

Waste to Energy from Flue Gas of Industrial Plants to Biodiesel: Effect of CO₂ on Microalgae Growth

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Abstract

Microalgae are a good source of lipid and other valuable chemicals which have applications in biodiesel production and food industry. Waste management using microalgae has recently gained attention since microalgae can grow by utilizing nutrient from waste resources. Carbon is quantitatively most important nutrient for cultivation of microalgae and can be supplied from flue gas of industrial plants. In this regard, selection of a suitable species of microalgae which has capability to grow using concentrated CO₂ from flue gas is an important consideration. In this study, the effect supplying two concentrations of CO₂ (5% and 15% (v/v)) during cultivation of two microalgae strains were investigated (*Chlorella vulgaris* and *Scenedesmus obliquus*). The results showed maximum biomass concentration of 2.59 g/L under 5.0% and 1.41 g/L under 15.0% CO₂ concentration for *Chlorella vulgaris*. However, the maximum biomass concentrations for *Scenedesmus obliquus* turned to be 30-60% lower. Also, the results indicated 40% and 130% higher maximum biomass productivity for *Chlorella vulgaris* under 5% and 15% CO₂ relative to *Scenedesmus obliquus*. Similarly, the maximum carbon dioxide fixation was shown to be significantly higher for *Chlorella vulgaris* relative to *Scenedesmus obliquus*. Overall our results indicated that *Chlorella vulgaris* is the more appropriate species to be used for cultivation using flue gas of industrial plants.

Keywords: Microalgae; Carbon management; CO₂ mitigation; Biomass production; Biofuel

Introduction

The vigorous increase in atmospheric content of greenhouse gas causes some environmental problems such as global warming and raise of ocean's water level [1]. CO₂ is considered as the major gaseous pollutant which is provided from burning fossil fuel. Among industries which produce high amount of carbon dioxide, power plants account for about 22% of global emission of carbon dioxide [2]. There are several methods for capturing carbon dioxide including chemical and physical methods. However these methods always impose some difficulties so that they cannot be considered as the ideal solutions. For instance, injection of carbon dioxide to the depth of ocean causes acidity of water which leads to devastation of aquatic environment. Other methods such as application of chemical catalysts for absorption of carbon dioxide require an extra location for storage of used catalysts [3]. Biofixation of carbon dioxide using microalgae is a promising alternative method for conventional sequestration techniques. 1 kg of algal dry cell weight utilizes around 1.83 kg of CO₂. Annually around 67.7 tonnes of CO₂ can be mitigated from raceway ponds according to dry weight biomass production of 30 to 37 tonnes per hectare in open ponds [4]. Microalgae can grow using wastewater and saline water as the nutrient and water source [5]. The application of saline water for microalgae cultivation can enhance the sustainability of the cultivation can address the severe lack of fresh water worldwide [6,7]. The biomass of microalgae can be used as the source of biofuel, biopolymers and nutraceutical products [8,9]. The recent researches for production of polymer can be employed for efficient conversion of algal organic acids to high quality products [10,11]. For biofuel production hydrodeoxygenation can efficiently convert biomass to bio-oil [12]. In this regard, the application of syngas under optimum temperature is recently proposed to further enhance the conversion process [13,14]. Despite recent advance in production of biofuel from microalgae, the cost of production need to be further reduced in order to compete with fossil fuel. In this regard, the construction of cultivation systems near to industrial plants can be a promising strategy since carbon, energy, water and nutrient sources can be inexpensively supplied from flue gas

and effluent of the plants [15,16]. However, the flue gas of power plant contains high concentrated CO₂ and other gaseous pollutants (e.g. SO_x and NO_x) which can inhibit the growth of microalgae by acidifying the cultures [17]. In this regard, the selection of an appropriate species that can tolerate the flue gas condition should be evaluated in advance.

In this study, the effect of 5 and 15% CO₂ on growth of two species of microalgae, *Chlorella vulgaris* and *Scenedesmus obliquus*, were investigated. The results of our study suggest that *Chlorella vulgaris* has the capability to utilize the CO₂ from flue gas of industrial plant.

Material and Methods

Microalgae cultivation

The microalgae used in this study were *Chlorella vulgaris* and *Scenedesmus obliquus*. The standard BG-11 medium [18] has been applied for cultivation of these species. The cultivation was conducted for 14 days in 250 mL Erlenmeyer flask (150 mL working volume) under 30°C temperate and continuous light irradiation. The pre-sterilized air with the rate of 20 ml/min was supplied continuously using an air compressor from the bottom of flask. The initial inoculation was 5% v/v and initial pH was 7. The pH variation during addition of flue gas of power plant is simulated by addition of a weak acid (citric acid). The inorganic carbon source is replaced by CO₂ [19]. Samples were daily collected to examine the cell concentration.

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Measurement of cell growth

The cell concentration was measured by measuring the optical density at 650 nm and comparison with calibration curves. For each species, the total suspended solid (TSS) of the standard solutions were measured and the calibration curve was the linear equation between TSS values and measured optical density at 650 nm [20].

To calculate the biomass productivity (Bp) (g/L/day) from change in biomass concentration during cultivation time, Equation 1 was used.

$$Bp = \frac{X_1 - X_0}{t_1 - t_0} \quad (1)$$

Where X_1 is final concentration of biomass, X_0 is initial concentration, t_1 is final time and t_0 is initial time.

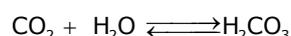
Specific growth rate μ (1/d) was calculated according to Equation 2.

$$\mu = \frac{\ln\left(\frac{X_1}{X_0}\right)}{t_1 - t_0} \quad (2)$$

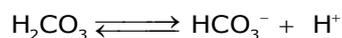
Experiments and procedure

Flue gas of power plants usually contains 13% CO₂, 60 ppm SO_x and 100 ppm NO_x [21]. To investigate the possibility CO₂ supplementation from flue gas for microalgae production, the cultivation of *Chlorella vulgaris* and *Scenedesmus obliquus* were simulated under concentrated CO₂ by addition of bicarbonate and decreasing the pH of the media.

If we consider the CO₂ reactions in water:



$$K = [\text{H}_2\text{CO}_3]/[\text{CO}_2] = 1.70 \times 10^{-3} \text{ at } 25^\circ\text{C}$$



$$K_a = 4.6 \times 10^{-7}; \text{p}K_a = 6.352 \text{ at } 25^\circ\text{C}$$

By considering CO₂ concentration as 5 and 15% (v/v), concentration of bicarbonate can be calculated using reactions' equilibrium equations [20]. In this regard, the accurate amount of bicarbonate was added to the culture throughout the cultivation experiment. Also, in order to evaluate the effect acidification of the cultures by addition of CO₂ supplementation, the pH of media were reduced according to reported data in literature throughout the experiment by using a weak acid (citric acid) [20].

In order to calculate the daily carbon dioxide fixation amount (R_f), according to microalgal growth, Equation 3 is suggested:

$$R_f = (X_1 - X_0) \times m_{cbm} \times V \times \left(\frac{m_{\text{CO}_2}}{m_c}\right) \quad (3)$$

Where m_{cbm} is carbon content ratio of microalgal species, V is cultural volume and m_{CO_2} and m_c are the molar mass of carbon dioxide and carbon. According to proposed general structure for microalgae CO_{0.48}H_{1.83}N_{0.11}P_{0.01} by Chisti [22], Equation 3 can be simplified to Equation 4.

$$R_f = 1.88 \times Bp \quad (4)$$

Where R_f is carbon dioxide biofixation ratio (g/L/day) and Bp is biomass productivity (g/L/day).

Results and Discussion

Biomass production and CO₂ fixation

The preliminary evaluation of the scrutinized microalgae showed

the higher growth and as a result higher biomass productivity and carbon dioxide fixation for *Chlorella vulgaris* at 5% carbon dioxide concentration. Although, while CO₂ concentration increases, the growth of microalgae is inhibited due to reduction of pH but *Chlorella vulgaris* still is considered as the main choice for utilizing the inorganic carbon of flue gas from power plant as a result of its higher biomass productivity and energy production. The higher lipid content of *Scenedesmus obliquus* made it more suitable choice for biofuel production. Figure 1 exhibits the biomass concentration (g/L) during cultivation time (day) for CO₂ concentration of 5%.

The results for change in biomass concentrations were shown in Figure 1. From results of biomass concentrations and by using Equation 1 the biomass productivity were calculated. Our results indicated the maximum biomass concentration of 2.58 g/L and 0.78 g/L during 12 d cultivation under 5% CO₂ for *Chlorella vulgaris* and *Scenedesmus obliquus*. Also, *Chlorella vulgaris* accounts for 40% higher maximum biomass productivity (0.39 g/L/day) relative to *Scenedesmus obliquus*. (0.28 g/L/day).

While by raise in concentration of CO₂ from 5% to 15%, a slight decrease in amount biomass concentrations is occurred, *Chlorella vulgaris* still has higher biomass concentration relative to *Scenedesmus obliquus* (Figure 2). By using Equation 1, maximum biomass productivity for *Chlorella vulgaris* and *Scenedesmus obliquus* were calculated as 0.37 g/L/day and 0.16 g/L/day, respectively.

Varied growth parameters including maximum biomass concentration, maximum specific growth rate and maximum biomass productivity in two concentration of CO₂ are compared for these two species in Table 1.

The significant change in the growth and productivity of *Scenedesmus obliquus* by increasing CO₂ concentration is observed in Table 1. However, for *Chlorella vulgaris* the inhibitory effect of increasing CO₂ concentration on growth is less drastic.

The effect of high and low concentration of nutrients on microalgae growth is consistently investigated in the literature [23,24]. While studies showed that increasing carbon source concentration in the media can be advantageous for microalgae production [25,26], the high concentration of CO₂ can decrease pH and inhibit growth of microalgae [20]. Some studies proposed application of media with high alkalinity for enhanced dissolution of CO₂ in media and productivity of microalgae [27]. In addition to CO₂, the modulation of other macronutrients and micronutrients can enhance the productivity of biomass and valuable products (e.g. lipid) from microalgae [28,29]. N in the media is shown to be a promising for production of carbon storage compounds which have application in biodiesel production [30]. In this regard, our results

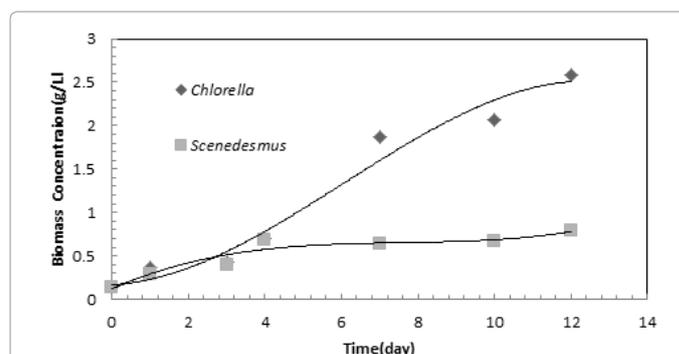


Figure 1: Change in biomass concentration during cultivation under 5% CO₂.

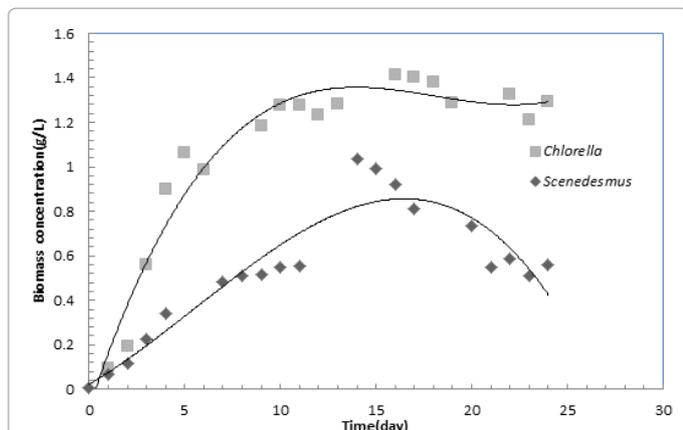


Figure 2: Change in biomass concentration during cultivation under 15% CO₂.

Microalgae species	Maximum biomass concentration(g/L)	Maximum specific growth rate, μ_{max} (1/day)	Maximum Biomass productivity, P_{max} (g/L/d)	CO ₂ concentration
<i>C. vulgaris</i>	2.587	0.44	0.39	5%
<i>S. obliquus</i>	0.78	0.34	0.28	5%
<i>C. vulgaris</i>	1.41	0.46	0.37	15%
<i>S. obliquus</i>	1.036	0.28	0.16	15%

Table 1: The summarized results for cultivation experiments under 5 and 15% CO₂.

suggest the cultivation of *Chlorella vulgaris* with 5% CO₂ and low N can lead to higher productivity of biodiesel from microalgae.

The carbon dioxide fixation ratio was calculated using Equation 4 and results were illustrated in Figure 3. The results showed a reduction in CO₂ fixation ratio from 0.73 to 0.69 g/L/day for *Chlorella vulgaris* and from 0.53 to 0.212 g/L/day for *Scenedesmus obliquus*, respectively, as CO₂ concentration increase from 5% to 15% maximum. If we compare maximum CO₂ fixation ratio (g/L/day) for *Chlorella vulgaris* and *Scenedesmus obliquus* in two different CO₂ concentrations it can be easily realized that higher CO₂ concentration impose less carbon dioxide mitigation. However, when the CO₂ concentration raises from 5% to 15% this decline in amount of carbon dioxide mitigation is not significant but in pure CO₂ injection the sequestration efficiency can be reduced vigorously.

Chlorella sp. is reported to be a suitable species to grow under harsh environmental condition [31]. Application of waste for production of valuable products can create opportunities for local and global improvement of economic and environmental sustainability [32,33]. Among different methods for management of waste, microalgae is specifically significant due to possibility of nutrient utilization from waste streams, high growth rate and capability for production of valuable products (e.g. lipid, protein, pigments) [8]. In this regard, various studies were conducted for use municipal, agricultural and industrial waste for cultivation of microalgae [34]. However, the prior dilution of waste resources is necessary to enhance the productivity and nutrient utilization rate of microalgae [35]. The results of our study suggest that *Chlorella vulgaris* is more suitable species for management of CO₂ from flue gas and production of valuable products (e.g. biomass).

Conclusion

In this study, two industrially important species of microalgae are

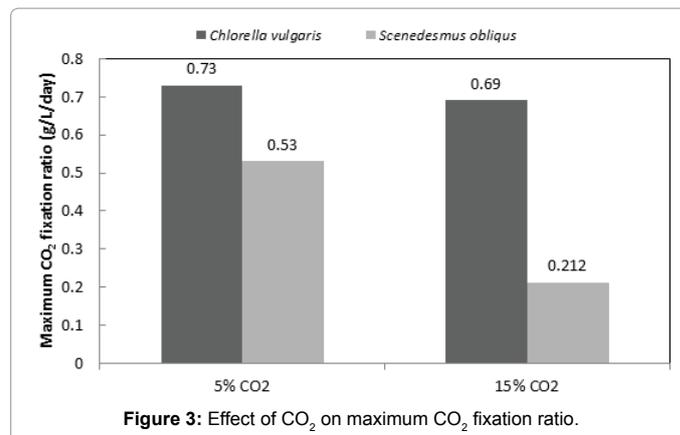


Figure 3: Effect of CO₂ on maximum CO₂ fixation ratio.

studied in two different conditions (5% and 15% CO₂). These conditions are simulated through experiment by replacement of carbon dioxide gas with sodium bicarbonate and simultaneous titration. Our results show higher biomass production and CO₂ fixation for *Chlorella vulgaris*. Our study suggests that cultivation of *Chlorella vulgaris* is a promising technique for CO₂ mitigation as well as production of biomass and other valuable compounds from flue gas of industrial plants.

References

- Hallenbeck PC, Benemann JR (2002) Biological hydrogen production; fundamentals and limiting processes. Int J Hydrog Energy 27: 1185-1193.
- Siegenthaler U, Stocker TF, Monnin E, Lüthi D, Schwander J, et al. (2005) Stable carbon cycle-climate relationship during the late Pleistocene. Science 310: 1313-1317.
- Lackner KS (2003) A guide to CO₂ sequestration. Science 300: 1677-1678.
- Brennan L, Owende P (2010) Biofuels from microalgae—a review of technologies for production, processing, and extractions of biofuels and co-products. Renew Sustainable Energy Rev 14: 557-577.
- Pittman JK, Dean AP, Osundeko O (2011) The potential of sustainable algal biofuel production using wastewater resources. Bioresour Technol 102: 17-25.
- Vaseghi G, Celik I, Apul D, Burian S (2017) Economic, environmental, and social criteria evaluation of rainwater harvesting system options for an office and lab building on the University of Utah Campus, frontiers in water savings in buildings. Bentham Science Publishers, pp: 117-155.
- Vaseghi G, Ghassemi A, Loya J (2016) Characterization of reverse osmosis and nanofiltration membranes: effects of operating conditions and specific ion rejection. Desalination and Water Treatment 57: 23461-23472.
- Borowitzka MA (2013) High-value products from microalgae-their development and commercialisation. J Appl Phycol 25: 743-756.
- Majdzadeh-Ardakani K, Zekriardehani S, Coleman MR, Jabarin SA (2017) A Novel Approach to Improve the Barrier Properties of PET/Clay Nanocomposites. Int J Polym Sci.
- Zekriardehani S, Jabarin SA, Gidley DR, Coleman MR (2017) Effect of chain dynamics, crystallinity, and free volume on the barrier properties of poly(ethylene terephthalate) biaxially oriented films. Macromolecules 50: 2845-2855.
- Liu X, Wu Y, Shmulyk R, Luo Y, Wang XA, et al. (2016) Developing a renewable hybrid resin system. Part I: Characterization of co-polymers of isocyanate with different molecular weights of phenolic resins. BioResources 11: 5300-5311.
- Luo Y, Hassan EB, Miao P, Xu Q, Steele PH (2017) Effects of single-stage syngas hydrotreating on the physical and chemical properties of oxidized fractionated bio-oil. Fuel 209: 634-642.
- Luo Y, Guda VK, Hassan EB, Steele PH, Mitchell B, et al. (2016) Hydrodeoxygenation of oxidized distilled bio-oil for the production of gasoline fuel type. Energ Convers Manage 112: 319-327.
- Luo Y, Hassan EB, Guda V, Wijayapala R, Steele PH (2016) Upgrading of syngas hydrotreated fractionated oxidized bio-oil to transportation grade

- hydrocarbons. *Energy Convers Manage* 115: 159-166.
15. Guzzon A, Bohn A, Diociaiuti M, Albertano P (2008) Cultured phototrophic biofilms for phosphorus removal in wastewater treatment. *Water research* 42: 4357-4367.
 16. Kumar A, Yuan X, Sahu AK, Dewulf J, Ergas SJ, et al. (2010) A hollow fiber membrane photobioreactor for CO₂ sequestration from combustion gas coupled with wastewater treatment: A process engineering approach. *J Chem Technol Biotechnol* 85: 387-394.
 17. Maeda K, Owada M, Kimura N, Omata K, Karube I (1996) CO₂ fixation from the flue gas on coal-fired thermal power plant by microalgae. *Energy Convers Manage* 36: 717-720.
 18. Hanifzadeh MM, Sarrafzadeh MH, Tavakoli O (2012) Carbon dioxide biofixation and biomass production from flue gas of power plant using microalgae. *Second Iranian Conference on Renewable Energy and Distributed Generation, IEEE*. pp: 61-64.
 19. De Morais MG, Costa JAV (2007) Isolation and selection of microalgae from coal fired thermoelectric power plant for biofixation of carbon dioxide. *Energy Convers Manage* 48: 2169-2173.
 20. Hanifzadeh M (2012) CO₂ capturing from flue gas of power plant using Microalgae. *Chemical Engineering, University of Tehran*. p. 132.
 21. Kumar K, Dasgupta CN, Nayak B, Lindblad P, Das D (2011) Development of suitable photobioreactors for CO₂ sequestration addressing global warming using green algae and cyanobacteria. *Bioresour Technol* 102: 4945-4953.
 22. Chisti Y (2007) Biodiesel from microalgae. *Biotechnol Adv* 25: 294-306.
 23. Nabati Z, Hanifzadeh M, Garcia EC, Viamajala S (2016) Investigation of *Chlorella* sp cultivation in low concentration of Mg and Ca. *38th Symposium on Biotechnology for Fuels and Chemicals*.
 24. Hanifzadeh M, Viamajala S (2014) Effects of nutrients on microalgae cultivation. *Annual Graduate Research Symposium*.
 25. Pendyala B, Hanifzadeh M, Viamajala S (2016) Enhanced biomass and lipid productivities of outdoor alkaliphilic microalgae cultures through increased media alkalinity.
 26. Garcia EC, Hanifzadeh M, Viamajala S (2015) The effect of bicarbonate on microalgae. *Midwest Graduate Research Symposium*.
 27. S Viamajala, B Pendyala, M Hanifzadeh (2016) Alkaliphilic algal cultivation as a means for improved productivity and stability of algae-based production systems. *Algae Biomass Summit*.
 28. Garcia EC, Hanifzadeh M, Viamajala S (2016) Effect of low concentration of Mg and Ca on microalgal biomass and lipid production. *Algal Biomass Organization Summit*.
 29. Hanifzadeh M (2015) Cultivation strategies for enhancement of valuable products yield from microalgae. *Science Alliance at the New York Academy of Sciences and PepsiCo*.
 30. Pendyala B, Hanifzadeh M, Viamajala S (2016) Cultivation of low nitrogen algal biomass for high-quality algal biofuels production. *Algal Biomass Organization Summit*.
 31. Pendyala B, Hanifzadeh M, Viamajala S (2017) Investigation of the effect of high salinity in media on Microalga *Chlorella sorokiniana* str. SLA04. *The 7th International Conference on Algal Biomass, Biofuels and Bioproducts, Elsevier*.
 32. Ovissipour M, Gholami S, Hanifzadeh M (2011) Achievement the knowledge of producing fluoro-protein for extinguishing fire in oil reservoirs.
 33. Hanifzadeh M, Nabati Z, Longka P, Malakul P, Apul D, et al. (2017) Life cycle assessment of superheated steam drying technology as a novel cow manure management method. *J Environ Manage* 199: 83-90.
 34. Pittman JK, Dean AP, Osundeko O (2010) The potential of sustainable algal biofuel production using wastewater resources. *Bioresour Technol* 102: 17-25.
 35. Bjornsson WJ, Nicol RW, Dickinson KE, McGinn PJ (2013) Anaerobic digestates are useful nutrient sources for microalgae cultivation: functional coupling of energy and biomass production. *J Appl Phycol* 25: 1523-1528.

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