

## The Fractal-like Complexity of Heart Rate Variability beyond Neurotransmitters and Autonomic Receptors: Signaling Intrinsic to Sinoatrial Node Pacemaker Cells

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### Abstract

The heart rate and rhythm are controlled by complex chaotic neural, chemical and hormonal networks which are not strictly regular, but exhibit fluctuations across multiple time scales. A careful assessment of the Heart Rate Variability (HRV) offers clues to this complexity. A reduction in HRV, specifically in advanced age, is associated with increase in morbidity and mortality. Mechanisms that induce this decrease, however, have not been fully elucidated. The classical literature characterizes changes in HRV as a result of changes in the balance of competing influences of the sympathetic and parasympathetic autonomic impulses delivered to the heart. It has now become clear, however, that the heart rate and HRV are also determined by intrinsic properties of the pacemaker cells that comprise sinoatrial node, and that these properties respond to autonomic receptor stimulation in a non-linear mode. That HRV is determined by both the intrinsic properties of pacemaker cells in the sinoatrial node and the competing influences of the two branches of the autonomic neural input to the cells requires an expansion of our perspective about mechanisms that govern HRV in the normal heart, and how HRV changes with aging in health and in heart diseases.

**Keywords:** Cardiac denervation; Coupled-clock pacemaker system; Fractal-like behavior of the heart rate

### Introduction

The healthy heart rapidly changes its rate in response to a broad range of integrated input signals that arise from diverse stimuli. Changes in heart rate are governed by robust control mechanisms involving regulatory networks of neural, chemical and hormonal pathways interacting through coupled feedback mechanisms that operate over a wide range of time periods, i.e., ultradian (from milliseconds, minutes to hours) to circadian (24 hours) and infradian (beyond days) [1]. Because these complex networks are chaotic, the periods of rhythms they regulate are never in a steady state, but continuously shift from one period to another. This chaotic complexity is inherent to the rate and rhythm of the healthy heart, and is the reason why the heart rhythm is not strictly regular, but exhibits fluctuations. Careful assessment of the Heart Rate Variability (HRV) is required to probe its complexity. Specifically, decoding the electrogram (ECG), permits access to complex harmonics imbedded in the heart rhythm. Loss of this complexity is manifest as a reduction in HRV, and is related to increases in both morbidity and mortality [2].

Changes in HRV have been mainly interpreted as the balance between the competing influences of the two branches of the autonomic nervous system: the sympathetic tone that accelerates the heart rate, and the parasympathetic (vagal) stimulation that slows the heart rate (Figure 1A). Based on this logic, the balance of flux between the two arms of the autonomic system determines both the heart rate and HRV. An imbalance between these two afferent systems has been thought to be the major mechanism for reduction in HRV associated with aging and cardiac diseases. Studies near the turn of this century, including recent discoveries [3-8], however, call attention to the importance of intrinsic properties of pacemaker cells within the sinoatrial node, the heart's primary pacemaker, and their responsiveness to autonomic signaling, as an important determinant of heart rate and HRV. Novel perspectives have emerged regarding the intrinsic properties of SinoAtrial Node

Cells (SANC) that determine the normal automaticity of spontaneous Action Potentials (APs) that emanate from the sinoatrial node (Figure 1B), and how signaling through autonomic receptors links to these intrinsic signaling pathways to alter normal automaticity (Figure 1C).

### Decoding the Complexity of HRV

If we place our finger on the radial artery to determine our heart rate, we are struck by the apparent regulatory of intervals at which the heart beat occur. In reality, however, a plot of the healthy heart beat-to-beat intervals determined from the ECG demonstrates substantial variation, even at rest in the absence of external stimuli (e.g. change in the nervous system input) [9]. Time-series analysis is the simplest means of evaluating HRV and to identify variation in beat-to-beat intervals over time. Common methods utilized are: standard deviation (SDNN) to measure global variation, the square root of mean squared differences of consecutive intervals (RMSSD) to measure short-time variation, and standard deviation of 5-min averages (SDANN) to evaluate long-term variation. Identical means and standard deviations in the time-domain measures, however, can be generated by different

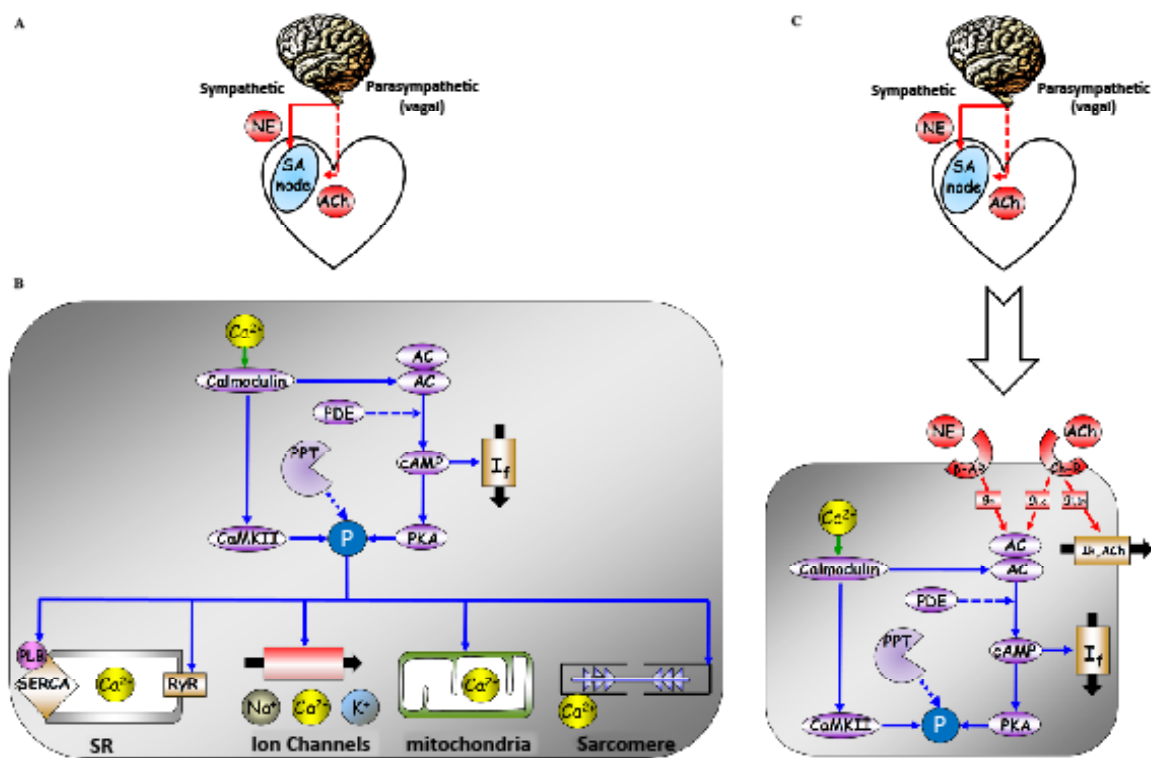
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**Figure 1:** The crosstalk between the autonomic nervous system and the sinoatrial node system. (A) In vivo, neural impulses via parasympathetic and sympathetic nerve and epinephrine (not shown) influence the heart rate and rhythm. (B) Intrinsic spontaneous AP firing by sinoatrial node cells, even in the absence of autonomic impulses, is driven by basal Ca<sup>2+</sup>-calmodulin activation of adenylyl cyclases (AC), which produce cAMP-PKA-dependent phosphorylation signaling. Ca<sup>2+</sup>- calmodulin activates, in parallel to AC, calmodulin-dependent kinase II (CaMKII) phosphorylation signaling. cAMP positively shifts the f-channel activation curve. Phosphodiesterases (PDE) degrade cAMP production, while protein phosphatase (PPT) degrade phosphorylation activity. PKA and CaMKII phosphorylate sarcoplasmic reticulum (SR) Ca<sup>2+</sup> cycling proteins (Ryanodine receptor (RyR), Phospholamban (PLB), which binds to and inhibit the sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase (SERCA)), surface membrane ion channel proteins, ATP production mechanisms in the mitochondria and sarcomere. (C) The neurotransmitters noradrenaline (NE) and acetylcholine (ACh) released from sympathetic or parasympathetic nerve terminals bind to  $\beta$ -adrenergic receptors ( $\beta$ -AR) or cholinergic receptors (Ch-R), respectively. Autonomic receptor signaling couples to G-proteins and leads to modulation of the same coupled-clock molecules that drive basal automaticity of SANC.

underlying rhythms in the time series of heart-rate intervals. Moreover, differences in the time series of heart-rate intervals lie not only in the distribution of the interbeat variations around the mean heart rate, but also in their time ordering: different dynamic patterns can occur at different time scales that correspond to the actual time scales of the underlying physiological process [10]. A nonlinear form and function has been introduced to examine these dynamic patterns. Fractal analysis of the heart beats is one of the method employed to decode the secrets hidden in HRV [11-14].

The idea of examining fractal-like behavior of a system to demonstrate its complexity was introduced by Mandelbort [15]. A system exhibits fractal-like temporal behavior if any time-based measurement segment within the system resembles the system behavior over the full time scale [15]. Fractal analyses of HRV have indicated that its control mechanisms are not linear, but typically exhibit chaotic behavior over a wide range of time scales that are slower than the heart rate per second. Further, application of fractal analysis to the physiology of HRV clearly demonstrates the presence of a high degree of chaos and entropy within the normal heart rhythm [2]. Moreover, the HRV components occur over a wide range of frequencies and exhibit inverse power-law scaling (1/f-like distribution) suggesting that fractal-like behavior regulates the HRV and therefore the heart rate [11]. Consequently, fractal-like behavior analyses can be used as a tool

to monitor heart pathophysiological changes in HRV that occur in a variety of settings.

Two quantitative methods have been employed to examine the fractal-like behavior of HRV. The first approach includes detection of the frequency harmonics embedded within the heart's rhythm by performing Fourier analysis on the inter heart beat intervals. For example, the frequency regimes embedded within the human heart rate include: 1) a high frequency band, between 0.15 and 0.4 Hz (between 2 to 6 heart cycles); 2) a low frequency band, between 0.04 and 0.15 Hz (between 6 to 25 heart cycles); and 3) a very low frequency band, below 0.04 Hz (more than 25 heart cycles) [16]. The slope of the line relating log frequency to log of the power spectrum density (derived from Fourier analysis of heart rate intervals) is the fractal scaling exponent,  $\beta$  (Figure 2A). The second method is the detrended fluctuation analysis (DFA), in which the degree of correlation among time scales embedded within the heart beat intervals is characterized [17]. In this method, the self-similarity of frequency regimes buried within the heart beat intervals assessed from ECG analysis is assumed to be bi-fractal, and is described by short- and long-term exponents,  $\alpha_1$  and  $\alpha_2$ , respectively (Figure 2B). Note that if  $\beta$  or  $\alpha$  is less than 0.5 the system dynamic is described as white noise (i.e., a random signal with a constant power spectral), and if  $\beta$  or  $\alpha$  equals 1.5, the system dynamic is described as

Brownian noise (i.e., an integral of white noise that has more energy at lower frequencies). That a reduced fractal slope can predict cardiac death [11] indicates that fractal correlation is a powerful tool to probe subtle alterations in heart rate dynamics and complexity.

Fractal scaling of HRV components is a universal organizing principle that unites diverse heart rates among hearts of diverse sizes in diverse members of the mammalian kingdom. When the log of power spectrum density of components that comprise the HRV for different mammals (dog, rat, hamster and human) is plotted against the log of frequency, a unique harmonically modulated inverse power-law dependence is observed [18,19]. Such log-log plots are represented by power spectra having a broad band of frequencies with a long low-amplitude, high-frequency tail and fractal scaling exponent,  $\beta$  approaching 1 (Figure 2, Table 1).

Fractal analyses have also been employed to decode changes in the heart's rhythm in response to pharmacological perturbations of  $\beta$  adrenergic receptor or cholinergic receptor signaling. Inhibition of the sympathetic or parasympathetic tone (vagal) increases or decreases the fractal-like behavior slope, respectively (Table 1). Therefore, parasympathetic receptor stimulation shifts the HRV fractal dynamics toward a white noise while the cardiac sympathetic receptor stimulation shifts the fractal dynamics toward a Brownian noise component. The conclusion that a shift in the ratio of sympathetic vs. parasympathetic tone toward sympathetic decreases HRV is supported by the observation that HRV is nearly suppressed in transgenic mice in which the  $\beta_1$ -adrenergic receptors are over expressed in the atria [20].

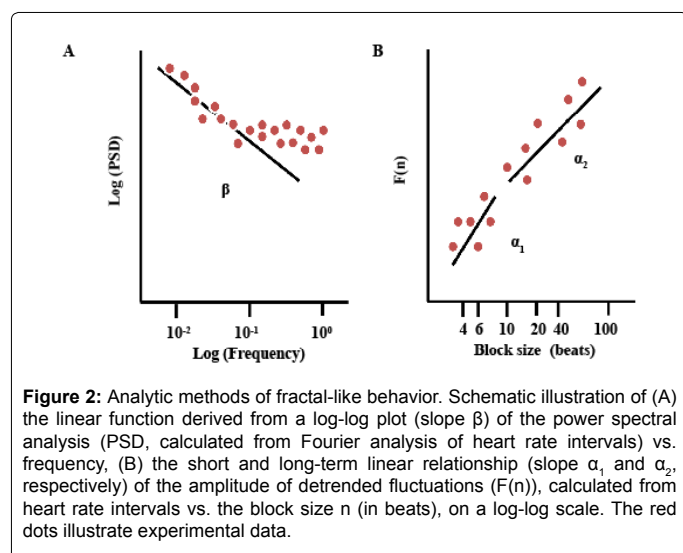
We contend that, fractal-like complexity of HRV goes beyond neuro-transmitters and autonomic receptors, and must include both the intrinsic properties within SANC and how these cell properties become organized within sinoatrial node tissue.

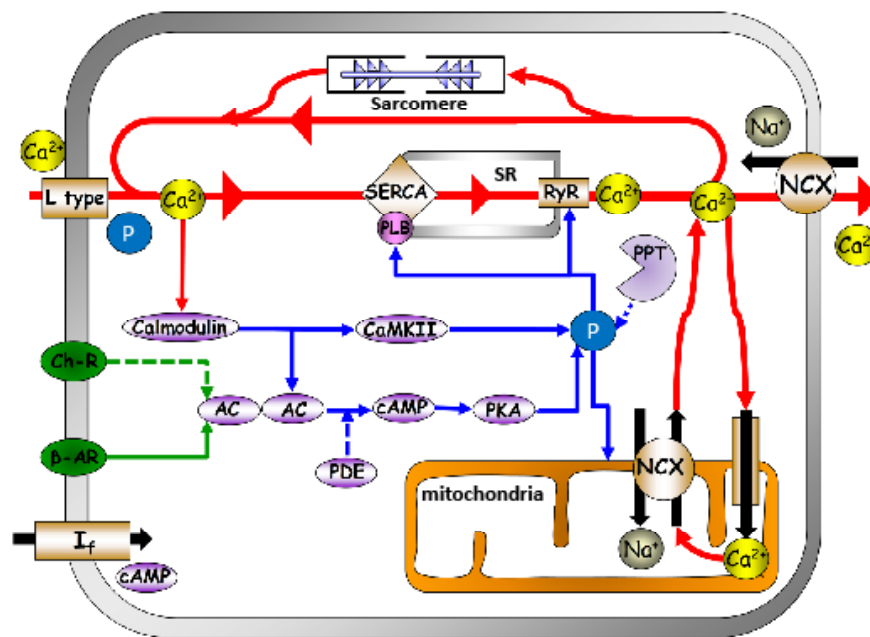
### Intrinsic Sinoatrial Node Cell Properties: A New Vista to Reveal the Mystery Embedded within the HRV

New experimental and numerical data support the hypothesis that, when isolated from the autonomic nervous system, the SANC AP firing rate is regulated by integrated functions within a system of two coupled clocks [21] (Figure 3): the sarcoplasmic reticulum is a  $\text{Ca}^{2+}$  clock that spontaneously generates roughly periodic local diastolic  $\text{Ca}^{2+}$  releases (LCRs) that activate an inward  $\text{Na}^+$ - $\text{Ca}^{2+}$  exchanger current, which

accelerates the rate of diastolic membrane depolarization.  $\text{Na}^+$ - $\text{Ca}^{2+}$  exchange current and the f-channel current, another member of the ensemble of sarcolemmal electrogenic molecules ("membrane clock"), concurrently drive the diastolic membrane depolarization to ignite the next AP. The two clocks are tightly coupled by common chemical axes:  $\text{Ca}^{2+}$ -calmodulin activated adenylyl cyclases (neuronal types 1 and 8) generate the second messenger cAMP, which in turn activates Protein Kinase A (PKA), and  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase II (CaMKII) (Figure 1B). Both kinases phosphorylate proteins of the "membrane clock" and of the " $\text{Ca}^{2+}$  clock"; cAMP positively shifts the f-channel activation curve [22]. Moreover, cAMP/PKA and CaMKII-dependent phosphorylation modulate mitochondrial ATP production [23]. The rate at which this intrinsic coupled-clock system ticks becomes further accelerated, or reduced, respectively, by adrenergic or cholinergic neurotransmitter receptor stimulation. Specifically, neurotransmitter-activated receptors signal via many of the same molecules that regulate basic automaticity. Autonomic receptors are coupled to G-proteins and to adenylyl cyclases (likely type 5 or 6) or to guanylyl cyclase, and this leads to activation or suppression of protein kinases that modulate the extent of protein phosphorylation of intracellular  $\text{Ca}^{2+}$  and surface membrane ion channel proteins that drives normal pacemaker clock activity (Figure 1C). Each one of these biochemical feedback mechanisms can operate over a wide range of time scales, and this heterogeneity, therefore, can induce fluctuations within different frequency regimes embedded within the heart rhythm. For example, in heart failure patients, ivabradine, a pharmacological  $I_f$  blocker (Figure 1C), induces a heart rate reduction and an increase in indices of heart rate variability [24]. But the resultant ivabradine-induced changes in heart rate and rhythm embody contributions of changes not only in  $I_f$ , but also of changes in  $\text{Ca}^{2+}$  cycling within SANC, because the membrane and  $\text{Ca}^{2+}$  clocks crosstalk (in response to any disturbance signal that directly perturbs either clock entrains a change in function of the other clock) [21].

The potential contribution of the intrinsic, coupled-clock signaling within SANC to the complexity of the heart rhythm mechanisms can be probed by deletion of autonomic input by complete cardiac denervation. Such denervation of the heart in situ dramatically reduces the HRV and therefore increases the fractal complexity of the heart rate in dogs [4]. However, even in the presence of this apparent complete denervation, the fractal-like behavior is still present [3], although the fractal-like slope ( $\beta$ ) is reduced. The continued presence of fractal-like behavior in the denervated heart suggests that the fractal complexity of the HRV is modulated not only by the autonomic nervous system, but also by intrinsic properties of the sinoatrial node and the cells within it (and also likely by epinephrine released from adrenal cortex). Interestingly, in isolated hearts (i.e., when the heart is completely detached from hormonal and neural input), there is a non-linear relationship between the HRV and heart beating rate in numerous species [8]. These findings are further supported by experiments in single SANC that has been isolated from the sinoatrial node after its removal from the heart. In isolated rabbit SANC, the AP firing rate exhibits a non-linear dependence on cholinergic receptor stimulation (by graded concentrations of acetylcholine) [5,6]. Moreover, there is a tight, but non-linear, relationship between the spontaneous AP firing rate and AP firing rate variation, indicating that changes in spontaneous AP rate and rhythm in single isolated SANC are not simply surrogates of autonomic receptor stimulation [5,6]. In other terms, a non-linear transformation of autonomic receptor signaling occurs within SANC, and this transformation is a determinant of changes in AP firing rate and rhythm initiated by receptor stimulation. Moreover, in isolated





**Figure 3:** The coupled clock system. Schematic illustration of the interplay of adenylyl cyclases (AC), phosphodiesterase (PDE) activity, protein phosphatase (PPT),  $\beta$  adrenergic receptor ( $\beta$ -AR) and cholinergic receptor (Ch-R) stimulation and cAMP/ protein kinase A (PKA)-dependent and  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase II (CaMKII) signaling to the sarcoplasmic reticulum (SR)  $\text{Ca}^{2+}$  cycling proteins (Ryanodine receptor (RyR), Phospholamban (PLB) that binds to the sarcoplasmic reticulum  $\text{Ca}^{2+}$ -ATPase (SERCA)), ion channels (funny channels ( $I_f$ ),  $\text{Ca}^{2+}$  L-type channels (L type),  $\text{Na}^+$ - $\text{Ca}^{2+}$  exchanger (NCX), sarcomere and mitochondria.

Species	Fractal slope in control	Intervention	Fractal slope after intervention	Reference
Rat (Wistar)	$\beta = -0.94$	$\beta$ -receptor blocker (propranolol)	$\beta = -0.75$	[34]
Rat (Wistar)	$\beta = -1.09$	$\alpha$ -receptor blocker (phentolamine)	$\beta = -1.14$	[34]
Rat (Wistar)	$\beta = -0.91$	Vagal blocker (atropine)	$\beta = -0.85$	[34]
Rabbit	$\beta = -1.8$	Vagal blocker (atropine)	Not fractal	[35]
Dog	$\beta = -0.86$	Denervation and $\alpha$ -receptor blocker (prazosin) or $\beta$ -receptor blocker (propranolol)	No additional effect after denervation	[36]
Human	$\beta = -1.15$	$\beta$ -receptor blocker (propranolol)	$\beta = -1.03$	[37]
Human	$\beta = -0.99$	$\beta$ -receptor blocker (propranolol)	$\beta = -0.73$	[38]
Human	$\beta = -0.99$	$\beta$ -receptor+ Vagal blockers (propranolol+atropine)	$\beta = -0.94$	[38]
Human with heart diseases	$\beta = -1.7$	$\beta$ -receptor blocker (atenolol)-long term	$\beta = -1.22'$	[39]
Dog	$\alpha_1 = 0.89$ $\alpha_2 = 0.99$	$\beta$ -receptor blocker (Nadolol)+ $\alpha$ -receptor blocker (Phenoxybenzamine)	$\alpha_1 = 0.8$ $\alpha_2 = 1.03$	[3]
Human	$\alpha_1 = 0.69$ $\alpha_2 = 0.84$	$\beta$ -receptor blocker (propranolol)	$\alpha_1 = 0.64'$ $\alpha_2 = 0.68'$	[40]
Human	$\alpha_1 = 0.69$ $\alpha_2 = 0.84$	Vagal blocker (atropine)	$\alpha_1 = 1.45'$ $\alpha_2 = 1.02'$	[40]
Human	$\alpha_1 = 0.86$	Vagal blocker (atropine)	$\alpha_1 = 1.4'$	[41]
Human	$\alpha_1 = 0.89$	Receptor stimulation (noradrenaline)	$\alpha_1 = 0.4'$	[41]

**Table 1:** The effect of autonomic receptor blockade on fractal-like behavior in different species. Changes in fractal-like behavior of the HRV in response to autonomic receptor blockade measured in different mammals. \* $p < 0.05$  compared to control.

rabbit SANC, beat-to-beat variations in the spontaneous AP firing rate are directly correlated with average variations in the period of the roughly periodic local  $\text{Ca}^{2+}$  releases that emanate from the sarcoplasmic reticulum via ryanodine receptors [7].

That HRV is determined by both the intrinsic properties of cells in the sinoatrial node and the competing influences of the two branches

of the autonomic neural input requires a broader interpretation regarding mechanisms that govern HRV in the normal heart, and how it changes with aging in health and in pathological conditions. Genetic manipulation of protein phosphorylation of intracellular  $\text{Ca}^{2+}$  and surface membrane ion channel proteins of the coupled-clock system (Figure 1B), i.e.,  $I_f$  [25], ryanodine receptors [26], and  $\alpha_1$  subunit of  $\text{Cav}_{1.3}$  [27] induce change in both heart rate and rhythm. In



addition, pharmacological interventions that impact on intrinsic SANC mechanisms (Figure 1), e.g., intracellular  $Ca^{2+}$  [28], can cause the AP firing rate to become irregular. It is also known that denervation of the autonomic nervous system decreases HRV [4]. These results indicate that under normal conditions the intrinsic properties of the sinoatrial node, per second; contribute Brownian noise-like components to the HRV fractal dynamics. In congestive heart failure syndrome, the HRV is reduced and exhibits a reduced fractal-like slope [29]. That congestive heart failure is associated with increased resting levels of sympathetic tone and with decreased parasympathetic tone can only partially explain the reduced HRV and fractal-like slope [29] because congestive heart failure is also associated with increased heart rate [30]. In other words, as demonstrated by Zaza and Lombardi [6], when the  $\beta$  adrenergic receptor is stimulated, the spontaneous AP firing rate in isolated SANC increases (i.e., the autonomic nervous system is not intact) and the HRV become reduced. Moreover, congestive heart failure is associated with intracellular structural remodeling, change in compartmentation of cAMP activity and alteration in neurotransmitter-activated receptor signaling via phosphorylation mechanisms [31]. Therefore, changes in the properties of cells that comprise the sinoatrial node are also likely implicated in the reduction of HRV and fractal-like slope observed in heart failure syndromes. Similarly, a reduction in HRV and fractal-like slope in advanced age [29] may not only be associated with increased resting levels of sympathetic tone and with decreased parasympathetic tone, and an exaggerated increase in plasma catecholamines in response to stress in the context of a reduced adrenergic receptor stimulation [32], but also to changes in intrinsic intracellular coupled-clock mechanisms that accompany advanced age. In fact, aging is associated with deficient intrinsic cAMP-PKA- $Ca^{2+}$  signaling [33]. Therefore, in this case, changes that occur in the properties of the cells in the sinoatrial node likely contribute to age-associated changes in both HRV and fractal-like behavior.

## Future Directions

In summary, the heart exhibits fractal-like behavior that is determined by intrinsic properties of sinoatrial node pacemaker cells in addition to the competing influences of the two branches of the autonomic neural input to the heart. Quantification of the fractal-like behavior at different complexity levels (intact heart, denervated heart, isolated sinoatrial node, phosphorylation networks within isolated SANC, and organization of SANC within the sinoatrial node) is required for a complete understanding of the mechanisms that regulate HRV and induce changes in HRV during aging in health and in the presence of cardiovascular diseases.

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## References

- Lakatta EG, Yaniv Y, Maltsev VA (2013) Minding the gaps that link intrinsic circadian clock within the heart to its intrinsic ultradian pacemaker clocks. Focus on "The cardiomyocyte molecular clock, regulation of Scn5a, and arrhythmia susceptibility". *Am J Physiol Cell Physiol* 304: C941-C944.
- Costa MD, Peng CK, Goldberger AL (2008) Multiscale analysis of heart rate dynamics: entropy and time irreversibility measures. *Cardiovasc Eng* 8: 88-93.
- Li D, Chiang AY, Clawson CA, Main BW, Leishman DJ (2008) Heartbeat dynamics in adrenergic blocker treated conscious beagle dogs. *J Pharmacol Toxicol Methods* 58: 118-128.
- Chiou CW, Zipes DP (1998) Selective vagal denervation of the atria eliminates heart rate variability and baroreflex sensitivity while preserving ventricular innervation. *Circulation* 98: 360-368.
- Rocchetti M, Malfatto G, Lombardi F, Zaza A (2000) Role of the input/output relation of sinoatrial myocytes in cholinergic modulation of heart rate variability. *J Cardiovasc Electrophysiol* 11: 522-530.
- Zaza A, Lombardi F (2001) Autonomic indexes based on the analysis of heart rate variability: a view from the sinus node. *Cardiovasc Res* 50: 434-442.
- Monfredi O, Maltseva LA, Spurgeon HA, Boyett MR, Lakatta EG, et al. (2013) Beat-to-Beat Variation in Periodicity of Local Calcium Releases Contributes to Intrinsic Variations of Spontaneous Cycle Length in Isolated Single Sinoatrial Node Cells. *PLoS One* 8: e67247.
- Monfredi OJ, Nirmalan M, Zhang H, Boyett MR (2013) Heart rate variability as a measure of autonomic nerve activity is fundamentally flawed. *Heart Rhythm*.
- Goldberger AL, Rigney DR, West BJ (1990) Chaos and fractals in human physiology. *Sci Am* 262: 42-49.
- Ivanov PCh, Rosenblum MG, Peng CK, Mietus JE, Havlin S, et al. (1998) Scaling and universality in heart rate variability distributions. *Physica A* 249: 587-593.
- Goldberger AL, Bhargava V, West BJ, Mandell AJ (1985) On a mechanism of cardiac electrical stability. The fractal hypothesis. *Biophys J* 48: 525-528.
- Mandel Y, Weissman A, Schick R, Barad L, Novak A, et al. (2012) Human embryonic and induced pluripotent stem cell-derived cardiomyocytes exhibit beat rate variability and power-law behavior. *Circulation* 125: 883-893.
- Noujaim SF, Berenfeld O, Kalifa J, Cerrone M, Nanthakumar K, et al. (2007) Universal scaling law of electrical turbulence in the mammalian heart. *Proc Natl Acad Sci U S A* 104: 20985-20989.
- Noujaim SF, Lucca E, Munoz V, Persaud D, Berenfeld O, et al. (2004) From mouse to whale: a universal scaling relation for the PR Interval of the electrocardiogram of mammals. *Circulation* 110: 2802-2808.
- Mandelbort B (1983) *The fractal geometry of nature*. Henry Holt and Company, San Francisco, USA.
- (1996) Heart rate variability: standards of measurement, physiological interpretation and clinical use. Task Force of the European Society of Cardiology and the North American Society of Pacing and Electrophysiology. *Circulation* 93: 1043-1065.
- Goldberger AL, Amaral LA, Glass L, Hausdorff JM, Ivanov PC, et al. (2000) PhysioBank, PhysioToolkit, and PhysioNet: components of a new research resource for complex physiologic signals. *Circulation* 101: E215-E220.
- West BJ, Bhargava V, Goldberger AL (1986) Beyond the principle of similitude: renormalization in the bronchial tree. *J Appl Physiol* 60: 1089-1097.
- Hu K, Scheer FA, Buijs RM, Shea SA (2008) The circadian pacemaker generates similar circadian rhythms in the fractal structure of heart rate in humans and rats. *Cardiovasc Res* 80: 62-68.
- Mansier P, Medigue C, Charlotte N, Vermeiren C, Coraboeuf E, et al. (1996) Decreased heart rate variability in transgenic mice overexpressing atrial beta 1-adrenoceptors. *Am J Physiol* 271: H1465-1472.
- Yaniv Y, Sirenko S, Ziman BD, Spurgeon HA, Maltsev VA, et al. (2013) New evidence for coupled clock regulation of the normal automaticity of sinoatrial nodal pacemaker cells: Bradycardic effects of ivabradine are linked to suppression of intracellular  $Ca^{2+}$  cycling. *J Mol Cell Cardiol* 62: 80-89.
- Lakatta EG, Maltsev VA, Vinogradova TM (2010) A coupled SYSTEM of intracellular  $Ca^{2+}$  clocks and surface membrane voltage clocks controls the timekeeping mechanism of the heart's pacemaker. *Circ Res* 106: 659-673.
- Yaniv Y, Spurgeon HA, Ziman BD, Lyashkov AE, Lakatta EG (2013) Mechanisms that match ATP supply to demand in cardiac pacemaker cells during high ATP demand. *Am J Physiol Heart Circ Physiol* 304: H1428-1438.
- Borer JS, Le Heuzey JY (2008) Characterization of the heart rate-lowering action of ivabradine, a selective I(f) current inhibitor. *Am J Ther* 15: 461-473.
- Netter MF, Zuzarte M, Schlichthorl G, Klocker N, Decher N (2012) The HCN4 channel mutation D553N associated with bradycardia has a C-linker mediated gating defect. *Cell Physiol Biochem* 30: 1227-1240.
- Neco P, Torrente AG, Mesirca P, Zorio E, Liu N, et al. (2012) Paradoxical effect of increased diastolic  $Ca^{2+}$  release and decreased sinoatrial node activity in a mouse model of catecholaminergic polymorphic ventricular tachycardia. *Circulation* 126: 392-401.
- Baig SM, Koschak A, Lieb A, Gebhart M, Dafinger C, et al. (2011) Loss of

- Ca(v)1.3 (CACNA1D) function in a human channelopathy with bradycardia and congenital deafness. *Nat Neurosci* 14: 77-84.
28. Yaniv Y, Maltsev VA, Escobar AL, Spurgeon HA, Ziman BD, et al. (2011) Beat-to-beat Ca(2+)-dependent regulation of sinoatrial nodal pacemaker cell rate and rhythm. *J Mol Cell Cardiol* 51: 902-905.
29. Peng CK, Havlin S, Hausdorff JM, Mietus JE, Stanley HE, et al. (1995) Fractal mechanisms and heart rate dynamics. Long-range correlations and their breakdown with disease. *J Electrocardiol* 28 Suppl: 59-65.
30. Tardif JC (2009) Heart rate as a treatable cardiovascular risk factor. *Br Med Bull* 90: 71-84.
31. Perera RK, Nikolaev VO (2013) Compartmentation of cAMP signalling in cardiomyocytes in health and disease. *Acta Physiol (Oxf)* 207: 650-662.
32. Xiao RP, Spurgeon HA, O'Connor F, Lakatta EG (1994) Age-associated changes in beta-adrenergic modulation on rat cardiac excitation-contraction coupling. *J Clin Invest* 94: 2051-2059.
33. Sirenko S, Shukla S, Liu J, Lakatta EG (2011) Age associated decrease in intrinsic action potential (AP) firing rate in sinoatrial node cells is linked to deficient intrinsic cAMP-PKA-Ca<sup>2+</sup> signaling. *Biophysical Journal* 100: 434a.
34. Beckers F, Verheyden B, Ramaekers D, Swynghedauw B, Aubert AE (2006) Effects of autonomic blockade on non-linear cardiovascular variability indices in rats. *Clin Exp Pharmacol Physiol* 33: 431-439.
35. Zwiener U, Hoyer D, Luthke B, Schmidt K, Bauer R (1996) Relations between parameters of spectral power densities and deterministic chaos of heart-rate variability. *J Auton Nerv Syst* 57: 132-135.
36. Wagner CD, Persson PB (1994) Two ranges in blood pressure power spectrum with different 1/f characteristics. *Am J Physiol* 267: H449-454.
37. Yamamoto Y, Hughson RL (1994) On the fractal nature of heart rate variability in humans: effects of data length and beta-adrenergic blockade. *Am J Physiol* 266: R40-49.
38. Hagerman I, Berglund M, Lorin M, Nowak J, Sylven C (1996) Chaos-related deterministic regulation of heart rate variability in time- and frequency domains: effects of autonomic blockade and exercise. *Cardiovasc Res* 31: 410-418.
39. Lin LY, Lin JL, Du CC, Lai LP, Tseng YZ, et al. (2001) Reversal of deteriorated fractal behavior of heart rate variability by beta-blocker therapy in patients with advanced congestive heart failure. *J Cardiovasc Electrophysiol* 12: 26-32.
40. Castiglioni P, Parati G, Di Rienzo M, Carabalona R, Cividjian A, et al. (2011) Scale exponents of blood pressure and heart rate during autonomic blockade as assessed by detrended fluctuation analysis. *J Physiol* 589: 355-369.
41. Tulppo MP, Makikallio TH, Seppanen T, Shoemaker K, Tutungi E, et al. (2001) Effects of pharmacological adrenergic and vagal modulation on fractal heart rate dynamics. *Clin Physiol* 21: 515-523.

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