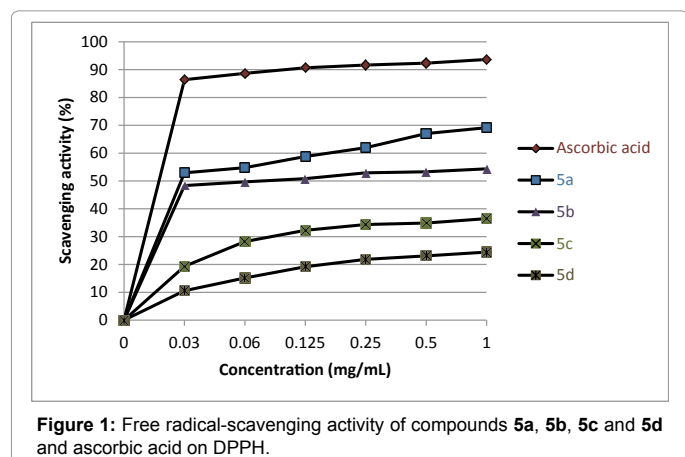**Table 2:** IC_{50} values of DPPH radical-scavenging activity and reducing power of compound **5a**, **5b**, **5c** and **5d**.

Figure 1: Free radical-scavenging activity of compounds **5a**, **5b**, **5c** and **5d** and ascorbic acid on DPPH.
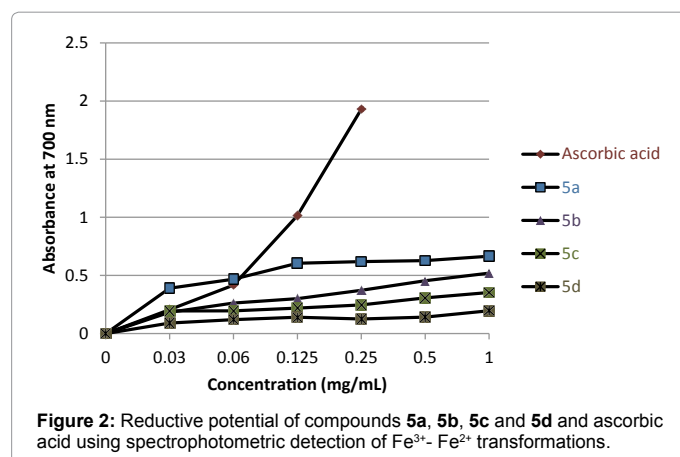

Figure 2: Reductive potential of compounds **5a**, **5b**, **5c** and **5d** and ascorbic acid using spectrophotometric detection of Fe^{3+}- Fe^{2+} transformations.

5c, d don't have any activity. So, compound 5a, b have the highest free radical scavenging activity, which was found to be comparable with that of ascorbic acid (IC_{50}=0.013 mg/mL) (Table 2).

In addition, Figure 2 shows the ability of compounds (5a, b, c, d) and ascorbic acid to reduce Fe^{3+} to Fe^{2+} at different concentration ranges. The reducing potential of compounds (5a, b, c, d) and ascorbic acid increased with increase of concentrations. Compound 5a had the highest reducing power, with IC_{50} = 0.042 mg/mL followed by compound 5b (IC_{50} = 0.065 mg/mL) (Table 2). The FRAP values of compounds 5a, b were comparable with that of ascorbic acid (IC_{50} = 0.025 mg/mL), however compounds 5c, d showed a lower reducing power (Table 2). These different values of IC_{50} indicated that the order of increasing reductive potential of ferric iron was 5b < 5a < ascorbic acid.

The reducing power of various compounds might be due to their hydrogen-donating ability as described by Shimada et al. [19]. So, compounds 5a, b is good electron donors and could terminate the free-radical chain reactions by converting free radicals to more stable products (Figure 2).

Anti-inflammatory activity: The anti-inflammatory activity of compounds 5a, b, c, d against acute pedal edema (induced by carrageenan) is shown in Table 3 and the results are comparable to that of the standard drug diclofenac, a potent inhibitor of cyclooxygenase 2 (Cox-2). Carrageenan induced paw edema remained even 6 h after its injection into the sub plantar region of rat paw. Diclofenac as a reference standard drug inhibited the edema formation due to carrageenan to an extent of 58.22% (at 3 h) at the dose of 25 mg/kg. The compounds 5a, b, c, d inhibited, significantly edema formation in rats (p< 0.01) in a dose dependent manner. Compound 5a at the dose of 50 mg/kg inhibited edema formation to the extent of 74.5% (at 3 h) and the edema was found to be reduced to 5.25 ± 2 10^{-2} mL (Figure 3). The compound 5b has also a height activity at a dose of 50 mg/

kg with a percentage of inhibition of edema of 65.54% (at 3 h), while compounds 5c and 5d at a dose of 50 mg/kg reduced edema with a percentage of 48.66 and 44.21%, respectively. The presence of edema is one of the prime signs of inflammation [20]. It has been documented that carrageenan induced rat paw edema is suitable *in vivo* model to study anti-inflammatory drugs both steroidal and non-steroidal since it involves several mediators [21]. This method was chosen for the present study since edema induced by carrageenan is the most prominent acute experimental model in search for new anti-inflammatory drugs [20]. Carrageenan induced edema has been commonly used as an experimental animal model for acute inflammation and is believed to be biphasic [22]. The early phase (1-2 h) of the carrageenan model is mainly mediated by histamine, serotonin and increased synthesis of prostaglandins in the damaged tissue surroundings. The late phase (2.5-5 h) is due to the over production of prostaglandin and nitric oxide with peak at 5 h, produced by inducible isoforms of cox (cox-2) and nitric oxide synthase (iNOS) [23].

However, treatment with compounds 5a, b, c, d significantly reduced carrageenan on induced inflammation in both the phases (1-5 h) of the experiment. Based on this, it may be that compounds 5a, b, c, d have a non-selective inhibiting effect on the release or actions of these mediators of inflammation and the suppression of the 1st phase may be due to inhibition of the release of early mediators, such as histamine and serotonin and the action in the 2nd phase may be explained by an inhibition of cyclooxygenase 2.

Gastroprotective activity: The results of gastroprotective activity of compounds 5a, b, c, d on gastric ulcer induced by HCl/ ethanol solution are shown in Table 4. Oral administration of the damaging agent to the control group clearly produced a mucosal damage characterized by

Treatment	Dose (mg/kg)	Edema (10⁻² mL) (mean ± S.E.M)			Edema inhibition (%)		
		1 h	**3 h**	**5 h**	**1 h**	**3 h**	**5 h**
Vehicle	-	25.5 ± 2.3	37.5 ± 2.2	42.6 ± 2.6	-	-	-
Diclofenac	25	13.22 ± 2.7*	15.67 ± 2.1*	20.05 ± 2.9*	48.15	58.22	52.94
5a	50	7.62 ± 1.4**	9.56 ± 1.6**	12.63 ± 2.2**	70.11	74.5	70.35
	100	3.75 ± 1.6**	5.25 ± 2**	7.75 ± 2.2**	85.29	86	81.8
5b	50	10.6 ± 2.2*	12.92 ± 2.3**	15.19 ± 1.8**	58.43	65.54	64.34
	100	6.75 ± 2.2**	8.75 ± 1.8**	10.25 ± 2.3**	73.52	76.66	75.93
5c	50	13.17 ± 2.4*	19.25 ± 1.6*	22.78 ± 2.4*	48.35	48.66	46.52
	100	9 ± 2.3**	13 ± 2.3**	16.25 ± 1.6**	64.7	65.33	61.85
5d	50	14.58 ± 1.8	20.92 ± 2.2*	25.3 ± 2.4	42.82	44.21	40.61
	100	8 ± 2.3**	11.75 ± 1.8**	14.2 ± 1.6**	68.62	68.66	66.66

Values are expressed as mean ± SEM; n=6 animals. *P<0.01, **P<0.001.
Vehicle: Ethanol: 5%; Tween 80: 5%; distilled water: 90%

Table 3: Anti-inflammatory effect of the intraperitoneal administration of compounds **5a**, **5b**, **5c** and **5d** and of reference drug (Diclofenac) in carrageenan-induced rat paw edema test.
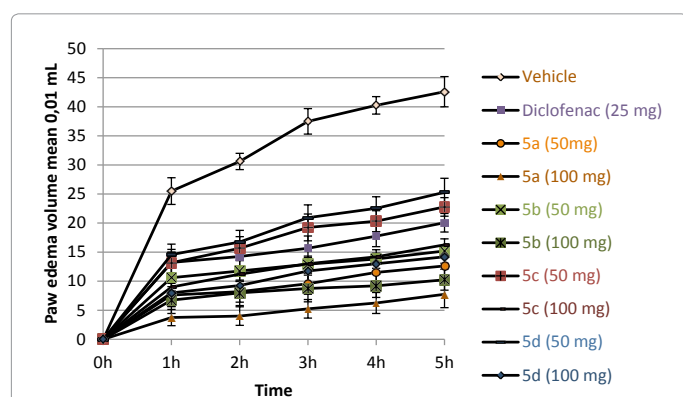

Figure 3: Effect of intraperitoneal administration of compounds **5a**, **b**, **c**, **d** on the carrageenan-induced rat paw edema. Values are mean ± SEM.

Treatment	Dose (mg/kg)	Ulcer index (mm)	Inhibition (%)
Vehicle	-	65.24 ± 4.9	-
5a	50	24.32 ± 1.6**	62.72
	100	15.31 ± 2,8**	76.53
5b	50	37.68 ± 1.4*	42.23
	100	22.52 ± 2.8**	65.48
5c	50	40.70 ± 3.2	37.61
	100	35.15 ± 3.6	46.11
5d	50	50.33 ± 1.4	22.84
	100	39.56 ± 1.6	39.36
Ranitidine	60	22.68 ± 2.9**	65.23

Values are expressed as mean ± SEM; n=6 animals. *P<0.01, **P<0.001.

Table 4: Effect of compounds **5a**, **5b**, **5c** and **5d**, and of reference drug (ranitidine) on gastric ulcer induced by HCl/ethanol in rats.
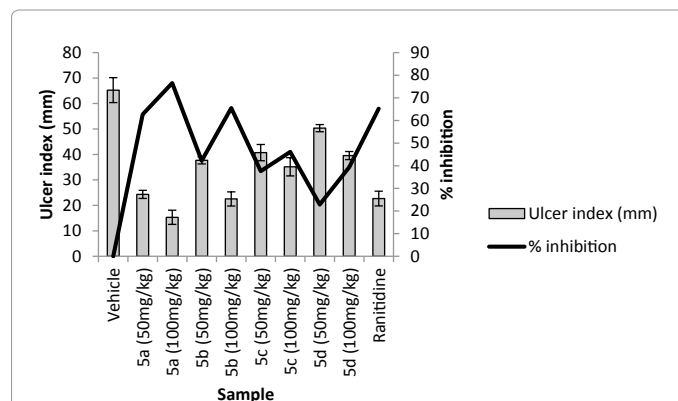
multiple haemorrhage red bands of different sizes along the long axis of the glandular stomach [9].

As shown in Table 4, the compound 5a significantly exhibited, at the dose of 100 mg/kg, the higher inhibition of gastric lesions (76.53%) than compound 5b at the same dose (65.48%). The gastroprotective effect of the two compounds 5a and 5b were similar to the effect produced by the reference drug, ranitidine, which exhibited 65.24% of inhibition. However, 5c and 5d were less effective than the reference drug; the percentage of inhibition of gastric lesions was 46.11% and 39.36%, respectively, at the dose of 100 mg/kg (Figure 4). As described by Jonsson et al. ethanol administration resulted in severe mucosal damage through an increase in reactive oxygen species generation and a decrease in the endogenous antioxidant defense mechanisms. So, oxygen-derived free radicals (ROS) may contribute to ethanol induced gastric mucosal lesions [24]. Thus, compounds 5a and 5b may function by decreasing the redox state in HCl/ethanol induced gastropathy (Figure 4).

Analgesic activity: In addition to the anti-inflammatory activity, the study of analgesic properties of compounds 5a, b, c, d were also evaluated. Among the several models of visceral pain, we use the acetic acid writhing test in mice. Acetic acid acts indirectly by inducing the release of endogenous mediator, which stimulates the nociceptive neurons sensitive to NSAIDs (nonsteroidal anti-inflammatory drugs) and/or opioids [25]. Injection of acetic acid into the control mice resulted in 72.44 writhes. Pretreatment with compounds 5a, b, c, d at doses 50 and 100 mg/kg reduced the number of writhes in a dose dependant manner. Interestingly, compounds 5a (16.82 writhes, 76.78%

Figure 4: Effect of compounds **5a, b, c, d** and the reference drug (Ranitidine) on gastric ulcer induced by HCl/Ethanol in rats. Data expressed as mean ± SEM (n=6). Significantly different from control group: *p<0.05; **p<0.01; ***p<0.001.

of inhibition) and 5b (20.84 writhes, 71.23% of inhibition) at a dose of 100 mg/kg registered higher levels of analgesic activity than 5c (39.22 writhes, 45.85% of inhibition) and 5d (32.56 writhes, 55% of inhibition) and approaches the activity of the standard drug ASL (20.24 writhes, 72% of inhibition) (Table 5).

Anticandidal activity: Anticandidal activity is reported as MIC and MFC (Table 6). All compounds showed significant antifungal activity against *Candida* strains. The best activity was observed with compounds 5a, b against *Candida albicans*. 5a had the highest anticandidal effect in all yeast strains with MIC ranged from 0.62 to 1.22 mg/ml, followed by 5b, d and c which demonstrated antifungal properties depending from the candidal strains. The MICs ranged from 0.84 to 3.42 mg/ml.

Treatment	Dose (mg/kg)	Number of writhes ± SEM	Inhibition of writhing (%)
Vehicle	-	72.44 ± 3.5	-
ASL	200	20.24 ± 1.6**	72.06
5a	50	22.52 ± 2.5**	68.91
	100	16.82 ± 1.4**	76.78
5b	50	28.64 ± 2.8*	60.46
	100	20.84 ± 3.2**	71.23
5c	50	46.82 ± 1.6	35.36
	100	39.22 ± 2.4	45.85
5d	50	42.14 ± 3.6	41.82
	100	32.56 ± 1.4*	55.05

Values are expressed as mean ± SEM; n=6 animals. *$P<0.01$, **$P<0.001$.
Vehicle: Ethanol: 5%; Tween 80: 5%; distilled water: 90%

Table 5: Analgesic effect of the subcutaneous administration of compounds **5a**, **5b**, **5c** and **5d** and of reference drug lysine acetylsalicylate (ASL) in the acetic acid 1% writhing test in mice.

	C. albicans ATCC 90028		*C. glabrata* ATCC 90030		*C. parapsilosis* ATCC22019	
	MIC	MFC	MIC	MFC	MIC	MFC
5a	0.62	1.54	1.22	3.25	0.85	2.32
5b	0.84	2.16	2.36	5.60	1.74	3.62
5c	2.56	6.23	3.42	6.84	2.30	5.24
5d	1.88	4.50	3.12	6.25	1.86	4.12

Positive control with Amphotericin B (MFC 0.5 µg/ml)

Table 6: Antifungal MIC (mg/ml) and MFC (mg/ml) of compounds **5a**, **5b**, **5c** and **5d**.

Either at low concentrations, these compounds showed an interesting anticandidal activity.

In conclusion, our results indicate that compounds 5a and 5b have potential anti-inflammatory and analgesic effects associated with antifungal and gastroprotective properties against HCl/ethanol induced gastric ulcer. The mechanism of 5a and 5b mediated protection may be related to decreases in free radical production. These observations raise the possibility of compounds 5a and 5b being used to treat inflammation and to improve resistance to gastric mucosal injury.

References

1. Hajhashemi V, Sajjadi SE, Heshmati M (2009) Anti-inflammatory and analgesic properties of Heracleum persicum essential oil and hydroalcoholic extract in animal models. J Ethnopharmacol 124: 475-480.

2. Ngo DH, Wijesekara I, Vo TS, Ta QV, Kim SK (2011) Marine food-derived functional ingredients as potential antioxidants in the food industry: an overview. Food Res Int 44: 523-529.

3. Ngo DH, Vo TS, Ngo DN, Wijesekara I, Kim SK (2012) Biological activities and potential health benefits of bioactive peptides derived from marine organisms. Int J Biol Macromol 51: 378-383.

4. Weitzel KW, Wickman JM, Augustin SG, Strom JG (2000) Zaleplon: a pyrazolopyrimidine sedative-hypnotic agent for the treatment of insomnia. Clin Ther 22: 1254-1267.

5. Gupta S, Rodrigues LM, Esteves AP, Oliveira-Campos AMF, José Nascimento MS, et al. (2008) Synthesis of N-aryl-5-amino-4-cyanopyrazole derivatives as potent xanthine oxidase inhibitors. Eur J Med Chem 43: 771-780.

6. Allouche F, Chabchoub F, Carta F, Supuran CT (2013) Synthesis of aminocyanopyrazoles via a multi-component reaction and anti-carbonic anhydrase inhibitory activity of their sulfamide derivatives against cytosolic and transmembrane isoforms. J Enzyme Inhib Med Chem 28: 343-349.

7. Kim JK, Noh JH, Lee S, Choi JS, Suh H, et al. (2002) The first total synthesis of 2,3,6-tribromo-4,5-dihydroxybenzyl methyl ether (TDB) and its anti-oxidant activity. Bull Korean Chem Soc 23: 661-662.

8. Zaouali Y, Bouzaine T, Boussaid M (2010) Essential oils composition in two Rosmarinus officinalis L. varieties and incidence for antimicrobial and antioxidant activities. Food Chem Toxicol 48: 3144-3152.

9. Deghrigue M, Festa C, Ghribi L, D'Auria MV, De Marino S, et al. (2014) Pharmacological evaluation of the semi-purified fractions from the soft coral Eunicella singularis and isolation of pure compounds. DARU J Pharm Sci 22: 64.

10. Ayed Y, Dellai A, Ben Mansour H, Bacha H, Abid S (2012) Analgesic and antibutyrylcholinestrasic activities of the venom prepared from the Mediterranean jellyfish Pelagia noctiluca (Forsskal, 1775). Ann Clin Microbiol Antimicrob 11: 15.

11. Hammer KA, Carson CF, Riley TV (1999) Antimicrobial activity of essential oils and other plant extracts. J Appl Microbiol 86: 985-990.

12. Petrie CR 3rd, Cottam HB, McKernan PA, Robins RK, Revankar GR (1985) Synthesis and biological activity of 6-azacadeguomycin and certain 3,4,6-trisubstituted pyrazolo[3,4-d]pyrimidine ribonucleosides. J Med Chem 28: 1010-1016.

13. Anderson JD, Cottam HB, Larson SB, Nord LD, Revankar GR, et al. (1990) Synthesis of certain pyrazolo[3,4-d]pyrimidin-3-one nucleosides. J Heterocycl Chem 27: 439-453.

14. Aggarwal R, Kumar V, Kumar R, Singh SP (2011) Approaches towards the synthesis of 5-aminopyrazoles. Beilstein J Org Chem 7: 179-197.

15. Booth BL, Costa FAT, Mahmood Z, Pritchard RG, Proença MF (1999) Synthesis of (Z)-N-(2-amino-,2-dicyanovinyl) formamide O-alkyloximes and a study of their cyclisation in the presence of base. J Chem Soc Perkin Trans 1 13: 1853-1858.

16. Oliveira-Campos AMF, Salaheldin AM, Rodrigues LM (2007) Synthesis of some novel pyrazolo[3,4-d]pyrimidine derivatives. Arkivoc 16: 92-100.

17. Bakavoli M, Bagherzadeh G, Vaseghifar M, Shiri A, Pordel M, et al. (2010) Molecular iodine promoted synthesis of new pyrazolo[3,4-d]pyrimidine derivatives as potential antibacterial agents. Eur J Med Chem 45: 647-650.

18. Zhang HY, Yang DP, Tang GY (2006) Multipotent antioxidants: from screening to design. Drug Discov Today 11: 749-754.

19. Shimada K, Fujikawa K, Yahara K, Nakamura T (1992) Antioxidative properties of xanthan on the autoxidation of soybean oil in cyclodextrin emulsion. J Agric Food Chem 40: 945-948.

20. Yonathan M, Asres K, Assefa A, Bucar F (2006) In vivo anti-inflammatory and anti-nociceptive activities of Cheilanthes farinosa. J Ethnopharmacol 108: 462-470.

21. Woldesellassie M, Eyasu M, Kelbessa U (2011) In vivo anti-inflammatory activities of leaf extracts of Ocimum lamiifolium in mice model. J Ethnopharmacol 134: 32-36.

22. William CM, Rao NV, Kumar BR, Mohan GK (2010) Anti-inflammatory and analgesic activities of methanolic extract of Kigelia pinnata DC flower. J Ethnopharmacol 130: 179-182.

23. Panthong A, Supraditaporn W, Kanjanapothi D, Taesotikul T, Reutrakul V (2007) Analgesic, anti-inflammatory and venotonic effects of Cissus quadrangularis Linn. J Ethnopharmacol 110: 264-270.

24. Matsumoto T, Moriguchi R, Yamada H (1992) Role of polymorphonuclearleucocytes and oxygen-derived free radicals in the formation of gastric lesions induced by HCl/ethanol, and a possible mechanism of rotection by antiulcer polysaccharide. J Pharm Pharmacol 45: 535-539.

25. Collier HO, Dinneen LC, Johnson CA, Schneider C (1968) The abdominal constriction response and its suppression by analgesic drugs in the mouse. Br J Pharmacol Chemother 32: 295-310.