

Targeting Obesity for the Treatment of Type 2 Diabetes Mellitus

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Abstract

The discovery of diabetes mellitus (simplified as “diabetes”) could be traced back to 1500 B.C. The first detailed description appeared in 1800, but in the last few decades, the prevalence of diabetes increases dramatically. Till present, according to WHO’s report, 247 million people worldwide have been diagnosed Diabetes, among which 3.47 million died from diabetes. Thus, WHO projects that diabetes will become the 7th leading cause of death in 2030. The typical feature of diabetes is hyperglycemia as a result of metabolic disorder. As the disease progress, complications develop including macrovascular and microvascular abnormality, leading to multiple organ failure and mortality. Apart from diabetes-associated vascular diseases, numerous studies have documented the association between hyperglycemia and cancer. There are three main types of diabetes: (1) type 1 diabetes mellitus (T1DM) in which pancreatic β cell death by immune attack; (2) type 2 diabetes mellitus (T2DM) in which pancreatic β cells fail to produce sufficient amount of insulin; and (3) gestational diabetes which occurs in pregnant women due to high demand for insulin production. Type 2 DM accounts for 80-90% of diabetes and obesity has been identified as a strong and modifiable risk factor in type 2 DM. In this review, we first provide a general introduction of T2DM in the aspects of diagnosis, pathogenesis and clinical characteristics of T2DM. We will focus on how obesity has the adverse impact on insulin resistance, pancreatic β cell death and dysfunction and the development of T2DM. Next, we will summarize the animal models of T2DM including their advantages and drawbacks when compared to their clinical relevance. Finally, we will summarize the interventions that have been applied to treat obesity and T2DM and the remaining problems.

Keywords: Obesity; Diabetes; Pancreatic β cells; Hepatocytes, Gastric bypass

General Introduction of T2DM

Definition and diagnosis criteria of T2DM

Hyperglycemia is the main feature of diabetes, thus, plasma glucose level remains the solo and golden standard in the diagnosis of the disease. According to WHO’s recent recommendations, the threshold of glucose level for diagnosis of diabetes, impaired glucose tolerance (IGT) and impaired fasting glucose (IFG) are summarized in Table 1. One whose fasting glucose level exceeds 7 mmol/l or 2-h oral glucose tolerance test (OGTT) exceeds 11.1 mmol/l is diagnosed as diabetes. Different from that, diagnosis of IGT and IFG require both criteria.

Pathogenesis of T2DM

In addition to diet and life style, both environment and genetic factors contribute to the manifestation of T2DM. Up to date, more than 36 genes have been identified to contribute to T2DM, most of which are involved in pancreatic β cell function [1]. As a result of diet and life style changes, obesity develops and becomes a strong

independent risk factor in T2DM. Despite the etiological factors are diverse and varied, the underlying pathogenesis of T2DM remains the same: insulin resistance, β cell hyperplasia and insulin insufficiency. Physically, plasma level of glucose increases after diet, which stimulates pancreatic beta cell to secrete insulin. Insulin, on one hand promotes glucose uptake and metabolism in the liver, skeletal muscle and fat tissues, on the other hand, enhances glycogen synthesis, leading to glucose homeostasis. In pathological conditions such as obesity, excess body fat, particularly visceral fat, releases large amount of free fatty acid into blood. The free fatty acid interferes with insulin signaling by (1) being absorbed by liver and skeletal muscle and thus becomes the main fuel for energy production; and (2) increased triglyceride storage in liver and skeletal and inhibition of glycogen synthesis. This reduced effect of liver and skeletal in responses to insulin is so called “insulin resistance”. The scheme of insulin resistance is illustrated in Figure 1.

To overcome sustained high level of glucose in blood, pancreatic β cells have to produce more insulin. The compensation response consists of both insulin production and increased β cell number so

	Diabetes	Impaired glucose tolerance (IGT)	Impaired fasting glucose (IFG)
Fasting	≥ 7 mmol/l	< 7 mmol/l	6.1-6.9 mmol/l
Plasma	(126 mg/dl)	(126 mg/dl)	(110-125 mg/dl)
Glucose			
2-h OGTT*	11.1 mmol/l	7.8-11.1 mmol/l	< 7.8 mmol/l
	(200 mg/dl)	(140-200 mg/dl)	(140 mg/dl)

*2-h OGTT: plasma glucose level at 2 hours after oral ingestion of 75 g glucose

Table 1: Plasma glucose level in the diagnosis of diabetes.

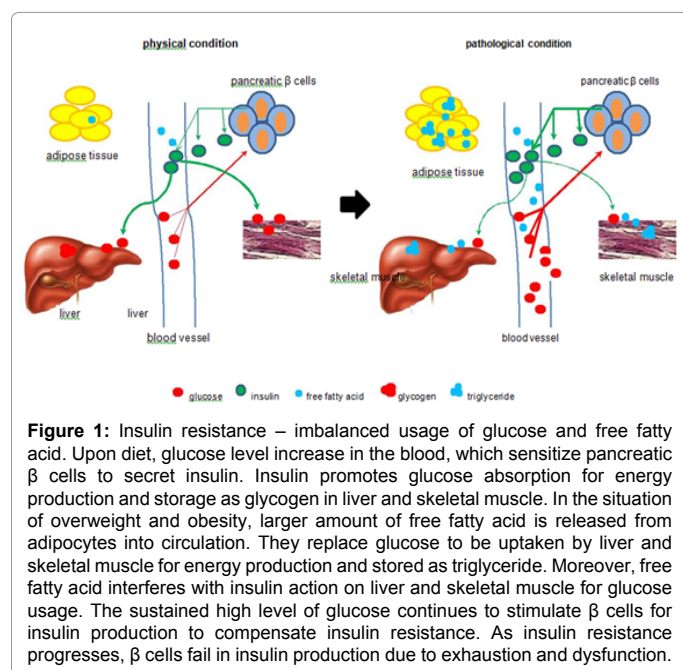
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called hyperplasia. Evolutionally, β cell replication occurs at high rate in neonatal period and grown young adult but decreases in adult stage [2]. To meet the increasing requirement of insulin, adult β cells could proliferate and generate new β cells for insulin production. Although it is still controversial whether β cell neogenesis or other pluripotent stem cells may also serve as sources for the formation of β cells, [3] by using lineage tracing technologies, more and more evidence collectively indicate that self-replication of pre-existing β cells remain the main mechanism in β cell hyperplasia [4-7].

As the pathological situation progresses, exhausted and dysfunctional β cell fail to produce adequate insulin in response to the metabolic demand. Following β cell hyperplasia, the state, of decreased insulin secretion, is namely “insulin sufficiency” that leads to diabetes.

Clinical features of T2DM

Hyperglycemia appears in the early stage of T2DM. Although mainly displayed in T1DM, features of excess thirst (polydipsia), frequent urination (polyuria), increased hunger (polyphagia) and loss of body weight can be also seen in T2DM in severe status. As most pathological progress involves in blood vessels of multiple organs, T2DM patients have multifaceted complications which are summarized below.

Macrovascular abnormality: Diabetes accounts for 50% of mortality in patients with cardiovascular disease [8,9]. Hyperglycemia, free fatty acid, dyslipidemia together with other factors may promote endothelium injury and platelet adhesion and aggregation to injured endothelium, leading to endothelial cell activation, inflammatory cell infiltration and then atherosclerotic plaque formation. The atherosclerotic plaques in T2DM patients do not differ from that of non-T2DM patients but they are more extensive.

Microvascular abnormality: Microvascular changes can be seen in almost all the tissues in T2DM, however, the clinical features that are frequently seen are retinopathy, nephropathy and neuropathy.

- a. Diabetic retinopathy: Approximately, about one third T2DM

patients develop retinopathy and the prevalence increases with the progression of diabetes and hypertension [10]. In American, retinopathy-induced blindness is the main cause of blindness than any other cause. Retinopathy is usually graded into background and proliferative retinopathy [11]. Although it is not clear how retinopathy develops, endothelial cell damage, thickening of basement membrane, tiny hemorrhages and microaneurysms are characteristic of background retinopathies. In progress, neovascularization and fibrovascula proliferation may occur in the progression of background retinopathy toward proliferation retinopathy. The new capillaries are more fragile with more leakage. It is only when pathological changes encroach on the macula that visual impairment and blindness occur.

- b. Diabetic nephrology: Diabetic nephrology affects about 33% of T2DM patients [12]. The earliest clinical sign of nephropthy in T2DM is the detection of small amount of proteins in the urine (termly microalbuminuria). Over decades, larger amount of proteinuria develops, leading to chronic renal failure.
- c. Diabetic peripheral neuropathy (DPN): DPN affects approximately 50% of diabetic patients, therefore, it becomes the most prevalent complication of diabetes [13]. Because of pathological change in the nerve, patients lose sensation of temperature but sense chronic pain, more severely recurrent foot ulceration. Amputation may be taken in pain relief and prevention of further ulceration and infections.

Diabetes-associated carcinoma: The diabetes-, obesity-related cancer was noticed in 1934. In these decades, extensive studies have been performed to interpret whether and how cancer occurs in response to diabetes, hyperinsulinmia and hyperglycemia. Table 2 provides an overview of the incidence of cancer in patients with obesity and diabetes. Although the mechanisms are not fully understood yet, the dysregulated insulin-insulin growth factor 1 (IGF-1) pathways and specific hormones have been demonstrated to play major roles in the occurrence of cancer in diabetes patients.

The Impact of Obesity on T2DM

The positive association between obesity and the prevalence of diabetes

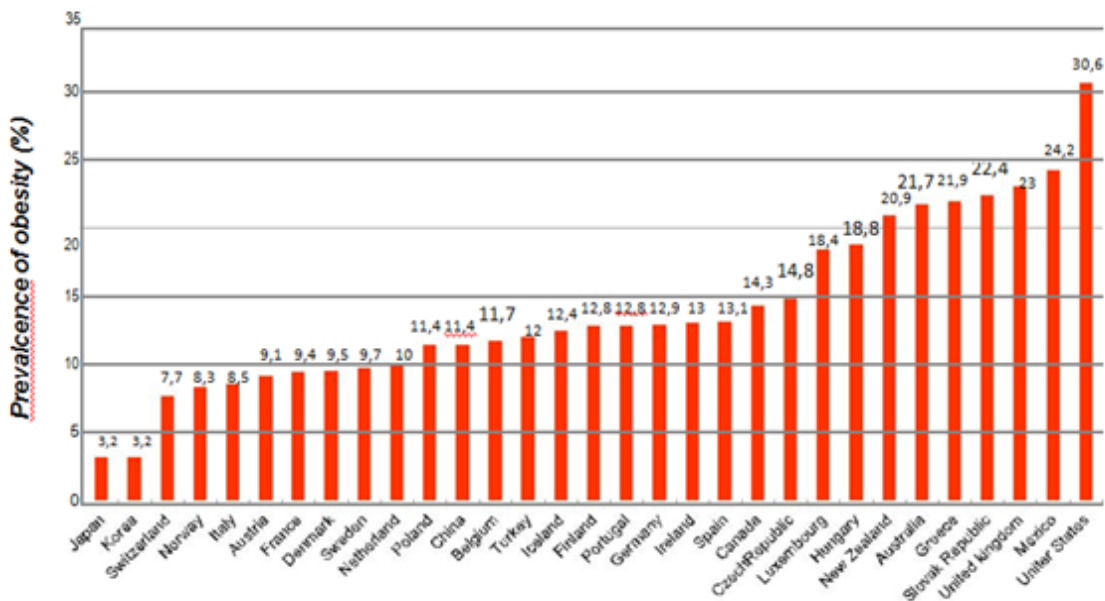
As a consequence of lifestyle changes, obesity becomes epidemic worldwide. The definition of overweight and obesity directly comes from body-mass index (BMI) while waist circumference serves as a reference. Definition of normal weight, overweight and obesity is shown in Table 3. To be addressed, the obesity criteria may slightly vary from ethic races. For example, in Asians, BMI 23 kg/m² can be used to define overweight and BMI 25 or 27 kg/m² can be used to indicate obesity. The worldwide prevalence of obesity is summarized in Figure 2. Accumulated evidence has identified that obesity is an independent risk factor in cardiovascular disease, [14-16] diabetes, [16-20] and hypertension [20,21]. Population-based studies consistently point out the intimate association between obesity and diabetes as a function of prevalence (Figure 3).

The molecular mechanisms underlining the adverse effect of obesity on T2DM

From a global view, adipocytes in obese subjects not only disturb glucose metabolism but also illicit inflammation cascade by producing

Type of study	Study size	Overall incidence of cancer or findings	Cancer subtype
Zhang et al. [79] cohort study	7,950 T2DM subjects	108.36/10 ⁵ subjects in men; 870.2/10 ⁵ subjects in women	Pancreatic cancer liver cancer; kidney cancer breast cancer
He et al. [80] Meta-analysis	2,596,856 T2DM Patients	66.68-84.51 per 100,000 person-years	Bladder cancer
Yang et al. [81] Meta-analysis	643,683 DM patients; 4,819,656 non-DM patients	Increased risk of cancer in DM patients compared to non-DM patients	Bladder cancer
Adami et al. [82] Cohort study	153, 852 T2DM patients	819 incident cancers in 1-24 years	Primary liver cancer, biliary tract cancer
Chari et al. [83] Cohort study	2122 diabetic patients	1% subjects diagnosed cancer in 3 years	Pancreatic cancer
Redaniel et al. [84] Cohort study	52,657 T2DM female patients	A 29% overall cancer risk in DM patients; Positive association between diabetes and breast cancer	Breast cancer
Park et al. [85] Retrospective study	1213 men in which 408 subjects were obese	Positive association between obesity and cancer	Prostate cancer
Larsson et al. [86] Meta-analysis	11,079 subjects	Obese persons with increased risk of cancer	Liver cancer
Olsen et al. [87] Meta-analysis	28 studies	Positive association between obesity and cancer	Ovarian cancer
Key et al. [88] Prospective study	624 obese subjects and 1669 controls	Positive association between obesity and cancer	Breast cancer

Table 2: Association between obesity, diabetes and cancer.



Adapted from Anon (2005) OECD Factbook; Xi et al, Obesity Reviews 2011.

Figure 2: Worldwide prevalence of obesity (BMI ≥ 30 kg/m²) in 2009. Adapted from Anon (2005) OECD Factbook; Xi et al. [76].

adipokines such as resistin and inflammatory cytokines such as TNF- α , monocyte chemoattractant proteins 1 (MCP-1) and interleukins. In this section, detailed overview is provided to understand the pathological machinery of obesity in T2DM.

Obesity and the fate of pancreatic β cells: In obesity, adipocytes produce and release huge amount of free fatty acid (FFA). Prolonged exposure of FFAs to β cells inhibits insulin synthesis [22] and impairs conversion of proinsulin to insulin [23,24]. In parallel, overloading FFAs to β cells promotes ER stress as evidenced by activation of ATF6 and XBP-1 and CHOP up-regulation. Exposure of high glucose and FFAs increases reactive oxygen species (ROS) production and leads to mitochondria dysfunction in β cells [25,26]. Resistin is specifically

expressed and secreted by adipocytes, whose expression is negatively regulated by β cells and by activation of PPAR γ [27,28]. Resistin antagonizes insulin action and reduces glucose uptake in peripheral tissue [28,29]. Apart from its negative effect on metabolism, it increases transcription of several inflammatory cytokines such as IL-1, TNF- α in an NF- κ B-mediated fashion [30,31]. Furthermore, it upregulates intercellular chemotactic molecule-1 (ICAM-1), vascular cell-adhesion molecule-1 and CCL12 expression on endothelial cells, leading to enhanced recruitment of leukocytes [32]. Leptin, a hormone secreted from adipocytes, restricts food intake and enhances energy consumption. Moreover, it is involved in the inhibition of insulin synthesis and secretion. The net effect of leptin tends to adapt glucose homeostasis to the amount of body fat. Released by

Category	BMI (kg/m ²)
Underweight	<18.5
Normal weight	18.5-24.9
Overweight	25.0-29.9
Obesity	≥30

Table 3: Classification of overweight and obesity.

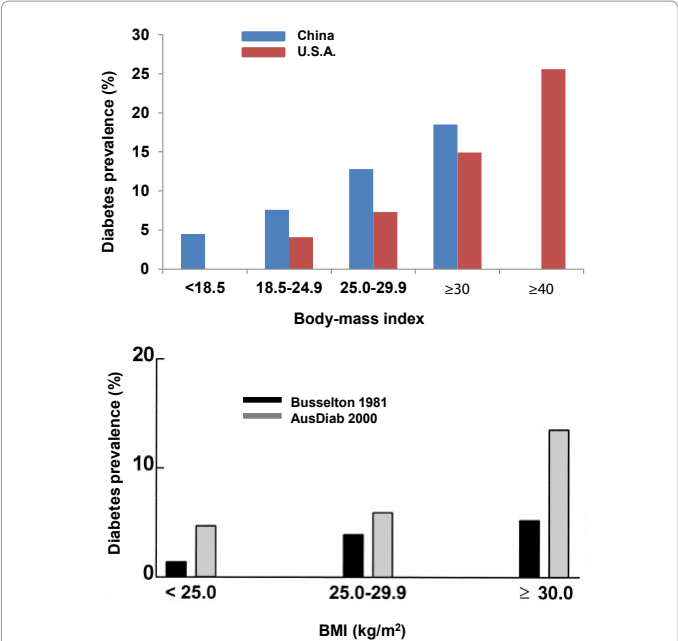


Figure 3: The percentage of diabetes in different categories of BMI. (A) In Chinese populations and Americans in 2009. Adapted from Mokdad et al. JAMA, 2003 [19] and Yang et al. N Engl J Med 2010 [77]. (B) In Austrian population in 1981 and 2000. Adapted from Dustan et al Diabetes Care 2002 [78].

adipocytes, transforming growth factors (TGF- β 1) inhibits insulin transcription via TGF-beta/Smad3 axis [33,34]. TGF- β 1, together with resistin and inflammatory cytokines TNF- α , plasminogen activator inhibitor-1 (PAI-1) and MCP-1 produced by adipocytes, they stimulate inflammation in both NF- κ B dependent and independent manners [26,35-37]. Adipocytes also produce adiponectin that has beneficial effect on insulin release and promote β cell survival [38,39]. Unfortunately, adiponectin secretion is dramatically reduced in obesity [40]. Ultimately, β cells are subjected to dysfunction and death. To be noted, different from these proinflammatory cytokines, IL-6 produced by adipocytes may have dual effects on β cells. Upon engagement of IL-6 to its receptors containing IL-6R and gp130, activation of Janus-associated kinases (JAK) and phospholipase C (PLC)-inositol triphosphate (IP₃) pathways has been reported, which is associated with inflammation and enhanced glucose-stimulated insulin secretion, respectively [41-44]. Thus, its function remains controversial. The description of link between adipocytes and β cells is illustrated in Figure 4.

To understand the pathological process of T2DM in context of obesity, we have to keep in mind that (1) adipocytes are not the only source of cytokine production. For instance, immune cells as well as β cells are reservoirs in cytokine production; (2) the effects of adipokines and cytokines could be different from tissue to tissue, organ to organ and low concentration to high concentration; and (3) The molecular basis of T1DM and T2DM is in large difference. Cytokines-promoted

NF- κ B plays a major role in β cell apoptosis in T1DM. However, FFAs-induced ER stress and β cell apoptosis does not require NF- κ B activation.

Obesity and the fate of hepatocytes: Glucose homeostasis is tightly controlled by three factors: (1) insulin secreted from pancreatic β cells; (2) glucose uptake by peripheral tissue and liver; and (3) regulation of hepatic glucose output. Therefore, hepatocytes are key players in glucose homeostasis. Before we discussed the adverse effect of obesity on peripheral tissue such as liver, we briefly illustrated the general action of insulin on cells. Following the binding of insulin to insulin receptor that is consisted of 2 α subunits and 2 β subunits, the complex phosphorylates insulin receptor substrates proteins (IRS proteins) at tyrosine residues. The effectors downstream of IRS proteins are phosphoinositide 3-kinase (PI 3-kinase), p42/p44 mitogen-associated protein kinase (MAPK), protein kinase C and a complex of proteins including cbl, caveolin and flotillin. These effectors facilitate gene transcription that regulates glycogen synthesis, glucose uptake, lipolysis, protein synthesis, cell proliferation, differentiation and survival. Moreover, as a very important function in response to insulin, glucose transporter 4 (Glut4) are transferred from intercellular compartment to cell membrane for glucose uptake. Of importance is the enhanced Glut4 mobilization to cell membrane in peripheral cells for its role in glucose uptake [45].

When obesity occurs, increased influx of FFAs to hepatocytes is oxidized for energy assumption and stored as triglyceride in liver. The fatty liver produces huge amount of very low density lipoprotein (VLDL), glucose, C-reactive protein (CRP), PAI-1, fibrinogen and coagulation factors, which are related to T2DM-associated cardiovascular

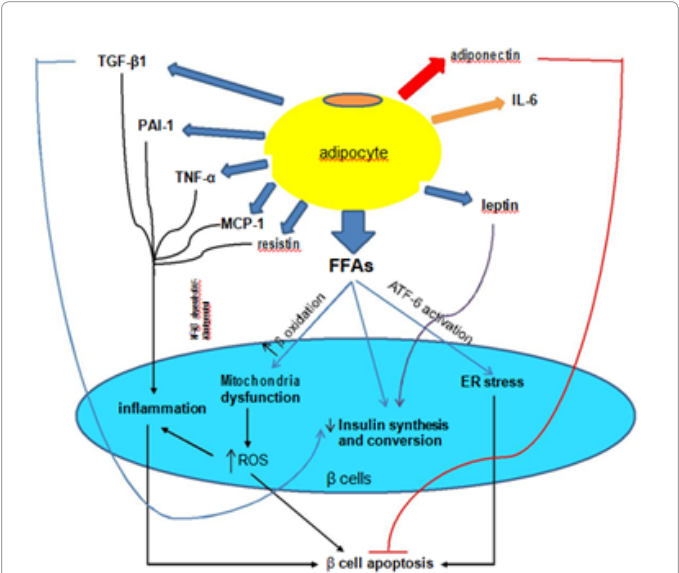


Figure 4: Link of adipocytes and pancreatic β cells. FFAs released by adipocytes suppress insulin synthesis and conversion of proinsulin to insulin, induce mitochondria dysfunction with increased ROS production and promote ER stress. Adipocytes produce proinflammatory cytokines including PAI-1, TNF- α , and MCP-1 and inflammatory adipokine resistin. TGF β 1 not only inhibits insulin transcription but also participates with proinflammatory cytokines and resistin in inflammation. The cytokines-mediated inflammation could be both NF- κ B dependent and independent. Leptin reduces insulin synthesis. Different from other adipokines and cytokines, adiponectin protects β cell death and stimulates insulin release. The role of IL-6 seems to have double sword effects that wait for further investigation.

complications. Moreover, FFAs repress Glut4 transcription and gene expression, resulting in reduced glucose uptake [46,47]. TGF- β 1 released from adipocytes triggers epithelial mesenchymal transition (EMT) and apoptosis, resulting in reduced glucose uptake and development of liver fibrosis and cancer [48,49]. Proinflammatory cytokines from adipocytes further accelerate hepatocyte apoptosis. Resistin interferes with insulin signaling on hepatocytes, resulting in reduced glycogen synthesis. Similar as FFAs, resistin stimulates hepatocytes to produce more apoB, leading to increased production of VLDL and LDL, accumulation of lipid content in hepatocytes and formation of steatosis. In addition, it could modulate hepatocyte stellate cell proliferation, migration and collagen I expression, which may contribute to liver fibronosis [50]. Leptin, at physically level, binds to its receptors on hepatocytes and induces insulin-like signaling pathways such as stimulating glycogen synthesis [51,52]. Adiponectin maintains its beneficial effect hepatocytes by inhibition of SREBP1 expression and lipogenesis, prohibition of hepatic gluconeogenic enzyme expression and thus reduction of endogenous glucose production and protection of hepatocytes from FFAs-induced apoptosis [53-56]. The effects of adipocytes on hepatocytes are demonstrated in Figure 5.

Obesity and other peripheral tissues: Similar as hepatocytes, impaired glucose uptake is seen in peripheral tissues such as skeletal muscles due to reduced glucose transporter GLUT4. The increased influx of FFAs to these cells results in fatty tissues due to triglyceride accumulation, ROS production and inflammation.

Animal Models Related to Obesity and T2DM

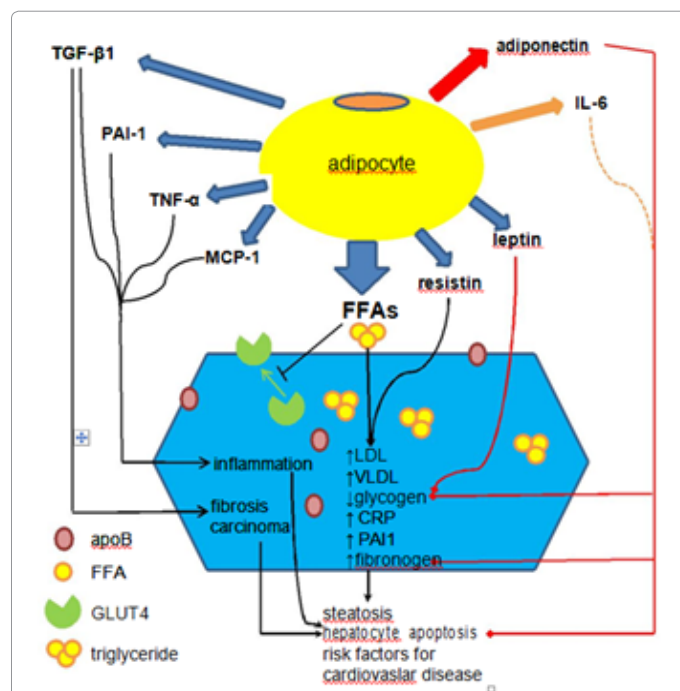


Figure 5: The impact of obesity on hepatocytes. This figure represents the adverse effects of adipocytes on hepatocytes. Influx of FFAs to hepatocytes result in reduced glucose uptake via impaired GLUT4 translocation, reduced glycogen synthesis and increased production of apoB-containing lipoproteins and accumulation of triglyceride. FFAs also stimulate inflammatory proteins and coagulation factors that increase the risk of vascular diseases. TGF- β 1 not only participates in fibrosis, but also potentiates other cytokines in inflammation. Resistin facilitates FFAs in lipoprotein and glucose metabolism whereas Leptin, adiponectin and IL6 exert protective effects on hepatocytes.

So far, we have delineated the pathogenesis of obesity on T2DM. In this section, we will summarize the animal models developed to study obesity and T2DM in Table 4. As you may notice from the table, the gap exists between animal models, obesity and T2DM patients. More specifically, partially due to short life span, animal models could not represent the diabetes-related complications, the evolution of β cell dysfunction and failure and the strong adverse impact of obesity on β cell. These also reflect the complexity of obesity and T2DM in human subjects.

Therapeutic Interventions of Obesity and T2DM

In the last section, we will discuss the interventions that have been carried out in the treatment of obesity and T2DM. Up to date, the main treatments include control of diet, lifestyle improvement, exercise, anti-diabetic medicines and insulin injection. On top of that, recently, gastrointestinal bypass has been applied more and more widely to obese subjects. Hereby, we focus on medications and surgeries interventions.

Anti-obesity medications

Sibutramine: Sibutramine inhibits serotonin-noradrenaline reuptake, promotes the feeling of satiety and decreases caloric uptake.

Orlistat: Orlistat is a potent gastric and pancreatic lipase inhibitor and reduces body weight. Nevertheless, its gastrointestinal side effects such as steatorrhea have been reported.

Rimonabant: It is by far, the first endocannabinoid CB1 receptor antagonist. Except weight loss, it decreases triglyceride level but increases HDL-cholesterol [57]. It was forced to be withdrawn in the United States for its severe side effect, i.e., the psychiatric problems.

Metformin and thiazolidinediones: AMP-activated protein lipase (AMPK) is a key regulator in glucose and energy homeostasis. It is ubiquitously expressed in tissues and organs. Activation of AMPK inhibits lipid and glucose metabolism via its regulation of enzymes involved in the metabolism. In details, (1) it decreases fatty acid and cholesterol synthesis by inhibition of ACC1 and HMG-CoA; [58,59] (2) it inhibits ACC2 to stimulate fatty acid oxidation; [58] (3) it inhibits lipolysis in adipocytes; [60] (4) it up-regulates GLUT4 expression and thus enhances its translocation to cell membrane for glucose uptake; [61] and (5) it prohibits hepatic gluconeogenesis by suppressing glucose-6-phosphatase (G6Pase) [62]. Both Metformin and thiazolidinediones have been shown to activate AMPK pathway, resulting in improved glucose homeostasis. Metformin also reduces body weight possibly via its regulation of ghrelin secretion, which acts as a hunger-stimulating peptide [63]. By contrast, thiazolidinediones increases body weight.

Glycogen-like peptide 1 (GLP-1) and GLP-1 agonist: GLP-1 is secreted from intestinal cells in response to nutrient uptake. It augments glucose-stimulated insulin secretion and reduces body weight by promoting satiety and decreasing caloric uptake [64]. Exenatide and liraglutide belong to GLP-1 agonist and share common features of GLP-1 in stimulation of insulin secretion after meal, suppression of glycogen release during meals, reduction of appetite and reduces fat content in hepatocytes [65,66]. Except its positive action on insulin secretion, increased β cell regeneration was also observed in rodents treated with GLP-1 mimetic peptide. Recent study of incretin therapy on diabetes patients has reminded us to stay cautious in incretin application. In this study, they did not detect human β cell replication when cultured with GLP-1. By contrast, because of GLP-1 receptor expression on endocrine and exocrine cells, the deleterious effect on the exocrine cells

Models	Animal species	Pathogenesis basis	phenotypes	Possible drawbacks
Streptozotocin in combination of fat diet [89,90]	Mouse, rat	Induced β cell damage	Hyperglycemia; obesity; reduced insulin secretion	Severe β cell damage and high mortality
Ob/ob (always in combination with high fat diet) [91]	Mouse	Leptin deficiency	obesity; Hyperglycemia and hyperinsulinemia; Developed peripheral neuropathy	No insulin insufficiency
DB/DB [92]	Mouse	Leptin receptor deficiency	obesity; Hyperglycemia and hyperinsulinemia; Developed peripheral neuropathy	β cell hyperplasia; No insulin insufficiency; reduced life span
Zucker (fa/fa) [93]	Rat	Leptin receptor deficiency	Body weight increase; Insulin resistance	No obvious hyperglycemia
KK-Ay (always in combination with high fat diet) [94]	Mouse	A glycoprotein gene mutation	Obesity; hyperglycemia; Insulin resistance; islet cell hyperplasia	
Goto Kakizaki (GK) [95,96]	Rat	Highest level of glucose challenge over many generations	resistance and impaired secretion; renal lesion, retina abnormality and nerve changes comparable to T2DM complications in human	relatively slim; no decreased β cell mass
NSY [97]	Mouse	Derived from NOD mice with spontaneously developed T2DM	Mild insulin resistance; impaired insulin secretion	No obesity; gender difference
OETF [98]	Rat	Selective for glucose tolerance	Obesity; Impaired glucose tolerance; Developed non-alcoholic fatty liver disease	Genetic variance that may have no causal relationship to diabetes itself
Irs1-/- Irs3-/- [99,100]	Mouse	Deficiency of insulin receptor substrate 1 and 3	Increased body weight; Hyperglycemia; hyperinsulinemia; Insulin resistance	No fatty liver

Table 4: Animal models of obesity and T2DM.

was noticed in diabetes subjected treated with incretin, as evidenced by increased exocrine cell proliferation and dysplasia. Thus, the safety and efficacy issues remain to be investigated when using incretin.

Repaglinide: Repaglinide stimulates insulin release via its inhibition of ATP-sensitive potassium channels. A mild weight change was observed in patients treated with Repaglinide.

Sitagliptin: Sitagliptin belongs to DPP4 inhibitor. By inhibiting DPP4 enzyme activity, it enhances GLP-1 expression. Despite it does not interfere with body weight, some adverse effects of DPP4 inhibitors have been observed in clinical studies.

Gastrointestinal bypass (GBP)

Mason and Ito proposed GBP treatment for severe obesity in 1967 [67]. The surgery techniques have been optimized and improved by clinicians through the years. In 2009, American Diabetes Association (ADA) includes GBP as one treatment for T2DM patients with obesity. In 2011, International Diabetes Foundation (IDF) indicated that GMP could be applied in early stage in T2DM patients whose BMI exceeds 35 kg/m² [68].

GBP surgery reduces stomach volume but increases the tension of stomach, therefore, food empties slowly. Due to slow digestion, satiety increases that leads to decreased energy intake. Thus it is not surprising to see the reduction of BMI and obesity after GBP surgery. Except that, although the mechanisms are not known, GBP surgeries have been shown to increase adiponectin [69] and PDX-1 [70] expression that improve β cell function. Clinical findings from different groups have demonstrated that GMP improved glucose metabolism and insulin resistance and lowered glucose level both in fasting state and OGTT test [71-73]. For instance, Buchwald et al performed a meta-analysis from 621 studies including 135246 T2DM patients. This study showed that GBP surgery resulted in 78.1% patients with complete remission, 86.6% patients with improved conditions. Two years after surgery, 62% patients were able to maintain diabetes remission [74]. In a retrospective cohort study from 9949 patients, after following-up of 7.1 years, long-term mortality in patients after GBP surgery was significantly reduced compared with control group [75]. However, the data of long-term effect of GBP is still very limited and therefore, it is dispensable to obtain the long-term data to assess the effect of GBP on obesity and T2DM.

Summary

The increasing incidence of obesity and diabetes has drawn more and more attention not only in disease treatment but also in disease prevention. Understanding the adverse effects of obesity on diabetes and the severity of obesity and diabetes will increase our awareness in improving diet control, lifestyle, exercise and medications against obesity and diabetes. To be addressed, there are many unsolved issues from basic to clinical research. For instance, better animal models are required to represent diabetes development and complications in humans. It still needs to define whether combinations of several medications will achieve better therapeutic effect and lower side effect than solo medication. Despite targeting obesity has shown successful and beneficial effect in diabetes patients, its long-term effect on diabetes waits for further investigations.

References

- Herder C, Roden M (2011) Genetics of type 2 diabetes: pathophysiologic and clinical relevance. Eur J Clin Invest 41: 679-692.
- Meier JJ, Butler AE, Saisho Y, Monchamp T, Galasso R, et al. (2008) Beta-cell replication is the primary mechanism subserving the postnatal expansion of beta-cell mass in humans. Diabetes 57: 1584-1594.
- Chung CH, Hao E, Piran R, Keinan E, Levine F (2010) Pancreatic β -cell neogenesis by direct conversion from mature α -cells. Stem Cells 28: 1630-1638.
- Dor Y, Brown J, Martinez OI, Melton DA (2004) Adult pancreatic beta-cells are formed by self-duplication rather than stem-cell differentiation. Nature 429: 41-46.
- Georgia S, Bhushan A (2004) Beta cell replication is the primary mechanism for maintaining postnatal beta cell mass. J Clin Invest 114: 963-968.
- Teta M, Rankin MM, Long SY, Stein GM, Kushner JA (2007) Growth and regeneration of adult beta cells does not involve specialized progenitors. Dev Cell 12: 817-826.
- Xiao X, Chen Z, Shiota C, Prasad K, Guo P, et al. (2013) No evidence for β cell neogenesis in murine adult pancreas. J Clin Invest 123: 2207-2217.
- Daga N, Nasir K, Hamirani Y, Tayek J, Bach P, et al. (2013) Prevalence and severity of coronary artery calcium in young persons with diabetes. J Cardiovasc Comput Tomogr 7: 241-247.
- Morrish NJ, Wang SL, Stevens LK, Fuller JH and Keen H (2001) Mortality and causes of death in the WHO Multinational Study of Vascular Disease in Diabetes. Diabetologia 44: S14-21.
- Ding J, Wong TY (2012) Current epidemiology of diabetic retinopathy and diabetic macular edema. Curr Diab Rep 12: 346-354.

11. Rosberger DF (2013) Diabetic retinopathy: current concepts and emerging therapy. *Endocrinol Metab Clin North Am* 42: 721-745.
12. Reutens AT and RC Atkins (2011) Epidemiology of diabetic nephropathy. *Contrib Nephrol* 170: 1-7.
13. O'Brien PD, Hinder LM, Sakowski SA, Feldman EL (2014) ER stress in diabetic peripheral neuropathy: A new therapeutic target. *Antioxid Redox Signal*.
14. The Global Burden of Metabolic Risk Factors for Chronic Diseases Collaboration (BMI Mediated Effects) (2013) Metabolic mediators of the effects of body-mass index, overweight, and obesity on coronary heart disease and stroke: a pooled analysis of 97 prospective cohorts with 1.8 million participants. *Lancet*.
15. Hubert HB, Feinleib M, McNamara PM, Castelli WP (1983) Obesity as an independent risk factor for cardiovascular disease: a 26-year follow-up of participants in the Framingham Heart Study. *Circulation* 67: 968-977.
16. Shimamoto Y, Mizukoshi M, Kuroi A, Imanishi T, Takeshita T, et al. (2013) Is visceral fat really a coronary risk factor? A multi-detector computed tomography study. *Int Heart J* 54: 273-278.
17. Chan JM, Rimm EB, Colditz GA, Stampfer MJ, Willett WC (1994) Obesity, fat distribution, and weight gain as risk factors for clinical diabetes in men. *Diabetes Care* 17: 961-969.
18. Colditz GA, Willett WC, Rotnitzky A, Manson JE (1995) Weight gain as a risk factor for clinical diabetes mellitus in women. *Ann Intern Med* 122: 481-486.
19. Mokdad AH, Ford ES, Bowman BA, Dietz WH, Vinicor F, et al. (2003) Prevalence of obesity, diabetes, and obesity-related health risk factors, 2001. *JAMA* 289: 76-79.
20. Yang WS, Shu XO, Gao J, Li HL, Cai H, et al. (2013) Prospective evaluation of type 2 diabetes mellitus on the risk of primary liver cancer in Chinese men and women. *Ann Oncol* 24: 1679-1685.
21. Kotchen TA (2010) Obesity-related hypertension: epidemiology, pathophysiology, and clinical management. *Am J Hypertens* 23: 1170-1178.
22. Gremlich S, Bonny C, Waeber G, Thorens B (1997) Fatty acids decrease IDX-1 expression in rat pancreatic islets and reduce GLUT2, glucokinase, insulin, and somatostatin levels. *J Biol Chem* 272: 30261-30269.
23. Björklund A, Grill V (1999) Enhancing effects of long-term elevated glucose and palmitate on stored and secreted proinsulin-to-insulin ratios in human pancreatic islets. *Diabetes* 48: 1409-1414.
24. Furukawa H, Carroll RJ, Swift HH, Steiner DF (1999) Long-term elevation of free fatty acids leads to delayed processing of proinsulin and prohormone convertases 2 and 3 in the pancreatic beta-cell line MIN6. *Diabetes* 48: 1395-1401.
25. Inoguchi T, Li P, Umeda F, Yu HY, Kakimoto M, et al. (2000) High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C-dependent activation of NAD(P)H oxidase in cultured vascular cells. *Diabetes* 49: 1939-1945.
26. Kharroubi I, Ladrière L, Cardozo AK, Dogusan Z, Cnop M, et al. (2004) Free fatty acids and cytokines induce pancreatic beta-cell apoptosis by different mechanisms: role of nuclear factor-kappaB and endoplasmic reticulum stress. *Endocrinology* 145: 5087-5096.
27. Kawashima J, Tsuruzoe K, Motoshima H, Shirakami A, Sakai K, et al. (2003) Insulin down-regulates resistin mRNA through the synthesis of protein(s) that could accelerate the degradation of resistin mRNA in 3T3-L1 adipocytes. *Diabetologia* 46: 231-240.
28. Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, et al. (2001) The hormone resistin links obesity to diabetes. *Nature* 409: 307-312.
29. Liu F, Yang T, Wang B, Zhang M, Gu N, et al. (2008) Resistin induces insulin resistance, but does not affect glucose output in rat-derived hepatocytes. *Acta Pharmacol Sin* 29: 98-104.
30. Milan G, Granzotto M, Scarda A, Calcagno A, Pagano C, et al. (2002) Resistin and adiponectin expression in visceral fat of obese rats: effect of weight loss. *Obes Res* 10: 1095-1103.
31. Silswal N, Singh AK, Aruna B, Mukhopadhyay S, Ghosh S, et al. (2005) Human resistin stimulates the pro-inflammatory cytokines TNF-alpha and IL-12 in macrophages by NF-kappaB-dependent pathway. *Biochem Biophys Res Commun* 334: 1092-1101.
32. Verma S, Li SH, Wang CH, Fedak PW, Li RK, et al. (2003) Resistin promotes endothelial cell activation: further evidence of adipokine-endothelial interaction. *Circulation* 108: 736-740.
33. Lin HM, Lee JH, Yadav H, Kamaraju AK, Liu E, et al. (2009) Transforming growth factor-beta/Smad3 signaling regulates insulin gene transcription and pancreatic islet beta-cell function. *J Biol Chem* 284: 12246-12257.
34. Scaglia L, Smith FE, Bonner-Weir S (1995) Apoptosis contributes to the involution of beta cell mass in the post partum rat pancreas. *Endocrinology* 136: 5461-5468.
35. Donath MY, Ehse JA, Maedler K, Schumann DM, Ellingsgaard H, et al. (2005) Mechanisms of beta-cell death in type 2 diabetes. *Diabetes* 54 Suppl 2: S108-113.
36. Fain JN (2006) Release of interleukins and other inflammatory cytokines by human adipose tissue is enhanced in obesity and primarily due to the nonfat cells. *Vitam Horm* 74: 443-477.
37. Wisse BE (2004) The inflammatory syndrome: the role of adipose tissue cytokines in metabolic disorders linked to obesity. *J Am Soc Nephrol* 15: 2792-2800.
38. Gu W, Li X, Liu C, Yang J, Ye L, et al. (2006) Globular adiponectin augments insulin secretion from pancreatic islet beta cells at high glucose concentrations. *Endocrine* 30: 217-221.
39. Wijesekara N, Krishnamurthy M, Bhattacharjee A, Suhail A, Sweeney G, et al. (2010) Adiponectin-induced ERK and Akt phosphorylation protects against pancreatic beta cell apoptosis and increases insulin gene expression and secretion. *J Biol Chem* 285: 33623-33631.
40. Weiss R, Dufour S, Groszmann A, Petersen K, Dziura J, et al. (2003) Low adiponectin levels in adolescent obesity: a marker of increased intramyocellular lipid accumulation. *J Clin Endocrinol Metab* 88: 2014-2018.
41. Kristiansen OP, Mandrup-Poulsen T (2005) Interleukin-6 and diabetes: the good, the bad, or the indifferent? *Diabetes* 54 Suppl 2: S114-124.
42. Ray P, Ghosh SK, Zhang DH, Ray A (1997) Repression of interleukin-6 gene expression by 17 beta-estradiol: inhibition of the DNA-binding activity of the transcription factors NF-IL6 and NF-kappa B by the estrogen receptor. *FEBS Lett* 409: 79-85.
43. Russell MA, Cooper AC, Dhayal S, Morgan NG (2013) Differential effects of interleukin-13 and interleukin-6 on Jak/STAT signaling and cell viability in pancreatic β -cells. *Islets* 5: 95-105.
44. Suzuki T, Imai J, Yamada T, Ishigaki Y, Kaneko K, et al. (2011) Interleukin-6 enhances glucose-stimulated insulin secretion from pancreatic beta-cells: potential involvement of the PLC-IP3-dependent pathway. *Diabetes* 60: 537-547.
45. Virkamäki A, Ueki K, Kahn CR (1999) Protein-protein interaction in insulin signaling and the molecular mechanisms of insulin resistance. *J Clin Invest* 103: 931-943.
46. Armoni M, Harel C, Bar-Yoseph F, Milo S, Karnieli E (2005) Free fatty acids repress the GLUT4 gene expression in cardiac muscle via novel response elements. *J Biol Chem* 280: 34786-34795.
47. Griesel BA, Weems J, Russell RA, Abel ED, Humphries K, et al. (2010) Acute inhibition of fatty acid import inhibits GLUT4 transcription in adipose tissue, but not skeletal or cardiac muscle tissue, partly through liver X receptor (LXR) signaling. *Diabetes* 59: 800-807.
48. Meyer C, Liu Y, Dooley S (2013) Caveolin and TGF- β entanglements. *J Cell Physiol* 228: 2097-2102.
49. Meyer C, Liu Y, Kaul A, Peipe I, Dooley S (2013) Caveolin-1 abrogates TGF- β mediated hepatocyte apoptosis. *Cell Death Dis* 4: e466.
50. Dong ZX, Su L, Brymora J, Bird C, Xie Q, et al. (2013) Resistin mediates the hepatic stellate cell phenotype. *World J Gastroenterol* 19: 4475-4485.
51. Aiston S, Agius L (1999) Leptin enhances glycogen storage in hepatocytes by inhibition of phosphorylase and exerts an additive effect with insulin. *Diabetes* 48: 15-20.
52. Zhao AZ, Shinohara MM, Huang D, Shimizu M, Eldar-Finkelman H, et al. (2000) Leptin induces insulin-like signaling that antagonizes cAMP elevation by glucagon in hepatocytes. *J Biol Chem* 275: 11348-11354.
53. Awazawa M, Ueki K, Inabe K, Yamauchi T, Kaneko K, et al. (2009) Adiponectin suppresses hepatic SREBP1c expression in an AdipoR1/LKB1/AMPK dependent pathway. *Biochem Biophys Res Commun* 382: 51-56.

54. Combs TP, Berg AH, Obici S, Scherer PE, Rossetti L (2001) Endogenous glucose production is inhibited by the adipose-derived protein Acrp30. *J Clin Invest* 108: 1875-1881.
55. Jung TW, Lee YJ, Lee MW, Kim SM, Jung TW (2009) Full-length adiponectin protects hepatocytes from palmitate-induced apoptosis via inhibition of c-Jun NH2 terminal kinase. *FEBS J* 276: 2278-2284.
56. Miller RA, Chu Q, Le Lay J, Scherer PE, Ahima RS, et al. (2011) Adiponectin suppresses gluconeogenic gene expression in mouse hepatocytes independent of LKB1-AMPK signaling. *J Clin Invest* 121: 2518-2528.
57. Fong TM, Heymsfield SB (2009) Cannabinoid-1 receptor inverse agonists: current understanding of mechanism of action and unanswered questions. *Int J Obes (Lond)* 33: 947-955.
58. López M, Lelliott CJ, Vidal-Puig A (2007) Hypothalamic fatty acid metabolism: a housekeeping pathway that regulates food intake. *Bioessays* 29: 248-261.
59. Woods A, Azzout-Marniche D, Foretz M, Stein SC, Lemarchand P, et al. (2000) Characterization of the role of AMP-activated protein kinase in the regulation of glucose-activated gene expression using constitutively active and dominant negative forms of the kinase. *Mol Cell Biol* 20: 6704-6711.
60. Daval M, Diot-Dupuy F, Bazin R, Hainault I, Viollet B, et al. (2005) Anti-lipolytic action of AMP-activated protein kinase in rodent adipocytes. *J Biol Chem* 280: 25250-25257.
61. Holmes BF, Kurth-Kraczek EJ, Winder WW (1999) Chronic activation of 5'-AMP-activated protein kinase increases GLUT-4, hexokinase, and glycogen in muscle. *J Appl Physiol* (1985) 87: 1990-1995.
62. Cool B, Zinker B, Chiou W, Kifle L, Cao N, et al. (2006) Identification and characterization of a small molecule AMPK activator that treats key components of type 2 diabetes and the metabolic syndrome. *Cell Metab* 3: 403-416.
63. Lee A, Morley JE (1998) Metformin decreases food consumption and induces weight loss in subjects with obesity with type II non-insulin-dependent diabetes. *Obes Res* 6: 47-53.
64. Barrera JG, Sandoval DA, D'Alessio DA, Seeley RJ (2011) GLP-1 and energy balance: an integrated model of short-term and long-term control. *Nat Rev Endocrinol* 7: 507-516.
65. Bunck MC, Diamant M, Cornér A, Eliasson B, Malloy JL, et al. (2009) One-year treatment with exenatide improves beta-cell function, compared with insulin glargine, in metformin-treated type 2 diabetic patients: a randomized, controlled trial. *Diabetes Care* 32: 762-768.
66. Ding X, Saxena NK, Lin S, Gupta NA, Anania FA (2006) Exendin-4, a glucagon-like protein-1 (GLP-1) receptor agonist, reverses hepatic steatosis in ob/ob mice. *Hepatology* 43: 173-181.
67. Mason EE, Ito C (1967) Gastric bypass in obesity. *Surg Clin North Am* 47: 1345-1351.
68. Dixon JB, Zimmet P, Alberti KG, Rubino F; International Diabetes Federation Taskforce on Epidemiology and Prevention (2011) Bariatric surgery: an IDF statement for obese Type 2 diabetes. *Arq Bras Endocrinol Metabol* 55: 367-382.
69. Shrestha C, He H, Liu Y, Zhu S, Xiong J, et al. (2013) Changes in Adipokines following Laparoscopic Roux-en-Y Gastric Bypass Surgery in Chinese Individuals with Type 2 Diabetes Mellitus and BMI of 22-30 kg·m⁻². *Int J Endocrinol* 2013: 240971.
70. Li Z, Zhang HY, Lv LX, Li DF, Dai JX, et al. (2010) Roux-en-Y gastric bypass promotes expression of PDX-1 and regeneration of beta-cells in Goto-Kakizaki rats. *World J Gastroenterol* 16: 2244-2251.
71. Ballantyne GH, Wasielewski A, Saunders JK (2009) The surgical Treatment of Type 2 Diabetes Mellitus: changes in HOMA Insulin resistance in the first year following laparoscopic Roux-en-Y gastric bypass (LRYGB) and laparoscopic adjustable gastric banding (LAGB). *Obes Surg* 19: 1297-1303.
72. Kashyap SR, Daud S, Kelly KR, Gastaldelli A, Win H, et al. (2010) Acute effects of gastric bypass versus gastric restrictive surgery on beta-cell function and insulinotropic hormones in severely obese patients with type 2 diabetes. *Int J Obes (Lond)* 34: 462-471.
73. Shah SS, Todkar JS, Shah PS, Cummings DE (2010) Diabetes remission and reduced cardiovascular risk after gastric bypass in Asian Indians with body mass index <35 kg/m². *Surg Obes Relat Dis* 6: 332-338.
74. Buchwald H, Estok R, Fährbach K, Banel D, Jensen MD, et al. (2009) Weight and type 2 diabetes after bariatric surgery: systematic review and meta-analysis. *Am J Med* 122: 248-256.
75. Adams TD, Gress RE, Smith SC, Halverson RC, Simper SC, et al. (2007) Long-term mortality after gastric bypass surgery. *N Engl J Med* 357: 753-761.
76. Xi B, Liang Y, He T, Reilly KH, Hu Y, et al. (2012) Secular trends in the prevalence of general and abdominal obesity among Chinese adults, 1993-2009. *Obes Rev* 13: 287-296.
77. Yang SH, Dou KF, Song WJ (2010) Prevalence of diabetes among men and women in China. *N Engl J Med* 362: 2425-2426.
78. Dunstan DW, Zimmet PZ, Welborn TA, De Courten MP, Cameron AJ, et al. (2002) The rising prevalence of diabetes and impaired glucose tolerance: the Australian Diabetes, Obesity and Lifestyle Study. *Diabetes Care* 25: 829-834.
79. Zhang PH, Chen ZW, Lv D, Xu YY, Gu WL, et al. (2012) Increased risk of cancer in patients with type 2 diabetes mellitus: a retrospective cohort study in China. *BMC Public Health* 12: 567.
80. He S, Tang YH, Zhao G, Yang X, Wang D, et al. (2013) Pioglitazone prescription increases risk of bladder cancer in patients with type 2 diabetes: an updated meta-analysis. *Tumour Biol*.
81. Yang XQ, Xu C, Sun Y, Han RF (2013) Diabetes mellitus increases the risk of bladder cancer: an updated meta-analysis. *Asian Pac J Cancer Prev* 14: 2583-2589.
82. Adami HO, Chow WH, Nyrén O, Berne C, Linet MS, et al. (1996) Excess risk of primary liver cancer in patients with diabetes mellitus. *J Natl Cancer Inst* 88: 1472-1477.
83. Chari ST, Leibson CL, Rabe KG, Ransom J, de Andrade M, et al. (2005) Probability of pancreatic cancer following diabetes: a population-based study. *Gastroenterology* 129: 504-511.
84. Redaniel MT, Jeffreys M, May MT, Ben-Shlomo Y, Martin RM (2012) Associations of type 2 diabetes and diabetes treatment with breast cancer risk and mortality: a population-based cohort study among British women. *Cancer Causes Control* 23: 1785-1795.
85. Park J, Cho SY, Lee SB, Son H, H Jeong (2013) Obesity is associated with higher risk of prostate cancer detection in a Korean biopsy population. *BJU Int*.
86. Larsson SC, Wolk A (2007) Overweight, obesity and risk of liver cancer: a meta-analysis of cohort studies. *Br J Cancer* 97: 1005-1008.
87. Olsen CM, Green AC, Whiteman DC, Sadeghi S, Kolahdooz F, et al. (2007) Obesity and the risk of epithelial ovarian cancer: a systematic review and meta-analysis. *Eur J Cancer* 43: 690-709.
88. Key TJ, Appleby PN, Reeves GK, Roddam A, Dorgan JF, et al. (2003) Body mass index, serum sex hormones, and breast cancer risk in postmenopausal women. *J Natl Cancer Inst* 95: 1218-1226.
89. Okamoto T, Kanemoto N, Ohbuchi Y, Okano M, Fukui H, et al. (2008) Characterization of STZ-Induced Type 2 Diabetes in Zucker Fatty Rats. *Exp Anim* 57: 335-345.
90. Zhang F, Ye C, Li G, Ding W, Zhou W, et al. (2003) The rat model of type 2 diabetic mellitus and its glycometabolism characters. *Exp Anim* 52: 401-407.
91. Drel VR, Mashtalir N, Illytska O, Shin J, Li F, et al. (2006) The leptin-deficient (ob/ob) mouse: a new animal model of peripheral neuropathy of type 2 diabetes and obesity. *Diabetes* 55: 3335-3343.
92. Shi TJ, Zhang MD, Zeberg H, Nilsson J, Grünler J, et al. (2013) Coenzyme Q10 prevents peripheral neuropathy and attenuates neuron loss in the db/db- mouse, a type 2 diabetes model. *Proc Natl Acad Sci U S A* 110: 690-695.
93. Tokuyama Y, Sturis J, DePaoli AM, Takeda J, Stoffel M, et al. (1995) Evolution of beta-cell dysfunction in the male Zucker diabetic fatty rat. *Diabetes* 44: 1447-1457.
94. Nakamura M, Yamada K (1967) Studies on a diabetic (KK) strain of the mouse. *Diabetologia* 3: 212-221.
95. Goto Y, Kakizaki M, Masaki N (1976) Production of spontaneous diabetic rats by repetition of selective breeding. *Tohoku J Exp Med* 119: 85-90.
96. Portha B, Lacraz G, Kergoat M, Homo-Delarche F, Giroix MH, et al. (2009) The GK rat beta-cell: a prototype for the diseased human beta-cell in type 2 diabetes? *Mol Cell Endocrinol* 297: 73-85.

97. Ueda H, Ikegami H, Kawaguchi Y, Fujisawa T, Yamato E, et al. (1999) Genetic analysis of late-onset type 2 diabetes in a mouse model of human complex trait. *Diabetes* 48: 1168-1174.
98. Rector RS, Uptergrove GM, Morris EM, Borengasser SJ, Laughlin MH, et al. (2011) Daily exercise vs. caloric restriction for prevention of nonalcoholic fatty liver disease in the OLETF rat model. *Am J Physiol Gastrointest Liver Physiol* 300: G874-883.
99. Laustsen PG, Michael MD, Crute BE, Cohen SE, Ueki K, et al. (2002) Lipoatrophic diabetes in *Irs1(-)/Irs3(-)* double knockout mice. *Genes Dev* 16: 3213-3222.
100. Terauchi, Iwamoto K, Tamemoto H, Komeda K, Ishii C, et al. (1997) Development of non-insulin-dependent diabetes mellitus in the double knockout mice with disruption of insulin receptor substrate-1 and beta cell glucokinase genes. Genetic reconstitution of diabetes as a polygenic disease. *J Clin Invest* 99: 861-866.